

Combined use of physical treatments and edible coatings in fresh produce: moving beyond

Cristian M Ortiz¹, Ariel R Vicente^{1,2*} and Adriana N Mauri¹

¹Centro de Investigación y Desarrollo en Criotecnología de Alimentos (CIDCA), CONICET, Universidad Nacional de La Plata, La Plata, Argentina

²Laboratorio de Investigación en Productos Agroindustriales (LIPA), Facultad de Ciencias Agrarias y Forestales, La Plata, Argentina

Purpose of the review: In this review we describe the general principles and recent trends in edible coating of fresh produce. We also discuss some features that may link coating applications with common postharvest physical methods (modified atmospheres, heat treatments and irradiation).

Main findings: Coatings of intact and fresh-cut produce must be considered a supplement to proper temperature control. Works aimed at determining the outcome of edible coatings must then compare their benefits under proper postharvest storage conditions (optimal temperatures and high relative humidity with minimal delays). Extensive research has been conducted in the last decade to generate material combinations that would meet the specific needs for coating fruits and vegetables. Substantial improvements in the materials properties (eg, through the evaluation of novel matrix constituents and functional ingredients, as well as by developing multilayer, blended or nanocomposite materials) are still possible. In spite of their relative long history as treatments to reduce deterioration and their joint categorization as environmentally-friendly approaches, coating technologies and postharvest physical methods have evolved quite independently. Combined developments may be useful to increase their positive effects. Rather than simply adding the improvements of different treatments applied sequentially, we argue that active collaboration between researchers working on postharvest physical treatments and edible coatings offers opportunities to develop better and innovative approaches that maximize the benefits on quality maintenance.

Directions for future research: Innovative approaches superseding the additive effects of combining coatings and physical treatments are needed. One area to be explored more deeply is the application of physical treatments during or after coating formation, which may improve the material's properties, while inducing hormetic responses in the commodities. Coating-induced modified atmospheres need to be revisited and improved. The feasibility of using mild post-coating physical treatments to induce custom changes in the materials and to facilitate the release of active principles also requires further work. Finally, research aimed at solving technical difficulties arising during scaling-up processes may help to facilitate the transfer of coating applications from academia to industry.

Keywords: coatings; biopolymers; physical treatments; postharvest; quality

Postharvest technology in the 21st century: revisiting and refining traditional tools

Postharvest science has greatly improved in recent years. Sequencing of some important fruit and vegetable species, development of powerful high-throughput analytical ('omic') approaches, and progress in deciphering the epigenetic regulation of plant development are contributing to increase our understanding of the physiology of harvested fruits and vegetables [1-4]. Some relatively new technologies such as the use of the ethylene action inhibitor 1-methylcyclopropene and the refinement of dynamic controlled atmospheres have provided alternative ways to control ripening and senescence [5-8]. In spite of this progress the main challenge on a global scale is still to develop sustainable tools to maintain the safety, nutritional and organoleptic quality of freshly harvested products and to reduce the unacceptably high losses occurring during distribution, storage and retail [9].

Recently there has been increased interest in revisiting the use of physical treatments (heat, irradiation) and natural edible coatings to prevent fruit and vegetable deterioration [10]. The fact that these methods have been envisioned as environmentally-friendly approaches to complement traditional postharvest handling methods [11] has raised the interest of industry and academia.

Postharvest physical treatments and edible coating applications have evolved relatively independently. However, they may have high potential as complementary approaches for postharvest management. Herein, we briefly describe the general features and trends in fresh produce edible coating applications. We also dis-

cuss the potential of combining coating technologies with appropriate postharvest physical methods. Active collaboration between researchers working on postharvest physical treatments and edible coatings offer opportunities to maximize their benefits on quality maintenance.

Edible coatings for fruits and vegetables: uses and formulations

Edible coatings consist of a thin layer of protective material added by dipping or spraying that can be consumed as a part of the product [12, 13]. The main interest in edible coatings is generally based on their potential to prevent quality loss and extend the shelf life of the commodities by modifying their surface properties. Coating applications have been used in fresh fruits and vegetables for a long time. Some records indicate that citrus fruits were waxed already in China in the 12th and 13th centuries [14]. Suspensions of oils or waxes in water were among the first fruit-coating formulations likely used to reduce dehydration. Since these early practices the aims of coating applications have expanded. Fruits and vegetables could be coated:

- to improve surface gloss,
- to reduce abrasion,
- to prevent the absorption of undesirable odors,
- to minimize solute leakage,
- to establish a barrier to moisture loss,
- to change CO₂ and O₂ levels inside the commodity, thereby modulating ripening and senescence,
- to retard decay, or

*Corresponding author: Ariel Vicente; Email: arielvicente@quimica.unlp.edu.ar

- to enhance produce stability and safety by including functional ingredients such as antioxidants and antimicrobials.

A variety of materials, including lipids, polysaccharides, and proteins, alone or in combination, have been used in edible coating formulations [12, 15]. Coating matrices generally comprise a major constituent, to which other components such as plasticizers, emulsifiers, reinforcements, additives and solvents (eg, water and alcohols) are added. The large battery of materials used and their possible combinations provide coating systems with a wide range of physical and chemical properties that could fit different applications.

The main matrix components used in coating applications are hydrocolloids, lipids and their mixtures (composites) [16*]. Polysaccharides (cellulose derivatives, alginate, pectin, starch, chitosan, carragenan and gums) and proteins (from vegetal origin like soy protein, wheat gluten, corn zein, and from animal sources such as gelatin, whey, casein and keratin) have been tested [17, 18]. Special interest has been devoted to the re-utilization of industrial byproducts (ie, gelatin, soy protein, whey and chitosan from the meat, oilseed, cheese and seafood industries, respectively) [19**].

