



## Instructions on how to make an Outbreak of American Cutaneous Leishmaniasis

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### Abstract

Outbreaks of American Cutaneous Leishmaniasis (ACL) are triggered by the confluence of multidimensional biological, climate, and social factors: the eco-epidemiological momentum. Despite the diversity of epidemiological scenarios, there are common “Ingredients for the recipe” to make an ACL outbreak. To describe the “Where”, “When”, and “Why” of this momentum could contribute to the understanding of the causes of epidemics, preventing their occurrence, and to define better strategies to control them. Typically, due to sylvatic-rural transmission, most urban ACL are still related in time and space to peripheral city deforestations, riparian forest, and green patches within the cities. Therefore, the “Where” of the ACL outbreaks could be characterized in the space as an edge effect, and afterwards the “When” of this edge could be categorized as an ephemeral, transient or permanent. The “Why” question is far more complex, as it includes the exposure of humans to vector due to anthropic activities in each edge scenario. The evidence for ACL outbreak control strategies: barricading the edge by chemical and physical barriers, environmental management, and individual prevention, were reviewed. Answers to questions regarding these “Where”, “When”, “Why”, and “How to control” guided the content of the questions to be asked in the new focus of ACL transmission, not only to mitigate current transmission and prevent future outbreaks, but also to highlight the biological factors that might contribute to the possibility of an epidemic, and those conditions that modulate its actual probability, the climate and the social determination of risk.

**Keywords:** Eco-Epidemiology; Edge Effect; Phlebotominae; Urban Transmission

### Introduction

Cutaneous leishmaniasis is a neglected tropical disease, categorized as emerging and uncontrolled, having the highest increase of prevalence among these diseases between 1990 and 2016 [1,2]. The estimated worldwide incidence is between 690,000 to 1,213,300 cases per year, including 48,915 reported cases in 17 countries of the Americas during 2016 [3,4].

American Cutaneous Leishmaniasis (ACL) is produced by several species of the genus *Leishmania*, transmitted by different species of Phlebotominae (Diptera: Psychodidae), with a broad spectrum of mammal reservoirs and epidemiological scenarios [5]. Despite this diversity in most of the cases, ACL transmission is restricted in space and time and reported as focal outbreaks, even in hyper-endemic areas, where it could be recurrent, but

with an irregular pattern different from a seasonal trend [6,7]. These outbreaks were generated by the coincidence of multiple factors, both biological and social, a “Perfect storm” or an eco-epidemiological momentum, in the multidimensional sense of the eco-epidemiology definition by Susser [8].

The retrospective eco-epidemiology at the first sub-national jurisdiction was already used to understand the change in the transmission of ACL related to changes in land use [9], but a prospective eco-epidemiology at smaller spatial scales could be used to understand the outbreak generation and guide the studies of epidemic foci to mitigate transmission and prevent future epidemic events. While the studies from regional to global scales contribute to program policy and general advocacy [10], this review focuses in scales from site up to village, which are more useful for operational purposes in high-risk areas [11].

### “Urban” ACL

Urban transmission of ACL has been proposed during the

last decades, mirroring the urbanization of the American Visceral Leishmaniasis [12]. Risk of urban transmission of ACL is sometimes associated with mobilization of vectors due to deforestation near city borders or the creation of peripheral new neighborhoods, which generate ACL “Hot spots” [13-25]. ACL urban transmission has also been associated with environmental heterogeneity as secondary forest edges, riparian forests with periodic overflows [26-30], forest fragments within cities that harbor vectors and infected reservoirs [31-42], and zoological gardens or cemeteries [43-45].

Consequently, the urban ACL risk seems more closely linked to the organization of urban space than to actual urban transmission cycles. Contributing factors include the cities green belt and green preserved patches, the changes in land value—peripheral expansion, displacement of old settlements—, and the trend of returning to nature [46-50]. The unplanned growth of the city and its new scenarios of vector-human contact involve, with the social determination as a driving factor, work relations, housing quality, public services, and health accessibility, as well as spatial segregation of people according to social status [51-53]. In turn, these situations pressure the vector-reservoir-parasite triad to adapt to new green-patched environments, but with a higher density of food sources for the vector and synanthropic reservoirs [54]. Further, the cases at urban borders could show a different spatial pattern from year to year, suggesting that transmission could be due to micro-local environmental interventions as deforestation in small areas or just around a new neighborhood, and so, in these foci, the urban peridomestic areas are not acting as a source population in the sense of the metapopulation dynamics [55-58]. However, in many cases city officials and politicians associated urban transmission with blanket spatial insecticide interventions over the whole city.

