



Ghrelin and memory: Differential effects on acquisition and retrieval

Valeria P. Carlini^{a,*}, Marisa Gherzi^a, Helgi B. Schiöth^b, Susana R. de Barioglio^a

^a Departamento de Farmacología, Facultad de Ciencias Químicas, UNC, Córdoba, Argentina

^b Uppsala University, Dept. of Neuroscience, Section of Pharmacology, Uppsala, Sweden

ARTICLE INFO

Article history:

Received 4 December 2009

Received in revised form 24 February 2010

Accepted 25 February 2010

Available online 7 March 2010

Keywords:

Ghrelin

Memory retention

Memory acquisition

Short-term memory

Long-term memory

ABSTRACT

In a previous paper we have demonstrated that the orexigenic peptide Ghrelin (Ghr), increases memory retention in rats and mice. In the present work we evaluated the Ghr effect when it was administered previous the training session or previous the test session (24 h after training) on the memory performance, using step-down test. The results showed that the intra-hippocampal Ghr administration previous the training session improved the long-term memory in this task, but did not modify the short-term memory. Nevertheless, when the Ghr was administrated previous the test session, no changes were observed in the memory performance. Taking into account these results and other previously published by our group, we could hypothesize that Ghr may modulate specific molecular intermediates involved in memory acquisition/consolidation but not in the retrieval.

© 2010 Elsevier Inc. All rights reserved.

1. Introduction

Ghrelin (Ghr) is a 28 amino acid peptide that induces robust feeding responses in different experimental models [29,36]. Although Ghr comes from both peripheral (stomach) and central sources, its hyperphagic properties, to a large extent, arise from activity at the brain level [22]. Ghrelin receptors (GHS-R) are expressed in several hypothalamic nucleus and other areas such as the hippocampus, substantia nigra, ventral tegmental area, and dorsal and median raphe nucleus [15,29,36].

In the mammals, the ability to seek and find food is crucial for survival. After finding the food and ingesting it, it is important to remember where this food can be found or perhaps – more importantly – to be able to retain the successful approach that was used to find it. Effective learning and memory are thus likely to be crucial for survival during periods of food shortage [30].

We have previously demonstrated that intracerebroventricular (icv) Ghr administration increased, in a dose-dependent manner, the memory retention (measured as the latency time in the step-down test) and induced anxiety-like behavior in rats. This was the first evidence showing that Ghr increases memory retention [9]. This finding prompted further studies where Ghr was administered into other brain regions, such as the hippocampus, amygdala and

dorsal raphe nucleus (DRN) [10]. In relation to memory, Diano et al. demonstrated that Ghr causes an increase in synaptic plasticity promoting new synaptic connections between hippocampal neurons [12]. Based on the above mentioned observations, the central Ghr receptor could be considered as a new drug target for therapeutic approaches to treat diseases that affect cognition [2].

Since the different memory processes including acquisition, consolidation and retrieval are well known [1], in the present work we extended previous experiments and analyzed the Ghr participation on memory acquisition/consolidation and retrieval.

2. Materials and procedure

2.1. Animals

Male Wistar rats weighting 260–290 g were maintained under controlled temperature at 21 ± 1 °C and a light/dark cycle (12 h light, 12 h dark) with food and water ad libitum. Rats were handled daily for 7 days before the experiments. All procedures were conducted according to the National Institutes Health (NIH) Guide for the Care and Use of Laboratory Animals (Publications No. 80-23, 1996) and approved by the local Animal Care and Use Committee. Every attempt was made to minimize the number of animals used and their suffering.

2.2. Surgery

The animals were anesthetized with 55 mg/kg Ketamine HCl and 11 mg/kg xylazine (both Kensol könig, Argentina) and placed in a

* Corresponding author at: Departamento de Farmacología, Facultad de Ciencias Químicas, Haya de la Torre y Medina Allende, Universidad Nacional de Córdoba, Ciudad Universitaria, 5016 Córdoba, Argentina. Tel.: +54 351 4334437; fax: +54 351 4334420.

