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Jorge Quarleri, Cintia Cevallos



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# Influence of aging on T cell response and renin-angiotensin system imbalance during SARS-CoV-2 infection

Jorge Quarleri<sup>1\*</sup>, Cintia Cevallos<sup>1</sup>, and M. Victoria Delpino<sup>2\*</sup>

<sup>1</sup> Instituto de Investigaciones Biomédicas en Retrovirus y Sida (INBIRS). Universidad de Buenos Aires. CONICET. Buenos Aires, Argentina.

<sup>2</sup> Instituto de Inmunología, Genética y Metabolismo (INIGEM). Universidad de Buenos Aires. CONICET. Buenos Aires, Argentina.

\*These authors contributed equally to the manuscript. Correspondence to Jorge F. Quarleri, quarleri@fmed.uba.ar, and M. Victoria Delpino, mdelpino@ffyb.uba.ar

## Highlights

- Th17 derived cytokines increase lung pathology in COVID-19 patients
- CD8+ T lymphocytes counts decrease in patients with severe COVID-19
- Checkpoint overexpression on T lymphocytes occurs in severe COVID-19 patients
- Increase of proinflammatory T cells appear in patients with low levels of Ang1-7
- ACE2 downmodulation is observed in patients with severe COVID-19

The viral clearance and the long-term antiviral immunity require an adequate T cell-mediated adaptive immune response. Paradoxically, they can also contribute to the cytokine storm observed in COVID-19 patients [1-3]. The cytokine storm during COVID-19 induces Th17 response promoting vascular permeability [4]. Additionally, Th1, NK, NKT, and CD8+ T lymphocytes along with other innate immune cells that target virus-infected cells may be overstimulated producing tissue damage [5]. Activated T lymphocytes express increased levels of inhibitory immune checkpoints TIM-3, CTLA-4, PD-1, and TIGIT [6]. The persistence of stimulation induces the expression of the inhibitory immune checkpoint sustained in the time with the progressive loss of T lymphocyte effector function, or exhaustion [7, 8]. Studies

performed in SARS-CoV-2 infected patients revealed the presence in the bloodstream of higher quantities of activated CD4+HLADR+CD38+ T cells and CD4+PD-1+CD57+ exhausted or senescent T cells with respect to healthy controls [9]. Accordingly, patients with severe COVID-19 have CD4+ T cells that express lower levels of IFN- $\gamma$ , TNF- $\alpha$ , and IL-2 than mild patients, or healthy controls [10]; and CD8+ T cells with high levels of CTLA-4, PD-1, TIGIT, granzyme B, and perforin were found in patients severely ill compared the mild group [10].

Older individuals are more predisposed to suffer infectious diseases. In effect, up to one-third of deaths in aging is a consequence of infectious diseases [11]. The persistence of viral infections throughout life can trigger monoclonal expansion of T cells resulting in low memory T cell variability that can lead to immune exhaustion [12]. This is a problem besides new infectious agents like SARS-CoV-2. Elevated levels of inflammation are characteristic of severe cases of COVID-19 can result in pneumonia with a compromise of the integrity and function of lung tissue. Furthermore, these exhausted T cells preferentially secrete pro-inflammatory cytokines such as TNF- $\alpha$  and IFN- $\gamma$  [12]. These, together with the cytokines secreted by cells of innate immunity, contribute to the mild inflammatory profile seen in elderly individuals [13].

During acute pulmonary infection diseases, the renin-angiotensin system (RAS) participates in the development of ARDS with consequent pulmonary fibrosis [14], a condition frequently observed in COVID-19 patients [15]. Angiotensin converting enzyme (ACE)-2 is the main receptor for SARS-CoV-2 and also a critical link between immune response and inflammation [16]. ACE2 cleaves angiotensin (Ang)II into Ang1-9 and Ang1-7, inactivating AngII [17]. Ang1-7 antagonizes inflammatory responses [18, 19]. It is possible that, under physiological conditions, ACE2 inhibits the production of IL-6 induced by AngII, with the consequent reduction of Th17 cells [20, 21]. Additionally, AngII induces TGF- $\beta$  expression [22, 23] involved in the differentiation of Th17 cells and lead to the production of massive quantities of proinflammatory cytokines and chemokines [4, 24]. It has been reported in COVID-19 associated with ARDS an increment of the highly proinflammatory CCR6+ Th17 subpopulation [25]. As well, the severity of lung injury shows a significant correlation with the amount of IL-17 [26].