Both protein and polysaccharide films and coatings generally exhibit excellent barrier properties against oxygen, and aroma, but have high water vapor permeability [19**]. A number of different lipid sources including waxes (beeswax, candelilla wax, carnauba wax), free fatty acids, fatty alcohols, fatty acid and sucrose esters, edible terpene resins, such as shellac and paraffins, have been used for fruit and vegetable coating [18, 20-22]. They usually increase surface gloss and limit water loss. Finally, composite coatings are produced by combining two or more constituents. They could be emulsions, multilayer coatings or blends [23, 24]. In emulsions and lipid-hydrocolloids composite coatings, the type of lipid, location, volume fraction, polymorphic phase, and drying conditions have significant impact on the final moisture-barrier properties [25]. The size of the dispersed component in an emulsion also has great effect on the coating characteristics. It can range from 0.2-50 μm in macro-emulsions to 10-100 nm in micro-emulsions [26-28]. Multilayer coatings are in general more effective water vapor barriers than emulsion films. However, the fact that they should be generated in various steps by sequential deposition of materials of interest in the commodity surface represents a technological drawback. An alternative promising strategy to improve the properties of edible coatings is biopolymer blending [29, 30].

As in the field of synthetic polymers the new generation of materials in the area of biopolymers is focusing on obtaining bio-nanocomposites. These materials consist of a biopolymer matrix reinforced with nanoparticles (particles having at least one dimension in the nanometer range 1-100 nm). They are expected to exhibit better mechanical and barrier properties, thermal stability, chemical resistance and surface appearance, even at low loading levels (5-10%), than traditional materials due to the high surface area of the nanoparticles, which allows them to interact strongly with the remaining components [31, 32]. The addition of nanoclays, like montmorillonite, to protein and polysaccharide formulations has enhanced the mechanical and barrier properties of biopolymeric films [33, 34]. Cellulose nanofibers, chitin and starch nanowhiskers have improved the mechanical properties of casein, starch and chitosan films [35-37]. Nanocomposites may also modulate the release of active ingredients incorporated in the polymeric matrix [28-41].

Different ingredients can be incorporated into edible coating matrices to improve or modify their functionality. Plasticizers are commonly added in polysaccharide and protein-based coatings to

decrease brittleness [42-44]. Common plasticizers used in edible coatings include water, glycerol, propylene glycol, sorbitol, sucrose, polyethylene glycol, fatty acids, and monoglycerides [42]. Added in different proportions (15-40% relative to the main matrix constituent) plasticizers may enhance film flexibility and susceptibility to humidity and decrease its strength and barrier properties against moisture and oxygen [45]. Hydrophobic plasticizers, such as citrate esters are also being studied [46].

The functionality of these main matrix constituents can be also modified by chemical, physical or enzymatic treatments [47-49]. Protein cross-linking may reduce the material solubility, generate stronger matrices and in some cases increase surface hydrophobicity [49-50]. Traditional chemical cross-linkers may not be suitable for edible materials due to their toxicity. In this case protein modifying-enzymes such transglutaminase, lipoxidase, lysyl-oxidase, polyphenol oxidase and peroxidase may be useful. Phosphorylation of soy proteins by a protein kinase can also be used to modulate the material solubility and emulsifying capacity [51]. Polysaccharide properties can be also modified by changing their degree of polymerization, side-chains, degree of acetylation or methylation [52].

Emulsifiers are surface-active agents of an amphiphilic nature and are able to reduce the surface tension of water-lipid or water-air interface. They can be also added to improve the formulation wetting, spreading and adhesion ability [53**]. Protein-lipid or polysaccharide-lipid composite coatings often require the addition of emulsifiers to facilitate the dispersion of the lipid component in aqueous media. Common emulsifiers include the polyethylene sorbitol esters, some fatty acids and salts, and phospholipids such as lecithin [46, 54, 55].

The functionality of edible coatings can be expanded by adding antioxidants, antimicrobials, colorants, flavors, nutrients, and spices to the formulation. These compounds could be retained on the food surface enhancing food quality, stability, and safety [56-60]. Common antimicrobial agents used in food systems, such as benzoic, propionic and sorbic acid salts, may be incorporated. Starch-based coatings containing potassium sorbate were applied on fresh strawberries to reduce decay [61]. Hydroxypropyl-methyl cellulose coatings containing ethanol inactivated *Salmonella montevideo* on fresh tomatoes [62]. Antimicrobial enzymes such as lysozyme have been incorporated into chitosan coatings to control *Escherichia coli* and *Streptococcus faecalis* [63]. Essential oils have been also widely investigated as natural antimicrobials for coated fruits and vegetables [64]. Interestingly, chitosan shows antimicrobial effects *per se* [65-67].

Early work by Swenson *et al.* [68] showed that coatings containing butylated hydroxyanisole (BHA), butylated hydroxy toluene (BHT) and citric acid reduced rancidity in nuts. Antioxidants such as ascorbic acid have been used to reduce browning in coated mushrooms [69]. Xanthan gum coatings mixed with α -tocopherol enhanced the nutritional quality and improved the surface color of peeled baby carrots [70]. Texture enhancers have been also incorporated into edible coatings. Hernández-Muñoz *et al.* [71] observed that the addition of calcium gluconate to the chitosan (1%) coating formulation increased the firmness of refrigerated strawberries. Flavor and coloring agents may also be added to edible coatings. However, very little has been reported regarding these applications. Finally, some researchers have endeavored to incorporate micronutrients and bioactive compounds [63-72].