In the same sense, alleged urbanization, with intra-domiciliary risk, could sometimes be related to towns of such size that have all the borders less than 250 m from the border with forested areas, relatively low captures of vectors, such as in Cochabamba, Bolivia [59], or villages promoted for eco-tourism or fishing close to reserves and hotels surrounded by forest [60-64]. Studies of vector distribution based on case-control designs or small sample sizes could also miss environmental associations when the cases are controlled by landscape or share a similar environment [65]. In larger cities, such as Manaus, Brazil, the ACL cases were clustered close to the Negro river and 76% lived within 500 m of forested areas inside the city, although the authors caution about that “It cannot be determined from these data whether transmission of these cases occurred within or outside the city of Manaus” [66]; further, as the surveillance notification systems usually records the residence or diagnosis of the cases site instead of the probable infection site.

The cases could be reported also as urban due to the residences recorded in their clinical files, but actually the cases are living in

illegally occupied lands, where they make small deforestations for subsistence farms and domestic animal breeding in the forest fringe [67,68]; or they all live in an urban neighborhood from where they are recruited together for forest-related work [20]. Domestic-peridomestic transmission assumed by the presence of endophagic vectors or the gender-age distribution of the cases does not mean neither urban transmission, as indoor transmission could happen in relation to closeness to edges [69-71], although the presence of vectors in human residential environments improves the probability of vector control [72], consistent with control guidelines [73,74].

This edge effect seems to explain many peri-urban ACL events, at the micro-scale. However, the edge effect could be extrapolated also to most of the ACL outbreaks as a conceptual frame of analysis? considering the broad spectra of interfaces between urban-rurban-rural-sylvatic landscapes, from indoor to extra-domestic transmission, from mixed urban and rural human practices [75-77]. To answer this question, and to also know what questions to ask in ACL focus studies for managing outbreaks, except atypical ACL due to *Leishmania infantum-Lutzomyia longipalpis* [12], I will discuss the “Where”, “When”, “Why”, and “How to control” described for ACL outbreaks. On the other hand, ACL urbanization is a circumstantial event by the effect of the pressure on vectors to be adapted to domestic environments, the selection and plasticity of species, and speciation trends within them, with different vectorial capacity.

### “Where”: The Edge Effect

As a general statement in anthropized landscapes with phlebotomine suitability, the species diversity decreases, while few species increase its relative and absolute abundance, mainly in domestic landscapes that could resemble forest microhabitats. Thus, the species composition varies according to the degree of fragmentation of the forest [39,41,78-94]. Phlebotomine collections in sylvatic environments could also have fewer collected individuals due to less blood availability close to the trap, as when light traps are used as the only method of capture [95].

Otherwise, when peridomestic environments still have great phlebotomine species diversity, the influence of nearby forest patches should be considered as population sources. In Brazil a higher species diversity in rural areas than in periurban areas was associated with the closeness to ecotopes, such as residual forests, rocky outcrops, and subsistence farms and its environmental gradients, together with the heterogeneity and density of sylvatic, synanthropic, and domestic animals that provide blood sources in the ecotone area [96,97]. This availability of blood sources and a vector-feeding preference threshold could in turn modulate the short-range dispersion from the edges [98,99].

Even small modifications in the landscape led to an increase in the number of individuals of vector competent species, as

was reported in a crop-forest interface in Argentina [100], or the generation of clustered ACL cases in forest intervened borders even in rural settlements [101,102]. These new edges, besides the concentrated source of blood and diversity of shelters, provide new breeding sites for vectors with soil enriched with fallen leaves and manure. Further, synanthropic mammals also attracted to crops or domestic garbage, facilitate the possibility of multi-host reservoirs or a reservoir community linking sylvatic and peridomestic parasite transmission cycles [103-106].

