

# Morphological and Chemical Analysis of the Molecularly-Imprinted Polymer-Based D-Arabinose Sensor on a Modified Electrode with Functionalized Carbon Nanotubes <sup>†</sup>

Miguel S. P. Sousa <sup>1</sup>, Cristián A. Ferretti <sup>2</sup>, María N. Kneeteman <sup>2</sup> and Leonardo L. Paim <sup>1,\*</sup>

<sup>1</sup> Engineering of Energy, Experimental Campus of Rosana, UNESP—São Paulo State University, 1881 dos Barrageiros, Av., São Paulo 19274-000, Brazil; miguel.sales@unesp.br

<sup>2</sup> Institute of Applied Chemistry of the Littoral (IQAL), Santa Fe, UNL-CONICET, Santiago del Estero 2654, Santa Fe S3000, Argentina; cferretti2371@gmail.com (C.A.F.); mkneeteman@fiq.unl.edu.ar (M.N.K.)

\* Correspondence: leonardo.paim@unesp.br; Tel.: +55-18-3284-9200

† Presented at the 23rd International Electronic Conference on Synthetic Organic Chemistry, 15 November–15 December 2019; Available online: <https://ecsoc-23.sciforum.net/>.

Published: 14 November 2019

**Abstract:** The sensors modified carbon nanotubes and molecularly imprinted polymers are an excellent choice for the determination of D-arabinose, because they have high selectivity and sensitivity for analysis. This allows the control and optimization of the second generation ethanol production process, given the clear understanding of the chemical changes that occur through the hydrolysis of lignocellulosic biomass. For this reason, the present work presents a chemical study of the functional groups of the chemical species present on the sensor surface by the Fourier transform infrared spectroscopy (FT-IR) technique and a morphological analysis by the scanning electron microscopy (SEM) technique.

**Keywords:** D-arabinose; electrochemical sensor; ethanol

## 1. Introduction

In order to promote the sustainability and independence of fossil fuels, first generation ethanol is an attractive option as it results in a net free emission of CO<sub>2</sub> due to the fixation of atmospheric CO<sub>2</sub> by plants during their growth [1]. The United States through corn and Brazil through sugarcane are the main producers of first generation ethanol. However, although the technology used for converting these raw materials is well established, they compete with the food chain, leading to the need to find other alternative sources of energy [2,3].

The second and third generation of ethanol are obtained from lignocellulosic biomass and algae, respectively [4]. Brazil has enormous potential to obtain second generation ethanol, given the high availability of this raw material, and even if the development of second generation (2G) production is not very advanced compared to the first one, it must be compensated. by scientific research that validates its use in energy vectors [4,5].

The production of 2G ethanol is mainly characterized by the preprocessing, hydrolysis and fermentation stages [6]. The hydrolysis process, which consists of the degradation of polysaccharides in fermentable sugars, plays an extremely important role for the final yield in 2G ethanol. Thus, the optimization of this process can lead to increased ethanol production [7].

In this sense, a clear understanding of the chemical changes that occur during lignocellulosic biomass hydrolysis is essential for process control and optimization. Through the use of analytical

techniques, process efficiency can be verified by compositional analysis of hydrolysates. The electrochemical technique is characterized by the acquisition of information in real time and in situ, through sensors (electrodes) that promote a fast and sensitive detection of the analyte, with low cost [8]. This makes them a great alternative to other analytical techniques, characterized by slow response and high cost. Electrochemical sensors can be chemically modified to improve their performance. These modifications can be made by introducing thin electroactive films and thick coatings, which would essentially improve greater selectivity and sensitivity of the analyte due to increased surface area [9].

Among the materials used for surface modification, carbon nanotubes stand out for their highly sensitive electrical properties, mechanical strength, chemical stability and volume-surface ratio, where their functionalization helps to improve specificity and interaction with the solution [10,11]. The surface modification can be performed by electrodeposition, whose process is controllable, simple, economical and fast, producing complex nanoscale structures that can be used as active sensor elements for the detection of various materials [12].

The use of nanomaterials in the development of molecularly imprinted polymer (MIP) based sensors is a current trend [13]. A synthesized polymer with predetermined selectivity for some analytes or a group of structurally related species has: high selectivity for the template molecule, chemical stability, low detection limit, easy preparation, low cost and possibility of reuse [14,15].

Electropolymerization of MIPs directly on the sensor surface provides good control of polymer film thickness. A well-established monomer is *o*-phenylenediamine (*o*-PD), where interactions with the template molecule are preserved in their spatial arrangement and subsequently stabilized by polymer crosslinking [13,16].