Challenges in edible coating technologies

Several works show that the benefits obtained by coating applica-

tions may be substantial [73-77]. However, compared to the large number of studies reported in literature relatively few applications have been adopted by the industry. Although various raw materials employed are industrial byproducts, the procurement, transport, distribution and storage costs of these materials as well as the infrastructure for large-scale coating production may be in some cases still too high. Modification of established procedures, changes in process operations and processing line layouts, and the need to implement specific training programs may have in some cases delayed technology transfer. Future studies aimed at helping to re-categorize coatings as a "widespread technology" should focus on:

- i. selecting the formulation, and coating-formation conditions that optimize the properties required for each commodity;
- ii. anticipating potential consumer concerns especially when complex activated formulations are tested. For example nanotechnology offers opportunities to develop novel packaging strategies [78]; but the debate about the potential benefits and risks of human consumption of some nanoparticles is still open and legislation on this issue is in many cases diffuse;
- iii. predicting potential difficulties during scaling-up processes may be highly useful. Simple procedures in lab settings such as generating continuous and even coatings, while avoiding physical damage and temperature abuses may be challenging in commercial settings.

The evaluation coatings at unacceptably high temperatures and low relative humidities for fruit and vegetable handling could overestimate their practical benefits. Technologically-oriented studies must evaluate coating applications under optimal storage conditions (temperature and relative humidity) since they should be considered a complement to proper refrigeration.

Intersecting fruit coating technology and postharvest physical methods

Physical treatments before or during coating formation

The application of physical methods to raw materials may modify the properties of the resulting coatings [79, 80]. Heat, high pressure and irradiation treatments have been shown to induce changes in the functionality of the resulting films. The effects are generally more marked for solubility and mechanical properties than for water vapor permeability [81-88].

Mild heat treatments are used in some cases to accelerate coating formation [89, 90]. The nature of the treatment applied may affect the physical-chemical properties of the coating itself [91]. The drying process applied to protein films and coatings may affect the type and proportion of covalent (S-S bonds) or non-covalent (hydrophobic interactions, ionic and hydrogen bonds) interactions and, consequently, material properties [92]. In addition, for polysaccharides the drying temperature may affect film performance [93]. Similarly, the relative humidity and pressure during the drying period has been also shown to affect the properties of soy and amaranth biopolymers [92, 94, 95].

While these physical treatments have been in many cases optimized based only on the material requirements, they may be potentially exploited to induce hormetic responses in the commodities, maximizing the final outcome of the combined treatment on quality maintenance. The presence of a protein, polysaccharide, lipid or composite coating on the commodity surface would affect the heat transfer properties and consequently the optimal treatment conditions will likely differ compared to the scheduled in uncoated products.

Physical treatments after coating formation

This approach includes: i) the modified atmosphere that would be

established in coated commodities due to the change in the fruit or vegetable surface permeability to gases (CO_2 , O_2 , ethylene); or ii) any change in the environment aimed in eliciting a desirable response in the commodity or in the coating material.

Coating-induced modified atmospheres

Once the coating is established the gas exchange between the fruit and the environment is affected, providing the opportunity to generate single fruit modified atmospheres. The equilibrium gas composition to be reached is determined by a number of product factors, coating properties and environmental conditions.

Product factors: The respiratory rate of the commodity is the most important factor affecting the modified atmosphere reached. It will depend on the species, cultivar and on its developmental stage [96]. Ethylene production and accumulation may be important especially in climacteric coated commodities in which it may increase respiration.

Coating properties: All factors affecting the mass transfer through the film and the diffusion process such as permeability to O_2 , CO_2 and thickness will determine the atmosphere established. Plasticizers, generally increase film permeability. The polymerization and/or cross-linking degree of the protein or polysaccharide as well as the functional groups/substituents added or removed would also contribute to the coating permeability. In general protein and polysaccharide coatings have high permeability to polar substances, such as water vapor, and low permeability to non-polar substances, such as oxygen. Protein coatings appear to have lower oxygen permeability than cellulose-based coatings. Besides the absolute permeability of the different coatings their selectivity is also a main determinant of the internal atmosphere of fruits [97, 98].

Environmental conditions: Coated fresh fruits and vegetables would be stored mostly at relative humidities between 85 and 95%. Under these conditions some coatings may hydrate, substantially changing their physical and chemical properties [28, 99]. In general, at low and intermediate relative humidity, the permeability to gases of protein and polysaccharide materials is much lower than that of polyethylene, which makes them interesting materials for certain applications. However, the permeability of polysaccharide and protein coatings are highly dependent on the relative humidity [100]. For instance, the permeability to oxygen and carbon dioxide may increase up to a thousand times when these materials are held at high relative humidity. In protein materials, this effect is much greater for the "hydrophilic" gases (CO_2) than for the "hydrophobic" gases (O_2) [101, 102]. It is important to highlight that many of the standard methods of material characterization use relative humidities which are far away (58%) from those in which they will be used in when coating fresh fruits and vegetables. The storage temperature also has a significant impact on the atmosphere reached, as it exponentially affects fruit and vegetable respiration rate. Variations in the cold chain may lead to fermentation in coated commodities [103]. The CO_2 and O_2 partial pressures in the storage atmosphere may also influence the levels of gases at steady state through alterations in the commodity respiration rate.