In hyper-endemic areas the abundance of individuals of phlebotomine species was also associated with primary or secondary forest edges and vegetation coverage in a buffer area of 100 to 250 m or within small local surveillance units [7,33,107-111]. The relative abundance of phlebotomine between preserved areas and neighborhoods with forest fragments in a locality of Mato Grosso, Brazil, was 32:1 [112], while in the three-country border of Brazil-Bolivia-Peru, this value was 1.7:1 in peridomestic captures performed 200 m from the riparian forest continuous with the rainforest [113]. In the Argentina-Brazil-Paraguay three-country border, ACL peridomestic vectors are associated with proximity to natural reserves, residual forest, and rural forested edges in periruban areas [114,115].

However, as a word of caution on the “Where” question, at the local of infection or the smallest spatial scale, the distribution of ACL cases may have some bias due to differences between the distribution of infection and the distribution of clinically expressed cases, as the latter could actually be related with the distribution of the people with other co-infections, such as helminth ones that could immunomodulate the pathogenesis of *Leishmania* [116-118]. Other factors that could create bias regarding the ACL distribution are the stochastic or unknown distribution determinants of reservoir, parasites, vectors, infected vectors and reservoirs, and the genetic human predisposition for clinical signs or clustering of social determinants, such as familiar mucosal cases [119], while the hypothesis of endemic transmission by human reservoirs should require further population-based epidemiological evidence [120].

At these micro-spatial levels, the intra-domestic or peridomestic human infection could be related to the actual distribution of vectors as in an ACL focus in Chapare, Bolivia, where 99.3% of phlebotomine were collected outdoors [121], but also to the hourly distribution of vectors according to the hourly distribution of human exposure activities [122]. In Chaparral, Colombia, the peridomestic outdoor risk is from 19:00 to 20:00 hours and the indoor risk is from 23:00 to 24:00 hours [123], while in other scenarios the ratio between outdoor indoor vector abundance showed a linear correlation and with ACL incidence [15]. Besides, the housing quality, as its openness and crevices, the peridomestic suitability for vector breeding, and practices related to vector exposition—sleeping outdoors, collecting water, domestic

or synanthropic and wild animals near the house—are factors associated with risk to infection, not only by vector accessibility to hosts, but also as it was stated above, to the social determination together with the closeness to woodland remnants [124-130].

### “When”: Ephemeral, Transient and Permanent Edges

The ACL transmission clusters in time, as in space, through extraordinary events, usually narrow peaks, even in endemic areas [131,132]. However, from the spatial conceptual frame of the edge effect described above, regarding the time the edge itself could be seen as ephemeral, transient, or permanent, and thus, the risk of transmission.

Ephemeral edges are associated with sporadic exposure, as is the case when humans in transit contact an endemic zoonotic cycle, the incidence is scattered in time and space, as it seems to be for ACL autochthonous cases in USA [133], or when researchers, students, officials, hunters, and eco-tourists visiting preserved areas [89,134-139]. ACL cases outside transmission areas, related to tourism, are frequently reported as exotic cases [140-143].

ACL outbreaks associated with military personnel, due to short-term training usually during nocturnal activities, battling, or after deployment, show a narrow shape-common-source epidemic curve [144-155], even reported in dogs involved in the training activities [156]. The larger number of articles about military cases reveals, not only an actual risk, but also better health accessibility than other populations at risk, and also, higher state-based social concern about these communities. In the city outbreaks associated with military deployment, although the number of cases could be as high as more than 3000 individuals, the estimated reproduction number close to one suggests the difficulties of sustaining the transmission cycle in the urban environment [157,158].

Transient edges are associated with temporary exposure due to provisional camps, occasional or ongoing deforestation as in “Fishbone” deforestation [159-161], and seasonal work or recreational activities in forest fragments concurrent with the seasonal peak of abundance of vectors [162,163]. New neighborhoods-related transmission became a transient edge if the incidence decreases after the deforestation event. Many times, these outbreaks are reported as sparse mini-epidemics, within an endemic area [164,165], or family clusters associated with secondary forest-unusable lands clearing for subsistence agriculture [166].

