Scientific Research Publishing

American Journal of Plant Sciences, 2020, 11, 861-868
https://www.scirp.org/journal/ajps
ISSN Online: 2158-2750
ISSN Print: 2158-2742

New 5-Alkenyl Resorcinols from *Lithraea molleoides* (Vell). Eng.

Alejandra Catalano[1,2], Beatriz Lantaño[3], Lucas Fabián[2,4], Paula López[1,2]

[1]Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Cátedra de Farmacognosia, Ciudad Autónoma de Buenos Aires, Argentina
[2]CONICET, Instituto de la Química y Metabolismo del Fármaco (IQUIMEFA), Cuidad Autónoma de Buenos Aires, Argentina
[3]Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Cátedra de Química Orgánica, Cuidad Autónoma de Buenos Aires, Argentina
[4]Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Cátedra de Química Medicinal, Cuidad Autónoma de Buenos Aires, Argentina
Email: vcatalano@ffyb.uba.ar

How to cite this paper: Catalano, A., Lantaño, B., Fabián, L. and López, P. (2020) New 5-Alkenyl Resorcinols from Lithraea molleoides (Vell). Eng. *American Journal of Plant Sciences*, **11**, 861-868.
https://doi.org/10.4236/ajps.2020.116062

Received: May 10, 2019
Accepted: June 26, 2020
Published: June 29, 2020

Copyright © 2020 by author(s) and Scientific Research Publishing Inc.
This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).
http://creativecommons.org/licenses/by/4.0/
Open Access

Abstract

Lithraea molleoides (Anacardiaceae) is a tree that grows in South America including Southern Brazil, Southern, and Eastern Bolivia, Southern Paraguay, Northern, and Central Argentina. Infusions, decoctions, or tinctures from its aerial parts (leaves, buds, and young stems) are employed in ethnomedicine mainly against respiratory, and digestive inflammations and illnesses. Antibacterial, antiviral, antioxidant, anti-inflammatory, and antinociceptive activities, among others, have been reported for *L. molleoides*. Many of its biological activities have been associated with the reported presence of 5-alkenyl resorcinols. Alkyl/alkenyl catechols and alkyl/alkenyl resorcinols are very common in members of the Anacardiaceae family and several activities have been attributed to them. This work describes the isolation and the structural elucidation of three new 5-alkenyl resorcinols isolated from *Lithraea molleoides* reported in nature for the first time.

Keywords

Lithraea molleoides, 5-Alkenyl Resorcinol,
(Z,Z)-5-(Pentadeca-6,9-Dienyl)-Resorcinol,
(Z,Z)-5-(Trideca-5,8-Dienyl)-Resorcinol,
(Z)-5-(Heptadec-6-Enyl)-Resorcinol

1. Introduction

The Anacardiaceae family comprises a great variety of medicinal plants from

which many biologically active substances have been isolated. Alkyl/alkenyl catechols and alkyl/alkenyl resorcinols, also phenolic lipids, are very common in members of this family and several activities have been attributed to them. The antifungal activity of 5-(12-heptadecenyl)-resorcinol and the anti-inflammatory activity of 5-(11'Z-heptadecenyl)-resorcinol and 5-(8'Z,11'Z-heptadecadienyl)-resorcinol, all compounds isolated from the peel of *Mangifera indica* L. (Anacardiaceae), have been informed [1]. Also, the resorcinol 5-((8Z,11Z,14Z)-hexatriaconta-8,11,14-trienyl) benzene-1,3-diol has been reported for its cytotoxic and apoptotic effects [2]. *Lithraea caustica* (Anacardiaceae) extract, characterized by the presence of long-chain catechols, has been demonstrated to promote an antitumoral response against M16 melanoma [3].

Lithraea molleoides (Vell.) Engl. (Anacardiaceae) is a tree that grows in Southern Brazil, Southern, and Eastern Bolivia, Southern Paraguay, Northern, and Central Argentina. In Argentina, it is a dominant orophilic species in the biogeographical Semi-arid Chacoan region. In the Paranaense and Humid Chacoan rainforests, *L. molleoides* is a member of the secondary succession community [4] [5] where is commonly named "chichita", "molle dulce" or ''molle de Córdoba''. Infusions, decoctions or tinctures from the aerial parts (leaves, buds, and young stems) are used by rural people of these countries for its antiarthritic, hemostatic, diuretic, and tonic properties [6] as well as used for the treatment of respiratory diseases [7] and digestive inflammations illnesses [8]. Previous investigations on different extracts of *L. molleoides* have reported antiviral [9], antimicrobial [10], anti-inflammatory [11], anti-ulcerogenic [12], antinociceptive [13], and antibacterial [14] activities.