Post-coating physical treatments

As previously indicated some physical treatments may be used to induce desirable changes in the vegetable physiology [103]. The evaluation of post-coating heat treatments has received little attention so far. Post-coating UV-irradiation may be expected to affect mostly the coating formulation given its low penetration. Some works have shown that the functional properties of biopolymers also can be improved if subjected to some physical treatments after their formation. The tensile strength of soy protein

films was improved when subjected to UV or γ radiation. This is probably due to the high content of tyrosine and phenylalanine, since these amino acids can cross-link when irradiated [104, 105]. Changes in the postharvest environment may also be potentially exploited to modulate the functional properties of activated coatings [106]. When incorporated into tomato fruit packages, soy protein pads absorbed water, facilitating the release of cyclodextrin- α -1-methylcyclopropene formulations which delayed ripening [107]. Commodity compatible heat treatments during storage may also be considered as a strategy to modify the diffusion rate of additives present in the coating matrix. In some cases they may also be envisioned as a strategy to increase the rate of temperature dependent reactions between coating ingredients (eg, enzymatic reactions). The release of active compounds added to the coating by photochemical means (eg, UV-vis radiation) may be another strategy to explore.

Concluding remarks

Edible coatings and postharvest physical methods have a relatively long history as treatments to reduce deterioration of stored fruits and vegetables and have been envisioned as environmentally-friendly approaches to supplement refrigeration. Although both strategies have evolved quite independently, combined developments may maximize the beneficial effects on quality maintenance of fresh produce. Coatings should be considered as a supplementary treatment to proper temperature control in intact and fresh-cut produce. Consequently, studies must compare their benefits against controls subjected to proper postharvest management and storage (refrigeration and high relative humidity with minimal delays). This will unequivocally determine their benefits relative to the recommended handling practices. Continuing the currently active work oriented to maximize coating functionality will be useful. Active collaboration between postharvest technologists and material scientists may reduce the boundaries between these two fields. Innovative approaches superseding the additive effects of combined coating-physical treatments application will be necessary. Some areas that may be explored include the application of physical treatments before or during coating formation which may improve the material properties while inducing hormetic responses in the commodities. Optimization of the single-fruit modified atmospheres generated after coating establishment is still needed. The feasibility of using mild post-coating physical treatments to make custom changes in the material properties and/or to facilitate the release active principles still needs to be determined. Finally, research aimed at solving technical difficulties arising during scaling-up processes may help to boost the transfer rate of coating applications from academia to industry.

References

Papers of interest have been highlighted as:

*Marginal importance

** Essential reading

- 1 Tikunov YM, Molthoff J, de Vos RCH, Beekwilder J, van Houwelingen A, van der Hoof JJJ, Nijenhuis-de Vries M, Labrie CW, Verkerke W, van de Geest H, Zamora MV, Presa S, Rambla JL, Granell A, Hall RD, Bovy AG. Non-smoky GLYCO-SYLTRANSFERASE₁ prevents the release of smoky aroma from tomato fruit. *Plant Cell* 2013; 25:3067-3078.
- 2 Molassiotis A, Tanou G, Filippou P, Fotopoulos V. Proteomics in the fruit tree science arena: New insights into fruit defense, development, and ripening. *Proteomics* 2013; 13:1871-1884.
- 3 Ay N, Janack B, Humbeck K. Epigenetic control of plant senescence and linked processes. *Journal of Experimental Botany* 2014; 65:3875-3887.
- 4 Karlova R, Chapman N, David K, Angenent GC, Seymour GB, De Maagd RA. Transcriptional control of fleshy fruit development and ripening. *Journal of Experimental Botany* 2014; 65:4527-4541.
- 5 Martínez-Romero D, Bailén G, Serrano M, Guillén F, Valverde JM, Zapata P, Castillo S, Valero D. Tools to maintain postharvest fruit and vegetable quality through the inhibition of ethylene action: A review. *Critical Reviews in Food Science and Nutrition* 2007; 47:543-560.
- 6 Sozzi GO, Beaudry RM. Current perspectives on the use of α -1-methylcyclopropene in tree fruit crops: An international survey. *Stewart Postharvest Review* 2007;3:8.
- 7 Watkins CB. The use of α -1-methylcyclopropene (1-MCP) on fruits and vegetables. *Biotechnology Advances* 2006; 24:389-409.
- 8 Rizzolo A, Grassi M, Vanoli M. α -1-Methylcyclopropene application, storage temperature and atmosphere modulate sensory quality changes in shelf-life of 'Abbé Fétel' pears. *Postharvest Biology and Technology* 2014; 92:87-97.
- 9 FAO (2013a) Food waste harms climate, water, land and biodiversity – new FAO report. September 2013. Available online at: <http://www.fao.org/news/story/en/item/196220/icode/>. Accessed 21 Oct 2014.
- 10 Pérez-Gago MB, Gonzales-Aguilar GA, Olivas GI. Edible coatings for fruits and vegetables. *Stewart Postharvest Review* 2010; 6:1-14.
- 11 Guilbert, S. Technology and application of edible protective films. In: Food packaging and preservation; Theory and practice. Mathlouthi M (Ed). Elsevier Applied Science, London, UK, 1986:371-394.
- 12 Ukai NY, Shingo I, Kurume TT, Fukuoka KM. Preservation of agricultural products, U.S. Patent, 1976: 3,997,674.
- 13 Gennadios A, Weller CL. Moisture adsorption by grain protein films. *American Society of Agricultural Engineers* 1994; 37:535-539.
- 14 Hardenburg RE. Wax and related coatings for horticultural products: a bibliography. *Agriculture, Agricultural Research Service Bulletin* 1967; 51:1-15.
- 15 Kester JJ, Fennema OR. Edible films and coatings: A review. *Food Technology* 1986; 40:47-59.
- 16 Poverenov E, Rutenberg R, Danino, S, Horev, B, Rodov, V. Gelatin-chitosan composite films and edible coatings to enhance the quality of food products: Layer-by-layer vs. Blended formulations. *Food and Bioprocess Technology* 2014; 7:3319-3327.
- * Gelatin and chitosan and their blends biopolymers generated and characterized. Fresh-cut melons coated with these formulation showed improved quality maintenance.
- 17 Rojas-Graü MA, Salvia-Trujillo L, Soliva-Fortuny R, Martín-Belloso O. Edible films and coatings. In: Decontamination of fresh and minimally processed produce, Gomex Lopez VM (Ed); 2012: 247-275.
- ** This chapter is a good source for researchers interested in understanding the general principles of edible coatings applications in fresh produce.
- 18 Park HJ, Byun YJ, Kim YT, Whiteside WS, Bae HJ. Processes and applications for edible coating and film materials from agropolymers. In: *Innovations in Food Packaging: Second Edition* 2013; 257-275.
- 19 Baldwin EA, Hagenmaier R, Bai J. *Edible Coatings and Films to Improve Food Quality*, Second Edition. Ed. CRC Press. Boca Raton, FL, USA, 2012.
- ** This book is a complete study of the coatings, films, wraps and surface treatments used for foods. It covers films and coating ingredients and additives, their permeability properties, applications, coating technologies and regulatory aspects.
- 20 Paredes-López O, Camargo-Rubio E, Gallardo-Navarro Y. Use of coatings of candelilla wax for the preservation of limes. *Journal of the Science of Food and Agriculture* 1974; 25:1207-1210.
- 21 Lawrence JF, Iyengar JR. Determination of paraffin wax and mineral oil on fresh fruits and vegetables by high temperature gas chromatography. *Journal of Food Safety* 1983; 5:119-129.
- 22 Warth AH. *The chemistry and technology of waxes*, Reinhold, New York, 1986:37-192.
- 23 Mantilla N, Castell-Perez ME, Gomes C, Moreira RG. Multilayered antimicrobial edible coating and its effect on quality and shelf-life of fresh-cut pineapple (*Ananas comosus*). *LWT-Food Science and Technology* 51: 2013:37-43.
- 24 Sipahi RE, Castell-Perez ME, Moreira RG, Gomes C, Castillo A. Improved multilayered antimicrobial alginate-based edible coating extends the shelf life of fresh-cut watermelon (*Citrullus lanatus*). *LWT - Food Science and Technology* 2013; 51:9-15.
- 25 Gontard N, Duche C, Cuq J, Guilbert S. Edible composite films of wheat gluten and lipids water vapor permeability and other physical properties. *International Journal of Food Science and Technology* 1994; 29:39-50.
- 26 Kamper SL, Fennema O. Water Vapor Permeability of Edible Bilayer Films. *Journal of Food Science* 1984; 49:1478-1481.
- 27 Kamper SL, Fennema O. Water Vapor Permeability of an Edible, Fatty Acid, Bilayer Film. *Journal of Food Science* 1984(b); 49:1482-1485.
- 28 Hagenmaier RD, Shaw PE. Moisture permeability of edible films made with fatty acid and (hydroxypropyl)methylcellulose. *Journal of Agricultural and Food Chemistry* 1990;38:1799-1803.
- 29 Ban W, Song J, Argyropoulos DS, Lucia LA. Improving the physical and chemical functionality of starch-derived films with biopolymers. *Journal of Applied Polymer Science* 2006; 100:2542-2548.
- 30 Famá L, Gerschenson L, Goyanes S. Starch-vegetable fiber composites to protect food products. *Carbohydrate polymers* 2009; 75:230-235.
- 31 Rhim JW, Perry KW. Natural biopolymer-based nanocomposite films for packaging applications. *Critical Reviews in Food Science and Nutrition* 2007; 47:1-24.
- 32 Zhao R, Torley P, Halley PJ. Emerging biodegradable materials: starch- and protein-based bio-nanocomposites. *Journal of Materials Science* 2008; 43:3058-3071.
- 33 Echeverría I, Eisenberg P, Mauri AN. Nanocomposites films based on soy proteins and montmorillonite processed by casting. *Journal of Membrane Science*