Permanent environmental edges after human settlements near the borders of the city, rural, or agricultural fields, from the “Back to nature” urban trend to smallholding farming, could generate endemic seasonal transmission, with outbreaks due to environmental modifications or exceptional climate events. The peridomestic or intra-domiciliary transmission depends on the edge closeness and the phlebotomine species present, as seen in

the north-central Pacific region of Ecuador, where more than 90% of ACL cases live in farms surrounded by secondary forests and 30% are children below 10 years of age [167], or in the Brazilian Amazon of Manaus, where the intra-domiciliary transmission could be associated with the structural organization of rural settlements [91].

Regarding the lag between the date of the event that contributes to the outbreak and the actual peak of clinical cases, there are considerations about the intrinsic incubation period, estimated from 8 to 22 weeks [123,168], but also periods from 3 months to 12 months due to long-term effects of rainfall on the population dynamics of the vector [15,169,170]. In this sense, extreme El Niño–Southern Oscillation (ENSO) years, with extreme rainfall, could be related to exceptional epidemic transmission, associated with the increase of the forest phlebotomine breeding and resting surfaces and forest growth getting closer the edge to human dwellings [171-176]. Furthermore, for major environmental modifications, migration and change of land use, the eco-epidemiological momentum for an ACL outbreak, could be reached after several years of the causative factors by a trigger threshold [177].

#### **“Why”: Living in the Edge of the Edge**

The presumed original mode of transmission in ephemeral edges with human contact, with sylvatic cycles for forest-related activities, is still one recurrent cause of ACL cases. In old reports, a rural infection in humans and dogs was acquired during excursions into the surrounding forest [178]. However, in recent reports, the visits to primary climax forest, many times after sunset, were usually associated with extractivism (i.e., lumbering, poaching, harvesting forest goods), recreational, training, research activities, and migrants in transit [179,180]. However, when these irregular behaviors change to regular work by farming or gold mining, the borders of the camps, agriculture fields, or settlements of landless workers became transient edges [49,94,132,159,181-193].

Regarding the agricultural frontier, the deforestation and consequent local micro-climatic changes could have the shape of a lineal front as seen in industrial deforestation [100]. Thus, the ACL risk is associated with the times when the people go to the forest fringes to relieve themselves, to rest or to wait until they are transported to their residences; otherwise, the individual deforestation for subsistence farming, many times has a sickle shape so the edge is close to the house in almost all directions [194,195].

It is also noteworthy that although the gender in the transmission associated with male activities is still an important factor to define peridomestic transmission [196], the incidence in women and children could be related to the observation that the entire family goes to the place of work, mainly with babies kept under the shadow of wooden fringes or making temporary

shelters in crop areas; in addition some male have the potential for a higher clinical susceptibility despite actual exposure [197-199], although in scenarios of high exposure this sex bias trends to decrease [139].

Besides, agriculture practices of cacao or coffee as soil-organic management, shadow and humidity distribution, and houses located close or inside the plantations seem to increase the risk of ACL [200-208], when the local Leishmanian ecological system is tolerant to the insertion of non native vegetation. For instance, in the case of Jari [Pará State, Brazil], insertion of *Gmelina* may produce suitable conditions for *L. amazonensis* enzootics and unsuitable for that of *L. guyanensis* [159]. Banana and sugar cane fields in many countries of South America were associated with larger populations of vectors of ACL that were explained by structural microhabitats to rest or carbohydrate food sources [209,210].

Changes in economic trends of global markets could promote the migration of immunological naïve people to endemic areas seeking economic improvement and so pioneering deforestation. Thus, ACL risk is associated with being a farmer migrant living close to the primary forest, in poor housing conditions with poultry and pigs near the house, and their average number of hours away from home, while the risk of ACL decreases when the plantations or nearby villages are far away of the forested areas [49,162,200,211-217]. On the other hand, the monoculture coffee crisis in Parana, Brazil, were associated with an increase in urban ACL reports due to rural to urban migration and seasonal migrant workers [218]. Even the ACL emergence in Ceará, Brazil, during the early twentieth century, was proposed to be due to migration to the Amazon region because of a catastrophic drought and smallpox epidemic, followed by the return of the people when the conditions were restored [219].

