

medicina

BUENOS AIRES Vol. 81 Supl. III - 2021



medicina

BUENOS AIRES, VOL. 81 Supl. III - 2021

COMITÉ DE REDACCIÓN

Sebastián F. Ameriso <i>FLENI, Buenos Aires, Argentina</i>	Caroline A. Lamb <i>Instituto de Biología y Medicina Experimental (IBYME), Buenos Aires, Argentina</i>
Pablo J. Azurmendi <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>	Oscar M. O. Laudanno <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>
Damasia Becú Villalobos <i>Instituto de Biología y Medicina Experimental-CONICET, Buenos Aires, Argentina</i>	Isabel A. Lüthy <i>Instituto de Biología y Medicina Experimental (IBYME), Buenos Aires, Argentina</i>
José H. Casabé <i>Instituto de Cardiología y Cirugía Cardiovascular, Hospital Universitario Fundación Favaloro, Buenos Aires, Argentina</i>	Jorge A. Manni <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>
Hugo N. Catalano <i>Hospital Alemán, Buenos Aires, Argentina</i>	Rodolfo S. Martin <i>Facultad de Ciencias Biomédicas y Hospital Universitario Austral, Buenos Aires, Argentina</i>
Eduardo L. De Vito <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>	Viviana Ritacco <i>Instituto Nacional de Enfermedades Infecciosas ANLIS-CONICET, Buenos Aires, Argentina</i>
Laura I. Jufe <i>Hospital General de Agudos J.M. Ramos Mejía, Buenos Aires, Argentina</i>	Guillermo B. Semeniuk <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>
Isabel Narvaiz Kantor <i>Organización Panamericana de la Salud (OPS/OMS), Argentina</i>	Oswaldo J. Stringa <i>Hospital de Clínicas José de San Martín, UBA, Argentina</i>
Basilio A. Kotsias <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>	
Gustavo Kusminsky <i>Hospital Universitario Austral, Buenos Aires, Argentina</i>	

MIEMBROS EMÉRITOS

Héctor O. Alonso <i>Instituto Cardiovascular Rosario, Santa Fe, Argentina</i>	Christiane Dosne Pasqualini <i>Academia Nacional de Medicina, Buenos Aires, Argentina</i>
María Marta de Elizalde de Bracco <i>IMEX-CONICET-Academia Nacional de Medicina, Buenos Aires, Argentina</i>	Rodolfo C. Puche <i>Facultad de Ciencias Médicas, Universidad Nacional de Rosario, Santa Fe, Argentina</i>
Guillermo Jaim Etcheverry <i>Facultad de Medicina, UBA, Argentina</i>	La Tapa Médanos <i>Daniela Kantor</i>
Daniel A. Manigot <i>Hospital San Juan de Dios, Buenos Aires, Argentina</i>	

MEDICINA (Buenos Aires) - Revista bimestral – ISSN 1669-9106 (En línea)

Registro de la Propiedad Intelectual N° 02683675
Personería Jurídica N° C-7497

Publicación de la Fundación Revista Medicina (Buenos Aires) Propietario de la publicación: Fundación Revista Medicina
Queda hecho el depósito que establece la Ley 11723

Publicada con el apoyo del Ministerio de Ciencia, Tecnología e Innovación Productiva.
MEDICINA no tiene propósitos comerciales. El objeto de su creación ha sido propender al adelanto de la medicina argentina.
Los beneficios que pudieran obtenerse serán aplicados exclusivamente a este fin.
Aparece en MEDLINE (PubMed), ISI-THOMSON REUTERS (Journal Citation Report, Current Contents, Biological Abstracts, Biosis, Life Sciences), CABI (Global Health), ELSEVIER (Scopus, Embase, Excerpta Medica), SciELO, LATINDEX, BVS (Biblioteca Virtual en Salud), DOAJ, Google Scholar y Google Books.
Incluida en el Núcleo Básico de Revistas Científicas Argentinas del CONICET.