The resorcinol 1,3-dihydroxy-5-(tridec-4',7'-dienyl)-benzene, is the most cited compound of *L. molleoides* as responsible for their biological activities. It has been isolated from this plant and showed cytotoxicity on human hepatocellular carcinoma cell line [15] as well as apoptosis induction [16]. The anti-inflammatory activity [8], the antioxidant and antiproliferative activity on tumor lymphocytes, and immunostimulant activity on normal lymphocytes [17] of the dichloromethane extract of *L. molleoides* were reported and identified this compound as the major component. The same compound has been identified in the ethanolic extract of *L. molleoides* and showed antibacterial activity against *Proteus mirabilis* [18]; it has been proposed as an effective tyrosinase inhibitor [19] to use in food preservation. Moreover, 13 carbon alkenyl resorcinols chains obtained from this species have shown nematicidal activity [20].

This work aims to isolate the minor resorcinols present in the *L. molleoides* dichloromethane extract and chemically identify them.

2. Materials and Methods

2.1. Plant Material

Leaves of *Lithraea molleoides* were collected in the district of Burruyacú, Sierra de Medina, province of Tucumán, Argentina, and botanically identified by Ph.D. Graciela Ponessa. Vouchers specimens are deposited in the Herbarium

Fundación Miguel Lillo, San Miguel de Tucumán, Tucumán, Argentina (Sierras de Medina, 26˚26'63"S 65˚01'53"W, 1112 MAMSL, 24-V-2014, A. Slanis, M. I. Mercado y G. I. Ponessa 530 (LIL); ibid., A. Slanis, M. I. Mercado y G. I. Ponessa 531 (LIL); ibid. A. Slanis, M. I. Mercado y G. I. Ponessa 532 (LIL).

2.2. Plant Extraction and Compound Isolation

Leaves of *L. molleoides* (70 g) were overnight macerated (6 × 150 ml) with dichloromethane (Cl_2CH_2) at room temperature. After vacuum filtration, the Cl_2CH_2 extracts were joined and taken to dryness under reduced pressure yielding 3.95 g of dried extract. The dry extract was separated on a Sephadex LH-20 column (50 cm × 5 cm) using a gradient of Cl_2CH_2 and methanol (MeOH) as solvents according to López *et al.* [16] obtaining 24 fractions. The fractions were monitored by TLC on Silicagel F_{254} plates using Cl_2CH_2-MeOH (95:5) as the mobile phase. Five fractions containing resorcinol derivatives were obtained.

The compounds were isolated from the resorcinol fractions by HPLC-UV DAD performed with a Varian® 9000 instrument using a diode array detector. An RP18 column (Gemini® 5 µm, 150 mm × 4.6 mm) and a mobile phase constituted by solvent A: H_2O/AcOH (98:2) and solvent B: MeOH/AcOH (98:2) was used. A gradient consisting in 70% B to 100% B in 30 minutes was applied. The flow rate was 1.2 ml/min. Eleven eluates were collected and taken to dryness in a Savant® concentrator. Each compound was numbered in order of increasing elution.

2.3. Compound Identification

Mass spectra were determined in an ESI Bruker micrOTOF-Q II™spectrometer. ^{1}H and ^{13}C-NMR spectra, homo- and heteronuclear correlation spectroscopy and NOESY experiments were recorded at ambient temperature in chloroform-d_3 ($CDCl_3$) with a Magneto Bruker Ultra Shield spectrometer operated at 600.13 MHz and 150.91 MHz for ^{1}H and ^{13}C nucleus respectively. Chemical shifts are expressed in ppm (relative to the solvent). Coupling constants (J) are in Hz.