E-mail address: vcarlini@mail.fcq.unc.edu.ar (V.P. Carlini).

stereotaxic apparatus. Then, rats were implanted bilaterally into the CA₁ hippocampus or intracerebroventricularly (icv) with steel guide cannula, according to the atlas of Paxinos [31]. The coordinates relative to bregma were anterior: -3.6 mm; lateral: ± 2.0 mm; vertical: -2.8 mm and anterior: -4.3 mm; vertical: -4.6 mm for hippocampus and icv, respectively. Cannulas were fixed to the skull surface with dental acrylic cement.

2.3. Drugs and infusion procedures

Animals were injected bilaterally with Ghr or artificial cerebrospinal fluid (ACSF) using a 10 μ l Hamilton syringe connected by Pe-10 polyethylene tubing to a 30-gauge needle. The infusion volume was 0.5 μ l bilaterally or 1 μ l for the intra-hippocampal and icv administration respectively, and it was delivered over a 1 min period. The Ghr peptide was purchased from Neosystem, France. Ghr was resuspended in ACSF, aliquoted to obtain final concentrations of 0.03, 0.3 and 3.0 nmol/ μ l and stored at -20°C until use. Injections were done between 11:00 am and noon in order to prevent variations induced by circadian rhythms.

Drug was administered 15 min previous the training session of the step down test or 15 min previous the test session (24 h after training) of this test in different animal groups.

2.4. Step-down test (inhibitory avoidance)

One-trial avoidance test in rats involves the activation of two molecular pathways in the CA₁ region of the dorsal hippocampus, one induces a short-term memory (STM) that lasts 3 h, and the other induces a long-term memory (LTM) that takes 3–6 h to be formed and lasts for many days or even months [14,16–20]. The apparatus was a 50 cm \times 25 cm \times 25 cm plastic box with a 2.5 cm high and 7.0 cm wide platform on the left of the training box apparatus. The animals were placed on the platform, and latency to step down by placing the four paws on the grid was measured. In the training session, immediately upon stepping down, the rats received a 0.4 mA, 2 s scrambled shock to the feet, and were then immediately removed from the training box and placed in their home cages. The test session was identical regarding the procedures, except that no shock was given. A ceiling of 180 s was imposed on the test measurement. Latency time was taken as an index of memory retention. The test session was carried out 1.5 and 24 h after training in order to measure STM and LTM respectively.

2.5. Histology

After the behavioral test, rats were anesthetized with chloral hydrate, cardially perfused with paraformaldehyde (4%) and their brains were removed. Frontal sections were cut in cryostat (Leica), and the cannula position was localized. Only results obtained from animals in which the tips of the cannulas were placed in the hippocampus or in the central ventricle were included.

2.6. Elevated plus-maze

Rats were tested in an elevated plus-maze, in order to determine if the Ghr promotes or reduce anxiety and locomotion under these experimental conditions (pro- or anti-conflict behaviors) [5–7,32]. Briefly, the elevated plus-maze consisted of a central platform (5 cm \times 5 cm), two open arms (50 cm \times 10 cm), and two closed arms (50 cm \times 10 cm \times 40 cm). The arms were arranged in such a way that two arms of each type were opposite to each other. The maze was 50 cm above floor level and tests were carried out under a dim red light 24 h after Ghr administration. Animals were placed individually on the central platform facing an open arm. The number of