Directores Responsables:

Basilio A. Kotsias, Eduardo L. De Vito, Isabel Narvaiz Kantor, Isabel Lüthy

Secretaría de Redacción: Ethel Di Vita, Instituto de Investigaciones Médicas Alfredo Lanari, Combatientes de Malvinas 3150,
1427 Buenos Aires, Argentina
Tel. 5287-3827 Int. 73919 y 4523-6619
e-mail: revmedbuenosaires@gmail.com – http://www.medicinabuenosaires.com

Vol. 81, Supl. III, Noviembre 2021

Diagramación y Diseño: Andrés Esteban Zapata - aez.sji@gmail.com

REUNIÓN DE SOCIEDADES DE BIOCENCIAS 2021

**LXVI REUNIÓN ANUAL DE LA
SOCIEDAD ARGENTINA DE INVESTIGACIÓN CLÍNICA (SAIC)**

**LXIX REUNIÓN ANUAL DE LA
SOCIEDAD ARGENTINA DE INMUNOLOGÍA (SAI)**

**LIII REUNIÓN ANUAL DE LA
ASOCIACIÓN ARGENTINA DE FARMACOLOGÍA EXPERIMENTAL (AAFE)**

**XI REUNIÓN ANUAL DE LA
ASOCIACIÓN ARGENTINA DE NANOMEDICINAS
(NANOMED-AR)**

17-20 de noviembre de 2021

EDITORES RESPONSABLES
Dr. Alejandro Curino Presidente
Dra. Mariana Maccioni
Dra. Paula Schaiquevich
Dra. Hebe Duran

ANNUAL MEETING OF BIOSCIENCE SOCIETIES 2021

**LXVI ANNUAL MEETING OF
SOCIEDAD ARGENTINA DE INVESTIGACIÓN CLÍNICA (SAIC)**

**LXIX ANNUAL MEETING OF
SOCIEDAD ARGENTINA DE INMUNOLOGÍA (SAI)**

**LIII ANNUAL MEETING OF
ASOCIACIÓN ARGENTINA DE FARMACOLOGÍA EXPERIMENTAL (AAFE)**

**XI ANNUAL MEETING OF
ASOCIACIÓN ARGENTINA DE NANOMEDICINAS
(NANOMED-AR)**

November 17-20, 2021

RESPONSIBLE EDITORS
Dr. Alejandro Curino Presidente
Dra. Mariana Maccioni
Dra. Paula Schaiquevich
Dra. Hebe Duran

LA TAPA

Daniela Kantor. Médanos, 2018

Técnica: Acrílico sobre cartón entelado. Medidas: 20x28 cm

Daniela Kantor nació el 23 de marzo de 1970. Es diseñadora gráfica (FADU-UBA), pintora, dibujante, historietista e ilustradora. Autora de la novela gráfica *Mujer Primeriza* (Ed. Burslesque, 2014) y *Aprendiza* (2019), *Naturella* (con guión de Arekasadaro, 2017) publicada en *Dis-Tinta* (Ed. Sudamericana, coordinado por Liniers y Martín Pérez). Con guion de Alejandro Fariás dibujó *Las moradas de Santa Teresa de Jesús* en historietas (Ed. Loco rabia + CCEBA Centro Cultural de España en Buenos Aires) y *Marilyn* (*Tren en movimiento*, 2019). Es miembro de la revista de historietas "El Tripero" fundada en 1993 junto al grupo de alumnos de Alberto Breccia. En el ámbito de la enseñanza es Jefa de Trabajos Prácticos en la materia Ilustración inicial, y docente en Ilustración Editorial en la Facultad de Arquitectura, Diseño y Urbanismo FADU/UBA. Dicta talleres sobre pintura e ilustración (C C Recoleta, 2019/ Quinta Trabucco, 2020/ taller particular junto a Daniel Roldán, 2019). Es maestra de niños y niñas en Dibujo e Historieta en Escuelas primarias, talleres (Filbita, Festival de literatura de Buenos Aires, 2018-9/ CCK, 2018/ taller propio desde 2014). Estudió Dibujo de Historieta con Alberto Breccia, Técnicas de Acuarela y Pastel con Carlos Nine, charlas sobre Historieta con José Muñoz, Curso de Color con Carlos Gorriarena, Clínica de Pintura con Mariano Sapia y Tulio de Sagastizábal, y Sumi-e en el Centro Okinawense. Trabaja para editoriales y revistas con ilustraciones e historietas (Ed. Troquel, Abran Cancha, Ed. Norma, Unicef, Barcelona, Crisis, Suplemento Ñ/ Clarín, Borges en la Biblioteca Nacional- Lectores de Borges). Fue invitada a la Feria del libro de los Universitarios de UNAM para presentar el libro "Palabra de ilustrador", y en 2019 ganó la Beca UBA Internacional en el marco de un programa de intercambio docente con la Universidad Regiomontana, Monterrey, México.

