# Tumor Growth is Stimulated by Muscarinic Receptor Agonism: Role of Autoantibodies from Breast Cancer Patients

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# Abstract

In developed countries, cancer has replaced infectious diseases as a major cause of death. Currently, efforts in the immunoprevention of cancer are beginning to resemble theter those presented by the prevention of infectious diseases by immunization a century ago. Breast cancer is the most frequent type of tumor in women and is the second leading cause of death by this illness, among them. Moreover, cancer incidence will grow during next years. Some findings in autoimmunity related to breast cancer in animal models have been important to clarify mechanisms that potentiate tumor growth. Clinical and experimental data now clearly indicate that chronic inflammation significantly contributes to cancer development. Emerging out of these studies is an appreciation that persistent humoral immune responses exacerbate recruitment and activation of innate immune cells in the neoplastic microenvironment where they regulate tissue remodeling, pro-angiogenic and pro-survival pathways that together potentiate cancer development. Generally, antigens involved in autoimmune response in breast cancer are modified selfproteins or over-expressed normal proteins that induce autoantibodies (autoAbs) formation, which exhibit tumor promoting actions. Very frequently muscarinic acetylcholine receptors (mAChR) are up-regulated in different types of tumors appearing in different species **Does this mean species of animal, or isoforms** of proteins?. mAChR have the ability to act as autoantigens for tumor bearers. This article will review recent results concerning to the ability of mAChR expressed in transformed breast cells to trigger auto-Abs formation either in experimental models or in breast cancer patients. We will also discuss the action of these antibodies as agonists of mAChR.

Keywords: Autoantibodies, muscarinic receptors, breast cancer, tumor growth.

Running title: Autoantibodies against mAChR and tumor growth

### AN OVERVIEW OF TUMOR IMMUNOLOGY

The effect of preventing tumorigenesis by the stimulation of immune system has been anxiously searched <del>by scientists</del> Scientists have long sought to prevent tumorigenesis by stimulation of the immune system in the same manner that microbiologists expected to immunize people against infective diseases a century ago. In spite of these efforts, many aspects of tumor immunology must be clarified to develop the most appropriate immunotherapeutic tools for cancer patients. It must be recognized that the immune system may be ineffective in preventing tumor growth. The theory of "immunosurveillance" that was proposed by Burnet in 1970, has a limited role since although immune cells are activated in tumor bearing individuals, they are not able to promote effective anti-tumor actions [1, 2]. The "cancer immunoediting" hypothesis has enriched classical immunosurveillance theory and comprises three phases. The first phase, referred to as "elimination", encompasses our traditional understanding of cancer immunosurveillance, in which cells of the innate and adaptive immune systems recognize and destroy developing tumors, protecting host against cancer. The second phase is "equilibrium" in which ongoing tumor growth and immunosurveillance enter into a dynamic balance with one another. The third phase is "escape", where tumor variants that can avoid immune mediated destruction emerge and develop into a clinical the stage neoplasm. Thus, in the long term tumor infiltration by immune cells, rather than exerting functions to protect the tumor host, produces great amounts of growth factors, oxidative molecules and pro-angiogenic mediators that can actually favor tumor development [3]. Although most tumors are antigenic, only a small fraction is spontaneously immunogenic. This is due to several escape mechanisms developed by tumor cells as alterations in processing or presentation of antigens, loss or down-regulation of class I and II major histocompatibility molecule expression, absence of B7 co-stimulatory molecules with consequential induction of anergy and/or tolerance [4]. Another important aspect of tumor escape either in animal models or in humans is the liberation of immunosuppressive cytokines such as prostaglandin  $E_2$ (PGE<sub>2</sub>), transforming growth factor  $\beta$ , interleukin-10 and vascular endothelial growth factor (VEGF), among others, that, prevent cytotoxic T cell specific activation against tumors [3]. In addition, VEGF inhibits antigen presentation by mature dendritic cells and stimulates angiogenesis and/or lymphangiogenesis triggering tumor growth [5].

Most of the antigens identified in human tumors are self proteins, without mutations but inappropriately expressed or over-expressed. As a consequence, instead of acting as tumor rejectionne antigens, they

induce tolerance or autoimmune responses to normal tissue [6-8]. In relation to the latter, **it has been** reported that the presence of antibodies against autoantigens during tumor development by B cells could be triggered by apoptotic tumor cells in situ, but they also can induce peripheral mechanisms that lead to tolerance. This sentence is not clear—does this refer to a B cell tumor, or to autoAbs produced by B cells? And how does the source of antigen relate to mechanisms of peripheral tolerance in the context of antibodies? The collective studies of T and B cell immunity in cancer patients<sup>1</sup> clearly demonstrate incomplete immunological tolerance against normal proteins. If genes are activated in tumors or expressed at unusually high levels, the gene products appear to gain access to the antigen presentation machinery leading to the induction of T cell immunity and the production of antibodies [9]. It has been postulated that B cell tolerance differs from T cell tolerance since the selection of autoreactive B clones is highly specific and appears when repetitive sequences are expressed in antigens, as has been observed for nuclear or neuronal proteins [10-12].