In this research work, the structural and morphological surface characterization of the electrochemical sensors was performed with the FT-IR and SEM techniques. The electrochemical sensors developed are graphite and paraffin compounds, modified with functionalized carbon nanotubes (FMWCNTs) and based on molecularly printed polymers (MIPs), for the determination of D-arabinose. These sensors are an excellent low cost alternative, featuring fast analyzes with high selectivity and good sensitivity, which is extremely important for the control and optimization of second generation ethanol production.

## 2. Materials and Methods

Potassium chloride (KCl, purity:  $\geq 99.0\%$ ), potassium ferrocyanide ( $K_3[Fe(CN)_6]$ , purity:  $\geq 99.5\%$ ) and histological paraffin (56–58 °C), were purchased from Synth. Acetic acid ( $CH_3COOH$ , purity: 97.3%), D-arabinose ( $C_5H_{10}O_5$ , purity:  $\geq 98.0\%$ ), *o*-Phenylenediamine ( $C_6H_8N_2$ , flaked, 99.5%), nitric acid ( $HNO_3$ , purity:  $\geq 98.0\%$ ), dimethylformamide ( $C_3H_7NO$ , purity:  $\geq 98.9\%$ ) and graphite ( $< 20\ \mu m$ , synthetic), were purchased from Sigma-Aldrich. Sodium acetate ( $C_2H_3NaO_2$ , purity:  $\geq 99.0\%$ ) was purchased from Dinamica Quimica Contemporanea Ltda (Diadema, Brazil). The oxidation probe solution was  $10.0 \times 10^{-3}\ mol\ L^{-1}\ K_3[Fe(CN)_6]$  in  $1.0\ mol\ L^{-1}\ KCl$ . Stock solution of FMWCNTs was prepared in  $0.55\ mol\ L^{-1}$  of  $HNO_3$ . Stock solution of  $3.0 \times 10^{-4}\ mol\ L^{-1}\ C_5H_{10}O_5$  and  $7.0 \times 10^{-3}\ mol\ L^{-1}\ C_6H_8N_2$ , were prepared in acetate buffer solution under pH 5.1 for electropolymerization.

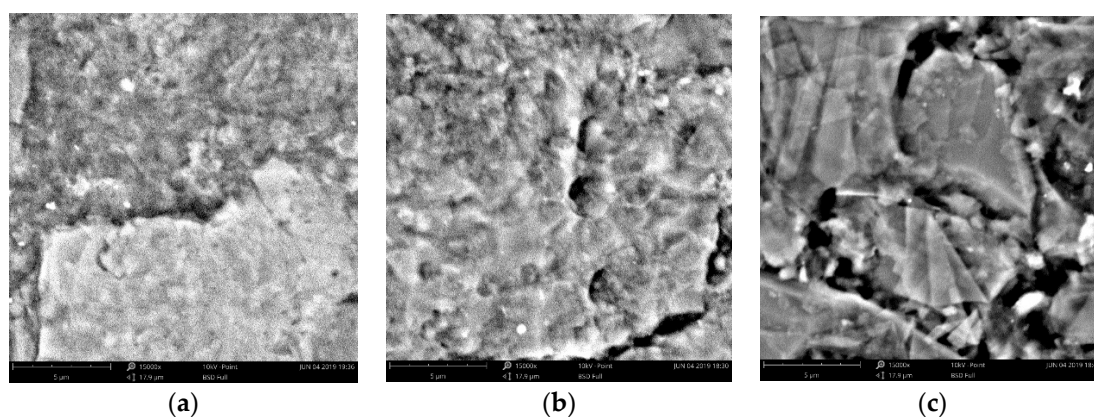
The electrochemical measurements were performed using a Potentiostat/Galvanostat 600E (CH Instruments, Inc.). A conventional three electrodes system was employed with a modified working electrode, a platinum electrode as counter electrode and Ag/AgCl as the referenced electrode. All experiments were carried out at room temperature. Fourier transform infrared spectroscopy (FT-IR) was performed using the IRPrestige-21 (Shimadzu, Japan) with the KBr sample pellet method, using approximately 1% of the sample relative to the mass of KBr per pellet. The analyzes were performed in the range of 400 to 4000  $cm^{-1}$  with a resolution of 4  $cm^{-1}$  and with 40 replicates per analysis. A scanning electron microscope (PHENOM WORLD Pro X, Thermo Fisher Scientific, Waltham, MA, USA) was used to obtain high resolution images on chemically modified sensor surfaces.