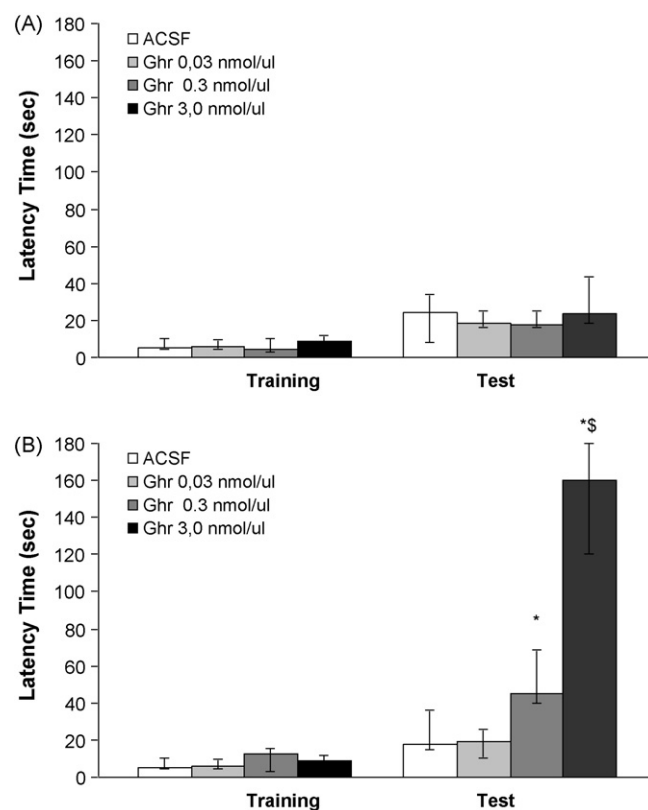


Fig. 1. Effect of Intra-hippocampal Ghr administration (previous the training session) upon memory performance in step down test: (A) short-term memory and (B) long-term memory. Animals were administrated with Ghr (0.03, 0.3 and 3.0 nmol/ μ l) previous the training session and evaluated 1 h (A) or 24 h (B) after training. The results are expressed as medians with the respective inter-quartile range. $n = 8-10$ animals/group. *Significant differences related to control animals (ACSF), $p \leq 0.05$. §Significant differences related to treated animals with Ghr 3.0 nmol/ μ l, $p \leq 0.05$.

entries in each arm, the time spent in the open and enclosed arms and the number of rearing were recorded during 5 min.

2.7. Statistics

Variables analyzed from data find avoidance being did not follow a normal distribution and its variance did not fulfil the assumption of homoscedasticity, these results were expressed as medians (inter-quartile range) and analyzed by non parametric tests (Mann–Whitney or Kruskal–Wallis).

Statistical inference of the data from plus-maze were expressed as mean \pm standard error (SEM) and analyzed by one-way multiple analysis of variance (MANOVA) followed by the post-hoc Hotelling T2 ($p \leq 0.05$).

3. Results

3.1. Effect Ghr infused previous the training session

Fig. 1 shows the effect of Ghr administered before the training session into the hippocampus on memory performance in a step down test. The Kruskal–Wallis ANOVA analysis revealed a significant Ghr effect only in LTM (chi-square = 23.01, $df = 3$, $p = 0.00$). As it can be seen in the figure Ghr enhanced LTM in a dose-dependent manner. (ACSF vs Ghr 0.03 nmol/ μ l, $p = 0.38$; ACSF vs Ghr 0.3 nmol/ μ l, $p = 0.00$; ACSF vs Ghr 3.0 nmol/ μ l, $p = 0.00$; Ghr 0.03 vs Ghr 0.3 nmol/ μ l, $p = 0.00$; Ghr 0.3 vs Ghr 3.0 nmol/ μ l, $p = 0.00$; Ghr 0.03 vs Ghr 3.0 nmol/ μ l, $p = 0.00$; $n = 8-10$

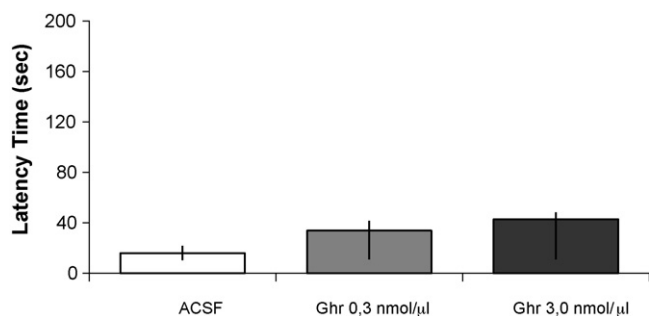


Fig. 2. Effect of Intra-hippocampal Ghr administration (previous the test session) upon memory performance in step down test. Animals were administrated with Ghr (0.3 and 3.0 nmol/μl) previous the test session (24 h after training). The results are expressed as medians with the respective inter-quartile range. $n=8-10$ animals/group.

animals/group). However Ghr did not induce changes in STM (chi-square = 4.92, $df=3$, $p=0.18$; $n=8-10$ animals/group).