Breast cancer is the most frequent class of tumor in women and is the second leading cause of death from cancer by this illness. Diagnosis is based mainly on clinical examination, mammography, cytology, and a few specific molecular markers, and there are no other conclusive methods for establishing diagnosis and/or prognosis for this pathology [13].

Several lines of evidence indicate that stromal cell responses in pre-malignant mammary tissue may "promote" progression to a malignant tumor. Cellular components of tumor stroma include (myo) fibroblasts, vascular cells, infiltrating leukocytes and specialized messenchymal support cells unique to each tissue microenvironment. A growing body of evidence has recently implicated tumor-infiltrating lymphocytes in tumor progression [14]. Hansen *et al.* [15] reported that tumor infiltrating B cells in patients with breast medullary carcinoma locally produce antibodies against autoantigens exposed in membranes as a consequence of cellular apoptosis. Moreover, several studies reviewed by Fernández Madrid [16] indicate that a plethora of autoantibodies (autoAbs) with different specificities has been found in breast cancer patients. These autoAbs directed to tumor associated antigens could be detected in the sera of patients months or years before clinical diagnosis and are potentially useful biomarkers for early detection of breast malignancies.

This article will **revise should be review?** recent results concerning the ability of muscarinic acetylcholine receptors (mAChR) expressed in transformed cells, to trigger autoAb

experimental models or in breast cancer patients. We will also discuss the action of these antibodies as agonists for mAChR functions.

### mAChR AND BREAST CANCER

The mAChRs mediate most of the actions of the neurotransmitter acetylcholine in the central nervous system (CNS) and peripheral nervous system, as well as in the end organs of parasympathetic nerves. In mammals, five distinct mAChR subtypes ( $M_1$ – $M_5$ ) have been identified, with each receptor subtype being the product of a different gene. The mAChRs belong to the superfamily of seven-transmembrane receptors, which activate signal transduction pathways through their interactions with heterotrimeric guanine nucleotide binding regulatory proteins (G proteins). The  $M_1$ ,  $M_3$ , and  $M_5$  receptor subtypes are efficiently coupled to the pertussis toxin-insensitive  $G\alpha q/11$  and  $G\alpha 13$  subtype G proteins, leading to, for example, activation of phospholipase (PL) C, PLA<sub>2</sub> and PLD<sub>7</sub>, while  $M_2$  and  $M_4$  receptors preferentially couple to pertussis toxin-sensitive Gi and Go proteins, leading to the inhibition of adenylyl cyclase and/or to calcium channel gating [17, 18]. mAChRs regulate a large number of central functions including cognitive, behavioral, sensory, motor and autonomic processes. Changes in mAChR levels and activity have been implicated in the pathophysiology of many major diseases of the CNS including Alzheimer's disease, Parkinson's disease, depression and schizophrenia. The actions of acetylcholine on mAChR expressed in effector tissues include, for example, decreases in heart rate (mediated by  $M_2$  receptors) and increases in smooth-muscle contractility and glandular secretion, predominantly mediated by  $M_3$  receptor subtype.

The components of cholinergic system<sup>4</sup> (mAChR, nicotinic receptors, acetylcholine, and the enzymes involved in its does "its" refer to Ach or other components of the system? synthesis and degradation) were **gradually** eventually detected in non-neuromuscular cells and tissues. Today the entire set of cholinergic molecules and their functions in cell-to-cell communication, mediated by intracellular ion dynamics, is almost entirely established. It has been demonstrated that mAChR expression and/or function are significantly increased in different types of tumor cells<sup>4</sup>, such as colon, lung and erythroleukemia, in comparison to cells derived from the same normal tissues [19]. On this point, we demonstrated that mAChRs are constitutively expressed in different murine breast cancer cell lines (LM2, LM3 and LMM3) derived from BALB/c **mee** mouse tumors, while they are absent in normal murine mammary cells [20]. We also showed that the expression of mAChRs increased with the metastatic ability of tumors, since LMM3 cells<sup>4</sup> that were obtained from a lung metastasis of M3 tumor<sup>4</sup> up-regulate mAChR expression by 50 fold in comparison with LM3 cells [21]. Moreover, we evaluated the functionality of mAChR in these cell lines and observed that the synthetic cholinergic agonist carbachol increased proliferation in all tumor cell lines. In LM3 cells, this agonist promoted proliferation via  $M_3$  receptor activation by inositol 1,4,5-triphosphate (IP3) and nitric oxide (NO) production. Carbachol-induced LM2 cells proliferation needed both  $M_2$  and  $M_1$  receptor activation increasing PGE<sub>2</sub> liberation and arginase catabolism respectively [22]. In addition, carbachol stimulated LM2 and LM3-induced angiogenesis and tumor growth. This activation followed different patterns. In LM2 tumor,  $M_1$  and  $M_2$  receptors activation stimulated neovascularization via arginase II and cyclooxygenase-2 (COX-2)-derived products, while  $M_1$  and  $M_3$  receptors mediated carbachol-induced tumor growth by the same effector enzymes. In LM3 tumor, we observe that  $M_1$  and  $M_2$  receptors are involved in agonist-stimulated angiogenesis by COX and nitric oxide synthase (NOS) 1-derived products while tumor growth is stimulated by  $M_3$  and  $M_2$  receptors activation and COX-2-derived prostanoids [22].

