Role of suspension feeders in antarctic pelagic-benthic coupling: Trophic ecology and potential carbon sinks under climate change

Gastón Alurralde, Verónica L. Fuentes, Tamara Maggioni, Juancho Movilla, Alejandro Olariaga, Covadonga Orejas, Irene R. Schloss, Marcos Tatián

Marine
Environmental
Research

F. Regoli, I. M. Sokolova

PII: S0141-1136(19)30367-8

DOI: https://doi.org/10.1016/j.marenvres.2019.104790

Reference: MERE 104790

To appear in: Marine Environmental Research

Received Date: 12 June 2019

Revised Date: 2 September 2019 Accepted Date: 7 September 2019

Please cite this article as: Alurralde, Gastó., Fuentes, Veró.L., Maggioni, T., Movilla, J., Olariaga, A., Orejas, C., Schloss, I.R., Tatián, M., Role of suspension feeders in antarctic pelagic-benthic coupling: Trophic ecology and potential carbon sinks under climate change, *Marine Environmental Research* (2019), doi: https://doi.org/10.1016/j.marenvres.2019.104790.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2019 Published by Elsevier Ltd.

- 1 Role of suspension feeders in Antarctic pelagic-benthic coupling: trophic ecology and
- 2 potential carbon sinks under Climate Change

3

- 4 Gastón Alurralde^{1,2}, Verónica L. Fuentes³, Tamara Maggioni^{1,2}, Juancho Movilla^{3,4}, Alejandro
- 5 Olariaga³, Covadonga Orejas⁵, Irene R. Schloss^{6,7,8}, Marcos Tatián^{1,2}

6

- 7 ¹ Universidad Nacional de Córdoba, Facultad de Ciencias Exactas, Físicas y Naturales,
- 8 Departamento de Diversidad Biológica y Ecología, Ecología Marina, Córdoba, Argentina.
- 9 ² Instituto de Diversidad y Ecología Animal (IDEA) CONICET, Córdoba, Argentina.
- 10 ³ Institut de Ciències del Mar (ICM-CSIC). Barcelona, Spain
- 11 ⁴ Instituto Español de Oceanografía, Centro Oceanográfico de Baleares, Estación de
- 12 Investigación Jaume Ferrer, Mahón, Spain
- 13 ⁵ Instituto Español de Oceanografía. Centro Oceanográfico de Baleares. Palma, Mallorca,
- 14 Spain
- 15 ⁶ Instituto Antártico Argentino, Ciudad de Buenos Aires, Argentina
- 16 ⁷ Centro Austral de Investigaciones Científicas, CONICET, Ushuaia, Argentina
- 17 ⁸ Universidad Nacional de Tierra del Fuego, Ushuaia, Argentina

18

19

20

21

- This research is an important contribution to previously unquantified aspects of the effects of climate change in species-mediated energy transfer between pelagic and benthic Antarctic coastal areas.
- 22 **Author Contributions:** GA, MT, VLF originally formulated the idea, GA, MT, VF, AO,
- 23 CO, IRS conceived and designed the experiments, AO, JM developed and constructed
- 24 experimental setup, GA, TM, JM conducted fieldwork and performed the experiments, GA
- 25 performed statistical analyses and developed the energy budget, GA wrote the manuscript,
- all the authors contributed with the interpretation of results and provided editorial advice.

HIGHLIGHTS

- Food processing and energy acquisition was studied on two Antarctic filter-feeder
 species subjected to different food sources and sediment concentrations.
- A general energy model was developed to assess the energy balance of an ascidian
 and to evaluate its potential as a carbon sink in coastal Antarctic ecosystems.
- Food characteristics and animal densities modulate the impact of ascidians on
 pelagic-benthic coupling and on carbon immobilisation.
 - Increasing inputs of sediment runoff would be a major constraint in coastal areas.

ABSTRACT

Sea-ice and coastal glacier loss in the Western Antarctic Peninsula open new ice-free areas. They allowing primary production and providing new seabed for colonisation, both acting as a negative feedback of climate change. However, the injection of sediment-laden runoff from the melting of land-terminating glaciers may reduce this feedback. Changes in particulate matter will affect nutrition and excretion (faeces stoichiometry and properties) of suspension feeders, reshaping coastal carbon dynamics and pelagic-benthic coupling. Absorption efficiency and biodeposition of *Euphausia superba* and *Cnemidocarpa verrucosa* were quantified for different food treatments and varying sediment concentrations. Both species showed high overall absorption efficiency for free-sediment diets, but were negatively affected by sediment addition. High sediment conditions increased krill biodeposition, while it decreased in ascidians. Energy balance estimation indicated high carbon sink potential in ascidians, but it is modulated by food characteristics and negatively affected by sediment inputs in the water column.

KEYWORDS Potter Cove, *Euphausia superba, Cnemidocarpa verrucosa*, absorption efficiency, faecal production, biodeposition, carbon sink

INTRODUCTION

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

While the rapid temperature increase in the Antarctic Peninsula seems to be paused since the late 1990s due to a short-term natural climate variability (Turner et al. 2016), evident impact of climate change is observed in the West Antarctic Peninsula (WAP) over the cryosphere (Rückamp et al. 2011, Cook et al. 2014, Gutt et al. 2015), the continental shelf and in particular the coastal area (Meredith and King 2005, Cook et al. 2016). Furthermore, climate model projections indicate that a large anthropogenic regional warming signal is likely to result in substantial warming across the Peninsula over the 21st century (Bracegirdle et al. 2008, Etourneau et al. 2019).

The loss of ice shelves and the retreat of coastal glaciers in Antarctica over recent decades, have created around 2.4 x 10⁴ km² of new open water, enhancing planktonic primary production (Peck et al. 2010, Moreau et al. 2015, Jansen et al. 2018a), but also new ice-free seabed areas prone to benthic colonisation (Bowden et al. 2006, Barnes et al. 2018, Jansen et al. 2018b). For instance, in Potter Cove (King George Island/Isla 25 de Mayo), high macroalgae and suspension feeder abundances have been reported in new ice-free areas (Quartino et al. 2013, Lagger et al. 2017, 2018). Similarly, in the Mertz Glacier Tongue (East Antarctic), there has been a large increase in suspension feeder abundance after massive ice loss (Jansen et al. 2018b). Such new production (planktonic and benthic) represents an important food source and an increase in energy and biomass for the whole ecosystem (Barnes et al. 2018, Jansen et al. 2018a). Furthermore, a high potential carbon (C) sink (blue carbon storage) could act as negative feedback to climate change due to water column C drawdown (Peck et al. 2010, Quartino et al. 2013, Barnes 2015, Barnes et al. 2018). Indeed, benthic secondary production would be an important C sink, as it involves long-term carbon storage in animal biomass (Rossi et al. 2012, Barnes 2015, Barnes et al. 2016).

Some WAP coastal areas are subjected to seasonal and sporadic injections of fresh water loaded with sediments due to melting of marine-terminating glaciers (Klöser et al. 1994, Monien et al. 2017, Meredith et al. 2018). In these areas phytoplankton and macroalgae growth seem to be arrested via light-limitation (Dierssen et al. 2002, Schloss et

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

al. 2002, 2012, Quartino et al. 2013, Deregibus et al. 2016). Furthermore, low concentrations of key nutrients and organic particulates occur when inorganic particle loads are high, which constrains food supplies for planktonic and benthic suspension feeding species (Klöser et al. 1994, Pakhomov et al. 2003, Müller-Navarra 2008, Fuentes et al. 2016). A high concentration of suspended sediments can also clog the filtering apparatus of these species, which further constraints physiological performance (Torre et al. 2012, 2014) and can even lead to massive mortality events (Pakhomov et al. 2003, Fuentes et al. 2016).

Suspension feeders are able to feed on a wide spectrum of food particle types and sizes (Riisgård and Larsen 2010). Through their trophic activity (ingestion and faecal production) these animals play a major role in pelagic-benthic coupling (PBC) and in the flow of C through marine shallow ecosystems, modulating the cycling of nutrients and moving C from the pelagic to the benthic realm and vice versa (Gili and Coma 1998). Sediment trap studies in shallow nearshore waters (5-25 m depth) indicate that faecal pellets from krill and other mesozooplanktonic consumers are important components of the organic matter (OM) flux (Schloss et al. 1999, Isla et al 2001, Alurralde 2018). Antarctic krill (Euphausia superba) is a key planktonic suspension feeder occurring in the Southern Ocean (Siegel 2016). High densities are frequent in the coastal areas of the WAP (Atkinson et al. 2009, Warren and Demer 2010, Steinberg et al. 2015, Belcher et al. 2016, 2017), where 66.7-96.3% of total particulate organic carbon (POC) flux in shallow waters (20-180 m depth), is represented by krill faeces (Isla et al. 2001, Belcher et al. 2016, 2017). In Potter Cove, krill annual abundances can be as high as ~3.000 ind.100 m⁻³ (Fuentes 2006), which promote high but seasonally variable inputs of faecal pellets (1.5 10³ -7.53 10⁵ mm³.m⁻².d⁻¹) sinking to the seafloor (Alurralde 2018). This enable benthic suspension feeders such as hydrozoans, corals, glass sponges and ascidians to feed directly on them and benefit from their organic contents (Orejas et al. 2000, Duineveld et al. 2004, Leys et al. 2004, Tatián et al. 2004). This functional group is the dominant component of the Southern Ocean shelf benthos (Gili et al. 2006, Gutt et al. 2013) and in contrast to the Arctic, nearshore biomass and diversity in fjords is much higher than the offshore sites (Grange and Smith 2013). Suspension feeders not

only enhance PBC in shallow waters (Ahn 1993, Tatián et al. 2004), but also develop complex three-dimensional biogenic structures particularly on soft bottoms (Tatián et al. 1998, Gili et al. 2006), where epibiosis is a common phenomenon especially over solitary ascidians (Rimondino et al. 2015). Solitary and compound ascidians comprise dominant representatives of Antarctic epifaunal macrobenthic communities on the shelf (Gutt et al. 2013, Segelken-Voigt et al. 2016), fjords and coastal areas (Grange and Smith 2013, Moon et al. 2015, Sahade et al. 2015, Lagger et al. 2018). In new ice-free areas of the WAP ascidians reach the higher abundances (Lagger et al. 2017, 2018), which could increase retention of a greater share of the productivity in their living structures for decades (Kowalke et al. 2001). Nevertheless, these organisms of apparently great C sink potential in shallow waters (Petersen and Riisgård 1992, Coppari et al. 2014) are generally neglected in the overall equation of C circulation in our changing world (Rossi et al. 2015).

The abundance and distribution of benthic suspension feeders depend on the quality of available food sources (Gardner 2000, Rossi et al. 2015, Jansen et al. 2018a,b). The degree to which the quantity and composition of accessible food meets the consumer's nutritional needs defines food quality (Müller-Navarra, 2008), while the characteristics of the diet define the way in which the digestion process takes place in suspension feeders (e.g. Jørgensen 1996, Mayer et al. 1997, Hawkins et al. 1998). Only part of the ingested food is absorbed (i.e., transported across the gut wall); the rest is expelled as faeces. A fraction of the absorbed food is diverted to growth and reproduction, while another fraction is used to maintain resting metabolism. If the energy expended in resting metabolism is lower than the energy gained through digestion, organisms are in a position of net energy gain, thus enabling biomass production that immobilises C in somatic tissues. But if absorption is low, a higher proportion of OM would be transferred to the seabed through faeces that will be remineralised by microbial activity and hence released back into the water column later (Claquin et al. 2010, Mermillod-Blondin 2014), buried in the sediments and/or consumed by detritivores, a process known as biodeposition (Ahn et al. 1993, Tatián et al. 2008, Mermillod-Blondin 2014). Consequently, the quantification of uptake efficiency for different food sources and of biodeposition are essential to assess energy flows through benthic andpelagic ecosystems.

Predicted changes in the properties of suspended particulate matter along the WAP (Montes-Hugo et al. 2009, Schofield et al. 2010) are expected to have profound implications for the trophic processes of suspension feeders, such as energy uptake (Robbins 1985, Wotton and Malmqvist 2001, Atkinson et al. 2012a), which will ultimately impact their net energy balance. The ecological consequences may be dramatic since the WAP supports a high abundance of both planktonic (Atkinson et al. 2012b, Puigcorbé et al. 2016) and benthic (Gutt et al. 2013) suspension feeders, and these species are the food supply to the scavengers and carnivores that form the rest of the benthic food web. Furthermore, faecal production may be affected by alterations in the rate, stoichiometry and properties of pellets (Ragueneau et al. 2005, Gergs et al. 2009), with cascade effects on ecosystem functioning (Claquin et al. 2010, Alcaraz et al. 2014, Coello-Camba et al. 2017). In the same way, high sediment inputs in marine coastal areas could drastically impact suspension feeders (Fuentes et al. 2016, Sahade et al. 2015), limiting blue carbon storage ecosystem services as it occurs with ice scour disturbance (Barnes 2017, Barnes et al. 2018).

In the present study, we first examined the absorption efficiency (AE) and biodeposition of *E. superba* and *C. verrucosa* subjected to different food conditions (diet composition and sediment concentrations). These abundant and conspicuous filter feeders are able to sieve particles from the water in a wide range of sizes (0.06 µm to >100 µm), overlapping with other suspension feeders (Riisgård and Larsen 2010, Suh and Nemoto 1987) making them good representatives of the suspension feeder guild. By means of a general energy budget, we assessed the energy balance of *C. verrucosa* under those food conditions to evaluate its potential as a C sink and the consequences of alternative food scenarios (i.e. phytoplankton- or macroalgal detritus-dominated seston, and increased suspended sediment concentration). Finally krill was considered in its role of food provider for benthic communities throughout biodeposition. To this end, we included krill faeces as food source for ascidians.

METHODS

Study area. Experiments were conducted at the Argentinean-German Dallmann Laboratory in the Argentinean Antarctic station Carlini (former Jubany) next to the study area in Potter Cove (62° 14' S, 58° 40' W), in King George Island (25 de Mayo), South Shetland Islands. The cove (Fig. 1), which is surrounded by the Fourcade Glacier on its north and east sides, is 4 km long and 2.5 km wide (Al-Handal and Wulff, 2008) with a maximum depth of 100 m. During the summer, melt water creeks carry large amounts of sediment from the land-terminating glacier (average maximum 284 mg L⁻¹), which are discharged into the cove by several creeks (Monien et al. 2017). A comprehensive description of the coastal ecosystem of Potter Cove can be found in Wiencke et al. (2008), and the hydrographical characteristics and environmental conditions for the last two decades are reported by Schloss et al. (2012) and Meredith et al. (2018).

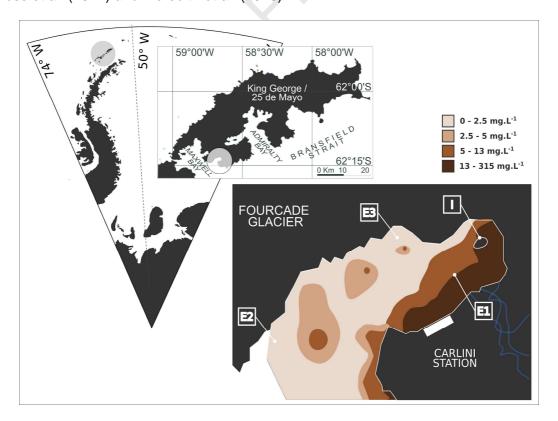


Figure 1. Location of Potter Cove, King George/25 de Mayo Island, South Shetland Islands, Antarctica. Average spatial distribution of total suspended particulate matter (TSPM)

concentration in the surface waters of Potter Cove during summer 2010/2011 (based on Figure 5a, Monien et al. 2017). E1, E2, E3, indicate the sampling stations mentioned by Sahade et al. 1998 and I, the ice-free area (Island) mentioned by Lagger et al. 2018.