3. Results and Discussion

Five 5-alkenyl resorcinols were obtained from the dichloromethane extract of *L. molleoides* (supplementary data) and after structural elucidation, they were identified as:

(Z,Z)-5-(trideca-4,7-dienyl)-resorcinol **3** [20] ^{1}H NMR (600 MHz): δ (ppm): 6.27 (2H, d, J = 2.3 Hz), 6.20 (1H, t, J = 2.3 Hz), 5.44 - 5.34 (4H, m), 2.79 (2H, t, J = 5.9 Hz), 2.51 (2H, t, J = 7.7 Hz), 2.10 (2H, q, J = 7.2 Hz), 2.06 (2H, q, J = 7.1 Hz), 1.66 (2H, quint., J = 7.6 Hz), 1.40 - 1.27 (6H, m), 0.90 (3H, t, J = 6.8 Hz). ^{13}C NMR (150.903 MHz): δ (ppm): 156.63, 145.73, 130.40, 129.46, 128.66, 127.80, 108.08, 100.27, 35.29, 31.51, 30.91, 29.32, 27.20, 26.76, 25.67, 22.57, 14.07.

(Z)-5-(trideca-4-enyl)-resorcinol **5** [20] 8) ^{1}H NMR (600 MHz): δ (ppm): 6.27 (2H, d, J = 2.3 Hz), 6.20 (1H, t, J = 2.3 Hz), 5.44 - 5.36 (2H, m), 2.52 (2H, t, J = 7.7 Hz), 2.08 (2H, q, J =7. 0 Hz), 2.02 (2H, q, J = 6.9 Hz), 1.66 (2H, quint., J = 7.6 Hz), 1.31 - 1.26 (14H, m), 0.90 (3H, t, J = 6.7 Hz). ^{13}C NMR (150.903 MHz): δ

(ppm): 156.63, 145.78, 130.60, 129.08, 108.07, 100.25, 35.32, 31.89, 31.01, 29.73, 29.50, 29.32, 29.29, 27.28, 26.78, 22.65, 14.06.

(Z,Z)-5-(pentadeca-6,9-dienyl)-resorcinol **6** ^{1}H NMR (600 MHz): δ (ppm): 6.26 (2H, d, J = 2.3 Hz), 6.20 (1H, t, J = 2.3 Hz), 5.40 - 5.33 (4H, m), 2.79 (2H, t, J = 6.8 Hz), 2.51 (2H, t, J = 7.8 Hz), 2.08 (4H, m), 1.61 (2H, m), 1.40 - 1.28 (10H, m), 0.90 (3H, t, J = 6.8 Hz). 13C NMR (150.903 MHz): δ (ppm): 156.61, 145.98, 130.26, 129.95, 128.14, 127.90, 108.03, 100.18, 35.77, 31.55, 30.91, 29.50, 29.33, 28.92, 27.21, 27.14, 25.64, 22.55, 14.03.

(Z,Z)-5-(trideca-5,8-dienyl)-resorcinol **9** ^{1}H NMR (600 MHz): δ (ppm): 6.26 (2H, d, J = 2.3 Hz), 6.19 (1H, t, J = 2.3 Hz), 5.43 - 5.33 (4H, m), 2.79 (2H, t, J = 6.8 Hz), 2.50 (2H, t, J = 7.8 Hz), 2.06 (4H, m), 1.60 (2H, m), 1.40 - 1.28 (6H, m), 0.90 (3H, t, J = 6.8 Hz). ^{13}C NMR (150.903 MHz): δ (ppm): 156.64, 146.08, 130.13, 130.05, 128.02, 127.94, 107.99, 100.13, 35.81, 31.53, 31.03, 29.23, 27.21, 25.64, 22.58, 14.08.

(Z)-5-(heptadec-6-enyl)-resorcinol **11** ^{1}H NMR (600 MHz): δ (ppm): 6.26 (2H, d, J = 2.3 Hz), 6.20 (1H, t, J = 2.3 Hz), 5.40 - 5.34 (2H, m), 2.54 (2H, t, J = 7.8 Hz), 2.04 (4H, m), 1.61 (2H, m), 1.38 - 1.25 (16H, m), 1.31 (4H, m), 0.90 (3H, t, J = 6.8 Hz). ^{13}C NMR (150.903 MHz): δ (ppm): 156.61, 146.11, 129.97, 129.81, 108.60, 100.17, 35.79, 31.89, 30.98, 29.76, 29.74, 29.50, 29.37, 29.31, 29.29, 29.22, 29.20, 27.22, 27.19, 22.66, 14.05.

Table 1 shows the spectroscopic data of compounds **6**, **9** and **11** (Figure 1).