The Ang1-7 controls the phosphorylation of extracellular signal-regulated kinase 1/2 (ERK1/2) [27], thus regulating the IL-10 production. This cytokine induces the differentiation of Th0 cells to Th2 type [28] and, additionally acts as anti-inflammatory factor preventing tissue damage [29, 30]. Th2 cells modulate the immune responses via the production of anti-

inflammatory cytokines such as IL-5, IL-4, IL-9, and IL-13 [31]. During COVID-19 disease the increase in the proinflammatory T cell response correlated with the low circulating plasma levels of Ang1-7 [32]. A recent molecular model of the impact of SARS-CoV-2 infection on RAS predicts a quantitative reduction in ARDS severity in COVID-19 patients, in agreement with the known anti-inflammation and anti-fibrosis nature of Ang1-7 [33].

Previous evidence obtained from SARS-CoV indicates that the infection can down modulate ACE2 expression on cells [14, 34-36]. This mechanism may be involved in COVID-19 and explains the multiple organ injury produced through an increased release of chemokines and proinflammatory cytokines with the increase of vascular permeability and consequent neutrophil migration to the lung [37]. It is still controversial the influence of aging on ACE2 expression. The increase in the susceptibility to cardiovascular disease and vascular injury that affects preferentially the elderly individuals could be explaining by the down regulated ACE2 in aging, as was suggested by several studies [38-40]. However, such age-dependent variation in the ACE2 levels can vary according to cell type. Thereby, an ACE2 depletion in the aortas and kidneys of aging female mice was observed. Similarly, this animal model has shown in the lungs the lowest level of ACE2 when compared with that in the heart, brain, and kidneys [41]. Other conditions such as dietary elements such as high potassium intake decreased ACE2 gene expression [42]. Moreover, the host genetic components that define ACE2 variants also influence its expression levels, which may also impact on COVID-19 outcome [43]. Other host-related conditions such as patients with type II diabetes with severe COVID-19 outcome have revealed diminished levels of ACE2 [44]. In contrast, those persons routinely treated with drugs belongs to Angiotensin-converting enzyme inhibitors (ACEi) and/or Angiotensin II type I receptor blockers (ARBs) have noticed a significant increase in ACE2 expression [46].

Although the expression of ACE2 could be biased by gender, no differences in the activity of this enzyme in the lungs have been demonstrated in the murine model [45]. Besides, in rats, the levels of ACE2 were dramatically reduced with aging in both genders, but with significantly higher expression in old female rats than male [40].

ACE2 protein is expressed in various human organs including oral and nasal mucosa, nasopharynx, lung, stomach, small intestine, colon, skin, lymph nodes, thymus, bone marrow, spleen, liver, kidney, and brain [47]. There was little to no expression of ACE2 on most of the human peripheral blood-derived immune cells including CD4<sup>+</sup> T, CD8<sup>+</sup> T, activated CD4<sup>+</sup>/CD8<sup>+</sup> T, Tregs, Th17, NKT, B, NK cells, monocytes, dendritic cells, and granulocytes. However, it was observed high expressions of ACE2 on human tissue macrophages, such as

alveolar macrophages, liver Kupffer cells, and microglial cells in steady state brain [48, 49]. Thus, it is relevant to consider potential unfavorable effects of SARS-CoV2 on macrophages, using these cells as a Trojan horse, enabling viral anchoring specifically within the pulmonary parenchyma, which may govern the severity of SARS-CoV-2 infection. Moreover, reallocation of viral-containing macrophages migrating out of the lung to other tissues is theoretically plausible in the context of viral spread with the involvement of other organs [50].

This absence of correlation between ACE2 expression and infection susceptibility appears to be contradictory since ACE2 is the entryway to SARS-CoV-2 indicating that the larger expression of ACE2 in the membrane is in line with larger infectivity. However, COVID-19 elderly patients have superior severity of lung damage and a higher lethality rate in comparison with young people [1]. Then, young individuals are more susceptible to have the infection while old people with lower ACE2 expression could have a more severe condition in the case of infection due to the exacerbated effect mediated by the inhibition of AngII processing [51] even with individual variations on its expression level [52].

Aging is also associated with exuberant inflammatory cytokines secreted by senescent nonlymphoid cells that may cause organ dysfunction in humans and animal models [53]. Such excessive inflammation can inhibit antigen-specific immunity in vivo. Since (mammalian target of rapamycin) mTOR pathway is involved in the increase of the expression of IL-6 receptor and IL-6 secretion [54], the negative impact of inflammation on immunity during aging can be reversed in part by treatment with mTOR inhibitor rapamycin [55].