Fuentes: <https://www.instagram.com/daniela.kantor.9/>; www.kantorconk.blogspot.com

CONSEJOS DIRECTIVOS

SAIC

Presidente
Alejandro Curino

Vicepresidente
Daniel Alonso

Secretario
Alejandro Urtreger

Tesorera
Laura Todaro

Prosecretaria
Stella Ranuncolo

Vocales
Evangelina Capobianco
María del Rocío Castilla Lozano
Pablo Gravina
Adriana Casas
Julieta Maymo
María Marta Amaral
Ricardo Cabrera
Sandra Ferreira
Marcela Bolontrade
Adriana Burgueño
Julia Halperin
Luis Di Ciano
María Laura Ruiz

Revisores de cuentas
Gabriela Lombardi
Mariela Pérez

SAI

Presidenta
Mariana Maccioni

Vicepresidente
Emilio Malchiodi

Secretaria
Silvia Correa

Tesorera
Mercedes Fuertes

Prosecretaria
Mariana Salatino

Protesorera
Marisa Castro

Vocales
Mercedes Borge
Karina Canziani
Esteban Grasso
Carolina Maldonado Galdeano
Gerardo Mirkin
Verónica Natoli
María Silvia Ventimiglia
Silvina Villar

AAFE

Presidenta

Paula Schaiquevich

Vicepresidente

Ventura Simonovich

Secretaria

Myriam Laconi

Tesorera

Susana Gorzalczany

Prosecretaria

Daniela Quinteros

Vocales

Carlos Reyes Toso

Fatima Nader

Santiago Palma

Revisores de cuentas

Héctor Alejandro Serra

María Victoria Aguirre

Revisores de cuentas

(suplentes)

Andrea Errasti

Ariel Perelsztein

NANOMED-ar

Presidenta

Hebe Durán

Vicepresidenta

Romina Glisoni

Secretaria

Leticia Higa

Tesorera

Julia Altube

Vocales

Dr. Eder Romero

Dra. Mariela Agotegaray

Vocal (suplente)

Dra. Priscila Schilreff

Revisora de cuentas

Dra. Marisa Taverna Porro

Revisora de cuentas

(suplente)

María José Morilla

**LAS SOCIEDADES QUE ORGANIZAN
ESTA REUNIÓN CONJUNTA
AGRADECEN EL APOYO DE**

INSTITUCIONES OFICIALES

**CONSEJO NACIONAL DE INVESTIGACIONES CIENTÍFICAS Y TÉCNICAS
MINISTERIO DE CIENCIA, TECNOLOGÍA E INNOVACIÓN PRODUCTIVA
AGENCIA NACIONAL DE PROMOCIÓN DE LA INVESTIGACIÓN,
EL DESARROLLO TECNOLÓGICO Y LA INNOVACIÓN**

OTRAS INSTITUCIONES Y AUSPICIANTES

**FUNDACIÓN CHERNY
FUNDACIÓN HONORIO BIGAND
LABORATORIO GADOR S.A.
ETC INTERNACIONAL S.A.
LABORATORIO DE HEMODERIVADOS-UNC
ARCOR S.A.
FUNDACIÓN JOSÉ A. BALSEIRO
FUNDACIÓN ARGENTINA DE NANOTECNOLOGÍA
ATOM-PROTECT®**

not show positive selection among all studied phylogenetic groups. These results strengthen the idea that ZPs are proteins that provide a more specific-specificity isolation than IZUMO1R, which is specialized in fusion events.