Target species and collection. During the summers of 2012/2013 and 2013/2014, we collected juvenile and adult *E. superba* from the outer part of Potter Cove using a 200 μ m mesh Nansen net with a 2 L cod end, towed vertically and obliquely (40 m below the surface). The contents of the cod end were immediately transferred to a 50 L plastic bucket filled with cooled (0°C) filtered seawater and then taken directly to a cool room (0 \pm 1°C), where the animals were placed in a 100 L container (further details in Fuentes et al. (2016)). Specimens of *C. verrucosa* were collected by SCUBA divers between 20 - 30 m depth in the inner part of Potter Cove (Fig. 1). Sampled individuals were immediately transported to the station aquaria in a 50 L plastic bucket filled with cooled (0 \pm 1°C) seawater and subsequently placed in a 100 L container after cleaning their tunics from large epibionts. Both species were kept in an open flow system with natural seawater (0 \pm 1°C) directly pumped from the cove for at least 10 days for acclimation (Torre et al. 2012, 2014). Prior to the experiments, krill specimens were starved for 48 h and ascidians for 5 days in filtered seawater (0.1 μ m) to allow the specimens to empty their stomachs.

Experimental setup. For both species, we used six independent cylindrical 8-L aquaria with individual closed circulation systems, placed in a 90 L running seawater bath for the incubation experiments at constant temperatures ($0 \pm 1^{\circ}$ C). We fitted a mesh (200 µm for krill and 1 mm for ascidians) 10 cm above the bottom of each cylindrical aquarium, allowing water to flow and avoiding the disaggregation of the faeces in the circulating system. For each treatment (see below) we used one control and five replicates. For krill incubation, we placed 12 juvenile krill specimens per aquarium (range size: 35-46 mm total length). For ascidian incubation, we placed 1 individual per aquarium (range size: 8-12 cm high).

Test Diets. Two types of diet were used (Table 1). Primary diets consisted of: 1) natural seston (S) present in surface seawater pumped directly from the cove; 2) natural

phytoplankton (P); 3) macroalgal detritus (D). Natural phytoplankton was collected by towing a 70 µm mesh net horizontally in the outer part of the cove to ensure the predominance of phytoplankton in the sample, which was transferred to a 60 L tank of 0.1 µm filtered seawater. Macroalgal detritus was collected by sieving debris directly from the water column on days following storms when there were significant amounts of resuspended detrital deposits. Macroalgal detritus was dried at 60°C, ground and sieved through a 50 µm mesh before being suspended in 0.1 µm filtered seawater at a final concentration of 5 mg L⁻¹. To evaluate the effect of suspended inorganic sediments, we used natural seston and natural seston with the addition of two suspended sediment concentrations: 1) 20 mg L⁻¹ (S and 20 mg of sediment per litre; S20); 2) 40 mg L⁻¹ (natural seston and 40 mg of sediment per litre; S40). These concentrations are within the range of the *in situ* concentrations recorded in Potter Cove (Pakhomov et al. 2003, Phillipps et al. 2011, Schloss et al. 2012, Sahade et al. 2015). The sediment added (<50 µm) in the experiments was obtained from the surface bottom sediments of the inner cove area (around 15 m depth, close to the mouth of the major creek), dried at 70°C and sieved through a 50 µm mesh, following Torre et al. (2012). Sediment OM content was gravimetrically estimated to correct subsequent calculations and to avoid biases in the estimation of trophic parameters.

231

232

233

234

235

230

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

Table 1: Details of the diet composition (food treatments) used in the feeding experiments and explanation of the abbreviations used. S, S20, S40, P were the primary diets offered to *Euphausia superba* and *Cnemidocarpa verrucosa*. FS, FS20, FS40, FP and FD were the secondary diets offered only to *C. verrucosa*.

236

Primary diets

- **S** Natural seston (running seawater pumped directly from the cove at 6 m depth)
- **S20** Natural seston with 20 mg L⁻¹ of sediment added
- **S40** Seston with 40 mg L⁻¹ of sediment added
- P Microalgae collected by towing of 50 µm net and diluted in GF/F filtered seawater

Detritus of macroalgae collected with a 1 mm sieve from the sea surface, dried, crushed and sieved through a 50 µm mesh. Concentration: 5 mg L⁻¹ in GF/F filtered seawater

Secondary diets

- Faecal pellets of krill fed on seston diet (S). Concentration: 5 mg L⁻¹ of faecal pellets suspended in GF/F filtered seawater
- FS20 Faecal pellets of krill fed on S20 diet. Concentration: 5 mg L⁻¹ of faecal pellets suspended in GF/F filtered seawater
- FS40 Faecal pellets of krill fed on S40 diet. Concentration: 5 mg L⁻¹ of faecal pellets suspended in GF/F filtered seawater
- FP Faecal pellets of krill fed on phytoplankton diet (P). Concentration: 5 mg L⁻¹ of faecal pellets suspended in GF/F filtered seawater
- Faecal pellets of krill fed on macroalgae detritus diet (D). Concentration: 5 mg L⁻¹ of faecal pellets suspended in GF/F filtered seawater

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

Secondary diets (only used to feed *C. verrucosa*) consisted of krill faeces obtained from krill fed on the same primary diets and sediment conditions described above. Hence, a total of five secondary diets were used to feed *C. verrucosa*: 1) faeces of krill obtained after animals fed on S (FS); 2) faeces of krill obtained after animals fed on S20 (FS20); 3) faeces of krill obtained after animals fed on P (FP); 5) faeces of krill obtained after animals fed on D (FD). Faeces from each condition were collected, dried and suspended in 0.1 µm filtered seawater at a final concentration of 5 mg L⁻¹.

Absorption Efficiency (AE). We estimated the AE of the organic fraction by means of the Conover ratio (Conover 1966), which assumes that only the organic component of the food is significantly affected by the digestive process. We calculated the AE as the difference of the ratio of mass loss after combustion of food and the corresponding percentage of mass loss after combustion of faeces, as follows:

252
$$AE= (F - E) / [(1 - E) * F]$$

- where F is the organic fraction of the food and E the organic fraction of the faeces. AE values are reported as percentages.
- To determine the organic fraction in food, at the beginning of each experiment (i.e.

each diet condition) we measured total suspended particulate matter (TSPM) concentration gravimetrically after filtering 0.25 – 2 L of aquaria seawater through combusted (500°C, 4.5 h) and pre-weighed 25 mm Whatman GF/F filters. After filtration, we rinsed filters twice with distilled water, dried them for 24 h at 60°C, and weighed them again to obtain the dry weight (DW) of the particles in the water samples. Filters were then burned at 500°C for 4.5 h to obtain ash weight of the sample, and by difference with the dry weight, the ash-free dry weight (AFDW). To estimate the organic fraction in the faeces, we used the same process as with the filters. In both cases, we measured the inorganic material directly as ash weight after ignition at 500°C, from which ash-free dry mass was estimated by subtraction from the dry weight values to determine the particulate organic matter (POM) and the particulate inorganic matter (PIM).

Faecal Production and Biodeposition Rates. We calculated the Faecal Production Rate

Faecal Production and Biodeposition Rates. We calculated the Faecal Production Rate (FPR) as the DW of faeces produced individual day, while we also presented the data as the Mass-specific Faecal Production Rate (MSFPR), which was the mass of faeces produced day divided by the animal body mass (not including the tunica in the ascidians), for comparison purposes with previous studies (e.g. Tatián et al. 2008, Torre et al. 2012, 2014). For experiments conducted with krill, faeces were collected every 30 minutes during the first h of incubation and hourly until 24 h incubation, following the protocol reported by Atkinson et al. (2012a). For experiments conducted with ascidians, we collected faeces every 24 h following the protocol of Tatián et al. (2008). After the experiments, we dried the organisms at 60°C, weighed them and then ignited them in a muffle furnace at 500°C for, 4.5 h, when they were weighed again. The organic fraction (AFDW) of each specimen was obtained from the difference between the ash weight after ignition and DW. When FPR was related to animal densities, we called this the Biodeposition Rate (BR).

Carbon Balance. To estimate the potential of *Cnemidocarpa verrucosa* as a C sink and its role in the PBC, we followed the approach of Coppari et al. (2014). Since ammonia excretion was not measured, the estimation of C balance is expected to be slightly elevated. The balanced C equation (without ammonium excretion) was considered as follows:

 $C_{j}=T+R+F$

where C_i is the total C ingested (as a function of the filtration rate (FR) and the C content of the food: FR*C_{food}), T is the production of somatic and reproductive tissues, R is the respiratory C loss, and F is the faeces C loss. Because the absorbed fraction (A) is the product of C_i and AE, T can be expressed as the difference between C absorbed from the food and C expenditure due to respiration (T= A - R). Hence T reflects the overall C balance and the values represent estimates of C gain or loss. Positive T values reflect the C which is available for biomass production (C sink potential) after meeting routine metabolic demands, whereas negative T values reflect an overall deficit of C.

Specific daily rates of filtration were calculated for each tested diet, based on the measurements of faecal pellet production by using the following relation:

$$FR = PR_{IF} / PIM = [FPR (1/E)] / PIM$$

where PR_{IF} is the inorganic fraction of Faecal Production Rate (FPR) and E is the organic content of faeces, as used in the Conover equation. This calculation (Iglesias et al. 1998) is correct on the only assumption that inorganic matter is a conservative tracer in the gut (no absorption of inorganics would occur). The oxygen consumption of *C. verrucosa* wasobtained from published values for standard conditions (Kowalke et al. 2001) and after sediment exposure to increased sediment loads (Torre et al. 2012). All data were standardised to 1 g dry tissue weight. Oxygen consumption was converted to carbon dioxide production using the respiratory quotient ($RQ = CO_2$ evolved/ O_2 consumed). We assumed the respiratory quotient (RQ) of 0.949 recommended for ammonotelic organisms as ascidians (Lampert 1984). In 3 water sample replicates (3 x 250 ml per replicate) collected at the beginning of the experiments for all treatments, we determined the C content of food (after removing calcium carbonate with hydrochloric acid) using a Carlo-Erba EA1108 analyser.

Known densities of *E. superba* (Fuentes 2006) and *C. verrucosa* for Potter Cove (Sahade et al. 1998, Lagger et al. 2017, 2018) and shelf areas of the WAP (Steinberg et al. 2015; Segelken-Voigt et al. 2016) were used to produce local scale estimates of the potential

312 biodeposition rates and C sinks.

Statistical analysis. We tested the normality of residuals using the Shapiro-Wilks test (Mahibbur Rahman and Govindarajulu 1997) and confirmed homoscedasticity by Levene's tests (Montgomery 1997). One-way ANOVA was employed to test the effects of the independent variables (primary and secondary diets) upon each separate trophic dependent variable (AE, FPR and MSFPR). The concentrations of S and P diets were uncontrolled, since both reflected the natural amounts in the local seawater. Moreover, those values may be related to the variables under study (especially for krill feeding). Thus, we used TSPM as a covariate. In the cases where the covariate was significant, we reported ANCOVA results. Finally, we performed one-way ANOVA to test differences in the carbon balance of *C. verrucosa* related to the different treatments. In all cases, when we encountered significant differences in ANOVA, we used the Tukey–Kramer method (Miller 1981) as a *post-hoc* test. All statistical analyses were performed using InfoStat v. 2016 (Di Rienzo et al. 2016).

RESULTS

Test Diets. The composition and concentration of the different treatments tested are summarized in Table 2. The initial TSPM of each treatment did not differ between experiments with krill or ascidians (P > 0.05), with the exception of the P treatment, which was higher in the case of ascidians (Table 2). However, the treatments showed differences between experiments with krill and ascidians in terms of initial PIM, POM and the organic fraction (P < 0.001). The organic component (POM) of S did not differ among krill and ascidian experiments (F = 0.34; P = 0.5737). Secondary diets (krill faeces) showed no difference in terms of initial TSPM nor PIM, but there was difference for the initial POM and for the organic fraction, f (P < 0.001). Food C content varied significantly among the different treatments (primary and secondary diets) for both species (P < 0.0001 in all cases, Table 2). The carbon to nitrogen ratio in primary diets was significantly higher for D (P = 0.014 for krill experiments, P = 0.027 for ascidian experiments), but no differences were observed for secondary diets (F = 2.42; P = 0.081).

Table 2. Composition and concentration of diet used in the feeding experiments conducted with *Euphausia superba* and *Cnemidocarpa verrucosa*. Values are given as mean ± standard deviation. TSPM: Total Particulate Matter (mg L⁻¹), PIM: particulate inorganic matter (mg L⁻¹), POM: particulate organic matter (mg L⁻¹), *f*: organic fraction (POM/TSPM), ChI a: chlorophyll a (μg L⁻¹), C:N: carbon and nitrogen ratio, POC: particulate organic carbon (mg L⁻¹).

| Species | Diet | TSPM (mg L ⁻¹) | PIM (mg L ⁻¹) | POM (mg L ⁻¹) | f (POM/TSPM) | C:N | POC (mg L ⁻¹) |
|--------------|------|-------------------------------|------------------------------|------------------------------|-----------------|-----|------------------------------|
| E. superba | S | 2.39±0.53 | 0.41±0.20 | 1.98±0.35 | 0.83±0.07 | 6.2 | 0.26±0.19 |
| | S20 | 22.30±1.60 | 15.16±1.11 | 7.14±0.48 | 0.32±0.03 | 6.1 | 1.14±1.08 |
| | S40 | 42.80±2.30 | 33.38±1.77 | 9.42±0.50 | 0.22±0.01 | 6.4 | 1.12±0.56 |
| | Р | 5.81±1.27 | 2.50±1.05 | 3.31±0.58 | 0.57±0.02 | 6.3 | 1.56±0.60 |
| | D | 4.72±0.39 | 1.65±0.56 | 3.07±0.30 | 0.65±0.10 | 7.6 | 0.94±0.23 |
| C. verrucosa | S | 2.64±0.94 | 1.00±0.55 | 1.64±0.65 | 0.62±0.12 | 5.0 | 0.09±0.03 |
| | S20 | 21.44±1.59 | 17.37±1.61 | 4.07±0.14 | 0.19±0.02 | 5.7 | 0.50±0.05 |
| | S40 | 39.92±1.06 | 32.73±7.74 | 7.19±1.36 | 0.18±0.02 | 6.5 | 0.63±0.10 |
| | Р | 17.73±3.48 | 10.29±3.14 | 7.44±0.72 | 0.42±0.08 | nd | nd |
| | D | 4.93±0.43 | 2.91±0.33 | 2.02±0.56 | 0.41±0.08 | 6.8 | 1.07±0.38 |
| C. verrucosa | FS | 4.93 ±0.77 | 3.50±0.64 | 1.430±0.28 | 0.29±0.05 | 8.8 | 0.23±0.05 |
| | FS20 | 4.69 ±0.34 | 3.80±0.38 | 0.891±0.09 | 0.19±0.02 | 8.0 | 0.17±0.04 |
| | FS40 | 4.67±0.17 | 3.92±0.25 | 0.75±0.09 | 0.16±0.02 | 6.7 | 0.16±0.04 |
| | FP | 4.74±0.21 | 3.46±0.12 | 1.280±0.14 | 0.27±0.02 | 7.2 | 0.11±0.02 |
| | FD | 4.71±0.26 | 3.39±0.20 | 1.32±0.13 | 0.28±0.02 | 7.8 | 0.11±0.01 |

Absorption Efficiency (AE). The mean AE was higher for S in both species (*Euphausia superba*: $88.29\% \pm 0.04$; *Cnemidocarpa verrucosa*: $86.05\% \pm 0.07$), reaching about 60% for the sediment-free primary diets (P and D); however, these were not significantly different (P > 0.05) (Fig. 2a, b). When S was compared to S20 and S40, the addition of sediment caused a 75% drop in AE in both species. Sediment concentration was inversely related to krill AE and differed significantly among the three tested concentrations (S, S20, S40) (F = 27.85, P < 0.0001) (Fig. 2a); the fall in AE was greater for S40 than for

S20 (P < 0.05; $15.75\% \pm 3.62$ and $21.44\% \pm 15.27$, respectively). The same response was observed for C. verrucosa (F = 9.17, P = 0.0045), where its AE was reduced to 23.83% ± 12.44 and 15.12% ± 7.42 in the S20 and S40 treatments respectively. However, no differences (P > 0.05) were detected between these two concentrations (Fig. 2b). For C. verrucosa fed on secondary diets, the maximum mean AE (52.6% ± 5.91) was recorded in the FD diet (F = 6.60; P = 0.0015), while the lowest mean AE (21.97% \pm 16.19) corresponded to the FS40 diet (Fig. 2c). In the remaining secondary diets (FS, FS20, FP) there were no significant differences.

