Accepted Manuscript

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PII:S1049-9644(13)00121-7DOI:http://dx.doi.org/10.1016/j.biocontrol.2013.06.002Reference:YBCON 2944To appear in:Biological ControlReceived Date:25 March 2013Accepted Date:10 June 2013



Please cite this article as: Vignale, M.V., Astiz-Gassó, M.M., Novas, M.V., Iannone, L.J., *Epichloid* endophytes confer resistance to the smut *Ustilago bullata* in the wild grass *Bromus auleticus* (Trin.), *Biological Control* (2013), doi: http://dx.doi.org/10.1016/j.biocontrol.2013.06.002

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Epichloid endophytes confer resistance to the smut *Ustilago bullata* in the wild grass *Bromus auleticus* (Trin.)

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15 Abstract

16 In this work it was studied for the first time whether asexual *Epichloë* (*Neotyphodium*)

17 endophytes of *Bromus auleticus*, protect their host plants against the pathogenic fungus

18 Ustilago bullata.

19 Seeds of two different ecotypes of *B. auleticus*, one of them infected with the endophyte

20 *Neotyphodium pampeanum* (NpE+) and the other infected with the endophyte *N*.

21 *tembladerae* (NtE+) and their respectively endophyte-free (NpE-/NtE-) counterparts were

22 used. Seeds of each ecotype and endophytic status were superficially disinfected and were

23 randomly assigned to different treatments named: S+ (smut fungus inoculated) and S-

24 (mock-inoculated). It was evaluated the effect of Ustilago bullata infection on plant

characteristics in every stage of their life cycle: seedling emergence, vegetative growth,

26 mortality and smut symptoms in the florets.

27 In NtE+ infected plants, smut disease was almost completely suppressed, whereas in their

- 28 endophyte-free counterparts (NpE-) the incidence of smut symptoms reached 64%. In
- 29 NpE+ infected plants smut incidence was significantly lower (7%) than in endophyte-free
- 30 plants (39%). Although U. bullata infection decreased the emergence rate of both
- 31 endophyte-infected and endophyte-free plants, neutral or protective effects of the

- 32 endophytes were observed in seedling development and survival. The survival during the
- 33 first year of NtE+ plants was higher than in their NtE- counterparts.
- These results indicate a strong beneficial effect of vertically transmitted endophytes againstthis pathogen.
- 36

37 Keywords

- 38 Defensive mutualism; Grass-endophytes; *Neotyphodium*; Smut disease; *Ustilago bullata*
- 39

40 **1. Introduction**

Some cool-season grasses (subfamily Pooideae) establish symbiotic associations with 41 endophytic fungi of the genus *Epichloë* Tul. and their asexual derivatives *Neotyphodium* 42 Glenn, Hanlin & Bacon (Clavicipictaceae, Hypocreales, Ascomycota). This association is 43 quite specific and so each endophytic species is able to colonize one or a few host species. 44 These fungi colonize the plant shoot meristems where they grow systemically in the 45 apoplast of developing leafs and culms obtaining nutrients (Kuldau and Bacon, 2008). 46 Since its growth is synchronized with the growth of the host plant and does not require the 47 degradation of cell walls of the host, no noticeable symptoms of endophytic infection are 48 produced (Christensen et al., 2008; Christensen and Voisey, 2007). Epichloë species 49 produce stromata with perithecia in the culms of reproductive tillers avoiding the 50 development of the flowers, causing total or partial sterility of the host plant (choke 51 *disease*). Ascospores produced in the perithecia are forcibly discharged and are responsible 52 for the infection of new plants. Some Epichloë species and most of Neotyphodium species 53 do not produce stromata. In these asexual species, hyphae colonize meristems of the 54 developing flowers and remain visible, in the mature seeds, between the aleurone cell layer 55 and the seed coat (Schardl et al., 2004; White, 1993). Thus, these endophytes are vertically 56 57 transmitted through the seeds of the host plant. 58 The associations between grasses and epichloid endophytes, mainly those established with

- 59 vertically transmitted endophytes, are considered in general as mutualists (Clay and
- 60 Schardl, 2002; Müller and Krauss, 2005; Schardl et al., 2004). The plant provides
- 61 photosynthates and shelter to the endophytes and they provide several benefits to the host