### 3. Results and Discussion

Electrochemical sensors composed of graphite/paraffin in proportion 7:3 (v/v) are initially modified by electrodeposition of the FMWCNTs in the range of  $-0.5$  V to  $1.0$  V for 15 consecutive cycles, with a sweep speed of  $50$  mVs $^{-1}$  [17–20]. And then they are modified by electropolymerization of the MIPs in the range of  $-0.4$  V to  $1.0$  V for 20 consecutive cycles at a sweep rate of  $50$  mVs $^{-1}$  in acetate buffer at pH 5.1 containing  $7.0 \times 10^{-3}$  mol L $^{-1}$  o-PD and  $3.0 \times 10^{-4}$  mol L $^{-1}$  D-arabinose (template molecule) [21].

In order to study the morphological specifications of the sensors, the scanning electron microscopy (SEM) technique is used. Figure 1a shows the surface of the graphite and paraffin composite electrode without any modification, while Figure 1b shows a change in its surface due to the electrodeposition of the FMWCNTs. Nanotubes have a highly tangled structure caused by  $\pi$ - $\pi$  stacking interactions, which increases the surface area of the electrode and allows the electroactive probes to diffuse easily [22,23].

In addition, functionalized carbon nanotubes cause an increase in effective binding sites, facilitating electron transfer and assigning better catalytic and conductive properties so that the sensor has a higher voltage [24]. Figure 1c shows the sensor morphology after electropolymerization of the MIPs. In this process, a complex is formed between the template molecule and the o-PD functional monomer. Thus, matrix-adapted binding sites are established, ensuring that the polymer has an affinity condition for the analytical object of interest [24].



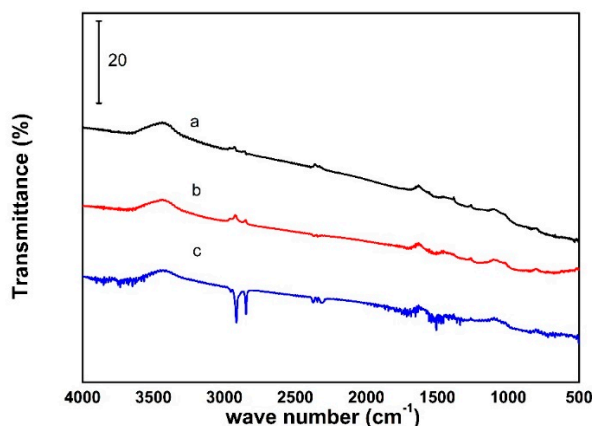
**Figure 1.** Scanning electron microscopy images of the graphite and paraffin composite electrode (a), after modification of its surface by electrodeposition of functionalized carbon nanotubes (FMWCNTs) (b), after further modification of its surface by electropolymerization of molecularly imprinted polymers (MIPs) (c).

The determination of the functional groups present on the electrode surface was performed by the Fourier Transform Infrared Spectroscopy (FT-IR) technique. In Figure 2, spectrum (a) represents graphite/paraffin substrate bands, where at the peak of  $3430$  cm $^{-1}$  an elongation of  $-OH$  associated with the absorbed water is identified. In the region corresponding to the frequencies of  $2965$ – $2850$  cm $^{-1}$  are the characteristic vibrations of the methyl and methylene group, which correspond to paraffin. In the region of  $1600$ – $1100$  cm $^{-1}$ , the flexion of  $OH$  and the elongations  $C=O$ ,  $C-OH$  and  $C-O$  are identified [25]. At peak  $2360$  cm $^{-1}$ , the  $CO_2$  absorbed by this material is observed [25,26].

In spectrum (b) a broad peak around  $3440$  cm $^{-1}$  is identified, attributed to the elongation of the vibrations of methyl and methylene groups produced in carbon nanotubes when functionalized with acid; in the region of the peaks  $1630$  cm $^{-1}$ ,  $1400$  cm $^{-1}$  and  $1260$  cm $^{-1}$  is identified  $C=O$ ,  $O-H$  and  $C-O$  of the stretching vibrations  $-COOH$  [27,28]. The peaks of  $1550$  cm $^{-1}$ ,  $1100$  cm $^{-1}$ ,  $800$  cm $^{-1}$  were attributed to the vibrations  $C=C$ ,  $C-O$  and  $C-H$ , respectively [28].