Faecal Production Rate. The mean FPR of *E. superba* did not differ between diets P (2.9 mg.ind $^{-1}$.d $^{-1}$ ± 1.2) and D (3.0 mg.ind $^{-1}$.d $^{-1}$ ± 0.9), but it was lower for S (1.4 mg.ind $^{-1}$.d $^{-1}$ ± 0.6) (F = 12.52, P < 0.0001). Sediment concentration in diets S, S20 and S40 was positively related (F = 82.75, P < 0.0001) to mean FPR, with a higher production (4.6 mg.ind $^{-1}$.d $^{-1}$ ± 0.3) feeding on S40 (Fig. 3a). The mean MSFPR of krill exhibited a trend similar to FPR (Fig. 3a) although no significant differences were detected among diets (F = 1.26, P = 0.32). Similarly in krill diets S, S20 and S40 did not produce any differences in MSFPR (F = 2.45, P = 0.1279).

Faecal Production Rates (FPR and MSFPR) of *C. verrucosa* were variable for both primary and secondary diets (Fig. 3b, c). For primary diets, the highest rates were recorded when animals were feeding on S (FPR: F = 5.06, P = 0.0056; MSFPR: F = 4.63, P = 0.0082) (Fig. 3b), while for secondary diets the highest rates were achieved under FP and FD diets (FPR: F = 13.16, P < 0.0001; MSFPR: F = 8.73, P = 0.0003) (Fig. 3c). Sediment addition was related negatively with the amount of faeces produced, and there were differences among all diets (S, S20, S40) (FPR: F = 9.87, P = 0.0029; MSFPR: F = 7.22, P = 0.0088). Sediment had no detectable effect among secondary diets FS, FS20 and FS40 (FPR: F = 0.91, P = 0.4269; MSFPR: F = 0.17, P = 0.8463).

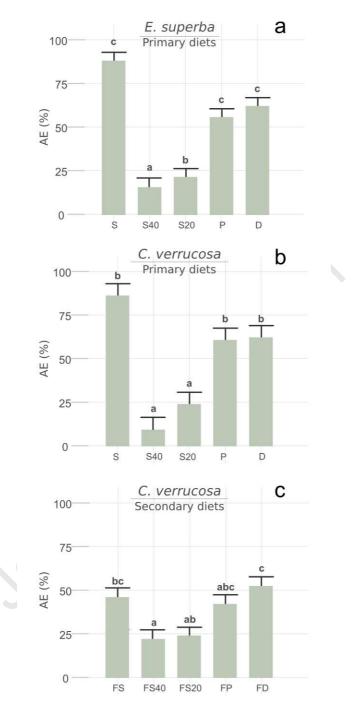


Figure 2. Absorption Efficiency (AE) (expressed in % as mean ± standard error) under the different treatments (see Table 1 for abbreviations). **a** *Euphausia superba* feeding on primary diets. **b** *Cnemidocarpa verrucosa* feeding on primary diets. **c** *C. verrucosa* feeding on secondary diets (krill faecal pellets). Different letters on the top of the bars indicate significant differences (P < 0.05). Data of *E. superba* AE on diets S, S20 and S40 are redrawn from Fuentes et al. (2016).

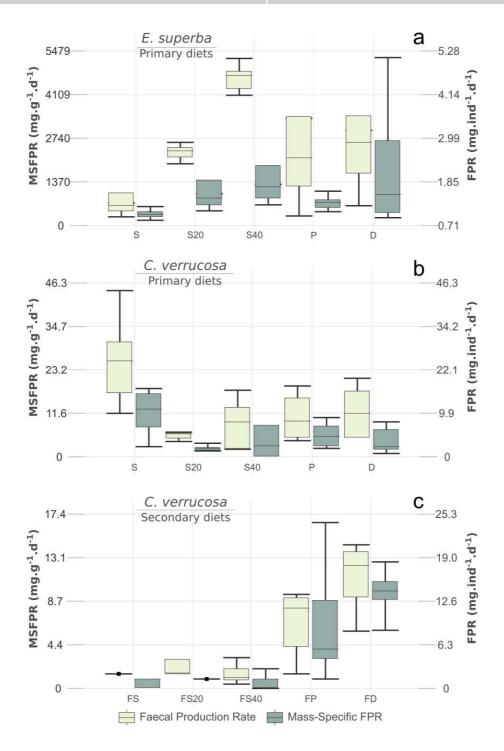


Figure 3. MassSpecific Faecal Production Rate (MSFR) expressed as mg faeces g.ind⁻¹ d⁻¹ and Faecal Production Rate (FPR) expressed as mg faeces ind⁻¹ d⁻¹. **a** *Euphausia superba* feeding on primary diets. **b** *Cnemidocarpa verrucosa* feeding on primary diets. c *C. verrucosa* feeding on secondary diets (krill faeces).

Carbon Balance. Based on the C content of the experimental diets and the AE determined for *C. verrucosa*, maximum mean C absorption (*A*) was obtained for S (11.69 \pm 8.75 mgC g⁻¹ d⁻¹) followed by D (5.97 \pm 3.46 mgC g⁻¹ d⁻¹), being higher than the other primary diets (F = 5.50, P =0.0104; Table 3). In contrast, for secondary diets, the highest mean *A* (11.38 \pm 2.82 mgC g⁻¹d⁻¹) was estimated for FD (F = 19.86, P < 0.0001; Table 3). Mean respiratory C loss (*R*) differed among primary diets (F = 214.01, P < 0.0001), and there was a negative effect of sediment addition. The highest sediment concentration (S40) generated the highest *R* (67.68 \pm 6.76mgC g⁻¹ d⁻¹), followed by the S20 diet where *R* was 55.43 \pm 8.08 mgC g⁻¹ d⁻¹. The remaining treatments had no detectable on R (Table 3), and *R* did not differ among secondary diets (P = 0.24).

Table 3. Estimated C ingested (C_i) (as a function of the filtration rate and the C content of the food), C absorbed (A) (the product of C_i and AE) and respiratory C loss (R) for Cnemidocarpa verrucosa (mgC.g⁻¹d⁻¹), considering the C content of the different diets. Values are expressed as mean \pm standard deviation.

| S 13.48±9.61 11.69±8.75 0.20±0.03 S20 0.51±0.19 0.12±0.07 55.43±9.78 S40 1.21±0.44 0.16±0.04 67.68±8.18 P nd nd nd D 10.08±6.34 5.97±3.46 0.25±0.05 Secondary diet C _i A R FS 2.74±2.07 1.42±1.42 0.21±0.06 FS20 2.43±1.89 0.62±0.55 0.18±0.05 |
|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| S40 1.21±0.44 0.16±0.04 67.68±8.18 P nd nd nd D 10.08±6.34 5.97±3.46 0.25±0.05 Secondary diet C _i A R FS 2.74±2.07 1.42±1.42 0.21±0.06 |
| P nd nd nd D 10.08±6.34 5.97±3.46 0.25±0.05 Secondary diet C _i A R FS 2.74±2.07 1.42±1.42 0.21±0.06 |
| D 10.08±6.34 5.97±3.46 0.25±0.05 Secondary diet C _i A R FS 2.74±2.07 1.42±1.42 0.21±0.06 |
| Secondary diet C _i A R FS 2.74±2.07 1.42±1.42 0.21±0.06 |
| FS 2.74±2.07 1.42±1.42 0.21±0.06 |
| |
| FS20 2.43±1.89 0.62±0.55 0.18±0.05 |
| |
| FS40 2.38±2.26 0.34±0.22 0.24±0.05 |
| FP 11.83±11.14 4.75±4.09 0.19±0.07 |
| FD 21.86±6.01 11.38±2.82 0.21±0.04 |

The overall C balance (7) after feeding on primary diets (Fig. 4) was positive for

animals feeding on S, and D, but there was a marked deficit under conditions where sediment was added (S20 and S40) (F = 158.45 P < 0.0001). The greatest C gain (11.45 \pm 8.76 mgC g⁻¹ d⁻¹) was estimated for *C. verrucosa* feeding on S while the largest C deficit (-81.76 \pm 1.05 mgC g⁻¹ d⁻¹) was for individuals feeding in the highest sediment concentration (S40), although this was not different (P >0.05) from S20 (-66.98 \pm 0.39 mgC g⁻¹ d⁻¹). Secondary diets lead to positive T (Fig.4), although gains from FS20 and FS40 were almost negligible (0.41 \pm 0.58 and 0.05 \pm 0.22, respectively). FD diet generated a C gain of 11.12 \pm 2.79 mgC g⁻¹ d⁻¹ that was significantly higher than the remaining diets (F =20.41, P < 0.0001)

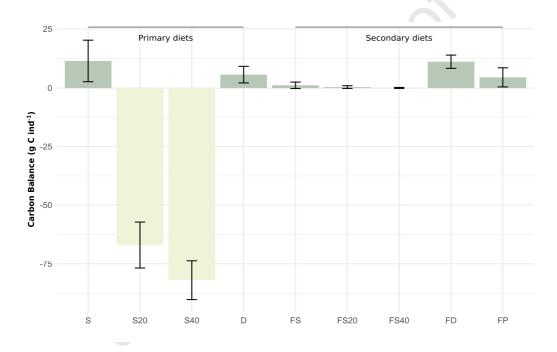


Figure 4. Carbon Balance of *Cnemidocarpa verrucosa* relating absorbed and respired carbon from different diets. Results are expressed as mean and standard deviation.

Biodeposition Rate and ascidian C sink potential. BR for krill and *C. verrucosa* (Tables 4 and 5 respectively), varied with population density at different sites and depths as well as the potential of ascidians to act as C sink (Fig. 5 and 6). The highest BR for krill identified here was in Potter Cove under a S40 diet (1226.4 \pm 92.24 mg.m⁻³.d⁻¹), and this decreased throughout the offshore sites where lower densities of krill were reported (Table 4). Also for *C. verrucosa*, the highest BR (4199.47 \pm 2036.26 mg.m⁻².d⁻¹) was calculated to

occur where there were high densities of ascidians, such as those reported for a new ice-free area in the inner part of Potter Cove, but when feeding on the S diet (Table 5). For the same diet, high BR (2755.9 mg.m⁻².d⁻¹ ± 1136.29) was calculated for Bransfield Strait, where high densities of *C. verrucosa* are also reported (Table 5). For secondary diets, the same trend was observed, but for the FP (12285 mg.m⁻².d⁻¹ ± 5365.17) and FD (18067 mg.m⁻².d⁻¹ ± 5315.36) diets (Table 5). Similarly, the potential of ascidians to act as a C sink varied with population density at different sites and depth in Potter Cove (Figure 5) and in Bransfield Strait (Fig. 6). The highest C sink potential was calculated to be in new ice-free areas in the inner part of Potter Cove (Fig. 5) followed by the Bransfield Strait (Fig. 6).

Table 4. Biodeposition Rate (BR, g.d⁻¹) for reported abundances of *Euphausia superba* in Potter Cove and the Western Antarctic Peninsula. BR is expressed as mean ± standard deviation.

| | POTT | ER COVE | WAP | | | | | |
|-----------------------------------------|-----------|-----------------|------------|-----------|------------|--|--|--|
| - | 2002 | 2003 | Coast | Shelf | Slope | | | |
| Abundance (ind.100 m ⁻³) | 264.1 | 3000 | 18.2 | 10.2 | 2 | | | |
| DIET | 10 | | | | | | | |
| S | 0.37±0.15 | 1044.17±466.06 | 0.03±0.01 | 0.01±0.01 | 0.003±0.00 | | | |
| S20 | 0.71±0.06 | 3396.40±2645.29 | 0.05±0.00 | 0.03±0.00 | 0.01±0.00 | | | |
| S40 | 1.23±0.09 | 4646.51±3639.44 | 0.08±0.01 | 0.05±0.00 | 0.01±0.00 | | | |
| Р | 0.77±0.31 | 2140.75±711.01 | 0.053±0.02 | 0.03±0.01 | 0.01±0.00 | | | |
| D | 0.79±0.24 | 5189.28±6145.21 | 0.05±0.02 | 0.03±0.01 | 0.01±0.00 | | | |

Table 5. Biodeposition Rate (BR, mg.m⁻².d⁻¹) for reported abundances of *Cnemidocarpa verrucosa* at different depths in Potter Cove and Bransfield Strait. BR is expressed as mean ± standard deviation.