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plant. Among these benefits, the most important are protection against herbivores, mediated 62 by the production of different fungal alkaloids including loline and peramine, mainly toxic 63 64 to insects, and lolitrems and ergot alkaloids that affect primarily cattle (Bacon, 1977; Clay and Schardl, 2002; Lane et al., 2000; Latch, 1993; Panaccione et al., 2006; Popay et al., 65 2009; Schardl et al., 2007; 2004; Schardl and Phillips, 1997; Torres et al., 2008). Increased 66 growth and drought resistance have also been attributed to these endophytes in agronomic 67 68 and native wild grasses (Clay, 1987; Iannone and Cabral, 2006; Novas et al., 2003). However, the endophyte may be detrimental under some environmental conditions and in 69 some host species (Cheplick and Faeth, 2009; Faeth et al., 2004). 70 Endophytes seem to protect their host against some fungal pathogens (Bonos et al., 2005; 71 Clarke et al., 2006; Gwinn and Gavin, 1992; Nan and Li, 2000; Yue et al., 2000) and also 72 73 to modulate positively or negatively the interaction between their hosts and arbuscular 74 mycorrhizal fungi (AM) (Chu-Chou et al., 1992; Guo et al., 1992; Liu et al., 2011; Mack and Rudgers, 2008; Müller, 2003; Novas et al., 2005; 2009; Omacini et al., 2006). 75 76 Smut fungi (Ustilaginales, Basidiomycota) are common pathogens of cereals and are studied because of their impact on agriculture worldwide (Agrios, 2005; Wilcoxson et al., 77 1996). These pathogens cause diseases and losses in crops (Martínez-Espinoza et al., 2002; 78 Wilcoxson et al., 1996) and also infect wild grasses, such as Festuca and Lolium (Durán 79 80 and Fischer, 1961; Vánky, 1994). Although several smut fungi species may present differences in their life cycles, all of them 81 cause sterility in their hosts. The ovary of the infected plants is replaced by the pathogen 82 that produces masses of spores, known as teliospores, in the sori within host tissues 83 (Martínez-Espinoza et al., 2002). Teliospores are resting spores that are spread by wind and 84 remain in the soil or attached to the lemma and palea or to the cariopses coat (Agrios, 85 2005). Dikariotic teliospores that undergo karyogamy, germinate along with the seed 86 forming a germ tube (promycelia) (Alexopoulos et al., 1996; Meyer et al., 2001). The 87 diploid nucleus migrates to the promycelium and undergoes meiosis forming four haploid 88 89 basidiospores. Basidiospores can either unite as compatible mating types producing the infection hypha, or they can proliferate mitotically to produce sporidia. Sporidia of 90 compatible mating types may then fuse to penetrate the host as a dikaryotic hypha (Agrios, 91 2005). 92

Ustilago bullata, the causal organism of head smut of grasses, is a highly polymorphic and
systemic smut fungus that infects its host soon after the emergence of the coleoptile from
the seed (Falloon, 1979; Fischer, 1940). The presence of the fungus in their host becomes
apparent at anthesis when the glumes and ovary of infected hosts are destroyed, being
replaced by a dark black mass of teliospores (Falloon and Hume, 1988). The effects of *U*. *bullata* on *Bromus* spp, invasive species in USA, or forage species have been extensively
studied by Falloon (1976; 1979); Falloon and Hume (1988); García-Guzmán et al. (1996);

100 Hirschhorn (1986); Meyer et al. (2001).

101 Bromus auleticus Trin., is a native perennial grass that inhabits grasslands of Argentina,

102 Uruguay and southern Brazil. In Argentina, B. auleticus is infected by two species of

103 endophytes with a frequency of infection higher than 95% in most of the studied

- populations (Iannone et al., 2009). This grass has been reported as host of the smut U.
- 105 *bullata* Berk (Astiz Gassó and Molina, 2010; Traverso, 2001). Field surveys carried out in

106 Argentina indicate that, infection of *B. auleticus* by *U. bullata* has not been very commonly

107 observed in nature, but in field assays, studying endophyte-free plants, smut symptoms

108 produced by *U. bullata* are usually observed (De Battista, personal communication).