In spectrum (c) MIPs bands are shown, peaks in the region of  $1630$ – $1614$  cm $^{-1}$  were attributed to elongation  $C=N$ , the peaks of  $1483$  cm $^{-1}$  and  $1466$  cm $^{-1}$  are attributed to the double  $C=C$  bonds of atoms present in the aromatic ring of the phenylenediamine molecule, the  $1261$  cm $^{-1}$  peak at  $C-N-C$

elongation in benzene units and the  $714\text{ cm}^{-1}$  and  $524\text{ cm}^{-1}$  peaks at the C-H plane vibrations in the phenylenediamine ring [29].



**Figure 2.** FT-IR spectra of graphite/paraffin (a), functionalized carbon nanotubes (FMWCNTs) (b), molecularly imprinted polymers (MIPs) (c).

#### 4. Conclusions

The morphological and chemical characterization of the molecularly imprinted polymer-based D-arabinose sensor on a modified electrode with functionalized carbon nanotubes was performed using the FT-IR and SEM techniques. Where, morphological change and the presence of new functional groups prove the success of the sensor surface modification, which allows high selectivity and good sensitivity when performing analyzes.

**Funding:** This research was funded by the São Paulo Research Foundation (FAPESP) (Proc.No.2017/09492-4; 2018/25830-0; 2017/09123-9).

**Conflicts of Interest:** The authors declare no conflict of interest.

#### References

1. Neamhom, T.; Polprasert, C.; Englande, A.J., Jr. Ways that sugarcane industry can help reduce carbon emissions in Thailand. *J. Clean. Prod.* **2016**, *131*, 561–571.
2. Bechara, R.; Gomez, A.; Saint-Antonin, V.; Schweitzer, J.-M.; Maréchal, F.; Ensinas, A. Review of design works for the conversion of sugarcane to first and second-generation ethanol and electricity. *Renew. Sustain. Energy Rev.* **2018**, *91*, 152–164.
3. Germeç, M.; Turhan, I. Ethanol production from acid-pretreated and detoxified tea processing waste and its modeling. *Fuel* **2018**, *231*, 101–109.
4. Aditiya, H.; Mahlia, T.; Chong, W.; Nur, H.; Sebayang, A. Second generation bioethanol production: A critical review. *Renew. Sustain. Energy Rev.* **2016**, *66*, 631–653.
5. Dechambre, D.; Thien, J.; Bardow, A. When 2nd generation biofuel meets water—The water solubility and phase stability issue. *Fuel* **2017**, *209*, 615–623.
6. Parsons, S.; McManus, M.C.; Taylor, C.M. Second-Generation Ethanol from Lignocellulose. *Greenh. Gases Balanc. Bioenergy Syst.* **2018**, *2018*, 193–206.
7. Laca, A.; Laca, A.; Díaz, M. Hydrolysis: From cellulose and hemicellulose to simple sugars. In *Second and Third Generation of Feedstocks*; Basile, A., Dalena, F., Eds.; Elsevier: Oviedo, Spain, 2019; pp. 213–240.
8. Dumitrescu, E.; Andreescu, S. Bioapplications of electrochemical sensors and biosensors. *Meth. Enzymol.* **2017**, *589*, 301–350.
9. Sajid, M.; Nazal, M.K.; Mansha, M.; Alsharaa, A.; Jillani, S.M.S.; Basheer, C. Chemically modified electrodes for electrochemical detection of dopamine in the presence of uric acid and ascorbic acid: A review. *TrAC Trends Anal. Chem.* **2016**, *76*, 15–29.
10. Baig, N.; Sajid, M.; Saleh, T.A. Recent trends in nanomaterial-modified electrodes for electroanalytical applications. *TrAC Trends Anal. Chem.* **2019**, *111*, 47–61.