Afterwards, we analyzed mAChR expression in human breast cancer tissue. Samples were surgically obtained from six patients with tumors corresponding to different histological grades. By Western blot using specific commercial anti-mAChR antibodies, in homogenates of breast tumor tissue, major expression of M<sub>2</sub> and M<sub>3</sub> receptor subtypes were was observed, and growing amounts increased levels of both proteins correlated with the malignancy and invasive capacity of those breast carcinomas. Breast samples obtained from patients with breast benign fibroadenoma (BFA) or free of illness from healthy donors lack of mAChR protein expression at all [23]. In addition, we provided direct evidence on the constitutive expression of functional mAChR in MCF-7 cells, a hormone-sensitive cell line derived from an invasive ductal human breast carcinoma that represents an early stage human breast cancer. We demonstrated for the first time, that MCF-7 cells express  $M_3$  and  $M_4$  receptor subtypes [24]. The activation of M<sub>3</sub> receptor with the cholinergic agonist carbachol increased MCF-7 celle proliferation since 4-DAMP, a selective  $M_3$  receptor antagonist, but not tropicamide ( $M_4$  selective antagonist), reduced carbachol action (Fig. 1A). Jiménez and Montiel [25] previously reported that in MCF-7 cells, mAChR stimulation with carbachol, induced **en** increased of protein synthesis and cell proliferation, and these effects were prevented by PD098059, a specific inhibitor of the mitogen activated kinase kinase (MAPKK). They also demonstrated that mAChR downstream effectors PKC-zeta, PI3K, and Src family of tyrosine kinases, but not intracellular-free Ca<sup>2+</sup> mobilization, are key molecules in the signal cascade leading to MAPK/ERK activation.

Taken together, these previous results reveal that the presence of mAChR in breast tumor cells and tissues, and the absence of these receptors in normal cells, could be assigning a central role to mAChR as promoters of malignant progression. In addition, the appearance of these receptors in transformed cells could be challenging the immune system to synthesize autoAbs. It has been described that mAChR can dimerize with nicotinic receptors **eon** forming an heterodimeric receptor that **should I** would say **"appears to be..."** be the target of autoAbs in patients with pemphigus vulgaris [26]. Also, M<sub>2</sub> and M<sub>3</sub> receptors can oligomerize forming dim**m**ers or trimers of higher molecular weight than native monomeric receptors [27]. Likewise the presence of heterodimeric mAChR with c-Myc protein that could increase its oncogenic and immunogenic ability has been described [28].

# ANTIBODIES AGAINST mAChR FROM BREAST CANCER PATIENTES ACT AS RECEPTOR<mark>S</mark> AGONIST<mark>S</mark> AND PROMOTE TUMOR PROGRESSION

The collective studies of T and B cell immunity in cancer patients clearly demonstrate incomplete immunological tolerance against normal proteins. If genes are activated in tumors or expressed at unusually high levels, the gene products appear to gain access to the antigen presentation machinery leading to the induction of T cell immunity and the production of antibodies. The antibody responses detected in cancer patients appear to be T cell dependent, since they are of IgG isotype and not of the T cell independent IgM isotype. An important question that has remained unresolved is the kinetics of immune responses against aberrantly expressed tumor antigens. Do they responses occur late in disease at a stage when tumor burden and antigen load is high, or do they occur early when the tumor load is small. A study by Zhang *et al.* [29] has monitored tumor immunity in patients before and after the diagnosis of hepatocellular carcinoma. A **survey** longitudinal study of 17 patients showed that the transition from chronic hepatitis/cirrhosis to malignancy was associated with the appearance of antibody responses against protein expressed in the G2 and M phases of the cell cycle (p62). Interestingly, in one case these antibodies were detected before the clinical diagnosis of hepatocellular carcinoma. These data suggest that immune responses against these proteins can occur at early stages of the malignant process prior to clinical disease manifestation.