Massive migration, unplanned urbanization and poverty, together with ACL emergence and spread, were also associated with developmental projects, such as dams, pipe-lines construction, or trans-oceanic highways across the forest [219-228]. Other sources of migration, together with military activity, included ephemeral transit through the forest, transient camps, and unhealthy periurban housing where the people were displaced due to social disturbances. At the national level when the conflicts were categorized worldwide, ACL incidence was 2.38 times greater between the countries with higher and the lower level of conflict, and it was also significantly different from the lower level to no-conflict countries [229].

In Acre, Brazil, from 2007-2013, the three patterns of ACL transmission were reported simultaneously in different space-time clusters: ephemeral edge by transmission due to extractive activities as chestnut, rubber-tree bleeding, and fishing; transient edge among small-scale farmers close to natural reserves or

riparian forest with peridomestic/intra-domestic occasional transmission; and a transitional situation between the sylvatic and the peridomestic cycle tending to be a permanent edge, the last one as the only growing high-risk cluster, while the reported urban cases were from visitors to the other clusters [130,230,231].

Therefore, answers to the “Why” question show that the anthropogenic drivers are those that modulates the ACL risk in space and time, besides the climate and the environmental vector-reservoir suitability [232]. So, only when social marginality and exclusion are controlled, the climate becomes the most significant factor for ACL [233]. These social determinants include changes in demography and migration, land use, and land value, resulting in landscape edges that are overlapped with social spatial segregation, ethnic discrimination, housing deficiencies, labor practices and unfair regulations, protein–calorie malnutrition, and difficulties accessing the health system, which worsened by ACL treatment itself. There are also macro-factors associated with the political and economic crises, the urban structure, and market fashion trends from gold to the Brazilian chestnut or açai, from periurban marginality to enclosed private neighborhoods in pristine landscapes [234-238]. Therefore, as it was noted above in the section about urban transmission, case-control studies, if paired spatially and controlled by factors actually associated with social determination, could be biased, highlighting secondary individual risk variables instead of collective ones [239].

### “How to Control”: Barricading the Edge

To understand the “Where”, “When”, and “Why” components that come together to generate the eco-epidemiological momentum of an ACL outbreak, at focus and site spatial scales, could contribute to improve the effectiveness of control strategies, by focusing the interventions in the weakest links of the epidemic production chain.

Until 1993 DDT and afterward pyrethroids were empirically used for blocking transmission of endophilic-endophagic vectors of leishmaniasis, or the impact on leishmaniasis’ vectors were “Collateral damage” of blanket spraying for other vector-borne diseases. Different degrees of success were attributed to these uncontrolled interventions [240-245]. Proposed outcomes ranged from dog ACL serology [246] to alleged cases for four years [247]. DDT was also tested outdoors due to its residual efficacy in tree buttresses, where the vector clustered to rest during the day, with a reduction in the vector population for three weeks [248], and in an oily emulsion to avoid the washing-out effect of tropical rains, by obtaining trunks not re-occupied for 11 months, but others re-colonized by vector immigration from nearby non treated areas [249]. Another report of interventions with an impact on ACL was helicopter-based insecticide campaigns, which lacked controls and protocol details [250].

For transient edges, mainly for military camps, some controlled trials that provided short-term chemical barriers in a buffer area (pushing away the edge) were tested. In the rain forest of Panama, the design involved experimental plots of 100 m in diameter, spaced 25 m apart from each other and periodical malathion spraying. However, the small and fluctuant amount of phlebotomine along the study relativized the results [251]. In another controlled trial, again with a buffer area of 100 m, backpack sprayed 25% cyfluthrin, in a palm oil carrier, significantly reduced phlebotomine from reaching the cantonment area, and mainly its center, for more than 80 days [252].