11. Kwon, Y.J.; Gil Na, H.; Kang, S.Y.; Choi, S.-W.; Kim, S.S.; Kim, H.W. Selective detection of low concentration toluene gas using Pt-decorated carbon nanotubes sensors. *Sens. Actuators B Chem.* **2016**, *227*, 157–168.
12. Morrin, A. Sensors: Chemically modified electrodes. In *Reference Module in Chemistry, Molecular Sciences and Chemical Engineering*; Worsfold, P., Townshend, A., Colin, P., Eds.; Elsevier: Dublin, Ireland, 2018; pp. 161–171.
13. Ahmad, O.S.; Bedwell, T.S.; Esen, C.; Garcia-Cruz, A.; Piletsky, S.A. Molecularly Imprinted Polymers in Electrochemical and Optical Sensors. *Trends Biotechnol.* **2019**, *37*, 294–309.
14. Gui, R.; Jin, H.; Guo, H.; Wang, Z. Recent advances and future prospects in molecularly imprinted polymers-based electrochemical biosensors. *Biosens. Bioelectron.* **2018**, *100*, 56–70.
15. Wei, X.; Xu, X.; Qi, W.; Wu, Y.; Wang, L. Molecularly imprinted polymer/graphene oxide modified glassy carbon electrode for selective detection of sulfanilamide. *Prog. Nat. Sci.* **2017**, *27*, 374–379.
16. Shumyantseva, V.V.; Bulko, T.V.; Sigolaeva, L.V.; Kuzikov, A.V.; Archakov, A.I. Electrosynthesis and binding properties of molecularly imprinted poly-o-phenylenediamine for selective recognition and direct electrochemical detection of myoglobin. *Biosens. Bioelectron.* **2016**, *86*, 330–336.
17. Alizadeh, T.; Ganjali, M.R.; Norouzi, P.; Zare, M.; Zeraatkar, A. A novel high selective and sensitive para-nitrophenol voltammetric sensor, based on a molecularly imprinted polymer–carbon paste electrode. *Talanta* **2009**, *79*, 1197–1203.
18. Du, D.; Chen, S.; Cai, J.; Tao, Y.; Tu, H.; Zhang, A. Recognition of dimethoate carried by bi-layer electrodeposition of silver nanoparticles and imprinted poly-o-phenylenediamine. *Electrochim. Acta* **2008**, *53*, 6589–6595.
19. Chen, L.; Tang, Y.; Wang, K.; Liu, C.; Luo, S. Direct electrodeposition of reduced graphene oxide on glassy carbon electrode and its electrochemical application. *Electrochem. Commun.* **2011**, *13*, 133–137.
20. Guzman, M.; Dille, J.; Godet, S. Synthesis of silver nanoparticles by chemical reduction method and their antibacterial activity. *Int. J. Mater. Metall. Eng.* **2008**, *2*, 91–98.
21. Wang, Q.; Paim, L.; Zhang, X.; Wang, S.; Stradiotto, N.R. An electrochemical sensor for reducing sugars based on a glassy carbon electrode modified with electropolymerized molecularly imprinted poly-o-phenylenediamine film. *Electroanalysis* **2014**, *26*, 1612–1622.
22. Rajalakshmi, K.; John, S.A. Highly sensitive determination of nitrite using FMWCNTs-conducting polymer composite modified electrode. *Sens. Actuators B Chem.* **2015**, *215*, 119–124.
23. Gomes, R.N.; Sousa, C.P.; Casciano, P.N.; Ribeiro, F.W.P.; Morais, S.; De Lima-Neto, P.; Correia, A.N. Dispersion of multi-walled carbon nanotubes in [BMIM]PF<sub>6</sub> for electrochemical sensing of acetaminophen. *Mater. Sci. Eng. C* **2018**, *88*, 148–156.
24. Beluomini, M.A.; Da Silva, J.L.; De Sá, A.C.; Buffon, E.; Pereira, T.C.; Stradiotto, N.R. Electrochemical sensors based on molecularly imprinted polymer on nanostructured carbon materials: A review. *J. Electroanal. Chem.* **2019**, *840*, 343–366.
25. Anirudhan, T.; Deepa, J.; Binussreejayan Electrochemical sensing of cholesterol by molecularly imprinted polymer of silylated graphene oxide and chemically modified nanocellulose polymer. *Mater. Sci. Eng. C* **2018**, *92*, 942–956.
26. Sun, Z.; Zhang, Y.; Zheng, S.; Park, Y.; Frost, R.L. Preparation and thermal energy storage properties of paraffin/calcined diatomite composites as form-stable phase change materials. *Thermochim. Acta* **2013**, *558*, 16–21.
27. Stobinski, L.; Lesiak, B.; Kövér, L.; Toth, J.; Biniak, S.; Trykowski, G.; Judek, J. Multiwall carbon nanotubes purification and oxidation by nitric acid studied by the FTIR and electron spectroscopy methods. *J. Alloys Compd.* **2010**, *501*, 77–84.
28. Ujjain, S.K.; Bhatia, R.; Ahuja, P.; Attri, P. Highly Conductive Aromatic Functionalized Multi-Walled Carbon Nanotube for Inkjet Printable High Performance Supercapacitor Electrodes. *PLoS ONE* **2015**, *10*, e0131475.
29. Duan, Y.; Luo, X.; Qin, Y.; Zhang, H.; Sun, G.; Sun, X.; Yan, Y. Determination of epigallocatechin-3-gallate with a high-efficiency electrochemical sensor based on a molecularly imprinted poly(o-phenylenediamine) film. *J. Appl. Polym. Sci.* **2013**, *129*, 2882–2890.