Oppositely, when Ghr was administrated icv both, STM and LTM were enhanced (chi-square = 13.54, $df=3$, $p=0.00$) (ACSF = 33.97 (18.53–44.38); Ghr 0.03 nmol/μl = 180 (95–180); Ghr 0.3 nmol/μl = 180 (130–180); Ghr 3.0 nmol/μl = 180 (142.36–180) for STM and ACSF = 20.57 (10.53–42.54); Ghr 0.03 nmol/μl = 115.38 (90–165); Ghr 0.3 nmol/μl = 165 (128–180); Ghr 3.0 nmol/μl = 180 (120–180) for LTM acquisition; $n=9-11$ animals/group).

3.2. Effect Ghr infused previous the test session

Fig. 2 shows the effect of Ghr administered before the test session into the hippocampus on memory performance in a step down test. As it can be seen retrieval was not affected by the intra-hippocampal peptide administration (chi-square = 2.57, $df=2$, $p=0.28$; $n=8-10$ animals/group).

The icv Ghr administration did not affect the LTM retrieval (chi-square = 5.21, $df=2$, $p=0.65$) (ACSF = 40.56 (20.18–45.21); Ghr 0.3 nmol/μl = 32.58 (15.42–41.52); Ghr 3.0 nmol/μl = 27.56 (11.52–33.85); $n=10-13$ animals/group).

3.3. Ghr effect on anxiety-like behavior

Table 1 shows the effects of different Ghr doses in the relative indexes of anxiety and number of total entries 24 h after administration. Intra-hippocampal treated rats did not display changes in any of the parameters measured. These results could suggest that their effects on LTM acquisition are not due to interferences with locomotion activity or anxiety (or prior anti-conflict behavior) levels. The MANOVA test showed that Ghr administration did not have a significant effect in the parameters related to the anxiety, as the number of entries into open arms, time spent in open arms, the percentage of open arms entries and percentage of time spent in

Table 1

Ghr effect 24 h after intra-hippocampal administration on elevated plus maze test. The results are expressed as mean \pm SEM. The numbers between parentheses indicates the numbers of animals.

Parameter	Control (12)	Ghr Hi 0,3 (9)	Ghr Hi 3,0 (9)
Number of entries to open arms	3.0 \pm 0.2	3.9 \pm 0.4	2.9 \pm 0.8
Number of entries to closed arms	5.0 \pm 0.5	5.4 \pm 0.9	6.0 \pm 0.5
Number of total entries (OA + CA)	8.0 \pm 0.7	9.3 \pm 0.3	8.9 \pm 0.5
Percent of entries to open arms	38.2 \pm 2.1	43.7 \pm 5.5	28.2 \pm 7.7
Number of rearings	8.1 \pm 1.1	7.7 \pm 1.1	8.4 \pm 0.7
Time spent on open arms (s)	60.8 \pm 3.6	70.6 \pm 12.8	56.9 \pm 16.8
Time spent on closed arms (s)	194.2 \pm 5.7	182.5 \pm 13.2	216.3 \pm 24.2
Percent of time spent on open arms	23.8 \pm 1.0	27.8 \pm 4.8	22.6 \pm 6.9
Risk-assessment	3.5 \pm 1.1	3.3 \pm 0.6	2.5 \pm 0.9
Grooming	4.3 \pm 0.7	3.4 \pm 0.4	3.2 \pm 0.4

open arms (Wilks = 0.02, $df=3$, $p>0.05$). In addition, Ghr did not have significant effects on the total number of entries, a measure of overall locomotor activity. Similar results were observed in icv treated rats (data not shown).