| | | | | | | | | РО | TTER | COVI | BRANSFIELD STRAIT | | | | | | | | |
|-----------------------------------|------------------|----------------|---------------|-----|-----|-----|-----|----------------|------|------|-------------------|---------------|-----------------|----------------|-----------------|----------------|----------------|----------------|----------------|
| | | E1 | | | | | E2 | | | | E3 | | | Isla | and | | Statio n I | Statio n II | StationII I |
| Depth (m) | 15 | 20 | 25 | 30 | 15 | 20 | 25 | 30 | 15 | 20 | 25 | 30 | 15 | 20 | 25 | 30 | 204 | 175 | 117 |
| Abundances (ind.m ⁻²) | 0.17 | 3.33 | 7.67 | 0 | 0 | 0 | 0 | 1.67 | 0 | 0 | 6.33 | 7.67 | 160 | 90 | 140 | 70 | 105 | 90 | 60 |
| Primary Diet | S | | | | | | | | | | | | | | | | | | |
| S | 0.004± 0.002 | 0.09± 0.04 | 0.20± 0.1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.04± 0.02 | 0.0 | 0.0 | 0.17± 0.08 | 0.20± 0.10 | 4.20± 2.04 | 2.36± 1.15 | 3.67± 1.78 | 1.84± 0.89 | 2.76± 1.15 | 2.36± 0.76 | 1.57± 0.00 |
| S20 | 0.001± 0.0002 | 0.01± 0.003 | 0.03± 0.01 | 0.0 | 0.0 | 0.0 | 0.0 | 0.01± 0.002 | 0.0 | 0.0 | 0.03± 0.01 | 0.03± 0.01 | 0.64± 0.15 | 0.36± 0.09 | 0.56± 0.13 | 0.28± 0.07 | 0.42± 0.10 | 0.36± 0.09 | 0.24± 0.06 |
| S40 | 0.002± 0.001 | 0.04± 0.01 | 0.09± 0.03 | 0.0 | 0.0 | 0.0 | 0.0 | 0.02± 0.01 | 0.0 | 0.0 | 0.08± 0.03 | 0.09± 0.03 | 1.97± 0.72 | 1.11± 0.40 | 1.72± 0.63 | 0.86± 0.31 | 1.29± 0.47 | 1.11± 0.40 | 0.74± 0.27 |
| Р | 0.002± 0.002 | 0.04± 0.03 | 0.10± 0.07 | 0.0 | 0.0 | 0.0 | 0.0 | 0.02± 0.02 | 0.0 | 0.0 | 0.08± 0.06 | 0.10± 0.07 | 2.02± 1.56 | 1.14± 0.88 | 1.77± 1.36 | 0.89± 0.68 | 1.33± 1.02 | 1.14± 0.88 | 0.76± 0.58 |
| D | 0.001± 0.001 | 0.03± 0.02 | 0.07± 0.05 | 0.0 | 0.0 | 0.0 | 0.0 | 0.02± 0.01 | 0.0 | 0.0 | 0.06± 0.05 | 0.07± 0.05 | 1.52± 1.14 | 0.86± 0.64 | 1.33± 1.00 | 0.67± 0.50 | 1.00± 0.75 | 0.86± 0.64 | 0.57± 0.43 |
| Secondary D | iets | | | | | | | | - | | | | 1 | ' | 1 | 1 | 1 | | 1 |
| FS | 0.002± 0.01 | 0.04± 0.01 | 0.09± 0.02 | 0.0 | 0.0 | 0.0 | 0.0 | 0.02± 0.01 | 0.0 | 0.0 | 0.07± 0.02 | 0.09± 0.02 | 1.82± 0.51 | 1.02± 0.29 | 1.59± 0.45 | 0.80± 0.22 | 1.20± 0.34 | 1.02± 0.29 | 0.68± 0.19 |
| FS20 | 0.004± 0.004 | 0.08± 0.08 | 0.19± 0.18 | 0.0 | 0.0 | 0.0 | 0.0 | 0.04± 0.04 | 0.0 | 0.0 | 0.15± 0.15 | 0.19± 0.18 | 3.90± 3.70 | 2.19± 2.08 | 3.41± 3.24 | 1.70± 1.62 | 2.56± 2.43 | 2.19± 2.08 | 1.46± 1.39 |
| FS40 | 0.002± 0.003 | 0.04± 0.05 | 0.10± 0.12 | 0.0 | 0.0 | 0.0 | 0.0 | 0.02± 0.03 | 0.0 | 0.0 | 0.08± 0.10 | 0.10± 0.12 | 2.14± 2.55 | 1.20± 1.44 | 1.87± 2.23 | 0.94± 1.12 | 1.40± 1.68 | 1.20± 1.44 | 0.80± 0.96 |
| FP | 0.02± 0.01 | 0.39± 0.27 | 0.90± 0.63 | 0.0 | 0.0 | 0.0 | 0.0 | 0.20± 0.14 | 0.0 | 0.0 | 0.74± 0.52 | 0.90± 0.63 | 18.72± 13.16 | 10.53± 7.40 | 16.38± 11.51 | 8.19± 5.76 | 12.29± 8.64 | 10.53± 7.40 | 7.02± 4.93 |
| FD | 0.03± 0.08 | 0.57± 0.17 | 1.32± 0.39 | 0.0 | 0.0 | 0.0 | 0.0 | 0.29± 0.08 | 0.0 | 0.0 | 1.09± 0.32 | 1.32± 0.29 | 27.53± 8.10 | 15.49± 4.56 | 24.09± 7.09 | 12.04± 3.54 | 18.07± 5.32 | 15.49± 4.56 | 10.32± 3.04 |



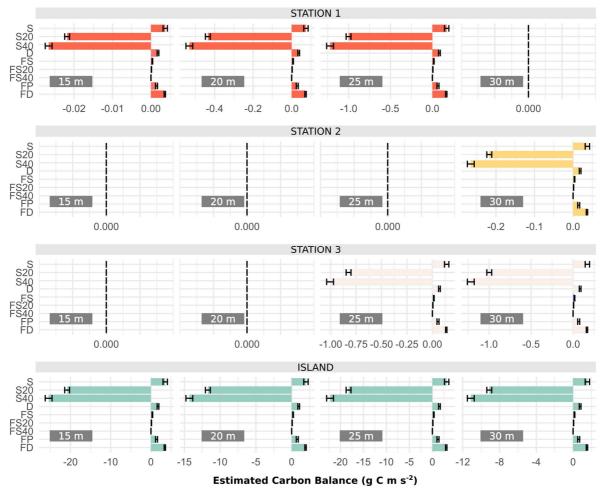


Figure 5. Estimated Carbon Balance of *Cnemidocarpa verrucosa* relating absorbed and respired carbon from different diets. Results based on abundances reported for different stations and depths in Potter Cove, are expressed as gC.m⁻² d⁻¹. Sampling stations: E1, E2, E3, Island. 15, 20, 25, 30 indicate the depth in meters.

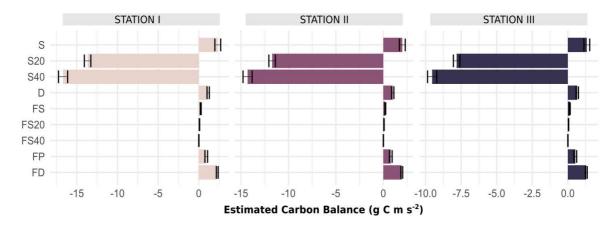


Figure 6. Estimated Carbon Balance of *Cnemidocarpa verrucosa* relating absorbed and respired carbon from different diets. Results based on abundances reported for selected stations (I, II, III) in Bransfield Strait, are expressed as gC.m⁻² d⁻¹. Sampling stations: I (204 m), II (175 m), III (117 m).

DISCUSSION

PBC intensity is directly linked to depth (Buesseler et al. 2007), hence PBC plays a major role in shallow coastal areas, determining the production and biological structure of these ecosystems (Sañe et al. 2012, Smith et al. 2012). Biodeposition, mediated by pelagic and benthic suspension feeders (especially active filter feeders) is the most recognized biological process promoting PBC in shallow waters (Gili and Coma 1998, Turner et al. 2015), since it can result in local downward flux of OM exceeding that of passive physical sedimentation (Ahn 1993, Tatián et al. 2008). Our results show that this process is dependent not only on animal densities, but on its interplay with species-specific AE and faecal production. For instance, a high AE leads to low OM in faeces, but this could be compensated by a high production of faeces due either to high animal specific production or high biodeposition, just because of high animal density.

Both studied species feed on POM that varies with shifts in coastal Antarctic environmental conditions. Because the direction of such change is unknown, we selected the most likely particles based on gut content observations performed in previous studies (for

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

491

492

493

494

495

496

497

498

499

500

501

502

krill: Schmidt et al. 2012, Fuentes et al. 2016; for ascidians: Tatián et al. 2004) and the composition of particulate matter collected in sediment traps (Alurralde 2018). Furthermore, we considered similar potential food sources for E. superba and C. verrucosa, since Potter Cove is a shallow fjord where the water column is generally well mixed (Schloss et al. 2002). Both target species displayed the expected high AE when fed primary diets free of sediment (Clarke et al. 1988, Tatián et al. 2008), in accordance with their omnivore or opportunistic feeding mode. Although krill AE varied among diets, this was not statistically significant, which was also reported in previous studies (Atkinson et al. 2012a). The AE of C. verrucosa did not differ among sediment free primary diets but values were higher than in previous reports (Tatián et al. 2002, 2008). Several studies in filter feeding species suggest that AE is strongly dependent upon food quality (Tito de Morais and Fiala-Médioni 1985, Navarro and Thompson 1996, Gardner and Thompson 2001), and is particularly linked to the ratio of organic to inorganic matter (Gardner 2000, Armsworthy et al. 2001). The variability observed in AE estimations is therefore in line with expectations, since Potter Cove exhibits considerable temporal and spatial variability in seston composition and organic to inorganic ratios of particulate matter (Schloss et al. 1999, 2012, Alurralde 2018). When fed on the macroalgal detritus diet, AE seems to be lower and two ascidian species, C. verrucosa and Pyura setosa, are reported to be able to use up to 40% of the organic content of macroalgal detritus (Tatián et al. 2008). In those experiments, Tatián et al. (2008) used a diet principally based on two Desmarestia species that could have been unpalatable for ascidians (Amsler et al. 2005). Macroalgal detritus can represent an important food source only after entry into the detrital pathway (Norderhaug et al. 2003). However, recent studies show that material derived from species in the order Demarestiales constitute only a minor food source, because they have a high content of phlorotannins (Iken et al. 2007), which prevents microbial degradation (Goecke et al. 2010, Dubois and Iken 2012, Sosik and Simenstad 2013). In the present study, macroalgal detritus was collected directly from the water column by sieving it after resuspension events. Consequently, other macroalgae components may had been present and likely had experienced a longer degradation process than in previous

studies, which improved AE due to an enhanced palatability (Amsler et al. 2012) after mechanical breakdown and trophic upgrading by microbial activity (Quartino et al. 2015). Thus different macroalgal degradation states likely affect AE in ascidians.

Under high suspended sediment conditions, we observed that both target species reduced their AE while FPR and MSFPR responded differently in both species, probably as a result of two different mechanisms for maximizing energy acquisition from low quality food. Reduced AE has been previously reported for krill feeding in sediment-laden coastal waters of the WAP (Clarke et al. 1988, Fuentes et al. 2016). For *C. verrucosa*, it has been suggested that these ascidians could meet their energy demand (given a net year-round AE of 70%) even in the presence of sediments (Tatián et al. 2002). However, our results show that AE drops drastically (more than 50%) when *C. verrucosa* is subjected to high sediment loads. It seems likely that the high efficiencies reported by Tatián et al. (2002) are due to sediment concentrations that were up to eight times lower than those used in our experiments and lower than recorded values in the area for the last decade (Schloss et al. 2012, Monien et al. 2017).

The faecal production rates (FPR and MSFPR) and OM content of krill faeces were inversely related with AE as observed in wild conditions by Atkinson et al. (2012a). We observed low rates of faecal production in sediment-free diets, but these increased progressively with higher amounts of sediment (Fuentes et al. 2016). Probably, when the amount of organic material to be digested is reduced and much diluted with sediment, krill shorten their gut residence time for food which allows more space to process new food by increasing faecal production (Le Vay et al. 2001), and thus favouring enzyme kinetics with enough substrate to act (Jumars et al. 1989). Ingested food is macerate and crushed by the gastric mill, which is located inside the stomach, and mixed with digestive enzymes (Suh and Toda 1992, Suh 1996). It is then pressed through a fine filter system which allows fine food particles (0.2 µm) to enter the midgut, where they are further digested and nutrients are absorbed. Coarse food particles, together with small inorganic, can be transported directly to the hindgut (Ullrich et al. 1991, Fuentes et al. 2016). Most probably, krill can sort and

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

558

accumulate inorganic material inside the stomach (Fuentes et al. 2016) exposing smaller amounts of food to high enzyme activity (Mayzaud et al. 1985, Saborowski and Buchholz 1999, Saborowski 2012) maximizing the absorption of nutrients (Jumars 2000).

On the contrary, digestion in ascidians may be related inversely to the amount of refractory material in the food (Karasov and Douglas 2013), as highly digestible food would require less time to be hydrolysed and absorbed, with consequent higher rates of faecal production (Willows 1992, Jumars 2000). Seston is easily digested by benthic filter feeders, which typically consists of a mixture of phytoplankton, bacteria, protozoa, detritus and some inorganic particles (Hawkins et al. 1996, Gardner 2000). Our faecal production and AE results were slightly higher when C. verrucosa fed on natural seston than on the other sediment-free diets, suggesting that a diet mainly composed of P or D may impose some constraints on the digestive process, possibly due to a high content of refractory components. Some benthic filter feeders have been observed to decrease their AE when fed on certain species of phytoplankton (Ren et al. 2006) or when phytoplankton have a high content of refractory material (Fernández-Reiriz et al. 2015). Despite tunicates possess cellulase enzymes (Davison and Blaxter 2005), the refractory composition of the substrate attacked by digestive enzymes may alter the efficiency of the digestive process. Hence, the low faecal production rates observed for C. verrucosa fed on P, D or diets with high sediment concentrations (S20 and S40), may be the result of keeping food longer in the digestive tract to maximize AE, as has been already observed in other filtering organisms such as the cockle Cerastoderma edule (Navarro et al. 2009, Arambalza et al. 2014).