This review constitutes the initial background to understand how SARS-CoV-2 infection deregulates RAS in severe cases, and the impact on T cells and ARDS will enable novel therapeutic strategies to control the disease progression (Figure 1).

## References

- [1] G. Chen, D. Wu, W. Guo, Y. Cao, D. Huang, H. Wang, T. Wang, X. Zhang, H. Chen, H. Yu, X. Zhang, M. Zhang, S. Wu, J. Song, T. Chen, M. Han, S. Li, X. Luo, J. Zhao, Q. Ning, Clinical and immunological features of severe and moderate coronavirus disease 2019, *J Clin Invest* 130(5) (2020) 2620-2629.
- [2] B. Diao, C. Wang, Y. Tan, X. Chen, Y. Liu, L. Ning, L. Chen, M. Li, Y. Liu, G. Wang, Z. Yuan, Z. Feng, Y. Zhang, Y. Wu, Y. Chen, Reduction and Functional Exhaustion of T Cells in Patients With Coronavirus Disease 2019 (COVID-19), *Front Immunol* 11 (2020) 827.
- [3] S.A. Vardhana, J.D. Wolchok, The many faces of the anti-COVID immune response, *J Exp Med* 217(6) (2020).
- [4] O. Pacha, M.A. Sallman, S.E. Evans, COVID-19: a case for inhibiting IL-17?, *Nat Rev Immunol* 20(6) (2020) 345-346.
- [5] J.J.M. Wong, J.Y. Leong, J.H. Lee, S. Albani, J.G. Yeo, Insights into the immuno-pathogenesis of acute respiratory distress syndrome, *Ann Transl Med* 7(19) (2019) 504.
- [6] L. Chen, D.B. Flies, Molecular mechanisms of T cell co-stimulation and co-inhibition, *Nat Rev Immunol* 13(4) (2013) 227-42.
- [7] F. Chiappelli, A. Khakshooy, G. Greenberg, CoViD-19 Immunopathology and Immunotherapy, *Bioinformatics* 16(3) (2020) 219-222.
- [8] A. Saeidi, K. Zandi, Y.Y. Cheok, H. Saeidi, W.F. Wong, C.Y.Q. Lee, H.C. Cheong, Y.K. Yong, M. Larsson, E.M. Shankar, T-Cell Exhaustion in Chronic Infections: Reversing the State of Exhaustion and Reinvigorating Optimal Protective Immune Responses, *Front Immunol* 9 (2018) 2569.
- [9] D. Weiskopf, K.S. Schmitz, M.P. Raadsen, A. Grifoni, N.M.A. Okba, H. Endeman, J.P.C. van den Akker, R. Molenkamp, M.P.G. Koopmans, E.C.M. van Gorp, B.L. Haagmans, R.L. de Swart, A. Sette, R.D. de Vries, Phenotype and kinetics of SARS-CoV-2-specific T cells in COVID-19 patients with acute respiratory distress syndrome, *Sci Immunol* 5(48) (2020).
- [10] H.Y. Zheng, M. Zhang, C.X. Yang, N. Zhang, X.C. Wang, X.P. Yang, X.Q. Dong, Y.T. Zheng, Elevated exhaustion levels and reduced functional diversity of T cells in peripheral blood may predict severe progression in COVID-19 patients, *Cell Mol Immunol* 17(5) (2020) 541-543.
- [11] K.A. Kline, D.M. Bowdish, Infection in an aging population, *Curr Opin Microbiol* 29 (2016) 63-7.
- [12] S. Brunner, D. Herndler-Brandstetter, B. Weinberger, B. Grubeck-Loebenstien, Persistent viral infections and immune aging, *Ageing Res Rev* 10(3) (2011) 362-9.
- [13] C. Franceschi, M. Capri, D. Monti, S. Giunta, F. Olivieri, F. Sevini, M.P. Panourgia, L. Invidia, L. Celani, M. Scurti, E. Cevenini, G.C. Castellani, S. Salvioli, Inflammaging and anti-inflammaging: a systemic perspective on aging and longevity emerged from studies in humans, *Mech Ageing Dev* 128(1) (2007) 92-105.
- [14] K. Kuba, Y. Imai, J.M. Penninger, Angiotensin-converting enzyme 2 in lung diseases, *Curr Opin Pharmacol* 6(3) (2006) 271-6.
- [15] J. Zhu, P. Ji, J. Pang, Z. Zhong, H. Li, C. He, J. Zhang, C. Zhao, Clinical characteristics of 3062 COVID-19 patients: A meta-analysis, *J Med Virol* (2020).
- [16] A.C. Mathewson, A. Bishop, Y. Yao, F. Kemp, J. Ren, H. Chen, X. Xu, B. Berkhout, L. van der Hoek, I.M. Jones, Interaction of severe acute respiratory syndrome-coronavirus and NL63 coronavirus spike proteins with angiotensin converting enzyme-2, *J Gen Virol* 89(Pt 11) (2008) 2741-2745.