Altogether, our findings indicate that sperm-oocyte interaction and fusion proteins lack the degree of diversification necessary to fix a prezygotic reproductive isolation barrier in felidae.

37. (498) PROTEOMIC STUDY OF BREAST CANCER CELL LINE AFTER HEMIN TREATMENT

Schweitzer K¹, Coló GP¹, Fernandez Chavez L¹, Alonso EG¹, Fässler R², Curino AC¹, Facchinetti MM¹

¹ Laboratorio de Biología del Cáncer – Instituto de Investigaciones Bioquímicas Bahía Blanca (INIBIBB) Universidad Nacional del Sur (UNSCONICET). Departamento de Biología, Bioquímica y Farmacia.

² Department of Molecular Medicine Max Planck Institute of Biochemistry (MPI), Martinsried, Germany.

Hemin is a ferriporphyrin (C₃₄H₃₂ClFeN₄O₄) with antitumoral effects on prostate, breast and colon cancer. It is widely used to increase the expression and activity of hemoxygenase-1 (HO-1). HO-1 is a microsomal enzyme that catalyses the degradation of heme group and can be translocated to subcellular compartments. Our laboratory, among others, has demonstrated that HO-1 regulates several processes related to cancer progression. The aim of this work was to study the protein expression modification due to hemin treatment in a murine breast cancer cell line (LM3). After hemin or vehicle (control) cell treatment, we performed Mass Spectrometry (MS) and Perseus proteome analyses. The proteome analysis showed that 1033 from 7292 proteins detected were modulated after hemin treatment. We observed that 595 proteins were significantly increased, including HO-1, and 353 proteins were significantly decreased in the group treated with hemin respect to their controls ($p < 0.05$). ANOVA significant proteins reveal an upregulated group of proteins related with lipid and iron metabolism. In the group of proteins whose expression decreased after hemin treatment, we found cytoskeleton-related proteins. The Perseus-MS-data analysis revealed that hemin treatment regulates adhesion proteins like vimentin and talin, actin and tubulin cytoskeletal proteins and their stabilizing proteins. In addition, from MS data after hemin treatment, we found an increase in some cancer suppressor proteins such as PTEN and SMAD2/3. Finally, we found that proteases involved in HO-1 nuclear translocation were upregulated after hemin treatment. We further corroborated some of the *in-silico* analysis in LM3 cell line by immunofluorescence and Western blot techniques. In addition, we used a syngeneic LM3 mice model to detect by immunohistochemistry some of the regulated protein. These results show the multiple physiological effects that pharmacological modulation with hemin has in a breast cancer cell line.

38. (501) ENZYMES INVOLVED IN EXTRACELLULAR MATRIX PROTEOGLYCAN'S SYNTHESIS AS A POTENTIAL THERAPEUTIC TARGET FOR COLORECTAL CANCER STEM CELLS

Ariadna BIROCCO¹, Agustín BLACHMAN¹, Nicole ZLOTOW¹, Sofia CURCIO¹, Graciela C. CALABRESE^{1,2}

¹ Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Departamento de Ciencias Biológicas, Catedra de Biología Celular y Molecular. ² IQUIFIB (UBA-Conicet)

Cancer Stem Cells (CSC) are characterized by self-renewal, differentiation, chemoresistance and phenotypic reversibility, which is associated with worse prognosis in tumors. Interaction with the micro-environment is one of the factors related with stemness. The aim of the present work is to analyze the expression of extracellular matrix proteoglycans in CSC derived from cell lines. CSC were enriched by scaffold free 3D culture (colonospheres) of human colorectal cancer cell line HCT116 in the presence of bFGF and EGF, employing ultra-low attachment plates in serum-free culture conditions. After 14 days of culture, microscopy studies were performed to assess colonosphere formation. Stemness was addressed by the expression of master genes SOX2, Nanog and CD44 by RT-PCR. Decorin and