Studies using isotopic tracers in shallow coastal waters like Potter Cove, indicate that benthic diatoms and microphytobenthos represent important components of suspension feeders diet, including *C. verrucosa* (Pasotti et al. 2015, Ha et al. 2019), since they are available after detachment from benthic substrates and resuspension into the water column (Ahn et al. 1997, 2016). It has been recently stated (Ha et al. 2019) that benthic diatoms are the primary food sources for filter-feeders in shallow coastal Antarctic areas. However, the filter-feeder fauna from Potter Cove seems to rely on a detrital-pool food supply were faecal

559

560

561

562

563

564

565

566

567

568

569

570

571

572

573

574

575

576

577

578

579

580

581

582

583

584

585

586

pellets are included (Pasotti et al. 2015). Krill is thought to play a major role in the biodeposition process by transferring high amounts of OM to the benthos that can then feed on it, because krill faeces are a dominant component of the total organic flux up to at least 170 m depth (Gleiber et al. 2012). In shallow coastal waters (20-180 m depth), krill would be equally relevant as benthic diatoms, once high krill densities were reported for several areas in the WAP (Warren and Demer 2010, Steinberg et al. 2015, Belcher et al. 2016, 2017), and due to the high contribution (66.7–96.3%) of krill faecal pellets to the total particulate organic carbon (POC) flux, even at very shallow depths of 20-50 m (Isla et al. 2001, Belcher et al. 2016, 2017). Our results indicate that krill faeces are likely to be a valuable food source for the benthos, considering that C. verrucosa reached 25-50% AE values that varied with faecal sediment content. Although this is low compared with primary diets, an AE of 25-50% is still a high value compared to several ascidian species (Klumpp 1984, Tito de Morais and Fiala-Medioni 1985, Armsworthy et al. 2001). Although, faecal pellets were not included so far in isotopes studies, we believe that they are an important secondary food source, equally available as benthic diatoms, since both have the same chance to be resuspended into the water column. Moreover, depending on the considered group of benthic diatoms (epilytic, epiphytic, epibionts, microphytobenthos, etc), to some extent faecal pellets would be easier to resuspend than diatoms since they tend to be loosely aggregated in the seafloor, while diatoms are able to attach to the surface where they grow (Al-Handal and Wulff 2008, Ahn et al 2016). Faecal pellets and macroalgal detritus were the major components (mean 91%) in gut contents year-round in C. verrucosa occurring at the inner part of Potter Cove, while diatoms were a low percentage (mean 4.5%) (Tatián et al. 2004). High mean annual krill abundance (from 264 to 3000 ind.100 m⁻³) has been estimated in Potter Cove, from nettowing samples (Fuentes 2006). The presence and relevance of krill in this shallow system is further suggested by the high occurrence in gut contents of *Notothenia coriiceps*, a dominant benthic fish in Potter Cove (Barrera-Oro and Casaux 1990) which feeds primarily on krill (Fanta et al. 2003), and also by frequent massive beaching of krill on the shoreline that has been observed for over a decade (Fuentes et al. 2016). Upscaling our experimental FPR to

the reported abundances by Fuentes (2006), we estimated a high potential input of krill faeces in Potter Cove (Table 4) which matches the high proportion of these particles collected in a four years (2012-2015) sediment traps study (Alurralde 2018). In that study, faecal pellets inputs to the sea floor were high but seasonally variable (1.5 10³ - 7.53 10⁵ mm³.m⁻².d⁻¹). It is interesting to note that the highest biodeposition calculated in the present study was for S40, and this, despite reducing krill AE, would result in a higher amount of OM carried in a larger volume of faeces. Therefore, our estimated FPR and the ability of *C. verrucosa* to feed on, highlight the role of krill in the pelagic-benthic coupling in Potter Cove.

The role of *Cnemidocarpa verrucosa* in the PBC and its potential as a C sink. Estimates of C held in coastal waters along the WAP suggest that, in newly opened areas (ice-free), about 20 gC m⁻² were accumulated through the development of benthic communities fuelled by new primary production (Peck et al. 2010). More recently, Barnes et al. (2018) argued that such estimation would be conservative and the immobilisation in coastal areas would be higher when iceberg calving is considered. In addition, strong links have been reported between suspended food and the abundance of sessile suspension feeders (Jansen et al. 2018a), where there is a strong increase in abundance close to glacier-influenced sites (Jansen et al 2018b).

Our estimates indicate a high C sink potential in new ice-free areas of Potter Cove and also in the Bransfield Strait, in case that ascidians receive a sediment-free, or low sediment content food supply. In such scenarios, our energy balance analyses indicate that *C. verrucosa* ingest more than the minimum C required to cover its energy demands (0.24 mgC L⁻¹ d⁻¹) (Kowalke et al. 2001), perhaps explaining the high abundance of ascidians in Potter Cove (Sahade et al. 1998, Lagger et al. 2017, 2018) and along the WAP (Segelken-Voigt et al. 2016). However, our results also indicate that this potential may be constrained by the effect of suspended sediments washed from the land. It is particularly interesting that high abundance of *C. verrucosa* (160 ind. m⁻²) was observed in a new ice-free area of Potter Cove (Lagger et al. 2017, 2018), close to the highest concentrations recorded of suspended sediments (Schloss et al. 2012, Monien et al. 2017). Under current summer sediment loads

615

616

617

618

619

620

621

622

623

624

625

626

627

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

in Potter Cove, C. verrucosa is able to actively respond to high sediment conditions whether performing squirting to expel inorganic material or closing their siphons while the exposure lasts (Torre et al. 2014). If the exposure persist, the oxygen consumption is triggered (Torre et al. 2012) inducing a high metabolic demand (Torre et al. 2014). This represents a high energy cost for C. verrucosa (Torre et al. 2012) and other ascidian species, exceeding what could be gained from a low-quality diet and a lower AE. According with our energy budget calculations, ascidians should not be successful in Potter Cove, because high sediment loads ultimately lead to a high C deficit, mediated by respiratory C loss that could scale up 200-fold, probably constraining animal growth (Robbins 1985, Petersen et al. 1995). However, these animals would shortly go through negative energy balance, when sediment loads are high in summer. The discharge and injection of sediments into the cove is strongly dependent upon meteorological control of glacial meltwater discharge linked to inter-annual climate variability driven by the Southern Annular Mode and the El Niño Southern Oscillation (Bers et al. 2013, Meredith et al. 2018). Sedimentation in Potter Cove is estimated to last 183 days during which average TSPM concentration is of 15 mg.l⁻¹, but episodic maximum peaks (up to 18 000 mg.l⁻¹) are observed throughout the summer (Monien et al. 2017). On the other hand, the highest densities of *C. verrucosa* are reported on hard rocky substrate with steep slopes, what could also reduce sedimentation effects on suspension feeders, reaching less amounts of sediments per square metre compared with horizontal substrates (Lagger et al. 2018). This would not impairs the OM supply which is lighter than inorganic material and can remain suspended for longer periods of time (Schloss et al. 1999). Hence, topographic and seasonal or interannual control on sediment injection would allow this species to get positive energy balances over time, and therefore be able to cope with high sediment loads. However, if this is not the case, it is possibly that this ascidian population will stand until a threshold limit has pushed the system to another stage where ascidians will not be dominant (Sahade et al. 2015).

Although these calculations are approximate, the estimated C sink potential stands as a first attempt at quantifying the functional role of ascidians in Antarctic waters. Peck (2018)

643

644

645

646

647

648

649

650

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669

670

highlighted that there have been very few energy budgets constructed for Antarctic marine species, and as he warns, some limitations and biases should be pointed out. In our energy budget, we were unable to directly measure filtration and respiration rates, and values were therefore computed from FPR and taken from the literature, respectively. Respiration were constrained to rates calculated for a seston diet (Kowalke et al. 2001) or sediment concentrations similar to those used in this study (Torre et al. 2012), but these rates were referred to body weight excluding the tunic. The tunic organic content certainly represents a significant part of the ascidian biomass (49.54±3.49 % in our study) and while our C sink potential would be underestimated, growth of body parts is much faster than the tunic growth (Petersen et al. 1995). Similarly, mucus production could be a relevant source of bias. It has been only exceptionally considered in ascidians (Goddard and Hogget, 1982), but in some species, such as gastropod molluscs, the losses to mucus can be 30% or more (Peck et al. 1997). Also, interannual variation in animal ecology and physiology might introduce other potential biases in our carbon energy balance (Petersen et al. 1995). In addition, diet (Fernández-Reiriz et al. 2015, Souster et al. 2018), temperature (Klumpp 1984, Jiang et al. 2008) and ascidian size (Jiang et al. 2008, Souster et al. 2018) have all been documented to alter ascidian filtration and respiration rates, biasing the potential C sink (Coppari et al. 2014) estimated here for C. verrucosa. Moreover, this potential is likely to also vary throughout the year (particularly in winter), as many Antarctic marine benthic species have lower winter resting metabolic requirements in response to a range of factors including temperature (Brockington and Clarke 2001, Peck 2018), reduced food supplies (Fraser et al. 2002, Auer et al. 2015) and reproductive investment (e.g. Grange et al. 2007). Although C. verrucosa reproduces during winter (Sahade et al. 2004), the species seems capable of feeding throughout the year even when food availability is low (Tatián et a. 2002, Lesser and Slattery 2015), as has also been observed in some other benthic suspension feeders, both in shallow coastal areas (Barnes and Clarke 1995, Ducklow et al. 2001) and in deeper areas of the Antarctic shelf (Orejas et al. 2000, Gili et al. 2001).

Future perspectives. PBC is usually examined in terms of nutrient exchange and

OM sedimentation (Buesseler et al. 2007). The variability of biological processes, however, is rarely assessed, which precludes an understanding of the whole system. Several biological processes are inherently species-specific and contribute to shaping ecosystem functioning (Barnes and Sands 2017). There are large gaps in our understanding of the response to climate-driven changes in the nearshore WAP, due to the complex interplay of negative and positive feedbacks that act simultaneously. The conceptual model outlined for Potter Cove (Fig. 7) illustrates the complex response of trophic processes involved in the PBC to changes in suspended particle composition envisaged to be some of the major consequences of climate change. Current climate conditions in the WAP are the primary cause of changes in TSPM and environmental settings. Ice retreat allows for both new areas for plankton production and new seabed available for benthic colonisation. Species-specific trophic ecology and the characteristics of the TSPM (composition and OM proportion) will determine the interplay between the intake of energy (high or low AE) and the rate of faecal production. The AE will determine the OM in faeces (the higher the AE, the lower the OM), but the net transfer of OM to the seabed will be modulated by the amount of faeces produced combined with the AE, which in turn will be determined by food processing (FPR and MSFPR) on the seabed and the density of the species involved. At the same time, food availability and the AE will regulate the energy balance of the species, determining energy uptake from the available food. Despite the many possible combinations of responses, when food supply has low sediment loads or consists of krill faeces, the overall energy balance for ascidians is positive. However, the melting of the land-terminating glacier in Potter Cove has increased the amount of sediment in the water column during summer, which has lowered the AE in both species studied, even if FPR and MSFPR are different. Lower AE's would result in a greater transfer of OM to the seabed and to sediments. For benthic animals such as ascidians, the ultimate effect would be to promote a highly negative energy balance that would eventually compromise the last benthic secondary production (somatic growth) and therefore C accumulation, if the exposure to suspended sediments becomes chronic.

671

672

673

674

675

676

677

678

679

680

681

682

683

684

685

686

687

688

689

690

691

692

693

694

695

696

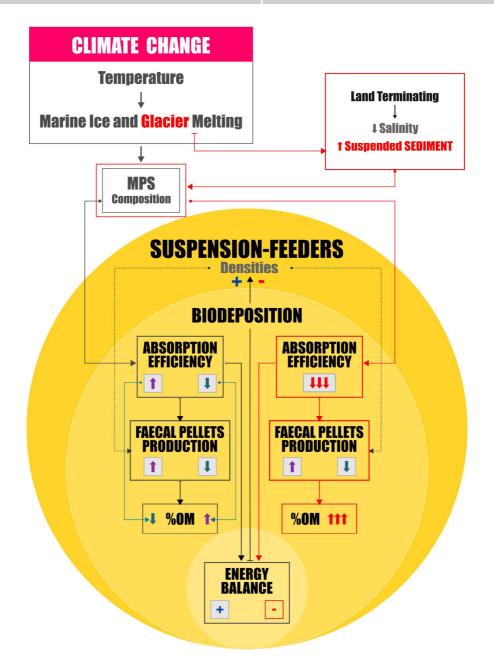


Figure 7. A conceptual model of the trophic processes involved in the PBC and their response to alternative suspended particle composition scenarios driven by climate change. PBC promoted by biodeposition mediated by suspension feeders is regulated by the characteristics of the diet, which determine different digestive processing (absorption efficiency and faeces production) in different species, which will regulate the amount of OM transferred, and finally by the densities of the species considered. A free-sediment food provision (black boxes) can increase (violet arrow) or reduce (green arrow) the AE and the faeces production. A high AE will determine a low content of OM in the faeces (%OM) and

vice versa (blue lines). The net transfer of OM in the faeces will therefore be regulated by the species-specific production and the animal densities (dashed lines). Under high sediment conditions (red boxes), AE is negatively affected in all cases (red arrows) transferring more OM in the faeces, but faecal pellet production differs according to the species. Independently of free-sediment diets, the energy balance would be positive (black line) assuring the survival of suspension feeders and even promoting their increase (positive effect on the ecosystem). Conversely, the increase of suspended sediment leads to a very negative energy balance that would drastically affect the suspension feeder community (negative effect on the ecosystem).

Despite a generalised marine and glacier ice loss, a high potential for benthic carbon accumulation and immobilization has been estimated as a negative feedback to climate change (Peck et al. 2010, Barnes 2015, Barnes and Sands 2017). The results presented here generally support this view and we are confident that they are realistic and accurate for the system under study in summer. They emphasise that the role of suspension feeders in the flow of energy between the water column and the benthos is higher than previously thought. Disentangling the effects of different pressures on the species-mediated energy transfer between pelagic and benthic habitats is challenging, because our understanding of how particular mechanistic processes and pressures may be modulated by environmental drivers is still limited. Recent research suggests that, if warming of the WAP resume from the current hiatus (Turner et al. 2016, Etourneau et al. 2019) and extensive fjordic areas follow the same trend (retreating landward), it is likely that such systems will face similar conditions to those reported here (Meredith et al. 2018). We highlight in this work the importance of an integrated perspective for understanding how coastal ecosystems can respond to climate drivers through their effects on PBC.

Notation

A Absorbed fraction: the product of C_i and AE

| 736 | AE Absorption Efficiency |
|-----|-----------------------------------------------------------------------------------------------|
| 737 | BR Biodeposition Rate |
| 738 | C carbon |
| 739 | C _i total C ingested |
| 740 | E Organic fraction of the faeces |
| 741 | F Organic fraction of the food |
| 742 | FPR Faecal Production Rate |
| 743 | MSFPR Mass-specific Faecal Production Rate |
| 744 | OM Organic Matter |
| 745 | PBC Pelagic-benthic coupling |
| 746 | PIM Particulate Inorganic Matter |
| 747 | POM Particulate Organic Matter |
| 748 | R Respiratory C loss |
| 749 | T Overall C balance |
| 750 | TSPM Total Suspended Particulate Matter |
| 751 | Conflict of Interest: The authors declare that they have no conflict of interest |
| 752 | Funding: This project benefited from the financial support of the Total Foundation |
| 753 | (ECLIPSE Project), Argentinean funds through PICT-Raíces 2011-1320 to IS, PICTO-DNA |
| 754 | Nº 119. It has been additionally supported by the European Commision under the 7th |
| 755 | Framework Programme through the Action – IMCONet (FP7 IRSES, action no. 319718). It is |
| 756 | a contribution to the Coastal Ecology Monitoring programme of Instituto Antártico |
| 757 | Argentino/Dirección Nacional del Antártico in Carlini Station and the research program |
| 758 | PACES II (topic 1, work package 5) of the Alfred Wegener Institute. GA and TM received a |
| 759 | PhD scholarship (CONICET) at the Universidad Nacional de Córdoba. |
| 760 | Acknowledgements: We are grateful to the scientific, logistic and diving groups of |
| 761 | Carlini Station-Dallmann Laboratory for their technical assistance during the Antarctic |
| 762 | expeditions. In addition, to Oscar Gonzales, Alejandro Ullrich and Silvia Rodriguez Instituto |
| 763 | Antártico Argentino (IAA). We especially thank Dr. Lloyd Peck, and two anonymous |

reviewers for their valuable reviews and comments on the manuscript that helped to improve it significantly. Joss Heywood reviewed the English. Logistic and financial support was provided by IAA, Alfred Wegener Institut (AWI), CONICET, FONCyT, SECyT-UNC.