109 In grasses infected simultaneously by vertically transmitted epichloid endophytes and smut

110 fungi, both fungi compete for the colonization of the ovary and require, in a different way,

111 the flower for their reproduction and dissemination. If the endophyte is able to avoid the

replacement of the ovary by the smut fungus, leading to the development of a normal seed,

both the host plant and the endophyte will be able to reproduce and disperse.

114 The triple interaction host plant-epichloid endophyte-smut fungus represents an interesting

model to study the effect of endophyte on pathogenic fungi that remains to be explored.

116 Thus, the aim of this study was to establish whether vertically transmitted endophyte

117 species confer resistance to the smut fungus in the pathosystem *Ustilago bullata-Bromus*

118 auleticus Trin.

119

120 **2. Materials and methods**

121 2.1. Plant and smut fungus material

122 Endophyte infected (E_{+}) and endophyte-free (E_{-}) seeds of two different ecotypes of B. auleticus, originally from Intendente Alvear, La Pampa province (LP ecotype), Argentina, 123 infected with Neotyphodium pampeanum Iannone & Cabral and from El Palmar (EP 124 ecotype), Entre Ríos province, Argentina associated with Neotyphodium tembladerae 125 Cabral & White (Iannone et al., 2009) were used. Endophyte-free seeds of each ecotype 126 were obtained in 2007 by loss of endophyte viability in long term stored seeds. Since 2007, 127 128 E+ and E- plants of each ecotype were grown in the field and seeds are collected every year. Seeds used for all the experiments described below were collected during December 129 from the previous year to each experiment described below. 130 Teliospores of U. bullata were collected from infected Bromus catharticus plants in 131 132 December 2008 and 2009. Diseased florets exhibiting fully ripen sori were collected and mildly ground in a mortar and a pestle to release the teliospores. The powder containing 133 134 teliospores and pieces of vegetal tissues was sieved in a 1mm sieve to remove plant tissues. Teliospores were kept dry at 4 °C and were used during the first 12 months after the 135 136 collection. For the taxonomic identification of Ustilago bullata, ITS region was amplified by PCR accordingly to White et al. 1990. PCR product was purified and sequenced in an 137 ABI 3730xl DNA Analyzer. Identification of the smut fungus was performed by means of 138 BLAST on the GenBank database and followed by phylogenetic analyses using Maximum 139 140 Parsimony (Winclada v0.9.9) (Nixon, 1999) and MrBayes algorithms (Mr. Bayes 3.2) (Ronquist et al., 2012) (not shown). 141

142

143 2.2. Endophyte detection

144 The endophytic status of the seed lots and plants was established by the examination of the endophyte in seeds previous to each experiment and in the seedlings or plants at the end of 145 each experiment. To confirm the presence of the endophyte in seeds, caryopses were 146 147 soaked for 5 h in a 10 % v/v aqueous solution of sodium hydroxide at room temperature 148 (22–24°C), and then rinsed and stained with aniline blue (0.1% aqueous) (Clark et al., 1983). Endophytic mycelia were visualized in parenchymal tissues within the culm pith or 149 150 in the parenchyma of peeled sheaths, aniline blue stained as mentioned and observed under a light microscope. Plants were considered as endophyte infected if a mass of dark blue 151

- 152 hyphae was observed between the aleurone cell layer and the seed coat or when
- 153 characteristic unbranched hyphae were observed in parenchymal tissues.
- 154