Analysis of ^{1}H and ^{13}C NMR spectra of the isolated compounds indicates the presence of a resorcinol subnit in all of them with different side chains with various insaturation degrees (Table 1). Negative HRMS (ESI) showed the unprotonated molecular ion at *m/z* 315 for compound **6**. The corresponding molecular formula $C_{21}H_{32}O_2$ and the overlapping signals of four olefinic protons δ_H 5.37 - 5.40 suggested a 15-carbon side chain with two double bonds. HSQC spectra allowed the assignment of the multiplicity of all protonated carbons, which confirmed that C_{15} side chain was linear. HMBC data established the position of the double bonds at $\Delta^{6'}$ and $\Delta^{9'}$ with a bisallylic methylene group at δ_H 2.79 and δ_C 25.64 ppm. The proton signals of the double bond are complex and appear overlapped, unable to determine the coupling constants that would allow the assignment of the double bond configuration. However, the ^{13}C NMR resonances δ_C 27.14 and 27.21 ppm for the two allylic methylenes at position 5' and 11', and δ_C 25.64 ppm for the bisallylic methylene C-8' are also in agreement with literature data reported for other resorcinols with Z geometry [21]. Taking into account the spectroscopic data, compound **6** was identified as (*Z,Z*)-5-(pentadeca-6,9-dienyl)-resorcinol.

Positive HRMS (ESI) of compound **9** showed molecular ion+Na at *m/z* 311 that correspond to a molecular formula $C_{19}H_{28}O_2$. Their ^{1}H and ^{13}C NMR spectra were similar to those of **6** and indicated C_{13} linear alkenyl resorcinol. ^{1}H NMR spectra revealed resonance of one bisallylic methylene group, suggesting that the double bonds were separated by methylene group. HSQC and HMBC experiments allowed the assignment of all protonated carbons and showed that the double bonds are placed at $\Delta^{5'}$ and $\Delta^{8'}$. The ^{13}C NMR resonances for the two allylic methylenes at position 4' and 10' have the same value δ_C 27.21 ppm and

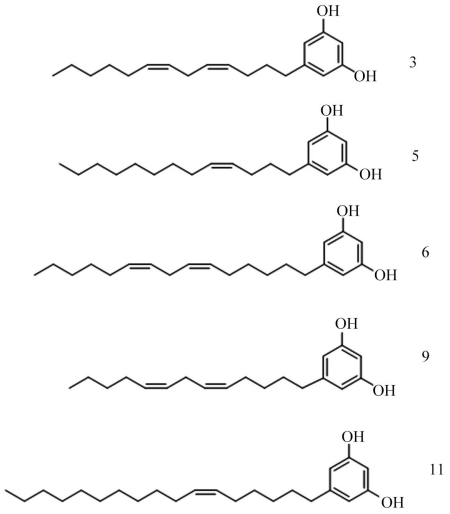

Figure 1. Structural formulas of the 5-alkenyl resorcinols isolated from *L. molleoides*.

Table 1. ^{13}C, ^{1}H NMR, HSQC, and HMBC Data of Compounds **6**, **9** and **11** in Cl_3CD[a].

	6			**9**			**11**		
position	δC	δH	HMBC	δC	δH	HMBC	δC	δH	HMBC
1	156.61			156.64			156.61		
2	100.18	6.20	1, 3, 4, 6	100.13	6.19	1, 3, 4, 6	100.17	6.20	1, 3, 4, 6
3	156.61			156.64			156.61		
4	108.03	6.26	2, 3, 6, 1’	107.99	6.26	2, 3, 6, 1’	108.60	6.26	2, 3, 6, 1’
5	145.98			146.08			146.11		
6	100.18	6.26	1, 2, 4, 1’	107.99	6.26	1, 2, 4, 1’	108.60	6.26	
1’	35.77	2.51	4, 5, 6, 2’, 3’	35.81	2.50	4, 5, 6, 2’, 3’	35.79	2.54	2’
2’	30.91	1.61	5, 1’, 3’, 4’	31.03	1.60	1’, 3’	30.98	1.61	1’, 3’
3’	28.92	1.37	4’, 2	29.23	1.33	2’, 4’	29.20	1.25 - 1.38	2’, 3
4’	29.33	1.37	5’	27.21	2.06	3’, 5’, 6’,	29.74	1.25 - 1.38	5’
5’	27.14[d]	2.08	6’, 7’, 9’, 10’, 4’	130.05[b]	5.33	4’, 7’,	27.19[d]	2.04	4’, 6’
6’	130.26[b]	5.40	8’	128.02[c]	5.34	4’, 7’	129.81[b]	5.40	5
7’	127.90[c]	5.37	8’	25.64	2.79	5’, 6’, 8’, 9’	129.97[b]	5.37	8’
8’	25.64	2.79	6’, 7’, 9’, 10’	127.94[c]	5.37	7’, 10’	27.22[d]	2.04	7’, 9’
9’	128.14[b]	5.37	8’	130.13[b]	5.38	7’, 10’	29.76	1.25 - 1.38	
10’	129.95[c]	5.40	8’	27.21	2.06	8’, 9’, 11’,	29.22	1.25 - 1.38	
11’	27.21[d]	2.08	6’, 7’, 9’, 10’, 12’, 13’	31.53	1.31	10’, 11	29.31	1.25 - 1.38	
12’	29.50	1.33		22.58	1.32	13’	29.29	1.25 - 1.38	
13’	31.55	1.34		14.08	0.90	11’, 12’	29.37	1.25 - 1.38	
14’	22.55	1.33	15’, 13’				29.50	1.25 - 1.38	
15’	14.03	0.90	14’, 13’				31.89	1.31	
16’							22.66	1.31	15’, 17’
17’							14.05	0.90	15’, 16’