767

768

LITERATURE

- 1. Ahn IY (1993) Enhanced particle flux through the biodeposition by the Antarctic suspension-feeding bivalve *Laternula elliptica* in Marian Cove, King George Island. J
 Exp Mar Biol Ecol, 17:75–90
- Ahn IY, Chung H, Kang JS, Kang SH (1997) Diatom composition and biomass
 variability in nearshore waters of Maxwell Bay, Antarctica, during the 1992/1993
 austral summer. Polar Biol 17:123–130
- 3. Ahn IY, Moon HW, Jeon M, Kang SH (2016). First record of massive blooming of benthic diatoms and their association with megabenthic filter feeders on the shallow seafloor of an Antarctic Fjord: Does glacier melting fuel the bloom? Ocean Science Journal, 51(2), 273–279. https://doi.org/10.1007/s12601-016-0023-y
- 4. Al-Handal AY, Wulff A (2008) Marine benthic diatoms from Potter Cove, King George
 Island, Antarctica. Bot Mar, 51, 51-68
- 5. Alcaraz M, Almeda R, Duarte CM, Horstkotte B, Lasternas S, Agustí S (2014)
 Changes in the C, N, and P cycles by the predicted salps-krill shift in the southern
 ocean. Front Mar Sci, 1–13. https://doi.org/10.3389/fmars.2014.00045
- 6. Alurralde G (2018) Enlace bento-pelágico en un ecosistema costero antártico afectado por el deshielo: una aproximación ecológica. Tesis Doctoral. Facultad de Ciencias Exactas, Físicas y Naturales. Universidad Nacional de Córdoba.
- 787
 Amsler CD, Iken K, Mcclintock JB, Amsler MO, Peters KJ, Hubbard JM, Furrow FB,
 788
 Baker BJ (2005) Comprehensive evaluation of the palatability and chemical defenses
 of subtidal macroalgae from the Antarctic Peninsula. Mar Ecol Progr Ser, 294, 141–
 159
- 791 8. Amsler CD, Mcclintock JB, Baker BJ (2012) Palatability of living and dead detached

| 792 | Antarctic | macroalgae | to | consumers. | Antarctic | Sci, | 24(6), | 589–590. |
|-----|-----------|------------|----|------------|-----------|------|--------|----------|
|-----|-----------|------------|----|------------|-----------|------|--------|----------|

- 793 <u>https://doi.org/10.1017/S0954102012000624</u>
- 9. Arambalza U, Ibarrola I, Navarro E, Urrutia MB (2014) Ingestion and absorption of
- 795 particles derived from different macrophyta in the cockle Cerastoderma edule: Effects
- 796 of food ration. J Comp Phys B, 184(2), 179–192. https://doi.org/10.1007/s00360-013-
- 797 <u>0789-5</u>
- 798 10. Armsworthy SL, MacDonald BA, Ward JE (2001) Feeding activity, absorption
- 799 efficiency and suspension feeding processes in the ascidian, *Halocynthia pyriformis*
- 800 (Stolidobranchia: Ascidiacea): responses to variations in diet quantity and quality. J
- 801 Exp Mar Biol Ecol, 260(1), 41–69
- 11. Atkinson A, Schmidt K, Fielding S, Kawaguchi S, Geissler PA (2012a) Variable food
- absorption by Antarctic krill: Relationships between diet, egestion rate and the
- composition and sinking rates of their faecal pellets. Deep-Sea Res II, 59-60:147-158
- 12. Atkinson A, Siegel V, Pakhomov E, Jessopp MJ, Loeb V (2009) A re-appraisal of the
- total biomass and annual production of Antarctic krill. Deep-Sea Res I, 56: 727–740,
- 807 https://doi.org/10.1016/j.dsr.2008.12.007
- 13. Atkinson A, Ward P, Hunt B, Pakhomov EA, Hosie GW (2012b). An overview of
- 809 Southern Ocean zooplankton data: abundance, biomass, feeding and functional
- relationships. CCAMLR Sci. 19, 171–218
- 14. Auer SK, Salin K, Rudolf AM, Anderson GJ, Metcalfe NB (2015) Flexibility in
- 812 metabolic rate confers a growth advantage under changing food availability. J Anim
- 813 Ecol, 84:1405–1411
- 15. Barnes DKA (2015) Antarctic sea ice losses drive gains in benthic carbon drawdown.
- 815 Curr Biol, 25(18), R789–R790. https://doi.org/10.1016/j.cub.2015.07.042
- 816 16. Barnes DKA (2017) Iceberg killing fields limit huge potential for benthic blue carbon in
- 817 Antarctic shallows. Glob Change Biol, 23, 2649–2659. https://doi:10.1111/gcb.13523
- 17. Barnes DKA, Clarke A (1995) Seasonality of feeding activity in Antarctic suspension
- 819 feeders. Polar Biol 15:335–340. https://doi.org/10.1007/BF00238483

| | Journal 110-proof |
|-----|----------------------------------------------------------------------------------------|
| 820 | 18. Barnes DKA, Fleming A, Sands CJ, Quartino ML, Deregibus D (2018) Icebergs, sea |
| 821 | ice, blue carbon and Antarctic climate feedbacks. Phil Trans R Soc A 376: 20170176. |
| 822 | http://dx.doi.org/10.1098/rsta.2017.0176 |
| 823 | 19. Barnes DKA, Ireland L, Hogg OT, Morley S, Enderlein P, Sands CJ (2016) Why is the |
| 824 | South Orkney Island shelf (the world's first high seas marine protected area) a carbon |
| 825 | immobilization hotspot? Global Change Biol, 22, 1110-1120 |
| 826 | 20. Barnes DKA, Sands CJ (2017) Functional group diversity is key to Southern Ocean |
| 827 | benthic carbon pathways. PLoS ONE, 12(6), 14. |
| 828 | https://doi.org/10.1371/journal.pone.0179735 |
| 829 | 21. Barrera-Oro ER, Casaux RJ (1990) Feeding selectivity in Nothotenia neglecta, |
| 830 | Nybelin, from Potter Cove, South Shetland Islands, Antarctica. Antarct. Sci. 2, 207- |
| 831 | 2013. |
| 832 | 22. Belcher A, Iversen M, Manno C, Henson SA, Tarling GA, Sanders R (2016) The role |
| 833 | of particle associated microbes in remineralization of fecal pellets in the upper |
| 834 | mesopelagic of the Scotia Sea, Antarctica. Limnology and Oceanography, 61(3), |
| 835 | 1049-1064. https://doi.org/10.1002/lno.10269 |
| 836 | 23. Belcher A, Tarling G, Manno C, Atkinson A, Ward P, Skaret G, Fielding S, Henson, |
| 837 | SA, Sanders R (2017) The potential role of Antarctic krill faecal pellets in efficient |
| 838 | carbon export at the marginal ice zone of the South Orkney Islands in spring. Polar |
| 839 | Biol. https://doi.org/10.1007/s00300-017-2118-z |
| 840 | 24. Bers AV, Momo F, Schloss IR, Abele D (2013). Analysis of trends and sudden |
| 841 | changes in long-term environmental data from King George Island (Antarctica): |
| 842 | relationships between global climatic oscillations and local system response. Climatic |
| 843 | Change, 116:789–803. https://doi.org/10.1007/s10584-012-0523-4 |
| 844 | 25. Bowden DA, Clarke A, Peck LS, Barnes DKA (2006) Antarctic sessile marine |
| 845 | benthos: colonisation and growth on artificial substrata over three years. Mar Ecol |

26. Bracegirdle TJ, Connolley WM, Turner J (2008(Antarctic climate change over the

Prog Ser, 316: 1–16. <u>https://doi.org/10.3354/meps316001</u>

846

847

- twenty first century. J Geophys Res Atmos, 113, DO3103.
- 849 <u>https://doi:10.1029/2007JD008933</u>
- 27. Brockington S, Clarke A (2001) The relative influence of temperature and food on the
- metabolism of a marine invertebrate. JEMBE, 258(1):87-99.
- 852 <u>https://doi.org/10.1016/S0022-0981(00)00347-6</u>
- 28. Buesseler KO, Lamborg CH, Boyd PW, Lam PJ, Trull TW, Bidigare RR, Bishop JKB,
- Casciotti KL, Dehairs F, Elskens M, Honda M, Karl DM, Siegel D, Silver M, Steinberg
- D, Valdes J, Van Mooy B, Wilson SE (2007). Revisiting carbon flux through the
- 856 Ocean's twilight zone. Science 316 (5824), 567–570.
- 29. Claquin P, Longphuirt SN, Fouillaron P, Huonnic P, Ragueneau O, Klein C, Leynaert
- A (2010) Effects of simulated benthic fluxes on phytoplankton dynamic and
- photosynthetic parameters in a mesocosm experiment (Bay of Brest, France). Estuar
- 860 Coast Shelf Sci, 86:93-101
- 30. Clarke A, Quetin LB, Ross RM (1988) Laboratory and field estimates of the rate of
- faecal pellet production by Antarctic Krill Euphausia superba. Mar Biol, 98(4), 557-
- 863 564.
- 31. Coello-Camba A, Llabres M, Duarte C, Agusti S (2017). Zooplankton excretion
- 865 metabolites stimulate Southern Ocean phytoplankton growth. Polar Biol.
- 866 https://doi.org/10.1007/s00300-017-2123-2
- 32. Conover RJ (1966) Assimilation of organic matter by zooplankton. Limnol Oceanogr.
- 868 11: 338-354
- 33. Cook A, Vaughan D, Luckman A, Murray T (2014) A new Antarctic Peninsula glacier
- basin inventory and observed area changes since the 1940s. Antarctic Sci, 26(6),
- 871 614-624. https://doi.org/10.1017/S0954102014000200
- 34. Cook A, Holland P, Meredith M, Murray T, Luckman A, Vaughan D (2016) Ocean
- forcing of glacier retreat in the western Antarctic Peninsula. Science, 353(6296).
- 874 https://doi.org/10.1126/science.aag0209
- 875 35. Coppari M, Gori A, Rossi S (2014) Size, spatial, and bathymetrical distribution of the

| 876 | ascidian Halocynthia papillosa in Mediterranean coastal bottoms: benthic-pelagic |
|--------|---------------------------------------------------------------------------------------------------------------------|
| 877 | coupling implications. Mar Biol, 161(9), 2079–2095. https://doi.org/10.1007/s00227- |
| 878 | <u>014-2488-5</u> |
| 879 36 | 6. Di Rienzo JA, Casanoves F, Balzarini MG, Gonzalez L, Tablada M, Robledo CW. |
| 880 | InfoStat versión 2016. InfoStat Group, Facultad de Ciencias Agropecuarias, |
| 881 | Universidad Nacional de Córdoba, Argentina. URL http://www.infostat.com.ar |
| 882 3 | 7. Davison A, Blaxter M (2005) Ancient origin of glycosyl hydrolase family 9 cellulase |
| 883 | genes. Mol Biol Evol, 22:1273–1284 |
| 884 38 | 3. Dierssen HM, Smith RĐ, Vernet M (2002) Glacial meltwater dynamics in coastal |
| 885 | waters west of the Antarctic peninsula. PNAS, 99(4), 1790-1795. |
| 886 39 | 9. Deregibus D, Quartino ML, Campana GL, Momo FR, Wiencke C, Zacher K (2016) |
| 887 | Photosynthetic light requirements and vertical distribution of macroalgae in newly ice- |
| 888 | free areas in Potter Cove, South Shetland Islands, Antarctica. Polar Biol, 39(1), 153- |
| 889 | 166. https://doi.org/10.1007/s00300-015-1679-y |
| 890 40 | D. Dubois A, Iken K (2012) Seasonal variation in kelp phlorotannins in relation to grazer |
| 891 | abundance and environmental variables in the Alaskan sublittoral zone. Algae 27: |
| 892 | 9–19 |
| 893 4 | 1. Ducklow HW, Steinberg DK, Buesseler KO (2001) Upper ocean carbon export and |
| 894 | the biological pump. Oceanography, 14, 50–58. |
| 895 | https://doi.org/10.5670/oceanog.2001.06 |
| 896 42 | 2. Duineveld GCA, Lavaleye MSS, Berghuis EM (2004) Particle flux and food supply to |
| 897 | a seamount cold-water coral community (Galicia Bank, NW Spain). Mar Ecol Prog |
| 898 | Ser, 277: 13-23. https://doi:10.3354/meps277013 |
| 899 43 | 3. Etourneau J, Sgubin G, Crosta X, Swingedouw D, Willmott V, Barbara L, Houssais |
| 900 | MN, Schouten S, Damsté JS, Goosse H, Escutia C, Crespin J, Massé G, Kim J-H |
| 901 | (2019) Ocean temperature impact on ice shelf extent in the eastern Antarctic |

| | Journal Pre-proof |
|-----|-----------------------------------------------------------------------------------------|
| 902 | Peninsula. Nature communications. ;10(1):304. https://doi.org/10.1038/s41467-018 |
| 903 | <u>08195-6</u> |
| 904 | 44. Fanta E, Sant'Anna Rios F, Donatti L, Cardoso WE (2003) Spatial and tempora |
| 905 | variation in krill consumption by the Antarctic fish Notothenia coriiceps, in Admiralty |
| 906 | Bay, King George Island. Antarctic Science 15, 458–462. |
| 907 | 45. Fernández-Reiriz MJ, Irisarri J, Labarta U (2015) Feeding behaviour and differentia |
| 908 | absorption of nutrients in mussel Mytilus galloprovincialis: Responses to three |
| 909 | microalgae diets. Aquaculture, 446, 42-47 |
| 910 | 46. Fraser KPP, Clarke A, Peck LS (2002) Feast and famine in Antarctica: seasona |
| 911 | physiology in the limpet, Nacella concinna (Strebel, 1908). Mar Ecol Prog Ser 242 |
| 912 | 169-177. |
| 913 | 47. Fuentes VL (2006) Estudio de la Comunidad Zooplanctónica de Caleta Potter y |
| 914 | Bahía Guardia Nacional (Isla 25 de Mayo): su Rol en la Dinámica del Carbono er |
| 915 | Zonas Costeras Antárticas. Tesis Doctoral. Universidad de Buenos Aires, p 249 |
| 916 | 48. Fuentes V, Alurralde G, Meyer B, Aguirre GE, Canepa A, Wölfl AC., Hass CH |
| 917 | Williams GN, Schloss IR (2016). Glacial melting: an overlooked threat to Antarctic |
| 918 | krill. Scientific Reports, 27234. https://doi.org/10.1038/srep27234 |
| 919 | 49. Gardner JPA (2000) Where are the mussels on Cook Strait (New Zealand) shores |
| 920 | Low seston quality as a possible factor limiting multi-species distributions. Mar Eco |
| 921 | Prog Ser, 194:123–132 |
| 922 | 50. Gardner, J.P.A., Thompson, R.J., 2001. Naturally low seston concentration and the |
| 923 | net energy balance of the greenshell mussel (Perna canaliculus) at Island Bay, Cool |
| 924 | Strait, New Zealand. New Zealand J Mar Freshw Res 35, 457–468. |
| 925 | 51. Gergs R, Rinke K, Rothhaupt KO (2009) Zebra mussels mediate benthic-pelagic |
| 926 | coupling by biodeposition and changing detrital stoichiometry. Freshw Biol 54(7) |
| 927 | 1379–1391. https://doi.org/10.1111/j.1365-2427.2009.02188.x |