4. Discussion

The main finding of this paper was that Ghr administration into the hippocampus of rats previous the training session in a step-down test improved LTM, without altering the STM. Nevertheless, when the Ghr was administrated previous the test session, not changes were seen in the memory performance.

It has been demonstrated that during the memory acquisition the animal associates the context (step-down box) and the shock. During consolidation, which can last from minutes to days, this memory is moved from a labile to a more persistent state. During retrieval, the animal is returned to the context where memory for the context–shock association was assessed [1].

Our results showing that the intra-hippocampal Ghr administration previous the training session enhanced LTM, but did not modify STM, could probably be attributed to the fact that the hippocampal mechanisms of STM and LTM in the step-down test can operate separately [19]; thus, STM can be selectively altered by many treatments that do not affect LTM [16–20].

It has been demonstrated that fear conditioning often results in a robust memory for the context–shock association after a single shock presentation, which allows isolation of the stages of memory along time. The pharmacological approaches offer a highest temporal specificity because they can be applied and removed from the system within a relatively short time window. Nevertheless, it is important to take in mind that manipulations before acquisition will affect early stages of consolidation, and manipulations before retrieval may affect late stages of consolidation or retention [1].

Previous results of our laboratory showed that the peptide administrated immediately after training session [11] increased both STM and LTM (for all doses tested) and the increments were higher than those when Ghr was administered previous the training session. Thus, these findings and those from Fig. 1 in this study are in accordance with the hypotheses that suggest that Ghr is more effective in the modulation of the memory consolidation rather than in the acquisition.

The present results also demonstrated that the icv Ghr administration induced enhancement of both STM and LTM. The difference found between the Ghr effects after icv or intra-hippocampal administration previous the training session on STM could be attributed to the participation of the other structure, such as the amygdala when Ghr was administrated icv. It has been demonstrated that the amygdala presents a specific role in memory acquisition but not in memory consolidation [1]. Furthermore, it is relevant for the formation of motivated memories [24,25] playing

a key role in the learning and memory processes occurring during emotional events [3,4,21,25–28].

Another interesting finding of the present study is that the increase in the latency time observed in the step-down test was no attributed to the anxiogenic effect of the peptide. It is well known that freezing is a common behavior observed in anxious rats exposed to a novel environment [23]. Our results showed that Ghr did not induce any anxiety-like behavior 24 h after administration. In addition, the intra-hippocampal Ghr administration induced anxiogenesis only at the highest dose tested (3.0 nmol/ μ l) 15 min after administration [10].

Our results also showed that Ghr could affect either the memory acquisition or consolidation but certainly did not affect the retrieval. The pharmacological findings have shown that NMDA receptors, which are critical for memory acquisition, are not involved in retrieval of previously established memories [34,35]. It has been suggested that the activation of hippocampal NMDA receptors are primarily related to the contextual aspects (formation of a representation of the training environment) whereas NMDA receptors in the amygdala are more related to the aversive aspects (footshock) of the task [13,33]. Similarly, protein-kinase A (PKA) plays an important role on acquisition, and it is critically involved in consolidation, but, appears to be not necessary for retrieval [8,35].

Taking into account these results and previous findings from our laboratory, we could hypothesize that Ghr may modulate specific molecular intermediates involved in the memory acquisition/consolidation processes but not in those related to retrieval.

Acknowledgements

This work was supported by grants from CONICET (Consejo Nacional de Investigación Científica y Técnica), SECyT (Secretaría de Ciencia y Técnica de la Universidad Nacional de Córdoba) and the Swedish Research Council (VR, medicine). Dr. Susana R. de Bariogio is members of CONICET and Dr. Valeria P. Carlini is a CONICET fellow.