- [17] K. Sriram, P.A. Insel, A hypothesis for pathobiology and treatment of COVID-19: The centrality of ACE1/ACE2 imbalance, *Br J Pharmacol* 177(21) (2020) 4825-4844.
- [18] J. Hrenak, L. Paulis, F. Simko, Angiotensin A/Alamandine/MrgD Axis: Another Clue to Understanding Cardiovascular Pathophysiology, *Int J Mol Sci* 17(7) (2016).
- [19] S.S. Karnik, K.D. Singh, K. Tirupula, H. Unal, Significance of angiotensin 1-7 coupling with MAS1 receptor and other GPCRs to the renin-angiotensin system: IUPHAR Review 22, *Br J Pharmacol* 174(9) (2017) 737-753.
- [20] J.M. Luther, J.V. Gainer, L.J. Murphey, C. Yu, D.E. Vaughan, J.D. Morrow, N.J. Brown, Angiotensin II induces interleukin-6 in humans through a mineralocorticoid receptor-dependent mechanism, *Hypertension* 48(6) (2006) 1050-7.
- [21] M.S. Madhur, H.E. Lob, L.A. McCann, Y. Iwakura, Y. Blinder, T.J. Guzik, D.G. Harrison, Interleukin 17 promotes angiotensin II-induced hypertension and vascular dysfunction, *Hypertension* 55(2) (2010) 500-7.
- [22] T. Ehanire, L. Ren, J. Bond, M. Medina, G. Li, L. Bashirov, L. Chen, G. Kokosis, M. Ibrahim, A. Selim, G.C. Blobbe, H. Levinson, Angiotensin II stimulates canonical TGF-beta signaling pathway through angiotensin type 1 receptor to induce granulation tissue contraction, *J Mol Med (Berl)* 93(3) (2015) 289-302.
- [23] T.V. Lanz, Z. Ding, P.P. Ho, J. Luo, A.N. Agrawal, H. Srinagesh, R. Axtell, H. Zhang, M. Platten, T. Wyss-Coray, L. Steinman, Angiotensin II sustains brain inflammation in mice via TGF-beta, *J Clin Invest* 120(8) (2010) 2782-94.
- [24] D. Wu, X.O. Yang, TH17 responses in cytokine storm of COVID-19: An emerging target of JAK2 inhibitor Fedratinib, *J Microbiol Immunol Infect* 53(3) (2020) 368-370.
- [25] Z. Xu, L. Shi, Y. Wang, J. Zhang, L. Huang, C. Zhang, S. Liu, P. Zhao, H. Liu, L. Zhu, Y. Tai, C. Bai, T. Gao, J. Song, P. Xia, J. Dong, J. Zhao, F.S. Wang, Pathological findings of COVID-19 associated with acute respiratory distress syndrome, *Lancet Respir Med* 8(4) (2020) 420-422.
- [26] Y. Liu, Y. Yang, C. Zhang, F. Huang, F. Wang, J. Yuan, Z. Wang, J. Li, J. Li, C. Feng, Z. Zhang, L. Wang, L. Peng, L. Chen, Y. Qin, D. Zhao, S. Tan, L. Yin, J. Xu, C. Zhou, C. Jiang, L. Liu, Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury, *Sci China Life Sci* 63(3) (2020) 364-374.
- [27] P.E. Gallagher, E.A. Tallant, Inhibition of human lung cancer cell growth by angiotensin-(1-7), *Carcinogenesis* 25(11) (2004) 2045-52.
- [28] C.F. Chang, W.N. D'Souza, I.L. Ch'en, G. Pages, J. Pouyssegur, S.M. Hedrick, Polar opposites: Erk direction of CD4 T cell subsets, *J Immunol* 189(2) (2012) 721-31.
- [29] M. Guilleams, K. Movahedi, T. Bosschaerts, T. VandenDriessche, M.K. Chuah, M. Herin, A. Acosta-Sanchez, L. Ma, M. Moser, J.A. Van Ginderachter, L. Brys, P. De Baetselier, A. Beschin, IL-10 dampens TNF/inducible nitric oxide synthase-producing dendritic cell-mediated pathogenicity during parasitic infection, *J Immunol* 182(2) (2009) 1107-18.
- [30] J.J. Haddad, N.E. Saade, B. Safieh-Garabedian, Interleukin-10 and the regulation of mitogen-activated protein kinases: are these signalling modules targets for the anti-inflammatory action of this cytokine?, *Cell Signal* 15(3) (2003) 255-67.
- [31] T.R. Mosmann, J.J. Kobie, F.E. Lee, S.A. Quataert, T helper cytokine patterns: defined subsets, random expression, and external modulation, *Immunol Res* 45(2-3) (2009) 173-84.
- [32] B.M. Henry, J.L. Benoit, B.A. Berger, C. Pulvino, C.J. Lavie, G. Lippi, S.W. Benoit, Coronavirus disease 2019 is associated with low circulating plasma levels of angiotensin 1 and angiotensin 1,7, *J Med Virol* (2020).
- [33] F. Pucci, P. Bogaerts, M. Rooman, Modeling the Molecular Impact of SARS-CoV-2 Infection on the Renin-Angiotensin System, *Viruses* 12(12) (2020).