928 52. Gili JM, Coma R (1998) Benthic suspension feeders: their paramount role in littoral 929 marine food webs. Trends Ecol Evol 13, 318-321.

| 930 | 53. Gili JM, Coma R, Orejas C, López-González PJ, Zabala M (2001) Are Antarctic |
|-----|--------------------------------------------------------------------------------------------|
| 931 | suspension-feeding communities different from those elsewhere in the world? Polar |
| 932 | Biol, 24:473-485, https://doi.org/10.1007/s003000100257 |
| 933 | 54. Gleiber MR, Steinberg DK, Ducklow HW (2012). Time series of vertical flux of |
| 934 | zooplankton faecal pellets on the continental shelf of the western Antarctic Peninsula. |
| 935 | Mar Ecol Prog Ser, 471: 23–36. https://doi.org/10.3354/meps10021 |
| 936 | 55. Goddard CK, Hoggett AK (1982) Gut contents of the ascidian <i>Pyura praeputialis</i> : |
| 937 | endogenous and exogenous components. Journal of Zoology 196.4: 489-497. |
| 938 | 56. Goecke F, Labes A, Wiese J, Imhoff JF (2010) Chemical interactions between marine |
| 939 | macroalgae and bacteria. Mar Ecol Prog Ser 409: 267-299 |
| 940 | 57. Grange LJ, Smith CR (2013) Megafaunal communities in rapidly warming fjords along |
| 941 | the West Antarctic Peninsula: hotspots of abundance and beta diversity. PloS One, |
| 942 | 8(12), e77917. https://doi.org/10.1371/journal.pone.0077917 |
| 943 | 58. Grange LJ, Tyler PA, Peck LS (2007). Multi-year observations on the gametogenic |
| 944 | ecology of the Antarctic seastar Odontaster validus Mar Biol, 153(1):15-23. |
| 945 | https://doi.org/10.1007/s00227-007-0776-z |
| 946 | 59. Gutt J, Barnes D, Lockhart SJ, van de Putte A (2013) Antarctic macrobenthic |
| 947 | communities: A compilation of circumpolar information. Nature Conservation, 4, 1-13. |
| 948 | https://doi.org/10.3897/natureconservation.4.4499 |
| 949 | 60. Gutt J, Bertler N, Bracegirdle TJ, Buschmann A, Comiso J, Hosie G, Isla E, Schloss |
| 950 | IR, Smith CR, Tournadre J, Xavier JC (2015). The Southern Ocean ecosystem under |
| 951 | multiple climate change stresses - an integrated circumpolar assessment. Global |
| 952 | Change Biology, 21(4), 1434–1453. https://doi.org/10.1111/gcb.12794 |
| 953 | 61. Ha SY, Ahn IY, Moon HW, Choi B, Shin KH (2019) Tight trophic association between |
| 954 | benthic diatom blooms and shallow-water megabenthic communities in a rapidly |

- 955 deglaciated Antarctic fjord. Estuarine, Coastal and Shelf Science. 218:258-67.
- 956 62. Hawkins AJS, Smith RFM, Bayne BL, Heral M (1996) Novel observations underlying
- the fast growth of suspension-feeding shellfish in turbid environments: *Mytilus edulis*.
- 958 Mar Ecol Progr Ser 131, 179–190
- 959 63. Hawkins AJ, Bayne B, Bougrier S, Héral M, Iglesias JI, Navarro E, Smith, RFM,
- 960 Urrutia M (1998). Some general relationships in comparing the feeding physiology of
- 961 suspension-feeding bivalve molluscs. J Exp Mar Biol Ecol, 219(1), 87–103.
- 962 <u>https://doi.org/10.1016/S0022-0981(97)00176-7</u>
- 963 64. Iglesias JIP, Urrutia MB, Navarro E, Ibarrola I (1998) Measuring feeding and
- absorption in suspension-feeding bivalve molluscs: an appraisal of the biodeposition
- 965 method, J Exp Mar Biol Ecol, 219, 71-86. https://doi.org/10.1016/S0022-
- 966 0981(97)00175-5
- 967 65. Iken K, Amsler CD, Hubbard JM, McClintock JB, Baker BJ (2007) Allocation patterns
- of phlorotannins in Antarctic brown algae. Phycologia 46:386-395.
- 969 66. Isla E, Palanques A, Alvà V, Puig P, Guillén J (2001) Fluxes and composition of
- 970 settling particles during summer in an Antarctic shallow bay of Livingston Island,
- 971 South Shetlands. Polar Biol 24:670-676
- 972 67. Jansen J, Hill NA, Dunstan PK, Mckinlay J, Sumner MD, Post AL, Eléaume MP,
- 973 Armand LK, Warnock JP, Galton-Fenzi BK, Johnson CR (2018a). Abundance and
- 974 richness of key Antarctic seafloor fauna correlates with modelled food availability.
- 975 Nat. Ecol. Evol. 2, 71–80. https://doi.org/10.1038/s41559-017-0392-3
- 976 68. Jansen J, Hill NA, Dunstan PK, Cougnon EA, Galton-fenzi BK, Johnson CR (2018b)
- 977 Mapping Antarctic Suspension Feeder Abundances and Seafloor Food-Availability,
- 978 and Modeling Their Change After a Major Glacier Calving. Front. Ecol. Evol. 6:94, 1-
- 979 11. https://doi.org/10.3389/fevo.2018.00094
- 980 69. Jiang AL, Jian L, Wang CH (2008) Physiological energetics of the ascidian Styela
- 981 clava in relation to body size and temperature. Comp Biochem Phys A 149:129–136
- 982 70. Jumars PA (2000) Animal guts as ideal chemical reactors: maximizing absorption

- 983 rates. The American Naturalist 155(4): 527-543. https://doi.org/10.1086/303333
- 71. Jumars PA, Penry DL, Baross JA, Perry MJ, Frost BW (1989). Closing the microbial
- 985 loop: dissolved carbon pathway to heterotrophic bacteria from incomplete ingestion,
- 986 digestion and absorption in animals. Deep-Sea Res, 36(4), 483–495
- 987 72. Jørgensen CB (1996) Bivalve filter feeding revisited. Mar. Ecol. Prog. Ser. 142:287–
- 988 302.
- 73. Karasov WH, Douglas AE (2013). Comparative Digestive Physiology. Compr Physiol,
- 990 3(2): 741–783. https://doi.org/10.1002/cphy.c110054
- 74. Klöser H, Ferreyra G, Schloss I, Mercuri G, Laturnus F, Curtosi A (1994) Hydrography
- of Potter Cove, a Small Fjord-like Inlet on King George Island (South Shetlands).
- 993 Estuar Coast Shelf Sci. 38:523-537
- 75. Klumpp DW (1984). Nutritional ecology of the ascidian *Pyura stolonifera*: influence of
- body size, food quantity and quality on filter-feeding, respiration, assimilation
- efficiency and energy balance. Mar Ecol Prog Ser, 19, 269–284.
- 997 76. Kowalke J, Tatián M, Sahade R, Arntz W, Kowalke J (2001). Production and
- 998 respiration of Antarctic ascidians. Polar Biol, 24(9), 663–669.
- 999 https://doi.org/10.1007/s003000100266
- 1000 77. Lagger C, Nime M, Torre L, Servetto N, Tatián M, Sahade R (2018). Climate Change,
- Glacier Retreat and a New Ice-Free Island Offer New Insights on Antarctic Benthic
- 1002 Responses. Ecography, (7491), 0–2. https://doi.org/10.1111/ecog.03018
- 1003 78. Lagger C, Servetto N, Torre L, Sahade R (2017b). Benthic colonization in newly ice-
- free soft-bottom areas in an Antarctic fjord.PLoS ONE 12 (11): e0186756.
- 1005 <u>https://doi.org/10.1371/journal.pone.0186756</u>
- 1006 79. Lampert W (1984) The measurement of respiration. A manual on methods for the
- 1007 assessment of secondary production in fresh waters. Blackwell Scientific, Oxford, pp
- 1008 413–468
- 1009 80. Le Vay L, Jones D, Puello-Cruz AC, Sangha RS, Ngamphongsai C (2001) Digestion
- 1010 in relation to feeding strategies exhibited by crustacean larvae. Comp Biochem

| | Journal Pre-proof |
|------|-----------------------------------------------------------------------------------------|
| 1011 | Physiol A: Mol Integr Physiol, 128, 621–628. https://doi.org/10.1016/S1095- |
| 1012 | <u>6433(00)00339-1</u> |
| 1013 | 81. Lesser MP, Slattery M (2015). Picoplankton consumption supports the ascidian |
| 1014 | Cnemidocarpa verrucosa in McMurdo Sound, Antarctica. Mar Ecol Progr Ser, 525, |
| 1015 | 117-126. doi:10.3354/meps11215 |
| 1016 | 82. Leys SP, Wilson K, Holeton C, Reiswig HM, Austin WC, Tunnicliffe V (2004) Patterns |
| 1017 | of glass sponge (Porifera, Hexactinellida) distribution in coastal waters of British |
| 1018 | Columbia, Canada. Mar Ecol Prog Ser, 283: 133-149. |
| 1019 | https://doi.org/10.3354/meps283133 |
| 1020 | 83. Mahibbur Rahman M, Govindarajulu Z (1997) A modification of the test of Shapiro |
| 1021 | and Wilk for normality. Journal of Applied Statistics 24(2): 219-236, |
| 1022 | https://doi.org/10.1080/02664769723828 |
| 1023 | 84. Mayer LM, Schick LL, Selfz RFL, Jumars PA, Findlay RH, Chen Z, Sampson S (1997) |
| 1024 | Digestive environments of benthic macroinvertebrate guts: Enzymes, surfactants and |
| 1025 | dissolved organic matter. J Mar Res, 55, 785-812. |
| 1026 | 85. Mayzaud P, Farber-Lorda J, Corre MC (1985) Aspects of the nutritional metabolism of |
| 1027 | two Antarctic euphausiids: Euphausia superba and Thysanoessa macrura. In: |
| 1028 | Siegried WR, Condy PR, Laws RM (eds) Antarctic nutrient cycles and food webs. |
| 1029 | Springer, Berlin, pp 330–338 |
| 1030 | 86. Meredith MP, King JC (2005) Rapid climate change in the ocean to the west of the |
| 1031 | Antarctic Penisula during the second half of the twentieth century. Geophys. Res. |
| 1032 | Lett. 32, L19604. https://doi.org/10.1029/2005GL024042 |
| 1033 | 87. Meredith MP, Falk U, Valeria A, Mackensen A, Schloss IR, Barlett ER, Jerosch K, |
| 1034 | Silva Busso A, Abele D (2018). Anatomy of a glacial meltwater discharge event in an |
| 1035 | Antarctic cove. Phil.Trans.R.Soc.A 376: 20170163. |
| 1036 | http://dx.doi.org/10.1098/rsta.2017.0163 |
| 1037 | 88. Mermillod-Blondin F (2014). The functional significance of bioturbation and |

biodeposition on biogeochemical processes at the water - sediment interface in

1038

| | Journal Pre-proof |
|------|--------------------------------------------------------------------------------------------|
| 1039 | freshwater and marine ecosystems. J N Amer Benthol Soc, 30(3), 770-778. |
| 1040 | https://doi.org/10.1899/10-121.1 |
| 1041 | 89. Miller R (1981). Simultaneous Statistical Inference (2 ed.). New York: Springer-Verlag |
| 1042 | 90. Monien D, Monien P, Brünjes R, Widmer T, Kappenberg A, Busso AAS, Schnetger B, |
| 1043 | Brumsack H (2017). Meltwater as a source of potentially bioavailable iron to |
| 1044 | Antarctica waters. Antarctic Sci, 15, 1–15. |
| 1045 | https://doi.org/10.1017/S095410201600064X |
| 1046 | 91. Montgomery DC (1997) Design and analysis of experiments. 4th ed. New York: John |
| 1047 | Wiley and Sons. |
| 1048 | 92. Montes-Hugo M, Doney SC, Ducklow HW, Fraser W, Martinson D, Stammerjohn SE, |
| 1049 | Schofield O (2009) Recent changes in phytoplankton communities associated with |
| 1050 | rapid regional climate change along the western Antarctic Peninsula. Science 323, |
| 1051 | 1470–1473 |
| 1052 | 93. Moon HW, Wan Hussin WMR, Kim HC, Ahn IY (2015). The impacts of climate |
| 1053 | change on Antarctic nearshore mega-epifaunal benthic assemblages in a glacial fjord |
| 1054 | on King George Island: Responses and implications. Ecological Indicators, 57, 280- |
| 1055 | 292. https://doi.org/10.1016/j.ecolind.2015.04.031 |
| 1056 | 94. Moreau S, Mostajir B, Bélanger S, Schloss IR, Vancoppenolle M, Demers S, Ferreyra |
| 1057 | GA (2015). Climate change enhances primary production in the western Antarctic |
| 1058 | Peninsula. Glob Chang Biol, 21(6), 2191–2205. http://doi.org/10.1111/gcb.12878 |
| 1059 | 95. Müller-Navarra DC (2008) Food web paradigms: the biochemical view on trophic |
| 1060 | interactions. Int. Rev. Hydrobiol. 93, 489–505. |
| 1061 | http://dx.doi.org/10.1002/iroh.200711046 |
| 1062 | 96. Navarro E, Mendez S, Ibarrola I, Urrutia MB (2009) Comparative utilization of |
| 1063 | phytoplankton and vascular plant detritus by the cockle Cerastoderma edule: |

97. Navarro JM, Thompson RJ (1996) Physiological energetics of the horse mussel

Aquat Biol,

6, 247–262.

digestive responses during diet acclimation.

https://doi.org/10.3354/ab00127

1064

1065

1066

- 1067 *Modiolus modiolus* in a cold ocean environment. Mar. Ecol. Prog. Ser. 138, 135–148
- 1068 98. Norderhaug KM, Fredriksen S, Nygaard K (2003) Trophic importance of Laminaria
- 1069 hyperborea to kelp forest consumers and the importance of bacterial degradation to
- 1070 food quality. Mar Ecol Prog Ser 255:135–144
- 1071 99. Orejas C, JM Gili, Alvà V, Arntz WE (2000) Predatory impact of an epiphytic
- hydrozoan in an upwelling area in the Bay of Coliumo (Dichato, Chile). J Sea Res, 44:
- 1073 209-220,https://doi.org/10.1016/S1385-1101(00)00057-5
- 1074 100. Orejas C, Gili JM, Arntz WE, Ros JD, López PJ, Teixido N, Filipe P (2000).
- 1075 Benthic suspension feeders, key players in Antarctic marine ecosystems?
- 1076 Contributions to Science, Vol.1, No.3, pp. 299–311.
- 1077 101. Pasotti F, Saravia LA, De Troch M, Tarantelli MS, Sahade R, Vanreusel A
- 1078 (2015) Benthic Trophic Interactions in an Antarctic Shallow Water Ecosystem
- 1079 Affected by Recent Glacier Retreat. PLoS ONE 10(11): e0141742.
- 1080 doi:10.1371/journal. pone.0141742
- 1081 102. Pakhomov E, Fuentes V, Schloss IR, Atencio A, Esnal GB (2003) Beaching of
- the tunicate Salpa thompsoni at high levels of suspended particulate matter in the
- Southern Ocean: implications for a "starvation in the midst of plenty" hypothesis.
- 1084 Polar Biol, 26:427-431
- 1085 103. Peck LS (2018). Antarctic marine biodiversity: adaptations, environments and
- responses to change. Oceanography and Marine Biology: An Annual Review, 56,
- 1087 105-236 Eds. SJ Hawkins, AJ Evans, AC Dale, LB Firth, and IP Smith.
- 1088 104. Peck LS, Barnes DK, Cook AJ, Fleming AH, Clarke A (2010). Negative
- feedback in the cold: ice retreat produces new carbon sinks in Antarctica. Glob.
- 1090 Chang. Biol. 16, 2614–2623. https://doi.org/10.1111/j.1365-2486.2009.02071.x
- 1091 105. Petersen JK. Riisgård HU (1992) Filtration capacity of the ascidian Ciona
- intestinalis and its grazing impact in a shallow fjord. Mar Ecol Progr Ser, 88:9-17
- 1093 106. Petersen JK, Schou O, Thor P (1995). Growth and energetics in ascidian
- 1094 Ciona intestinalis. Mar Ecol Progr Ser, 120(1-3), 175-184.