155 *2.3. Treatments*

156 For all the experiments discussed below, seeds of each ecotype and endophytic status (N. pampeanum-infected (NpE+); N. pampeanum-free (NpE-); N. tembladerae-infected (NtE+) 157 158 and N. tembladerae-free (NtE-)) were superficially disinfected by consecutive washes as follows: ethanol 50%, 1 minute; sodium hypochlorite 2%, 5 minutes and ethanol 50%, 1 159 minute. E+ and E- seeds of each ecotype were randomly assigned to the different 160 treatments named: S+ (smut fungus inoculated) and S- (mock-inoculated). To achieve this, 161 162 seeds assigned to S+ treatments were placed in Petri dishes and a powder of teliospores $(0.15 \text{ mg teliospores.seed}^{-1})$ was poured on them. For control treatments, a mock-163 inoculation with heat inactivated teliospores (180°C for 4 hours) was done. The Petri dishes 164 were closed and gently shaken for 5 minutes to obtain a homogeneous spore distribution on 165 the seeds. In this way, 4 treatments were established for each ecotype named as follows: 166 NpE+S+; NpE+S-; NpE-S+; NpE-S-; NtE+S+; NtE+S-; NtE-S+ and NtE-S-. Before the 167 inoculation, the viability of the teliospores was evaluated by preparing a suspension of 168 teliospores in water $(1.5 \times 10^8 \text{ spores.ml}^{-1})$. Fifty µl of the solution were spread in Petri 169 dishes with water agar 2% and incubated 6 hours in darkness at 24°C. Spores able to 170 germinate (producing a germinating tube) were considered as viable and the percentage of 171 germination was registered. Teliospores viability ranged between 30 and 60%, and in those 172 inactivated for the S- treatments the percentage of germination was zero (even when the 173 inactivated teliospores were re-checked after 72 hours of incubation). 174

175

176 2.4. Effect of Ustilago bullata on seedling emergence and plant development

In order to determine the effect of infection by *U. bullata* on *B. auleticus* seedling
emergence and development, 150 seeds of each treatment and ecotype were sown in ten
Petri dishes (ten replicates with fifteen seeds/dish in each treatment) filled with sterilized
sand and incubated in a growing chamber at 22 °C under 12 hours photoperiod. The
percentage of seed germination was recorded and shoot length was measured after 15 days

6

182 from the sowing. Results were analyzed by a two way ANOVA (p<0.05) for each ecotype

183 where the inoculation with the smut fungus and the endophytic status were the main

184 factors. All data analyses were performed using the Infostat software (Di Rienzo et al.,

 185
 2011).

186

187 2.5. Evaluation of plant survival and smut symptoms development

188 One hundred and fifty seeds of each ecotype and endophytic status were inoculated with 189 teliospores as described above. Seeds of each treatment were germinated in trays filled with sterilized sand in a growth chamber at 22 °C under 12 hours photoperiod. Two-month-old 190 191 seedlings were transplanted individually to 25 cm deep x 15 cm in diameter pots, filled with commercial garden soil: sand: perlite 3:1:1 and transferred outdoors to the experimental 192 193 field of the Facultad de Ciencias Exactas y Naturales, University of Buenos Aires where they were allowed to grow and produce flowers. During this period, the survival of the two-194 195 month-old seedlings (before being transplanted to pots in the field), plant survival before flowering and the incidence of the disease in NpE+/NtE+ or NpE-/NtE- plants were 196 197 evaluated. The incidence of the disease in each treatment was evaluated as the number of plants with symptoms (flowers with sori/number of flowered plants). For each ecotype, the 198 differences among treatments in seedling and plant survival and disease incidence were 199 compared by means of a *Chi-square* test of homogeneity of proportions and the Marascuilo 200 201 procedure was used to make comparisons between all pairs of groups (Marascuilo and 202 McSweeney, 1977).

203

204 2.6. Vertical transmission of the endophyte via seeds

In those plants that produced seeds, the transmission of the endophyte was evaluated bychecking the presence of endophyte in the seeds, as previously described.

207

208 **3. Results**

209 *3.1. Seedling emergence*

- 210 The inoculation with teliospores of *U. bullata* in seeds decreased the overall percentage of
- emergence of *B. auleticus* (in LP ecotype $F_{1;36}$ =63.46 *P*<0.0001 and in EP $F_{1;36}$ =5.28
- 212 P=0.0275). The presence of the endophyte did not affect seedling emergence (LP:

213 $F_{1;36}=0.40 P=0.5293$; EP: $F_{1;36}=2.98 P=0.0930$) (Fig.1).

- In LP ecotype the seedling emergence in NpE+S+ treatment was 49% lower than in
- 215 NpE+S- treatment, whereas in NpE-S+ treatment was 35% lower than in NpE-S- treatment,
- but the difference in the emergence between NpE+S+ and NpE-S- treatment was not
- statistically significant (Fig. 1A). In EP ecotype seedling emergence decreased 16% in
- 218 NtE+ seeds while in NtE- seeds the germination was 23% lower than in the control (Fig.