Regarding transient to permanent edges, lambda-dacyhalothrin 10%, at a standard dose of 25 mg/m<sup>2</sup> as Indoor Residual Spraying, was evaluated in El Ingenio, Venezuela, between clustered houses according to its structure, showed short-term residually on walls and a short-term effect on the phlebotomine species *Pintomyia ovallesi* up to 79 days, where the results improved if indoor females and the short season of highest vector abundance were considered [253]. Controlled by locality [nine houses], deltamethrin sprayed on indoor and outdoor walls, roofs, and premises around 10 m at the same 25 mg/m<sup>2</sup> dose reduced the indoor abundance of the species *Nyssomyia intermedia* 32% and *Migonemyia migonei* 42%, significant only in some months, while the intervention had no impact in peridomestic populations, and the pretreatment abundance was more than 50% higher than the highest abundance in the control houses during the study, which weakened the results on effectiveness [242].

Furthermore, any extrapolation should be taken with caution due to species-specific response, as in an ACL focus in Bolivia where the indoor-outdoor walls sprayed with deltamethrin had a beneficial impact against the vectors associated with visceral leishmaniasis, but not on the more exophilic vectors of ACL [254]. In the same sense, with indoor-outdoor thermal fogging with deltamethrin, controlled by house quality and seasonality, the overall indoor phlebotomine decreased from 90% to 40%, but one species, *Ny. trapidoi*, actually increased after fogging 5%, while the quality of the house was the most important factor related to the insecticide effect that lasted for 4 months [255,256]. The substrate factor is also essential for the impact on highly endophilic vectors in the Peruvian Andes scenarios, where lambda-dacyhalothrin sprayed at 34 mg/m<sup>2</sup> every six months, with a follow up of two years, reduced the indoor abundance of the phlebotomine species of interest to 78%-83%, and the susceptible incidence of householders to 81% [202,257]. As a particular case of indoor intervention, the entomopathogenic fungus *Beauveria bassiana* did not demonstrate effectiveness in field applications (coffee plantation) [258]. Nevertheless, when the levels of evidence for different interventions were discriminated by a systematic review, only the Indoor Residual Spraying had moderate level of evidence, while the other control approaches had low level of evidence [259].

Physical and chemical barriers were applied together to “Block the edge” effect in an ACL outbreak in a new village—created by clearing the forest in French Guiana shore—where the sylvatic enzootic cycle had been already described. The actual fringe with active transmission was identified as 12 ha of peripheral residual forest and riparian vegetation, although all the houses of the locality are located less than 250 m from these borders. The physical barrier was a deforested belt of 200 m, together with a chemical barrier of dimethyl-1, 2-dibromo-2, 2-dichloro-ethyl phosphate. No human cases were recorded during the deforestation, or during the next season, while the abundance of phlebotomine and potential reservoirs dropped dramatically inside the village and the borders [260,261].

Regarding environmental management, with or without insecticide spraying, it is usually focused in the micro-environmental management of the household unit. However, as with chemical interventions, the environmental interventions should consider a buffer area according to the estimated dispersal of the vectors, usually taken as a radius of 100 to 250 m. Unfortunately, many times the actual impact of the interventions on the landscape and related cultural practices are not assessed [262]. Further, the environmental management protocols are discontinuous and unstandardized so the results are negligible, and even contradictory as in a 7-year follow up where the vector relative abundance increased [263,264]. In micro-scale studies, such as one conducted in Jacarepaguá, Rio de Janeiro, Brazil, the reduction of animals in the peridomicile and the improvement in the housing were the main interventions that reduced indoor collections of *Ny. intermedia* and *Mg. migonei* [265]. When the cost-effectiveness of different intervention measures were estimated environmental management has a larger initial cost to be instrumented, the larger lag until the results were observed, but the higher and more sustainable impact [266].

Regarding barriers for ephemeral edges, DEET on net jackets or permethrin-treated clothing did not provide protection in skin uncovered areas, while soap with DEET 20% and 0.5% permethrin had short-term effects but indistinguishable from the placebo [267,268]. Conversely, permethrin uniforms at 850 mg/m<sup>2</sup> reduced the incidence of leishmaniasis from 12% in the control group to 3% in uncovered skin areas of the experimental group [269]. Further, pre-exposure preventive education for military personnel (insect repellents, long-sleeve clothing, sleeping in protected areas) lowered the incidence up to no-transmission at all [154,270], while planning the training to avoid the vector’s most abundant season also show effectiveness [271].