In breast cancer, while it is clear that acute activation of B cells may play a role in eradicating early neoplastic cells, or may participate in spontaneous regression of tumors through classical and well-studied immunoglobulin-mediated mechanisms, it has been postulated that chronic activation of B cells may paradoxically play a role in potentiating carcinoma development [8, 12]. During breast carcinogenesis,

mature B cells (including naïve cells, and activated cells) can be found in secondary lymphoid tissues as well as in tumor-associated stroma. As compared with healthy patients without evidence of cancer the sentinel lymph nodes of breast cancer patients contain proliferating and mature IgG+ B lymphocytes [30]. The percentage of B cells present in sentinel and axillary lymph nodes in these patients, reveals that their numbers correlate with disease stage when comparing stage I vs. stage II, and against total tumor burden [31, 32]. Similarly, B cells present in breast tumor-associated stroma may also play a role in disease progression. Several studies have reported that infiltrating B lymphocytes represent the predominant lymphoid population (in excess of T lymphocytes) in pre-malignant breast tissue, including hyperplasia and early ductal carcinoma in situ. Although the presence of chronically activated B cells and autoAbs in breast cancer patients is clear, the role of the latter in this disease progression is not clear. We had previously documented the presence of autoAbs in LM3 murine tumor bearers. LM3 is a malignant cell line derived from a murine mammary adenocarcinoma that spontaneously over-expressed mAChR. We described that IgG purified from early and late tumor bearers, displaced tritiated quinuclidinyl benzilate binding to LM3 tumor cells, confirming the antibod<mark>⊭ies'</mark> interaction<mark>s</mark> with mAChR, while IgG from normal mice did not modify the antagonist binding to receptors [33]. In addition, Abs detected a protein of 70 kDa on immunoblots en of murine tumor celle lysates and on heart homogenates; that a similar band was also recognized by a specific anti-M<sub>2</sub> receptor monoclonal antibody, and The autoAbs also IS **THAT CORRECT?** gave a positive immunofluorescence staining on LM3 cells membrane [34]. We also observed that IgG purified from early tumor bearers stimulated LM3 cells proliferation in a more effective manner than the muscarinic agonist carbachol. This effect was also observed when the IgG molecule was cleaved from its Fc fragment by pepsin digestion (unpublished data). The specificity of IgG action on mAChR was confirmed to prevent proliferative effects with the muscarinic antagonist atropine [33]. THIS NEEDS CLARIFICATION—DOES ATROPINE REDUCE PROLIFERATION OF LM3 CELLS, AND THE AutoAbs BLOCK THIS EFFECT?

Recently, we demonstrated the presence of autoAbs in the IgG fraction obtained from blood samples of breast cancer patients in stage I (T1N0Mx: tumor size <2 cm, without axillary node metastases) that exhibit the ability to activate mAChR expressed in MCF-7 cells, promoting cells proliferation, while IgG from BFA bearers or from healthy donors patients free of illness were enable unable to produce this action (Fig. 1B and C) [35]. The action proliferative stimulus of IgG from patients with T1N0Mx stage

breast tumors (T1N0Mx-IgG) on MCF-7 cells was reverted by 4-DAMP and tropicamide, revealing the participation of  $M_3$  and  $M_4$  receptors in this effect (Fig. 1B).

The presence of circulating antibodies **AGAINST TUMOR AGS?** in breast cancer patient s' sera has been generally linked to a negative prognosis in this illness. By immunocytochemistry and Western blot it was demonstrated that Fab fragments of IgG from breast cancer patients, recognize different mammary proteins in MCF-7 and MDA-MB-231 cells that were not recognized by the same fragment derived from normal IgG. Fab fragments from serum IgG from patients with breast cancer also stimulate tumor cell proliferation *via* ERK2 [36].

It is well known, that  $M_3$  receptor activation leads to Gq/11 coupling and to IP3 formation and calcium mobilization via PLC. Regarding the latter, we observed that the proliferative action exerted by carbachol on MCF-7 cells was reverted when cells were treated previously with the PLC inhibitor 2-nitro-4carboxyphenyl N,N-diphenylcarbamate (NCDC), N<sup>G</sup>monomethyl-L-arginine (L-NMMA), a non selective inhibitor of NOS) or H-7 (inhibitor of PKC) (Fig. 2A). In addition we confirmed that MCF-7 cells express PLCβ2 enzyme (Fig 2B). PLC enzymatic activity was measured by determining IP1 levels, the final product derived from IP3, with a commercial ELISA kit in MCF-7 cells treated with T1N0Mx-IgG or carbachol, to verify this enzyme participation in their **NOT CLEAR WHICH ARE "THEIR" EFFECTS AT THIS POINT** effects. As it is shown in Figure 3A either T1N0Mx-IgG or carbachol increased IP1 formation by more than 100% in comparison to control (untreated tumor cells) (Fig. 3A). In addition, it has been demonstrated that PLC is involved in other steps of tumorigenesis. You et al. [37] reported that PLC is involved in tumor cells migration, by analyzing different human breast cancer cell lines, with distinct metastatic potential. Although we described the expression of PLC $\beta$ 2 in MCF-7 cells, we used a nonselective PLC inhibitor to test IP1 formation induced by carbachol or IgG, so other isoforms of this enzyme may be involved. Precisely, Alvarez-Breckenridge et al. [38] reported the expression of PLC $\gamma$  in MCF-7 cells and pointed to the involvement of its products in tumor progression. The presence of calcium-dependent isoforms NOS1 and NOS3 was documented by us in MCF-7 cells [35]. NO is considered an important mediator in carcinogenesis and depending on its levels it can exert opposite actions. NO liberated by calcium-dependent isoforms NOS1 and NOS3 promotes tumorigenesis, while NOS2-derived NO generally induces antitumor actions, cell cycle arrest, and apoptosis [39]. When NOS activity was quantified by determining nitrite<mark>s</mark> (NO<sub>2</sub><sup>-</sup>) production by Griess reaction, we observed that both T1N0Mx-IgG and carbachol increased NO formation. This effect was not