[a]Correlations between ^{1}H and ^{13}C based on HSQC spectra. [b,c,d]Assignments interchangeable.

are in agreement with Z configuration. Based on the spectroscopic data compound **9** was identified as (*Z*,*Z*)-5-(trideca-5,8-dienyl)-resorcinol.

Their ^{1}H and ^{13}C NMR spectra of **11** were similar to those of **6** and indicated C_{17} linear alkenyl resorcinol. Negative HRMS (ESI) showed the unprotonated molecular ion at *m/z* 345 that corresponds to a molecular formula $C_{23}H_{38}O_2$. A multiplet integrating for two protons (δ_H 5.40 - 5.37 ppm) is observed in ^{1}H NMR spectrum. HSQC and HMBC experiments unambiguously allowed the assignment of a $\Delta^{6'}$ double bond. ^{13}C NMR chemical shifts of 27.19 and 27.22 ppm for allylic C-5' and C-8' confirmed the *Z* geometry in compound **11**. Based on the spectroscopic data compound **11** was identified as (*Z*)-5-(heptadec-6-enyl)-resorcinol.

The major alkenyl resorcinols **3** and **5** were previously isolated from *L. molleoides* and associated with nematicidal [20] and cytotoxic activities [15] [16]. Compound **2** was a mixture of isomers
(Z,Z,Z)-5-(trideca-4,7,10-trienyl)-resorcinol and
(Z,Z,E)-5-(trideca-4,7,10-trienyl)-resorcinol that could not be separated by the HPLC system used and these had already been reported by Valcic *et al.* [20].

This is the first report in the nature of the 5-alkenyl resorcinols **6**, **9** and **11**.

The knowledge of the bioactive compounds present in extracts obtained from medicinal plants is of great importance and contributes to the characterization of species with medicinal potential. Over the last decades, natural products have been shown to be very varied sources of compounds that served to search for new molecules with biological activities of importance in human medicine [22]. Given previous reports and the promissory activity of 5-alkyl/alkenyl resorcinols further studies must be conducted to test the biological activity of the new alkenyl resorcinols isolated from *L. molleoides*.

Acknowledgements

This work was supported by the University of Buenos Aires (UBACYT 2002017 0200222BA).

Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

References

[1] Knödler, M., Conrad, J., Wenzig, E.M., Bauer, R., Lacorn, M., Beifuss, U., Carle, R. and Schieber, A. (2008) Anti-Inflammatory 5-(11'Z-heptadecenyl)- and 5-(8'Z,11'Z-heptadecadienyl)-resorcinols from Mango (*Mangifera Indica* L.) Peels. *Phytochemistry*, **69**, 988-993. https://doi.org/10.1016/j.phytochem.2007.10.013

[2] Ediriweera, M.K., Tennekoon, K.H., Samarakoon, S.R., Adhikari, A., Thabrew, I. and de Silva, E.D. (2017) Isolation of a New Resorcinolic Lipid from *Mangifera zeylanica* Hook. f. Bark and Its Cytotoxic and Apoptotic Potential. *Biomedicine & Pharmacotherapy*, **89**, 194-200. https://doi.org/10.1016/j.biopha.2017.01.176

[3] Robles, P.C., Michelson, S.A., Mena, J., Escrig, D., Rojas, J.L., Sanchez, G.G., Hernández, R., Barrera, A.C., Rojo, L.E., Sauma, D., Kalergis, A.M., Imarai, M.,

Fernández, R., Robles, C.A., Leiva, S.E., Santander, R., Escobar, A. and Acuña, C.C. (2019) *Lithraea caustic* (Litre) Extract Promotes an Antitumor Response against B16 Melanoma. *Frontiers in Pharmacology*, **10**, 1201. https://doi.org/10.3389/fphar.2019.01201

[4] Muñoz, J.D.D. (2000) Anacardiaceae Lindley. In: Huzinker, A.T., Ed., *Flora Fanerogámica Argentina*, Ed. IMBIV-CONICET, Córdoba, 65, 1-29.