- [34] S. Haga, N. Yamamoto, C. Nakai-Murakami, Y. Osawa, K. Tokunaga, T. Sata, N. Yamamoto, T. Sasazuki, Y. Ishizaka, Modulation of TNF-alpha-converting enzyme by the spike protein of SARS-CoV and ACE2 induces TNF-alpha production and facilitates viral entry, *Proc Natl Acad Sci U S A* 105(22) (2008) 7809-14.
- [35] I. Glowacka, S. Bertram, P. Herzog, S. Pfefferle, I. Steffen, M.O. Muench, G. Simmons, H. Hofmann, T. Kuri, F. Weber, J. Eichler, C. Drosten, S. Pohlmann, Differential downregulation of ACE2 by the spike proteins of severe acute respiratory syndrome coronavirus and human coronavirus NL63, *J Virol* 84(2) (2010) 1198-205.
- [36] G.Y. Oudit, Y. Imai, K. Kuba, J.W. Scholey, J.M. Penninger, The role of ACE2 in pulmonary diseases--relevance for the nephrologist, *Nephrol Dial Transplant* 24(5) (2009) 1362-5.
- [37] W. Ni, X. Yang, D. Yang, J. Bao, R. Li, Y. Xiao, C. Hou, H. Wang, J. Liu, D. Yang, Y. Xu, Z. Cao, Z. Gao, Role of angiotensin-converting enzyme 2 (ACE2) in COVID-19, *Crit Care* 24(1) (2020) 422.
- [38] G. Jia, A.R. Aroor, C. Jia, J.R. Sowers, Endothelial cell senescence in aging-related vascular dysfunction, *Biochim Biophys Acta Mol Basis Dis* 1865(7) (2019) 1802-1809.
- [39] T. Chen, Z. Dai, P. Mo, X. Li, Z. Ma, S. Song, X. Chen, M. Luo, K. Liang, S. Gao, Y. Zhang, L. Deng, Y. Xiong, Clinical Characteristics and Outcomes of Older Patients with Coronavirus Disease 2019 (COVID-19) in Wuhan, China: A Single-Centered, Retrospective Study, *J Gerontol A Biol Sci Med Sci* 75(9) (2020) 1788-1795.
- [40] X. Xie, J. Chen, X. Wang, F. Zhang, Y. Liu, Age- and gender-related difference of ACE2 expression in rat lung, *Life Sci* 78(19) (2006) 2166-71.
- [41] E. Bartova, S. Legartova, J. Krejci, O.A. Arcidiacono, Cell differentiation and aging accompanied by depletion of the ACE2 protein, *Aging (Albany NY)* 12(22) (2020) 22495-22508.
- [42] C.P. Vio, P. Gallardo, C. Cespedes, D. Salas, J. Diaz-Elizondo, N. Mendez, Dietary Potassium Downregulates Angiotensin-I Converting Enzyme, Renin, and Angiotensin Converting Enzyme 2, *Front Pharmacol* 11 (2020) 920.
- [43] S. Choudhary, K. Sreenivasulu, P. Mitra, S. Misra, P. Sharma, Role of Genetic Variants and Gene Expression in the Susceptibility and Severity of COVID-19, *Ann Lab Med* 41(2) (2021) 129-138.
- [44] Y. Cao, L. Li, Z. Feng, S. Wan, P. Huang, X. Sun, F. Wen, X. Huang, G. Ning, W. Wang, Comparative genetic analysis of the novel coronavirus (2019-nCoV/SARS-CoV-2) receptor ACE2 in different populations, *Cell Discov* 6 (2020) 11.
- [45] J. Liu, H. Ji, W. Zheng, X. Wu, J.J. Zhu, A.P. Arnold, K. Sandberg, Sex differences in renal angiotensin converting enzyme 2 (ACE2) activity are 17beta-oestradiol-dependent and sex chromosome-independent, *Biol Sex Differ* 1(1) (2010) 6.
- [46] R. Parit, S. Jayavel, Association of ACE inhibitors and Angiotensin type II blockers with ACE2 overexpression in COVID-19 comorbidities: a pathway-based analytical study, *Eur J Pharmacol* (2021) 173899.
- [47] I. Hamming, W. Timens, M.L. Bulthuis, A.T. Lely, G. Navis, H. van Goor, Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis, *J Pathol* 203(2) (2004) 631-7.
- [48] X. Song, W. Hu, H. Yu, L. Zhao, Y. Zhao, X. Zhao, H.H. Xue, Y. Zhao, Little to no expression of angiotensin-converting enzyme-2 on most human peripheral blood immune cells but highly expressed on tissue macrophages, *Cytometry A* (2020).
- [49] C. Wang, J. Xie, L. Zhao, X. Fei, H. Zhang, Y. Tan, X. Nie, L. Zhou, Z. Liu, Y. Ren, L. Yuan, Y. Zhang, J. Zhang, L. Liang, X. Chen, X. Liu, P. Wang, X. Han, X. Weng, Y. Chen, T. Yu, X. Zhang, J.