modified by preincubating cells with aminoguanidine, a NOS2 selective inhibitor, confirming the absence of this isoform in MCF-7 cells. It is important to note that T1N0Mx-IgG was more potent to stimulate NOS activity (7 fold) than carbachol (2 fold **increment** increase in NO<sub>2</sub><sup>-</sup> formation) (Fig. 3B). The latter could be due to different mechanisms of action; **that** these could include different coupling to effectors which trigger this signal transduction pathway, and/or the dimerization/oligomerization of receptors with stabilization and permanent activation, promoting tumor celle migration. Schulze W. et al. [40] have reviewed the binding of autoAbs to epitopes of  $\beta_1$  or M<sub>2</sub> receptors that belong to the GPCR family in various cardiovascular diseases. In spite of this While interaction is not completely understood, it has been shown that agonistic stimulation by autoAbs shifts the receptor to a dimeric state, stabilizes this conformation, and activates the signal cascade.

# CONCLUSIONS

In recent years many scientists have been attempting to explain the mechanisms underlying the interactive relation between immune cells and tumor progression. Abundant evidence indicates that the immune system plays a major role in this interaction and under chronic tumor stimulation, begins to facilitate tumor growth. AutoAbs against mAChR have been described in other pathologies such as HIV infection, Chagas disease, Sjögrem Syndrome, and pemphigus vulgaris, all of them with an important autoimmune component [41-43, 26]. Humoral responses against mAChR have not been described in other tumors, but in breast cancer anti-mAChR antibodies reveal that autoimmunity is present in this illness, promoting tumor progression by producing agonistic actions that favors different steps of tumorigenesis. The presence of these antibodies at early stages of disease could predict a worse prognosis in this illness. Whether these autoAbs "appear" prior to tissue malignancy requires further investigation in order to achieve a clear predictive value for them.

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# REFERENCES

[1] Hwu, P. Treating cancer by targeting the immune system. N. Engl. J. Med., 2010, 363, 779-781.

[2] Burnet, F. The concept of immunological surveillance. Prog. Exp. Tumor Res., 1970, 13: 1-27.

[3] Schreiber, R.D.; Old, L.J.; Smyth, M.J. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science*, **2011**, 331,1565-1570.

[4] Khong, H.T.; Restifo, N.P. Natural selection of tumor variants in the generation of "tumor escape" phenotypes. *Nat. Immunol.*, 2002, 3, 999–1005.

[5] Sheng, K.C.; Wright, M.D.; Apostolopoulos, V. Inflammatory mediators hold the key to dendritic cell suppression and tumor progression. *Curr. Med. Chem.*, **2011**, 18, 5507-5518.

[6] Ryan, S.O.; Turner, M.S.; Gariépy, J.; Finn, O.J. Tumor antigen epitopes interpreted by the immune system as self or abnormal-self differentially affect cancer vaccine responses. *Cancer Res.*, **2010**, 70: 5788-5796.

[7] Abken, H.; Hombach, A.; Heuser, C.; Kronfeld, K.; Seliger, B. Tuning tumor-specific T-cell activation: a matter of costimulation? *Trends Immunol.*, **2002**, 23,240-245.

[8] Nelson, B.H. CD20+ B cells: the other tumor-infiltrating lymphocytes. J. Immunol., 2010, 185, 4977-4982.

[9] Stauss, H.J. Benign autoimmunity to combat malignancy. Clin. Exp. Immunol., 2001, 125, 1-2.

[10] Torchilin, V.; Iakoubov, L.; Estrov, Z. Antinuclear autoantibodies as potential antineoplastic agents. *TRENDS Immunol.*, 2001, 22, 424-427.

[11] Bazhin, A.V.; Shifrina, O.N.; Savchenko, M.S.; Tikhomirova, N.K.; Goncharskaia, M.A.; Gorbunova, V.A.; Senin, I.I.; Chuchalin, A.G.; Philippov, P.P. Low titre autoantibodies against recoverin in sera of patients with small cell lung cancer without a loss of vision. *Lung Cancer*, **2001**, 34, 99-104.