- 2014: 449:15–26.
- 34 Quilaqueo Gutiérrez M, Echeverría I, Ihl M, Bifani V, Mauri AN. Carboxymethylcellulose-montmorillonite nanocomposite films activated with murta (*Ugni molinae* Turcz) leaves extract. *Carbohydrate Polymers* 2012; 87:1495-1502.
- 35 Pereda M, Amica G, Rácz I, Marcovich NE. Structure and properties of nanocomposite films based on sodium caseinate and nanocellulose fibers. *Journal of Food Engineering* 2011; 103:76-83.
- 36 Xie F, Pollet E, Halley PJ, Avérous L. Starch-based nano-biocomposites. *Progress in Polymer Science* 2013; 38:1590-1628.
- 37 Rubenthaler V, Ward TA, Chee CY, Tang CK. Processing and analysis of chitosan nanocomposites reinforced with chitin whiskers and tannic acid as a crosslinker. *Carbohydrate Polymers* 2015; 115:379-387.
- 38 Tunc C, Angellier H, Cahyana Y, Chalier P, Gontard N, Gastaldi E. Functional properties of wheat gluten/montmorillonite nanocomposite films processed by casting. *Journal of Membrane Science* 2007; 289:159-168.
- 39 Mascheroni E, Chalier P, Gontard N, Gastaldi E. Designing of a wheat gluten/montmorillonite based system as carvacrol carrier: Rheological and structural properties. *Food Hydrocolloids* 2010; 24:406-413.
- 40 Cortez-Vega WR, Pizato S, De Souza JTA, Prentice C. Using edible coatings from Whitemouth croaker (*Micropogonias furnieri*) protein isolate and organo-clay nanocomposite for improve the conservation properties of fresh-cut 'Formosa' papaya. *Innovative Food Science and Emerging Technologies* 2014; 22:197-202.
- 41 Zambrano-Zaragoza ML, Mercado-Silva E, Ramirez-Zamorano P, Cornejo-Villegas MA, Gutiérrez-Cortez E, Quintanar-Guerrero D. Use of solid lipid nanoparticles (SLNs) in edible coatings to increase guava (*Psidium guajava* L.) shelf-life. *Food Research International* 2013; 51:946-953.
- 42 Sothornvit R, Krochta JM. Plasticizers in edible films and coatings. In: *Innovations in Food Packaging*. Elsevier, CA, 2005: 403-433.
- 43 Han JH, Gennadios A. Edible films and coatings: a review. In: *Innovations in Food Packaging*. Elsevier, CA, 2005:239-262.
- 44 Oz AT, Ulukanli Z. Application of edible starch-based coating including glycerol plus oleum Nigella on arils from long-stored whole pomegranate fruits. *Journal of Food Processing and Preservation* 2012; 36:81-95.
- 45 Vanin FM, Sobral PJA, Menegalli FC, Carvalho RA, Habitante AMQB. Effects of plasticizers and their concentrations on thermal and functional properties of gelatin-based films. *Food Hydrocolloids* 2005; 19:899-907.
- 46 Andreuccetti C, Carvalho RA, Grosso CRF. Effect of hydrophobic plasticizers on functional properties of gelatin-based films. *Food Research International* 2009; 42:1113-1121.
- 47 Audic J-L, Chaufer B. Influence of plasticizers and crosslinking on the properties of biodegradable films made from sodium caseinate. *European Polymer Journal* 2005; 41:1934-1942.
- 48 Vaz CM, De Graaf LA, Reis RL, Cunha AM. In vitro degradation behaviour of biodegradable soy plastics: Effects of crosslinking with glyoxal and thermal treatment. *Polymer Degradation and Stability* 2003; 81:65-74.
- 49 Galietta G, Di Gioia L, Guilbert S, Cuq B. Mechanical and thermomechanical properties of films based on whey proteins as affected by plasticizer and crosslinking agents. *Journal of Dairy Science* 1998; 81:3123-3130.
- 50 Tang C-H, Jiang Y, Wen Q-B, Yang X-Q. Effect of transglutaminase treatment on the properties of cast films of soy protein isolates. *Journal of Biotechnology* 2005; 120:296-307.
- 51 Campbell NF, Shih FF, Marshall WE. Enzymatic phosphorylation of soy protein isolate for improved functional properties. *Journal of Agricultural and Food Chemistry* 1992; 40:403-406.
- 52 Dastidar TG, Netravali AN. Improving resin and film forming properties of native starches by chemical and physical modification. *Journal of Biobased Materials and Bioenergy* 2012; 6:1-24.
- 53 Krochta JM. Proteins as raw materials for films and coatings: definitions, current, status, and opportunities. In: Gennadios, A. (ed.), *Protein-based films and coatings*. CRC Press, Boca Raton, USA, 2002:1-42.
- **This chapter represents a good reference source covering the main aspects of protein films and coatings.
- 54 Bravin B, Peressini D, Sensidoni A. Influence of emulsifier type and content on functional properties of polysaccharide lipid-based edible films. *Journal of Agricultural and Food Chemistry* 2004; 52:6448-6455.
- 55 Rodriguez M, Osés J, Ziani K, Maté JI. Combined effect of plasticizers and surfactants on the physical properties of starch based edible films. *Food Research International* 2006; 39:840-846.
- 56 Mehryar GF, Al-Qadiri HM, Swanson BG. Edible coatings and retention of potassium sorbate on apples, tomatoes and cucumbers to improve antifungal activity during refrigerated storage. *Journal of Food Processing and Preservation* 2014; 38:175-182.
- 57 Martínez-Avila GCG, Aguilera AF, Saucedo S, Rojas, R, Rodríguez, R, Aguilar, CN. Fruit wastes fermentation for phenolic antioxidants production and their application in manufacture of edible coatings and films. *Critical Reviews in Food Science and Nutrition* 2014; 54:303-311.
- 58 Karaca H, Pérez-Gago MB, Taberner V, Palou L. Evaluating food additives as antifungal agents against *Monilinia fructicola* in vitro and in hydroxypropyl methylcellulose-lipid composite edible coatings for plums. *International Journal of Food Microbiology* 2014; 179:72-79.