biglycan proteoglycan's core protein expression was also analyzed by RT-PCR. Moreover, glycosaminoglycan and protein quantification was addressed by ionic exchange chromatography of the culture medium followed by colorimetric determination. RT-PCR was performed for the study of glycosaminoglycan synthesis enzyme expression. Bright light microscopy showed colonospheres around 50-100µm. The expression of master genes was heterogeneous among cultures correlated with an heterogeneous expression of decorin (number of experiments, $n = 3$). On the other hand, no biglycan expression was detected among different colonospheres ($n = 3$). No differences were registered in glycosaminoglycan/protein ratio among spheres ($0,274 \pm 0,127$). Nevertheless, Chondroitin-4-O-Sulfotransferase (C4ST) expression was detected in colonospheres while no expression was observed for Dermatan-4-O-Sulfotransferase (D4ST). The heterogeneity presented by 3D cultures represents the heterogeneity reported for CSC within tumors and C4ST and D4ST pattern suggests differences in GAG chain's quality. Therefore, colonospheres are a suitable model for the study of GAG enzymes as potential therapeutic targets.

39. (522) A PROTEOMIC STUDY REVEALS NEW MARKERS OF PROLACTIN-MODULATED OVARIAN FUNCTION IN PLAINS VICZACHAS

Cortasa, Santiago^{1,2}, Feehan, Kevin¹, Schmidt, Alejandro^{1,2}, Vitullo, Alfredo^{1,2}, Dorfman, Verónica^{1,2}, Halperin, Julia^{1,2}

¹ Centro de Estudios Biomédicos Básicos, Aplicados y Desarrollo (CEBBAD), Universidad Maimónides, Buenos Aires, Argentina

² Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Argentina

Prolactin (PRL) modulates the expression of the LH receptor in the ovary and, thus, the cascades of steroidogenic enzymes that synthesize and produce ovarian steroids. When pathological hyperprolactinemia occurs, the pulsatile release of GnRH decreases and alters the pituitary production of FSH and LH. Furthermore, it directly impairs the endocrine activity of ovarian follicles. To analyze which ovarian factors, aside to the aforementioned enzymes, respond to a high PRL environment, an ovarian proteomic study of viczachas with sulpiride-induced hyperprolactinemia was performed. For this, ovarian protein extracts from hyperprolactinemic (HPRL) and control (CTL) females ($n = 5$ per group) were used. Briefly, equal amounts of protein were analyzed using MALDI-TOF/MS and then, LC-ESI/MS (Orbitrap). The resulting peptides were identified with Proteome Discoverer Software using the Rodentia UniProt Database, and functional enrichment analysis was performed using DAVID, STRING and FunRich softwares. Proteins differentially expressed in each treatment were depicted in a volcano plot (t -test, $p < 0.05$). Functional enrichment analysis showed that 24 proteins were differentially expressed in HPRL ovarian tissue compared to that of CTLs. Among those, some cytoskeleton regulation markers such as annexin 2 (ANXA2), Actin related protein 2/3 complex subunit 5 like (Arpc5l), and Myosin regulatory light chain 12A (MYLC12A) prevailed in HPRL-ovaries, while other markers related to mitochondrial function as Dynamin-1-like protein (DNM1L), Succinate-CoA ligase (SUCLG2), and Mitochondrial fission 1 protein (FIS1) were down-regulated. In addition, the interactomes showed different network topology with different nodal peptides in HPRL vs CTL treatments. The present work showed an ovarian expression profile that significantly varies under a hyperprolactinemic environment. Finally, this report provides new markers for future investigations on PRL-dependent modulation of ovarian function.

40. (565) ANALYSIS OF REGULATORY CIS-ELEMENTS AND FOXA1/GATA2 TRANSCRIPTION FACTORS BINDING BEHIND MRP4/ABCC4 LEVELS IN PANCREATIC CANCER

Samanta N Gancedo¹, Ana Sañores¹, Natalia Gómez¹, Maximiliano De Sousa-Ferro¹, Carlos Davio¹, and Betina González¹

¹ Instituto de Investigaciones Farmacológicas, Consejo Nacional de Investigaciones Científicas y Técnicas- Universidad de Buenos Aires (CONICET-UBA), Buenos Aires, Argentina.