- Cai, R. Chen, Z.L. Shi, X.W. Bian, Alveolar macrophage dysfunction and cytokine storm in the pathogenesis of two severe COVID-19 patients, *EBioMedicine* 57 (2020) 102833.
- [50] Z. Abassi, Y. Knaney, T. Karram, S.N. Heyman, The Lung Macrophage in SARS-CoV-2 Infection: A Friend or a Foe?, *Front Immunol* 11 (2020) 1312.
- [51] M. AlGhatrif, O. Cingolani, E.G. Lakatta, The Dilemma of Coronavirus Disease 2019, Aging, and Cardiovascular Disease: Insights From Cardiovascular Aging Science, *JAMA Cardiol* 5(7) (2020) 747-748.
- [52] J. Chen, Q. Jiang, X. Xia, K. Liu, Z. Yu, W. Tao, W. Gong, J.J. Han, Individual variation of the SARS-CoV-2 receptor ACE2 gene expression and regulation, *Aging Cell* 19(7) (2020).
- [53] R.P.H. De Maeyer, R.C. van de Merwe, R. Louie, O.V. Bracken, O.P. Devine, D.R. Goldstein, M. Uddin, A.N. Akbar, D.W. Gilroy, Blocking elevated p38 MAPK restores efferocytosis and inflammatory resolution in the elderly, *Nat Immunol* 21(6) (2020) 615-625.
- [54] S. Wullschleger, R. Loewith, M.N. Hall, TOR signaling in growth and metabolism, *Cell* 124(3) (2006) 471-84.
- [55] J.B. Mannick, M. Morris, H.P. Hockey, G. Roma, M. Beibel, K. Kulmatycki, M. Watkins, T. Shavlakadze, W. Zhou, D. Quinn, D.J. Glass, L.B. Klickstein, TORC1 inhibition enhances immune function and reduces infections in the elderly, *Sci Transl Med* 10(449) (2018).

## Figure Legend

**Figure 1:** RAS and T cell response following SARS-CoV-2 infection during controlled (A) and uncontrolled (B) disease.

Angiotensin-converting enzyme 2 (ACE2) cleaves angiotensin II (Ang II) into Ang 1-7, inactivating Ang II. Ang 1-7 via MAS receptor antagonizes inflammatory response allowing an effective Th1 response (A). During acute pulmonary infection diseases, RAS participates in the development of ARDS with consequent pulmonary fibrosis. The unbalance between AngII and Ang 1-7 inducing the differentiation of Th17 cells that secrete numerous cytokines. In particular, IL-17 per se induces the production of massive quantities of proinflammatory cytokines and chemokines (B).