| 1095 | https://doi.org/10.3354/meps120175 |
|------|---------------------------------------------------------------------------------------|
| 1096 | 107. Philipps EER, Husmann G, Abele D (2011) The impact of sediment deposition |
| 1097 | and iceberg scour on the Antarctic soft shell clam Laternula elliptica at King George |
| 1098 | Island, Antarctica. Antarctic Sci 23: 127-138 |
| 1099 | 108. Puigcorbé V, Roca-Martí M, Masqué P, Benitez-Nelson CR, Rutgers vd. Loeff, |
| 1100 | M, Laglera LM, Bracher A, Cheah W, Strass V, Hoppema M, Santos-Echeandía J, |
| 1101 | Hunt BPV, Pakhomov EA, Klaas C (2016) Particulate organic carbon export across |
| 1102 | the Antarctic Circumpolar Current at 10°E: Differences north and south of the |
| 1103 | Antarctic Polar Front. Deep Sea Research Part II: Topical Studies in Oceanography, |
| 1104 | (May), 1–16. https://doi.org/10.1016/j.dsr2.2016.05.016 |
| 1105 | 109. Quartino ML, Vazquez SC, Latorre GEJ, Mac Cormack WP (2015) Possible |
| 1106 | role of bacteria in the degradation of macroalgae Desmarestia anceps Montagne |
| 1107 | (Phaeophyceae) in Antarctic marine waters. Revista Argentina de Microbiologia, |
| 1108 | 47(3), 274–276. https://doi.org/10.1016/j.ram.2015.04.003 |
| 1109 | 110. Quartino ML, Deregibus D, Campana GL, Latorre GEJ, Momo FR (2013). |
| 1110 | Evidence of macroalgal colonization on newly ice-free areas following glacial retreat |
| 1111 | in Potter Cove (South Shetland Islands), Antarctica. PloS One, 8(3), e58223. |
| 1112 | https://doi.org/10.1371/journal.pone.0058223 |
| 1113 | 111. Ragueneau O, Chauvaud L, Moriceau B, Leynaert A, Thouzeau G, Donval A, |
| 1114 | Le Loc'h F, Jean F (2005) Biodeposition by an invasive suspension feeder impacts |
| 1115 | the biogeochemical cycle of Si in a coastal ecosystem (Bay of Brest, France). |
| 1116 | Biogeochemistry 75:19–41 |
| 1117 | 112. Ren JS, Ross AH, Hayden BJ (2006) Comparison of assimilation efficiency on |
| 1118 | diets of nine phytoplankton species of the greenshell mussel Perna canaliculus. J |
| 1119 | Shellf Res, 25(3), 887–892. <u>https://doi.org/10.2983/0730-</u> |
| 1120 | 8000(2006)25[887:COAEOD]2.0.CO;2 |
| 1121 | 113. Robbins IJ (1985) Food passage and defaecation in Ciona intestinalis (L.); the |
| 1122 | effects of suspension quantity and quality. J Exp Mar Biol Ecol, 89:2-3, 247-254 |

| | Journal Pre-proof |
|------|-----------------------------------------------------------------------------------|
| 1123 | Rossi S, Bramanti L, Broglio E, Gili JM (2012) Trophic impact of long-lived |
| 1124 | species indicated by population dynamics in a short-lived hydrozoan, Eudendrium |
| 1125 | racemosum. Mar Ecol Prog Ser, 467:97–111 |
| 1126 | Rossi S, Coppari M, Viladrich N (2015) Benthic-Pelagic Coupling: New |
| 1127 | Perspectives in the Animal Forests. Marine Animal Forests. in: Marine Animal |
| 1128 | Forests: The Ecology of Benthic Biodiversity Hotspots, Ed. 1, Springer. pp. 1-31. |
| 1129 | https://doi.org/10.1007/978-3-319-17001-5_23-1 |
| 1130 | Rückamp M, Braun M, Suckro S, Blindow N (2011) Observed glacial changes |
| 1131 | on the King George Island ice cap, Antarctica, in the last decade. Global Planet |
| 1132 | Change. 79, 99-109 |
| 1133 | Saborowski R (2012) Related antipodes: a comparative study on digestive |
| 1134 | endopeptidases from northern krill and Antarctic krill (Euphausiacea). Polar Biol |

- endopeptidases from northern krill and Antarctic krill (Euphausiacea). Polar Biol 35:1003–1012
- 1136 118. Sabarowski R, Buchholz F (1999) A laboratory study on digestion processes
 1137 in the Antarctic krill, *Euphausia superba*, with special regard to chitinolytic enzymes.
 1138 Polar Biol 21:295–304
- 1139 119. Sahade R, Lagger C, Torre L, Momo F, Monien P, Schloss I,Barnes DKA,

 Servetto N, Tarantelli S, Tatián M, Zamboni N, Abele D (2015) Climate change and

 glacier retreat drive shifts in an Antarctic benthic ecosystem. Science Advances,

 1142 1(10), e1500050
- 120. Sahade R, Tatián M, Esnal G (2004) Reproductive ecology of the ascidian

 Cnemidocarpa verrucosa at Potter Cove, South Shetland Islands, Antarctica. Mar

 Ecol Progr Ser, 272, 131–140. https://doi.org/10.3354/meps272131
- 121. Sahade R, Tatián M, Kowalke J, Kühne S, Esnal GB (1998) Benthic fauna associations of soft substrates at Potter Cove, King George Island, Antarctica. Polar Biol. 19:85-91
- 1149 122. Sañé E, Isla E, Gerdes D, Montiel A, Gili J-M (2012) Benthic macrofauna 1150 assemblages and biochemical properties of sediments in two Antarctic regions

| 1151 | differently | affected | by | climate | change. | Continental | Shelf | Research, | 35, | 53–63 |
|------|-------------|----------|----|---------|---------|-------------|-------|-----------|-----|-------|
|------|-------------|----------|----|---------|---------|-------------|-------|-----------|-----|-------|

- 1152 <u>https://doi.org/10.1016/j.csr.2011.12.008</u>
- 1153 123. Schloss IR, Ferreyra GA (2002) Primary production, light and vertical mixing in
- Potter Cove, a shallow coastal Antarctic environment. Polar Biol. 25:41-48
- 1155 124. Schloss IR, Abele D, Moreau S, Demers S, Bers V, González O, Ferreyra GA
- 1156 (2012) Response of phytoplankton dynamics to 19-year (1991–2009) climate trends
- in Potter Cove (Antarctica). J Mar Syst. 92:53–66
- 1158 125. Schmidt K, Atkinson A, Venables HJ, Pond DW (2012) Early spawning of
- Antarctic krill in the Scotia Sea is fuelled by 'superfluous' feeding on non-ice
- associated phytoplankton blooms. Deep-Sea Res II 59-60:159-172
- 1161 126. Schofield O, Ducklow HW, Martinson DG, Meredith MP, Moline MA, Fraser
- WR (2010) How do polar marine ecosystems respond to rapid climate change?
- 1163 Science 328, 1520–1523
- 1164 127. Segelken-Voigt A, Bracher A, Dorschel B, Gutt J, Huneke W, Link H,
- Piepenburg D (2016) Spatial distribution patterns of ascidians (Ascidiacea: Tunicata)
- on the continental shelves off the northern Antarctic Peninsula. Polar Biol, 39(5),
- 1167 863–879. https://doi.org/10.1007/s00300-016-1909-y
- 1168 128. Siegel V (2016) Biology and Ecology of Antarctic Krill. (V. Siegel, Ed.).
- 1169 Springer Berlin Heidelberg. https://doi.org/10.1007/978-3-319-29279-3
- 1170 129. Smith CR, De Master D, Thomas C, Srsen P, Grange L, Evrard V, DeLeo F
- 1171 (2012). Pelagic-Benthic Coupling, Food Banks, and Climate Change on the West
- 1172 Antarctic Peninsula Shelf. Oceanography, 25(3), 188–201.
- 1173 <u>http://doi.org/10.5670/oceanog.2012.94</u>
- 1174 130. Sosik EA, Simenstad CA (2013). Isotopic evidence and consequences of the
- role of microbes in macroalgae detritus-based food webs. Marine Ecology Progress
- 1176 Series, 494, 107–119. https://doi.org/10.3354/meps10544
- 1177 131. Steinberg DK, Ruck KE, Gleiber MR, Garzio LM, Cope JS, Bernard KS,
- 1178 Stammerjohn SE, Schofield OME, Quetin LB, Ross RM (2015). Long-term (1993–

| 1179 2013) changes in | macrozooplankton off | f the Western A | Antarctic Peninsula. | Deep-Sea |
|-----------------------|----------------------|-----------------|----------------------|----------|
|-----------------------|----------------------|-----------------|----------------------|----------|

- 1180 Res I, 101, 54–70. https://doi.org/10.1016/j.dsr.2015.02.009
- 1181 132. Souster TA, Morley SA, Peck LS (2018). Seasonality of oxygen consumption
- in five common Antarctic benthic marine invertebrates. Polar Biol, 41 (5), 897-908.
- 1183 <u>https://doi.org/10.1007/s00300-018-2251-3</u>
- 1184 133. Suh HL (1996) The gastric mill in euphausiid crustaceans: a comparison of
- eleven species. Hydrobiologia 321:235–244
- 1186 134. Suh HL, Nemoto T (1987) Comparative morphology of filtering structure of five
- species of Euphausia (Euphausiacea, Crustacea) from the Antarctic Ocean. Proc
- 1188 NIPR Symp Polar Biol 1:72–83
- 1189 135. Suh HL, Toda T (1992) Morphology of the gastric mill of the genus *Euphausia*
- 1190 (Crustacea, Euphausiacea). Bull. Plankton Soc. Japan. 39, 17-24.
- 1191 136. Tatián M, Sahade R, Kowalke J, Kivatinitz SC, Esnal GB (2002) Food
- availability and gut contents in the ascidian *Cnemidocarpa verrucosa* at Potter Cove,
- 1193 Antarctica. Polar Biol, 25: 58-64
- 1194 137. Tatián M, Sahade R, Esnal GB (2004) Diet components in the food of
- Antarctic ascidians living at low levels of primary production. Ant Sci, 16(2): 123-128
- 138. Tatián M, Sahade R, Mercuri G, Fuentes VL, Antacli JC, Stellfeldt A, Esnal GB
- 1197 (2008) Feeding ecology of benthic filter-feeders at Potter Cove, an Antarctic coastal
- 1198 ecosystem. Polar Biol, 31: 509-517
- 1199 139. Tito de Morais A, Fiala-Médioni A (1985) Influence of the composition of
- 1200 natural seston upon absorption efficiency of the ascidian *Phallusia mammillata*
- 1201 (Cuvier, 1815). J Exp Mar Biol Ecol, 87:47–54
- 1202 140. Torre L, Servetto N, Eöry ML, Momo F, Tatián M, Abele D, Sahade R (2012)
- 1203 Respiratory responses of three Antarctic ascidians and a sea pen to increased
- sediment concentrations. Polar Biol. 35 (11): 1743-1748
- 1205 141. Torre L, Abele D, Lagger C, Momo F, Sahade R (2014) When shape matters:
- 1206 Strategies of different Antarctic ascidians morphotypes to deal with sedimentation.

| 1207 | Mar Env Res, 99, 179–187. <u>https://doi.org/10.1016/j.marenvres.2014.05.014</u> |
|------|---------------------------------------------------------------------------------------------------|
| 1208 | Turner JT (2015). Zooplankton faecal pellets, marine snow, phytodetritus and |
| 1209 | the ocean's biological pump. Progr Ocean, 130:205-248. |
| 1210 | https://doi.org/10.1016/j.pocean.2014.08.005 |
| 1211 | Turner J, Lu H, White I, King JC, Phillips T, Hosking JS, Bracegirdle TJ, |
| 1212 | Marshall GJ, Mulvaney R, Deb P (2016) Absence of 21st century warming on |
| 1213 | Antarctic Peninsula consistent with natural variability. Nature, 535(7612):411-415. |
| 1214 | https://doi.org/10.1038/nature18645 |
| 1215 | 144. Ullrich B, Storch V, Marschall HP (1991) Microscopic anatomy, functional |
| 1216 | morphology, and ultrastructure of the stomach of Euphausia superba Dana |
| 1217 | (Crustacea, Euphausiacea). Polar Biol 11:203–211 |
| 1218 | 145. Warren JD, Demer DA (2010). Abundance and distribution of Antarctic krill |
| 1219 | (Euphausia superba) nearshore of Cape Shirreff, Livingston Island, Antarctica, during |
| 1220 | six austral summers between 2000 and 2007. Can J Fish Aquat Sci, 67, 1159-1170 |
| 1221 | https://doi.org/10.1139/F10-042 |
| 1222 | 146. Wiencke C, Ferreyra GA, Abele D, Marenssi S (2008) The Antarctic |
| 1223 | ecosystem of Potter Cove, King-George Island (Isla 25 de Mayo). Synopsis of |
| 1224 | research performed 1999-2006 at the Dallmann Laboratory and Jubany Station |
| 1225 | Berichte zur Polar- und Meeresforschung 571 |
| 1226 | 147. Willows RI (1992) Optimal digestive investment: A model for filter feeders |
| 1227 | experiencing variable diets, Limnol Oceanogr, 37, |
| 1228 | https://doi.org/10.4319/lo.1992.37.4.0829 |
| 1229 | 148. Wotton RS, Malmqvist B (2001) Feces in Aquatic Ecosystems. BioScience, |
| 1230 | 51(7):537-544. doi: http://dx.doi.org/10.1641/0006- |
| 1231 | 3568(2001)051[0537:FIAE]2.0.CO;2 |
| 1232 | |
| 1233 | |

HIGHLIGHTS

- Food processing and energy acquisition was studied on two Antarctic filter-feeder species subjected to different food sources and sediment concentrations.
- A general energy model was developed to assess the energy balance of an ascidian and to evaluate its potential as a carbon sink in coastal Antarctic ecosystems.
- Food characteristics and animal densities modulate the impact of ascidians on pelagic-benthic coupling and on carbon immobilisation.
- Increasing inputs of sediment runoff would be a major constraint in coastal areas.