References

- Abel T, Lattal KM. Molecular mechanisms of memory acquisition, consolidation and retrieval. *Curr Opin Neurobiol* 2001;11(2):180–7. Review.
- Atcha Z, Chen WS, Ong AB, Wong FK, Neo A, Browne ER, et al. Cognitive enhancing effects of ghrelin receptor agonists. *Psychopharmacology (Berl)* 2009;206(3):415–27.
- Bianchin M, Walz R, Ruschel AC, Zanatta MS, Da Silva RC, Bueno e Silva M, et al. Memory expression is blocked by the infusion of CNQX into the hippocampus and/or the amygdala up to 20 days after training. *Behav Neural Biol* 1993;59:83–6.
- Bianchin MM, Spanis CW, Roesler R, McGaugh JL, Izquierdo I. (\pm)- α -Methyl-4-carboxyphenylglycine, a metabotropic glutamate receptor blocker, impairs retention of an inhibitory avoidance task in rats when infused into the basolateral nucleus of the amygdala. *Brain Res* 2000;852(2):436–43.
- Barros DM, Izquierdo LA, Mello e Souza T, Ardenghi PG, Pereira P, Medina JH, et al. Molecular signalling pathways in the cerebral cortex are required for retrieval of one-trial avoidance learning in rats. *Behav Brain Res* 2000;114(1–2):183–92.
- Barros DM, Mello e Souza T, de Souza MM, Choi H, DeDavid e Silva T, Lenz G, et al. LY294002, an inhibitor of phosphoinositide 3-kinase given into rat hippocampus impairs acquisition, consolidation and retrieval of memory for one-trial step-down inhibitory avoidance. *Behav Pharmacol* 2001;12(8):629–34.
- Barros DM, Izquierdo LA, Medina JH, Izquierdo I. Pharmacological findings contribute to the understanding of the main physiological mechanisms of memory retrieval. *Curr Drug Targets CNS Neurol Disord* 2003;2(2):81–94. Review.
- Bourtchouladze R, Abel T, Berman N, Gordon R, Lapidus K, Kandel ER. Different training procedures recruit either one or two critical periods for contextual memory consolidation, each of which requires protein synthesis and PKA. *Learn Memory* 1998;5:365–74.
- Carlini VP, Monzón ME, Varas MM, Cragolini AB, Schiöth HB, Scimionelli TN, et al. Ghrelin increases anxiety-like behavior and memory retention in rats. *Biochem Biophys Res Commun* 2002;299(5):739–43.
- Carlini VP, Varas MM, Cragolini AB, Schiöth HB, Scimionelli TN, de Bariogio SR. Differential role of the hippocampus, amygdala, and dorsal raphe nucleus in regulating feeding, memory, and anxiety-like behavioral responses to ghrelin. *Biochem Biophys Res Commun* 2004;313(3):635–41.
- Carlini VP, Gaydou RC, Schiöth HB, de Bariogio SR. Selective serotonin reuptake inhibitor (fluoxetine) decreases the effects of ghrelin on memory retention and food intake. *Regul Pept* 2007;140(1–2):65–73.
- Diano S, Farr SA, Benoit SC, McNay EC, da Silva I, Horvath B, et al. Ghrelin controls hippocampal spine synapse density and memory performance. *Nat Neurosci* 2006;9(3):381–8.
- Fanselow MS, Kim JJ, Yipp J, De Oca B. Differential effects of the *N*-methyl-aspartate antagonist-2-amino-5-phosphonovalerate on acquisition of fear of auditory and contextual cues. *Behav Neurosci* 1994;108:235–40.
- Gold PE. The use of avoidance training in studies of modulation of memory storage. *Behav Neural Biol* 1986;46(1):87–98.
- Guan XM, Yu H, Palyha OC, McKee KK, Feighner SD, Sirinathsinghji DJ, et al. Distribution of mRNA encoding the growth hormone secretagogue receptor in brain and peripheral tissues. *Brain Res Mol Brain Res* 1997;48(1):23–9.
- Izquierdo I, Barros DM, Mello e Souza T, de Souza MM, Izquierdo LA, Medina JH. Mechanisms for memory types differ. *Nature* 1998;393(6686):635–6.