[5] Catálogo de las Plantas Vasculares de la Flora del Cono Sur (2020) Instituto de Botánica Darwinion, Buenos Aires, Argentina. http://www.darwin.edu.ar

[6] Toursarkissian, M. (1980) Plantas Medicinales de la Argentina. Ed Hemisferio Sur, Buenos Aires, 4.

[7] Ratera, E.L. and Ratera, M.O. (1980) Plantas de la flora Argentina empleadas en medicina popular. Ed. Hemisferio Sur., Buenos Aires, 127.

[8] Rondina, R.V., Bandoni, A.L. and Coussio, J.D. (2003) In: OEA-CYTED Editores, Plantas Silvestres Argentinas con Reconocidas Propiedades Medicinales o Tóxicas. Buenos Aires.

[9] Kott, V., Barbini, L., Cruañes, M.C., Muñoz, J.D., Cruañes, J., Vivot, *et al.* (1999) Antiviral Activity in Argentine Medicinal Plants. *Journal of Ethnopharmacology*, **64**, 79-84. https://doi.org/10.1016/S0378-8741(98)00098-1

[10] Penna, C., Marino, S., Vivot, E., Cruañes, M.C., Muñoz, J.D., Cruañes, J., Ferraro, G., Gutkind, G. and Martino, V. (2001) Antimicrobial Activity of Argentine Plants Used in Treatment of Infectious Diseases. Isolation of Active Compounds from *Sebastiania brasiliensis*. *Journal of Ethnopharmacology*, **77**, 37-40. https://doi.org/10.1016/S0378-8741(01)00266-5

[11] Gorzalczany, S., López, P., Acevedo, C. and Ferraro, G. (2011) Anti-Inflammatory Effect of *Lithraea molleoides* Extracts and Isolated Active Compounds. *Journal of Ethnopharmacology*, **133**, 994-998. https://doi.org/10.1016/j.jep.2010.11.031

[12] Araujo, C.E.P., Bela, R.T., Bueno, L.J.F., Rodrigues, R.F.O. and Shimizu, M.T. (2006) Anti-Ulcerogenic Activity of the Aerial Parts of *Lithraea Molleoides*. *Fitoterapia*, **77**, 406-407. https://doi.org/10.1016/j.fitote.2006.05.011

[13] Morucci, F., López, P., Miño, J., Ferraro, G. and Gorzalczany, G. (2012) Antinociceptive Activity of Aqueous Extract and Isolated Compounds of *Lithrea molleoides*. *Journal of Ethnopharmacology*, **142**, 401-406. https://doi.org/10.1016/j.jep.2012.05.009

[14] Garro, M., Salinas, I.A., Vega, A., Arismendi, S.A., Pelzer, L. and Saad, J.M.A. (2015) Gastroprotective Effects and Antimicrobial Activity of *Lithraea molleoides* and Isolated Compounds against *Helicobacter pylori*. *Journal of Ethnopharmacology*, **176**, 469-474. https://doi.org/10.1016/j.jep.2015.11.009

[15] Barbini, L., López, P., Ruffa, J., Martino, V., Ferraro, G., Campos, R. and Cavallaro, L. (2006) Induction of Apoptosis on Human Hepatocarcinoma Cell Lines by an Alkyl Resorcinol Isolated from *Lithraea molleoides*. *World Journal of Gastroenterology*, **12**, 5959-5963. https://doi.org/10.3748/wjg.v12.i37.5959

[16] López, P., Ruffa, M.J., Cavallaro, L., Campos, R., Martino, V. and Ferraro, G. (2005) 1,3-Dihydroxy-5-(tridec-4',7'-dienyl)benzene: A New Cytotoxic Compound from *Lithraea molleoides*. *Phytomedicine*, **12**, 108-111. https://doi.org/10.1016/j.phymed.2003.07.013

[17] López, P., Hidalgo