[12] DiLillo, D.J.; Matsushita, T.; Tedder, T.F. B10 cells and regulatory B cells balance immune responses during inflammation, autoimmunity, and cancer. *Ann. N. Y. Acad. Sci.*, **2010**, 1183, 38-57.

[13] Armanasco, E.; Azar, M.E.; Gonzalez, E.; Cresta Morgado, C.; Montoya, D.; Noblia, C.; Ipiña, M.
In: Pautas en Oncología. Diagnóstico, tratamiento y seguimiento del cáncer. Instituto Angel H. Roffo.
UBA. Tumores de mama; Roth, B.; Kirchuk, R.; García Salas, C. Ed. Inst. de Oncologia A.H. Roffo
2008, 202-238.

[14] DeNardo, D.G.; Coussens, L.M. Balancing immune response: crosstalk between adaptive and innate immune cells during breast cancer progression. *Breast Cancer Res.*, **2007**, 9,212.

[15] Hansen, M.; Ostenstad, B.; Sioud, M. The tumor infiltrating B cell response in medullary breast cancer is oligoclonal and directed against the autoantigen actin exposed on the surface of apoptotic cancer cells. *Proc. Natl. Acad. Sci. USA*, **2001**, 98, 12659-12664.

[16] Fernandez Madrid, F. Autoantibodies in breast cancer sera: candidate biomarkers and reporters of tumorigenesis. *Cancer Lett.*, **2005**, 230,187-198.

[17] van Coppen, C.J.; Kaiser, V. Regulation of muscarinic acetylcholine receptors signaling. *Pharmacol. Ther.*, **2003**, 98, 197-200.

[18] Eglen, RM. Muscarinic receptor subtypes in neuronal and non-neuronal cholinergic function. *Auton. Autacoid Pharmacol.*, **2006**, 26, 219-233.

[19] Paleari, L.; Grozio, A.; Cesario, A.; Russo, P. The cholinergic system in cancer. *Sem. Cancer Biol.*, 2008, 18, 211-217.

[20] Español, A.; Eiján, A.; Mazzoni, E.; Davel, L.; Jasnis, M.A.; Sacerdote de Lustig, E.; Sales, M.E. Nitric Oxide Synthase, Arginase and Cyclooxygenase are Involved in Muscarinic Receptors Activation in Different Murine Mammary Adenocarcinoma Cell Lines. *Int. J. Mol. Med.*, **2002**, 9, 651-657.

[21] Rimmaudo, L.; de la Torre, E.; Sacerdote de Lustig, E.; Sales, M.E. mAchR are involved in murine mammary adenocarcinoma cells LMM3 proliferation and angiogenesis. *Biochem. Biophys. Res. Commun.*, 2005, 334, 1360-1365.

[22] Español, A.; de la Torre, E.; Fiszman, G.; Sales, M.E. Role of non-neuronal cholinergic system in breast cancer progression. *Life Sci.*, **2007**, 80, 2281-2285.

[23] Fiszman, G.L.; Sales M.E. Antibodies against muscarinic receptors in breast cancer: agonizing tumor growth. *Curr Immunol Rev.*, **2008**, 4, 176-182.

[24] Fiszman, G.L.; Middonno, M.C.; de la Torre, E.; Farina, M.; Español, A.J.; Sales, M.E. Activation of muscarinic cholinergic receptors induces MCF-7 cells proliferation and angiogenesis by stimulating nitric oxide synthase activity. *Cancer Biol. Ther.*, **2007**, 6, 1106-1113.

[25] Jiménez, E.; Montiel, M. Activation of MAP kinase by muscarinic cholinergic receptors induces cell proliferation and protein synthesis in human breast cancer cells. *J. Cell. Physiol.*, **2005**, 204,678-686.

[26] Grando, S.A. Pemphigus autoimmunity: hypotheses and realities. Autoimmunity., 2012, 45,7-35.

[27] Goin, J.C.; Nathanson, N. Quantitative analysis of muscarinic acetylcholine receptor homo- and heterodimerization in live cells: regulation of receptor down-regulation by heterodimerization. *J. Biol. Chem.*, **2006**, 281, 5416-5425.

[28] Park, P.S.; Wells, J. Oligomeric potential of the M2 muscarinic cholinergic receptor. *J. Neurochem.*,2004, 90, 537-548.

[29] Zhang, J.Y.; Zhu, W.; Imai, H.; Kiyosawa, K.; Chan, E.K.; Tan, E.M. De-novo humoral immune responses to cancer-associated autoantigens during transition from chronic liver disease to hepatocellular carcinoma. *Clin. Exp. Immunol.*, **2001**, 125,3-9.

[30] Lu, P.; Zhu, X.Q.; Xu, Z.L.; Zhou, Q.; Zhang, J.; Wu, F. Increased infiltration of activated tumor-infiltrating lymphocytes after high intensity focused ultrasound ablation of human breast cancer. *Surgery*, 2009, 145, 286-293.