219

220

221 *3.2. Seedling growth*

1B).

No significant differences were observed in the shoot length between E+ and E- plants of 222 each ecotype (LP: F_{1:36}=1.31, P=0.2592; EP: F_{1:36}=2.43, P=0.1278) (Fig. 2). However, in 223 both ecotypes, seedlings were negatively affected by the presence of the smut fungus (LP: 224 $F_{1:36}=138.14$, P<0.0001; EP: $F_{1:36}=39.46$, P<0.0001). Ustilago bullata effects were more 225 evident in LP ecotype where NpE+S+ and NpE-S+ plants were 46% and 43% smaller 226 227 respectively than their S- counterparts (Fig. 2A). In EP ecotype, NtE-S+ seedlings were 228 39% smaller than the NtE-S- ones, whereas NtE+S+ seedlings were only 29% smaller than their NtE+S- counterparts (Fig. 2B), but this difference was not statistically significant. 229

230

231 3.3. Plant survival

- 232 The inoculation of seeds with teliospores of U. *bullata* decreased the seedlings survival of
- both ecotypes, during the first 60 days of growth (LP: $\chi^2_{0.95;3}$ =32.02; *P*<0.0001and EP:
- 234 $\chi^2_{0.95;3}$ =61.04; *P*<0.0001) (Fig. 3A and B). However, while in LP ecotype no differences
- 235 were observed due to the endophytic status among the smut inoculated plants, in NtE+S+
- seedlings was significantly higher than in the NtE- ones.
- Among the plants grown to evaluate the development of the disease at the flowering time,
- the percentage of survival during the first year of growth in the field was significantly

higher in plants grown from S- seeds (Fig. 3C and D) (LP: $\chi^2_{0.95;3}$ =74.67; P<0.0001 and 239 EP: $\chi^2_{0.95;3}$ =78.32; *P*<0.0001). 240

In LP ecotype, even though only the 15% of the NpE-S+ plants survived, this value was not 241

significantly different from the 31% of survival presented by the NpE+S+ ones (Fig. 3C). 242

On the other hand in EP ecotype the 65.8 % of survival presented by the NtE+S+ plants 243

was significantly higher than that observed in the NtE-S+, where only the 3% of the plants 244 245 survived (Fig. 3D). 5CK

246

3.4. Development of smut symptoms in field 247

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The presence of smut disease symptoms in the florets was evaluated in one or two year old 249 plants grown in pots at field conditions. Disease incidence was almost totally suppressed or 250 significantly diminished in E+ plants of both ecotypes (LP: $\chi^2_{0.95;1}$ =12.67; P=0.0004 and 251 EP: $\chi^2_{0.95;l}$ =78.21; P<0.0001) (Fig. 4). None of the control plants (mock-inoculated) 252 presented symptoms of disease (not shown in figure 4). In the plants that presented smut 253 254 symptoms all the flowers were destroyed by the pathogen.

255

3.5. Vertical transmission of the endophyte 256

None of the NpE+ or NpE- smut-symptomless plants (from LP ecotype) produce fully ripen 257 seeds. In EP ecotype 11 plants produced fully ripen seeds, but only two to five seeds were 258 produced by each plant. The analysis of the presence of the endophytes in the seeds showed 259 that all the seeds were endophyte infected; indicating that in EP ecotype, the inoculation 260 261 with Ustilago bullata did not affected the transmission of the endophyte to the seeds.

262

4. Discussion 263

264 The present work, to our knowledge, is the first report of protective effect of *Neotyphodium* 265 endophytes against a systemic pathogen like U. bullata that produces castration of the

plants. Our findings suggest that plants of *Bromus auleticus* associated with *Neotyphodium tembladerae* or *N. pampeanum* were more resistant to the "head smut" of grasses produced
by *Ustilago bullata* than endophyte-free plants.