- 59 Fagundes C, Pérez-Gago MB, Monteiro AR, Palou L. Antifungal activity of food additives in vitro and as ingredients of hydroxypropyl methylcellulose-lipid edible coatings against *Botrytis cinerea* and *Alternaria alternata* on cherry tomato. *International Journal of Food Microbiology* 2013; 166:391-398.
- 60 Freitas IR, Cortez-Vega WR, Pizato S, Prentice-Hernández C, Borges CD. Document Xanthan gum as a carrier of preservative agents and calcium chloride applied on fresh-cut apple. *Journal of Food Safety* 2013; 33:229-238.
- 61 García MA, Martino MN, Zaritzky NE. Plasticized starch-based coatings to improve strawberry (*Fragaria x Ananassa*) quality and stability. *Journal of Agricultural and Food Chemistry* 1998; 46:3758-3767.
- 62 Zhuang R-Y, Beuchat LR. Effectiveness of trisodium phosphate for killing *Salmonella montevideo* on tomatoes. *Letters in Applied Microbiology* 1996; 22:97-100.
- 63 Park S-I, Zhao Y. Incorporation of a high concentration of mineral or vitamin into chitosan-based films. *Journal of Agricultural and Food Chemistry* 2004; 52:1933-1939.
- 64 Novaes Azevedo A, Ribeiro Buarque P, Oliveira Cruz OM, Fitzgerald Blank A, Barreto Alves P, Nunes ML, Lins de Aquino Santana LC. Response surface methodology for optimisation of edible chitosan coating formulations incorporating essential oil against several foodborne pathogenic bacteria. *Food Control* 2014; 43:1-9.
- 65 Kannat SR, Rao MS, Chawla SP, Sharma A. Effects of chitosan coating on shelf-life of ready-to-cook meat products during chilled storage. *LWT - Food Science and Technology* 2013; 53:321-326.
- 66 Bordenave N, Grelier S, Coma V. Hydrophobization and antimicrobial activity of chitosan and paper-based packaging material. *Biomacromolecules* 2010; 11:88-96.
- 67 Papineau AM, Hoover DG, Knorr D, Farkas DF. Antimicrobial effect of water-soluble chitosans with high hydrostatic pressure. *Food Biotechnology* 1991; 5:45-47.
- 68 Swenson HA, Miers JC, Schultz TH, Owens HS. Pectinate and pectate coatings. II. Application to nut and fruit products. *Food Technology* 1953; 7:232-235.
- 69 Nisperos-Carriedo MO, Buslig BS, Shaw PE. Simultaneous detection of dehydroascorbic, ascorbic, and some organic acids in fruits and vegetables by HPLC. *Journal of Agricultural and Food Chemistry* 1992; 40:1127-1130.
- 70 Mei Y, Zhao Y, Yang J, Furr HC. Using edible coating to enhance nutritional and sensory qualities of baby carrots. *Journal of Food Science* 2002; 67:1964-1968.
- 71 Hernández-Muñoz P, Almenar E, Valle VD, Velez D, Gavara R. Effect of chitosan coating combined with postharvest calcium treatment on strawberry (*Fragaria x ananassa*) quality during refrigerated storage. *Food Chemistry* 2008; 110:428-435.
- 72 Han C, Zhao Y, Leonard SW, Traber MG. Edible coatings to improve storability and enhance nutritional value of fresh and frozen strawberries (*Fragaria x ananassa*) and raspberries (*Rubus ideaus*). *Postharvest Biology and Technology* 2004; 33:67-78.
- 73 Gennadios A, Weller CL, Gooding CH. Measurement errors in water vapor permeability of highly permeable, hydrophilic edible films. *Journal of Food Engineering* 1994; 21:395-409.
- 74 Gennadios A, Hanna MA, Kurth LB. Application of edible coatings on meats, poultry and seafoods: A review. *LWT - Food Science and Technology* 1997; 30:337-350.
- 75 Miller KS, Krochta JM. Oxygen and aroma barrier properties of edible films: A review. *Trends in Food Science and Technology* 1997; 8:228-237.
- 76 Baldwin EA, Nisperos MO, Chen X, Hagenmaier RD. Improving storage life of cut apple and potato with edible coating. *Postharvest Biology and Technology* 1996; 9: 151-163.
- 77 Li P, Barth MM. Impact of edible coatings on nutritional and physiological changes in lightly-processed carrots. *Postharvest Biology and Technology* 1998; 14:51-60.
- 78 Kalia, A., Parshad, V.R. Novel trends to revolutionize preservation and packaging of fruits/fruit products: Microbiological and nanotechnological perspectives. *Critical Reviews in Food Science and Nutrition* 2015; 55: 159-182.
- 79 Vu KD, Hollingsworth RG, Salmieri S, Takala PN, Lacroix M. Development of bioactive coatings based on γ -irradiated proteins to preserve strawberries. *Radiation Physics and Chemistry* 2012; 81:1211-1214.
- 80 Junqueira-Gonçalves MP, Tapia A, Rodríguez C, Roschztzardt FI, Valenzuela X, Aguirre P. Extension of strawberry shelf-life by irradiated edible coating. *Italian Journal of Food Science* 2011; 23:125-130.
- 81 Koehler P, Kieffer R, Wieser H. Effect of hydrostatic pressure and temperature on the chemical and functional properties of wheat gluten III. Studies on gluten films. *Journal of Cereal Science* 2010; 51:140-145.
- 82 Mauri AN, Anón MC. Effect of solution pH on solubility and some structural properties of soybean protein isolate films. *Journal of the Science of Food and Agriculture* 2006; 86:1064-1072.
- 83 Mauri AN, Anón MC. Mechanical and physical properties of soy protein films with pH-modified microstructures. *Food Science and Technology International* 2008; 14:119-125.
- 84 Rangavajhyala N, Ghorpade V, Hanna M. Solubility and Molecular Properties of Heat-Cured Soy Protein Films. *Journal of Agricultural and Food Chemistry* 1997; 45:4204-4208.
- 85 Rhim JW, Gennadios A, Handa A, Weller CL, Hanna MA. Solubility, tensile, and color properties of modified soy protein isolate films. *Journal of Agricultural and Food Chemistry* 2000; 48:4937-4941.
- 86 Condés MC, Anón MC, Mauri AN. Amaranth protein films from thermally treated proteins. *Journal of Food Engineering* 2013; 119:573-579.