[31] Morton, B.A.; Ramey, W.G.; Paderon, H.; Miller R.E. Monoclonal antibody defined phenotypes of regional lymph node and peripheral blood lymphocyte subpopulations in early breast cancer. *Cancer Res.*, **1986**, 46, 2121-2126.

[32] Wernicke, M. Quantitative morphologic assessment of immunoreactivity in regional lymph nodes of patients with carcinoma of the breast. *Surg. Gynecol. Obstet.*, **1975**, 140,919-924.

[33] Fiszman, G.; Cattaneo, V.; de la Torre, E.; Español, A.; Colombo, L.L.; Sacerdote de Lustig, E.; Sales, M.E. Muscarinic receptors autoantibodies purified from mammary adenocarcinoma bearing mice sera stimulate tumor progression. *Int. Immunopharmac.*, **2006**, 6,1323-1330.

[34] Fiszman, G.; Cattaneo, V.; de la Torre, E.; Colombo, L.; Middonno, M.C.; Sacerdote de Lustig, E.;
Sales, M.E. Autoanticuerpos contra receptores muscarínicos en cáncer de mama. *Química Viva*, 2005, 4, 1-17.

[35] Negroni, M.P.; Fiszman, G.L.; Azar, M.E.; Cresta Morgado, C.; Español, A.J.; Pellegrina, L.T.; de la Torre, E. Sales, M.E. mAChR activity is modulated by auto-antibodies from breast cancer patients in MCF-7 cells. *J. Clin. Immunol.*, **2010**, 30, 474-484.

[36] Wen, Y.; Mancino, A.; Pashov, A.; Whitehead, T.; Stanley, J.; Kieber-Emmons, T. Antigen binding of human IgG Fabs mediates ERK-associated proliferation of human breast cancer cells. *DNA Cell. Biol.*, 2005, 24,73-84.

[37] You, J.; Mi, D.; Zhou, X.; Qiao, L.; Zhang, H.; Zhang, X.; Ye, L. A positive feedback between activated extracellularly regulated kinase and cyclooxygenase/lipoxygenase maintains proliferation and migration of breast cancer cells. *Endocrinology*, **2009**, 150, 1607-1617.

[38] Alvarez-Breckenridge, C.A.; Waite, K.A.; Eng, C. PTEN regulates phospholipase D and phospholipase C. *Hum. Mol. Gen.*, **2007**, 16, 1157-1163.

[39] Lancaster, J.R.; Xie, K. Tumors face NO problems? Cancer Res., 2006, 66, 6459-6462.

[40] Schulze, W.; Kunze, R.; Wallukat, G. Pathophysiological role of autoantibodies against G-proteincoupled receptors in the cardiovascular system. *Exp. Clin. Cardiol.*, **2005**, 10, 170–172.

[41] Sales, M.E.; Español, A.J.; Sterin-Borda, L.; Borda, E.; de Bracco, M.M. Protein kinase C regulates NO-cGMP pathway in muscarinic receptor activation by HIV+-IgA. *Int. J. Mol. Med.*, **1999**, 3, 633–637.

[42] Hernandez, C.C.; Nascimento, J.H.; Chaves, E.A.; Costa, P.C.; Masuda, M.O.; Kurtenbach, E.; Campos, D.E.; Carvalho, A.C.; Gimenez, L.E. Autoantibodies enhance agonist action and binding to cardiac muscarinic receptors in chronic Chagas' disease. *J. Recept. Signal. Transduct. Res.*, **2008**, 28, 375–401.

[43] Wu, Y.; Li, M.T.; Zhao, Y.; Feng, S.; Su, L.; Dong, Y. The clinical significance of autoantibodies against acetylcholine muscarinic 3 receptor in primary Sjögren's syndrome. *Zhonghua Nei Ke Za Zhi.*, 2008, 47, 563–565.

## **Legends of Figures**

Fig. (1): Action of the maximal effective concentration of carbachol or T1N0Mx-IgG on MCF-7 cells proliferation. MCF-7 cells were treated with (A) carbachol ( $10^{-9}$  M), (B) T1N0Mx-IgG (2 mg/ml), (C) normal or benign fibroadenoma (BFA) IgG (2 mg/ml) **ABs IN FIG ARE LISTED IN MICROGRAMS/ML** in the absence or presence of  $10^{-6}$  M: atropine (AT; **mo** non-selective mAChR antagonist), 4-DAMP (M<sub>3</sub> receptor selective antagonist), or tropicamide (TROP; M<sub>4</sub> selective antagonist). Values are mean ± SEM of three experiments performed in triplicate. ###p<0.001, ##p<0.01 vs. cut-off value (represented by a dotted line). \*\*p<0.001, \*p<0.01 vs. carbachol or T1N0Mx-IgG **HOW WAS PROLIFERATION MEASURED? WHAT IS THE CUT-OFF VALUE, AND HOW IS TT DETERMINED? ALSO, GIVEN THIS PRESENTATION, IT SEEMS LIKE CARBACHOL AND THE PATIENT ABs ACTUALLY INHIBIT CELL GROWTH COMPARED TO CONTROL?** 