- Izquierdo I, Izquierdo LA, Barros DM, Mello e Souza T, de Souza MM, Quevedo J, et al. Differential involvement of cortical receptor mechanisms in working, short-term and long-term memory. *JH Behav Pharmacol* 1998;9(5–6):421–7.
- Izquierdo I, Medina JH, Izquierdo LA, Barros DM, de Souza MM, Mello e Souza T. Short- and long-term memory are differentially regulated by monoaminergic systems in the rat brain. *Neurobiol Learn Mem* 1998;69(3):219–24.
- Izquierdo I, Medina JH, Vianna MR, Izquierdo LA, Barros DM. Separate mechanisms for short- and long-term memory. *Behav Brain Res* 1999;103(1):1–11. Review.
- Izquierdo LA, Barros DM, Vianna MR, Coitinho A, de David e Silva T, Choi H, et al. Molecular pharmacological dissection of short- and long-term memory. *Cell Mol Neurobiol* 2002;22(3):269–87. Review.
- Jerusalinsky D, Ferreira MB, Walz R, Da Silva RC, Bianchin M, Ruschel AC, et al. Amnesia by post-training infusion of glutamate receptor antagonists into the amygdala, hippocampus, and entorhinal cortex. *Behav Neural Biol* 1992;58:76–80.
- Kojima M, Kangawa K. Ghrelin: structure and function. *Physiol Rev* 2005;85(2):495–522. Review.
- Lát J. The analysis of habituation. *Acta Neurobiol Exp (Wars)* 1973;33(4):771–89.
- Maren S. Synaptic transmission and plasticity in the amygdala. An emerging physiology of fear conditioning circuits. *Mol Neurobiol* 1996;13:1–22.
- Maren S, Aharonov G, Stote DL, Fanselow MS. *N*-methyl-aspartate receptors in the basolateral amygdala are required for both acquisition and expression of conditional fear in rats. *Behav Neurosci* 1996;110:1365–74.
- McGaugh JL, Cahill L, Roozendaal B. Involvement of the amygdala in memory storage: interaction with other brain systems. *Proc Natl Acad Sci USA* 1996;93:13508–14.
- Meschers MH, Bianchin M, McGaugh JL. The effects of intra-amygdala infusion of the AMPA receptor antagonist CNQX on retention performance following aversive training. *Neurobiol Learn Mem* 1996;66:324–40.
- Miserendino MJ, Sananes CB, Melia KR, Davis M. Blocking of acquisition but not expression of conditioned fear-potentiated startle by NMDA antagonists in the amygdala. *Nature* 1990;345:716–8.
- Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K, et al. A role for ghrelin in the central regulation of feeding. *Nature* 2001;409(6817):194–8.
- Olszewski PK, Schiöth HB, Levine AS. Ghrelin in the CNS: from hunger to a rewarding and memorable meal? *Brain Res Rev* 2008;58:160–70. Review.
- Paxinos G, Watson C. The rat brain in stereotaxic coordinates. San Diego: Academic Press; 1987.
- Pellow S, Chopin P, File SE, Briley M. Validation of open/closed arm entries in an elevated plus-maze as a measure of anxiety in the rat. *J Neurosci Methods* 1985;14(3):149–67.
- Roesler R, Schröder N, Vianna MR, Quevedo J, Bromberg E, Kapczinski F, et al. Differential involvement of hippocampal and amygdalar NMDA receptors in contextual and aversive aspects of inhibitory avoidance memory in rats. *Brain Res* 2003;975(1–2):207–13.
- Steele RJ, Morris RG. Delay-dependent impairment of a matching-to-place task with chronic and intrahippocampal infusion of the NMDA-antagonist D-AP5. *Hippocampus* 1999;9(2):118–36.
- Szapiro G, Vianna MR, McGaugh JL, Medina JH, Izquierdo I. The role of NMDA glutamate receptors, PKA, MAPK, and CAMKII in the hippocampus in extinction of conditioned fear. *Hippocampus* 2003;13(1):53–8.
- Tschöp M, Lahner H, Feldmeier H, Grasberger H, Morrison KM, Janssen OE, et al. Effects of growth hormone replacement therapy on levels of cortisol and cortisol-binding globulin in hypopituitary adults. *Eur J Endocrinol* 2000;143(6):769–73.