In this work we found that, whereas in endophyte-free plants the incidence of the disease 269 reached 39 to 64%, in endophyte-infected plants disease incidence ranged from 1 to 7%. In 270 those plants that presented smut symptoms seed production was totally suppressed 271 272 producing sterility in the affected plants. Thus, our results show that the endophytes prevent castration of the host plant, ensuring sexual reproduction of the host. Although the amount 273 of fully ripen seeds produced by control or symptomless plants, in the S+ treatment, and 274 checked for endophyte infection, was not enough to evaluate accurately the efficiency of 275 276 the transmission of the endophyte through the seeds; our results also showed that the vertical transmission of the endophyte is not affected by the inoculation of the smut fungus. 277 278 In vitro assays, performed in our laboratory, showed that teliospore germination is inhibited by N. pampeanum and N. tembladerae (Iannone et al., 2012b). Protective effects of 279 280 epichloid endophytes against plant fungal pathogens such as Laetisaria fuciformis (Bonos et al., 2005), Alternaria alternata, Fusarium (Nan and Li, 2000), Cercospora, 281 Cryphonectria parasitica (in vitro) (Yue et al., 2000), Sclerotinia homeocarpa (Clarke et 282 al., 2006), Rhizoctonia zeae (Gwinn and Gavin, 1992) have been also reported. All 283 284 together, these results are in agreement with the hypothesis of the defensive mutualism suggested for the grass-endophyte associations (Clay, 1988; 1989; Saikkonen et al., 2010). 285 In spite of the beneficial effects observed in E+ plants with respect to prevention of smut 286 disease development, the endophytes had neutral effects on seedling emergence and 287 growth, since these variables where similarly (negatively) affected by the presence of the 288 289 smut fungus both in the E+ as in the E- treatments. Considering that U. bullata requires flower production for its dissemination, negative effects on plant survival and development 290 291 should not be expected. However, these kind of effects produced by this pathogen on its host plants were also reported in Bromus catharticus (Falloon, 1976; García-Guzmán et al., 292 293 1996). In addition, we consider that the amount of teliospores used in each experiment was 294 significantly higher than that expected to be found in nature since after the inoculation the seeds remained totally covered by a black coat of spores. Thus, detrimental effects of the 295

smut fungus could have been enhanced and some of the protective effects of the endophyte could have been masked in our experiments. Protective effects of the endophyte could be even more important in natural conditions where the charge (inoculum) of teliospores is expected to be lower.

300 Different behaviors were observed between plants of different ecotypes, whereas smut

development was almost totally suppressed in NtE+ plants (EP ecotype; *N. tembladerae* infected), in NpE+ plants (LP ecotype; *N. pampeanum*-infected) the incidence of the

disease was diminished but not so drastically as in NtE+ plants. In the presence of the

pathogen, survival of NtE+ plants was higher than in the NtE- ones, but no differences were

305 observed between NpE+ and NpE- plants. These differences observed in plant survival and

306 disease incidence between the E+ plants of each ecotype seem to indicate that the

307 protective effects of *N. tembladerae* against this pathogen are stronger than those conferred

308 by *N. pampeanum*. However, we cannot discard that the observed differences could be due

to differences in the susceptibility of each plant ecotype. Supporting our hypothesis of a

310 higher protective capacity of *N. tembladerae*, there are *in vitro* studies that showed that *N*.

311 *tembladerae* presented the highest inhibitory capacity against several fungal plant

312 pathogens with respect to other *Epichloë/Neotyphodium* species (Yue et al., 2000) and

against *U. bullata* (Iannone et al., 2012b).

The protective effects shown in the E+ plants against the head smut of grasses disease could be due to, 1) the endophytes preventing the infection by *U. bullata* at seedling stage or 2) the endophytes preventing the colonization of the ovary by the pathogen. The

317 detrimental effects of smut fungus inoculation observed on seedling emergence, seedling

survival and development in E+ and E- treatments would support hypothesis 2, indicating

that the smut fungus is able to infect the seedlings of *B. auleticus* irrespectively of their

320 endophytic status.

321 The higher survival and resistance of endophyte-infected plants to *U. bullata*, in addition to 322 other beneficial properties observed in endophyte infected plants (Iannone et al., 2012a)

323 could explain the higher incidence of endophytes in populations (smut-symptomless) of this

host in nature. Endophyte infected plants produced more seeds than E- ones (Iannone et al.,

325 2012a) and seed production was suppressed in E- plants when infected with U. bullata.