Fig. (2): Participation of phospholipase C (PLC)/ protein kinase C/ Nitric oxide synthase (NOS) in the proliferative action of carbachol on MCF-7 cells. A) Cells were treated during 1 h with carbachol  $(10^{-9} \text{ M})$  in the absence or presence of 2-nitro-4-carboxyphenyl N,N-diphenylcarbamate (NCDC)  $(10^{-6}\text{ M})$ , N<sup>G</sup>monomethyl-L-arginine (L-NMMA)  $(10^{-4}\text{ M})$  or H-7  $(10^{-7}\text{ M})$ . Values are mean  $\pm$  SEM of 5 experiments performed in triplicate. \* p<0,05; \*\* p<0.01 vs. control (cells without treatment) SHOULDN'T THE CARBACHOL BAR GO ALL THE WAY ACROSS? AND AGAIN, THIS

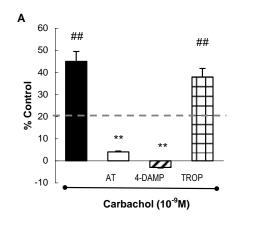
PRESENTATION STYLE MAKE IT LOOK LIKE CARBACHOL IS IHNIBITORY. B) Western

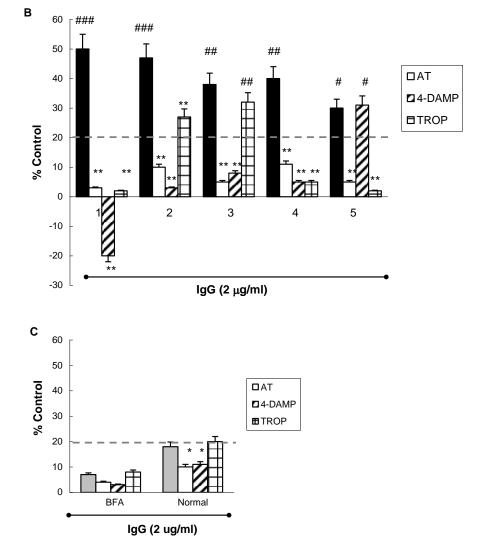
blot assays to detect PLCβ2 and NOS isoforms. Molecular weights of proteins are indicated on the right. One representative experiment of three is shown. glyceraldehyde-3-phosphate dehydrogenase (GAPDH) that was used as loading control.

Fig. (3): A) Inositol monophosphate (IP1) production by MCF-7 cells. Cells were treated with carbachol (CARB) or T1N0Mx-IgG in the absence or presence of  $10^{-5}$ M 2-nitro-4carboxyphenyl N,N-diphenylcarbamate (NCDC) or  $10^{-9}$ M atropine (AT) or 4-DAMP. Values are mean **PRESUME THIS IS +/-?** SEM of three experiments performed in duplicate. \*p<0.001 vs. control (cells without treatment). B) Nitrite (NO<sub>2</sub><sup>-</sup>) production by MCF-7 cells. Cells were treated with carbachol (CARB) or T1N0Mx-IgG in the absence or presence of  $10^{-4}$ M N<sup>G</sup>monomethyl-L-arginine (L-NMMA),  $10^{-3}$ M aminoguanidine (AG) or  $10^{-9}$ M atropine (AT) or 4-DAMP. Values are mean SEM of three experiments performed in duplicate. \*p<0.001 vs. control (cells without treatment).

Fig. (4): Model of anti-muscarinic acetylcholine receptors (mAChR) antibodies action on tumor cells. CARB (carbachol) stimulates mAChR and via Gq/11 protein increases phospholipase C (PLC) activity. **These** This enzyme hydrolyzes phosphoinositides producing myoinositol 1,4,5-trisphosphate (IP3) and finally IP1. The latter increases intracellular calcium levels that in turn activates nitric oxide synthase (NOS) activity that produces nitric oxide (NO). The agonist and autoantibodies (auto-Abs) also increase the synthesis and liberation of vascular endothelial growth factor-A (VEGF-A). AutoAbs could resemble agonist action.

Fig.1





A в 100 90 80 70 % of Control 60 GAPDH 50 40 NOS1 30 20 -160 kDa \*\* 10 0  $\sim$ GAPDH -10 L NCDC H-7 L-NMMA Carbachol 10<sup>-9</sup> M

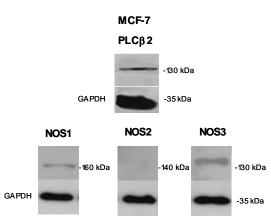


Fig. 2

Fig. 3