- 87 Ruan RR, Xu L, Chen PL. Water vapor permeability and tensile strength of cellulose-based composite edible films. *Applied Engineering in Agriculture* 1998; 14:411-413.
- 88 Cieřla K, Salmieri S, Lacroix M. γ -irradiation influence on the structure and properties of calcium caseinate-whey protein isolate based films. Part 2. Influence of polysaccharide addition and radiation treatment on the structure and functional properties of the films. *Journal of Agricultural and Food Chemistry* 2006; 54:8899-8908.
- 89 Arnon H, Granit R, Porat R, Poverenov E. Development of polysaccharides-based edible coatings for citrus fruits: A layer-by-layer approach. *Food Chemistry* 2015; 166:465-472.
- 90 Badwaik LS, Borah PK, Deka SC. Antimicrobial and enzymatic antibrowning film used as coating for bamboo shoot quality improvement. *Carbohydrate Polymers* 2014; 103:213-220.
- 91 Tapia-Blácido DR, Sobral PJA, Menegalli FC. Effects of drying temperature and relative humidity on the mechanical properties of amaranth flour films plasticized with glycerol. *Brazilian Journal of Chemical Engineering* 2005; 22:249-256.
- 92 Denavi G, Tapia-Blácido DR, Añón MC, Sobral PJA, Mauri AN, Menegalli FC. Effects of drying conditions on some physical properties of soy protein films. *Journal of Food Engineering* 2009; 90:341-349.
- 93 Flores S, Famá L, Rojas AM, Goyanes S, Gerschenson L. Physical properties of tapioca-starch edible films: Influence of filmmaking and potassium sorbate. *Food Research International* 2007; 40:257-265.
- 94 Tapia-Blácido DR, Sobral PJA, Menegalli FC. Effect of drying conditions and plasticizer type on some physical and mechanical properties of amaranth flour films. *LWT - Food Science and Technology* 2013; 50:392-400.
- 95 Thakhiew W, Devahastin S, Soponronnarit S. Effects of drying methods and plasticizer concentration on some physical and mechanical properties of edible chitosan films. *Journal of Food Engineering* 2010; 99:216-224.
- 96 Fonseca SC, Oliveira SAR, Brecht JK. Modelling respiration rate of fresh fruits and vegetables for modified atmosphere packages: a review. *Journal of Food Engineering* 2002; 52:99-119.
- 97 Guilbert S, Gontard N, Gorris LGM. Prolongation of the shelf-life of perishable food products using biodegradable films and coatings. *LWT - Food Science and Technology* 1996; 29:10-17.
- 98 Barron C, Varoquaux P, Guilbert S, Gontard N, Gouble B. Modified atmosphere packaging of cultivated mushroom (*Agaricus bisporus* L.) with hydrophilic films. *Journal of Food Science* 2002; 67:251-257.
- 99 Hagenmaier RD, Shaw PE. Permeability of shellac coatings to gases and water vapor. *Journal of Agricultural and Food Chemistry* 1991; 39:825-829.
- 100 Cisneros-Zevallos L, Krochta JM. Internal modified atmospheres of coated fresh fruits and vegetables: Understanding relative humidity effects. In: *Innovations in Food Packaging*, Elsevier, CA, 2005:173-184.
- 101 Guilbert S, Cuq B. Material formed from proteins. In: *Handbook of biodegradable polymers*. Ed. Bastioli, C. Rapra Technology Limited, Shawbury, Shrewsbury, Shropshire, Inglaterra: Smithers. 2005:339-384.
- 102 Farber JN, Harris LJ, Parish ME, Beuchat LR, Suslow TV, Gorny JR, Garrett EH, Busta FF. Microbiological safety of controlled and modified atmosphere packaging of fresh and fresh-cut produce. 2003. In: E. Allen Foegeding, E.A. (Ed.). *Comprehensive Reviews in Food Science & Food Safety* 2003; 142-160.
- 103 Lurie S. Postharvest heat treatments. *Postharvest Biology and Technology* 1998; 14:257-269.
- 104 Gennadios A, Rhim JW, Handa A, Weller CL, Hanna MA. Ultraviolet radiation affects physical and molecular properties of soy protein films. *Journal of Food Science* 1998; 63:225-228.
- 105 Gueguen J, Viroben G, Noireaux P, Subirade M. Influence of plasticizers and treatments on the properties of films from pea proteins. *Industrial Crops and Products* 1998; 7:149-157.
- 106 Pinheiro AC, Bourbon AI, Quintas MAC, Coimbra MA, Vicente AA. K-carrageenan/chitosan nanolayered coating for controlled release of a model bioactive compound. *Innovative Food Science and Emerging Technologies* 2012; 16:227-232.
- 107 Ortiz CM, Mauri AN, Vicente AR. Use of soy protein based 1-methylcyclopropene-releasing pads to extend the shelf life of tomato (*Solanum lycopersicum* L.) fruit. *Innovative Food Science and Emerging Technologies* 2013; 20:281-287.