However, considering that *B. auleticus* is a highly perennial plant, more long term studies

11

- are necessary in order to evaluate the importance of *Ustilago bullata* and the effects of both
 symbionts on the dynamics of the populations of this host.
- 329 Although more research should be done in order to establish the mechanism through which
- both fungal symbionts interact in the host plant so that the incidence of the disease is lower
- in E+ plants; our findings are relevant for a better understanding of the biology of the grass-
- endophyte symbiosis and could be also applied in grass breeding programs. Currently,
- studies are being conducted in our laboratory in order to evaluate the effect of the
- endophyte-smut fungus interactions in plant competition, and seed production in field.
- Finally, O'Hanlon et al. (2012) stated that more attention should be paid to dissecting the
- potential of fungal endophytes as biological control agents against cereal pathogens. In this
- sense, our studies and results on smut resistance should be expanded to other endophyte-
- 338 infected grasses, mainly wild barley species as *Hordeum bogdanii*, *H. brevisubulatum* and
- *H.comosum.*
- 340

341 Acknowledgments

- 342 This research was supported by the University of Buenos Aires (grant UBACyT
- 20020090300118), CONICET (grant PIP 1482) and ANPCyT (grant PICT-PAE N°58;
- 344 PICT 2011-1527; PICT 2008-670). PROPLAME-PRHIDEB-CONICET: Publication n°
- 195. We thank Dr. Miguel A. Galvagno for critical review and comments.
- 346

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497	Fig. 1. Effect of Ustilago bullata and Neotyphodium pampeanum (Np) (A) or N. tembladerae (Nt)
498	(B) endophytic status on seedling emergence of Bromus auleticus. Endophyte infected
499	(NpE+/NtE+) and endophyte free (NpE-/NtE-) seeds, inoculated (S+) or mock-inoculated (S-) with
500	<i>Ustilago bullata</i> . Different letters indicate significant differences (<i>P</i> <0.05).
501	
502	Fig. 2. Effect of Ustilago bullata and Neotyphodium pampeanum (Np) (A) or N. tembladerae (Nt)
503	(B) on <i>Bromus auleticus</i> seedlings shoot length (cm). Endophyte infected (NpE+/NtE+) and
504	endophyte free (NpE-/NtE-) seeds, inoculated (S+) or mock-inoculated (S-) with Ustilago bullata.
505	Data are means; SE. Different letters indicate significant differences ($P < 0.05$).
506	
507	Fig. 3. Effect of Ustilago bullata and Neotyphodium pampeanum (Np) (A, C) or N. tembladerae
508	(Nt) (B, D) on <i>Bromus auleticus</i> seedlings survival during the first two months of growth (A, B)
509	and plants survival during the first year of growth under field conditions (C, D). Endophyte infected
510	(NpE+/NtE+) and endophyte free (NpE-/NtE-) seeds, inoculated (S+) or mock-inoculated (S-) with
511	Ustilago bullata. Different letters indicate significant differences (P<0.05).
512	
513	Fig. 4. Percentage of Bromus auleticus plants inoculated with Ustilago bullata with smut symptoms
514	in the florets. NpE+: Neotyphodium pampeanum infected; NpE-: N. pampeanum free; NtE+: N.
515	tembladerae infected and NtE-: N. tembladerae free. Different letters indicate significant
516	differences within each ecotype ($P < 0.05$). The plants in those treatments inoculated with inactive
517	teliospores (S-) did not present smut disease symptoms (not shown in the figure).
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Figure1









*Graphical Abstract (for review)



- 525 Highlights
- 526 Some grasses are usually co-infected by smut fungi and mutualist epichloid endophytes.
- 527 Endophytes are transmitted via seeds and smut fungi replace the seeds with teliospores.
- The endophyte and the smut fungus compete in a race for the colonization of the ovary. 528
- 529 The effect of *Neotyphodium* spp. against head smut of grasses was evaluated.

- Disease incidence was diminished in endophyte-infected Bromus auleticus plants. 530
- 531