The difficulties of some phlebotomine to bite through fabrics were reported by cloth protection [272], but also encouraged the testing of nets and curtains as alternative preventive tools for ephemeral to permanent edges. Pyrethroid-Long Lasting Insecticidal Nets reduced vector landing rates, increased mortality of females in contact with the net, and induces exophily by repellency

[273,274]. Pyrethroid impregnated curtains that did not cover all the phlebotomine entrances, reduced the number of vectors that entered to experimental hen houses, and even non-impregnated curtains had a significant effect with fewer phlebotomine trapped than in the control chicken coops without curtains [275]. In the same sense, in trials controlled by clusters of city sectors, polyester curtains (mesh 0.05 mm) impregnated twice in a year with 12.5 mg/m<sup>2</sup> of lambda-cyhalothrin and loosely hanged, reduced vector indoor trapping, even with open doors, and the incidence after 12 months in the control houses with non-impregnated curtains was 8% while in experimental ones was 0% [276,277]. In another experiment, deltamethrin (26 mg a.i./m<sup>2</sup>) impregnated bed nets and curtains showed fewer phlebotomine caught by human bait under the nets (0.14 phlebotomine/man-hour) than outside the nets in the same room (1.91 phlebotomine/man-hour) or in unprotected rooms (3.29 phlebotomine/man-hour) [278]. A trail controlled by village clustering, which involved nets impregnated with deltamethrin, repellent (20% DEET and 0.5% permethrin) delivered to each residence, whitewash painting of tree bases up to 50 m from the house, and health education, showed the greatest effect in lowering the incidence in children younger than 10 years old and people living on the periphery of the village, but the low incidence of cases also in the control group during the follow up period reduced the statistic significance of the results [279]. Hence, the effectiveness of a net-curtain strategy to lowering the ACL incidence depends on the degree of coverage of the whole human population at risk with intact nets, it requires relatively negligible transmission out of the net during the hours of vector highest activity, and hinges on the total abundance of vectors during peaks, so the proportion that can cross the fabric.

This last issue about the amount of biting females during the epidemic peaks relativizes the extrapolation of modeling or control trials based on abundance data during endemic transmission periods. The population explosions of vectors that produce outbreaks are usually outlier peaks with different dynamics and behavior than those computed from inter-epidemic parameters. Furthermore, the actual abundance of vectors and their infection rates during these outbreaks, despite a proportional reduction by control measures, could still be above the transmission threshold. During inter-epidemic periods in rural areas surrounded by forest in the Orinoquia Region of Colombia, the ratio between indoor:outdoor:forest phlebotomine abundance was 1:7.8:18.3 so a peak of the forest population could increase the transmission despite the reduction of the domestic ones by any intervention, as in Panama where after deltamethrin fogging the vector infection rates increased while the blood sources for the phlebotomine were sustained [280,281]. However, by modeling we can forecast the “Where”, “When”, and “Why” the transmission could take place, which would allow us to focus regular surveillance protocols, in space and time, instead of blanket surveillance strategies what are impractical due to the lack of financial, human, or logistical

resources.

### **Larger Scales: Diluting the Edge Effect**

At the second and third subnational jurisdictional level in Colombia, ACL cases could still be correlated with natural vegetation, mainly, due to riparian forest from the hydrographic corridors and residual forest, permanent crops, and heterogeneous agricultural zones, while urbanization and shrub coverage was negatively associated with ACL prevalence [237,282]. However, at larger modeling scales, the aggregated data and variables that correlated with opposite plus/minus signs could compensate the actual micro-focal trends of individual risk in a kind of “Ecological fallacy”, while big extensions of new agroecosystems or forested areas can dilute the edge effect. Therefore, at these scales, climate could be the driving factor that better explains the distribution of transmission or climate together with vegetation indexes [283-286], rather than anthropic or cultural factors [175].

In the conceptual frame of larger scales, the deforestation would reduce ACL transmission, and so happen during the first half of the twentieth century. Nonetheless, ACL reemerged during 1970-1980, mainly in rural foci along new edges with clustered sources of blood [244,287,288]. Further, climate change forecasting models for the Neotropical region predict that some phlebotomine vector species will reduce their spatial distribution but increase their presence at higher altitudes [289], while other phlebotomine vector species will expand their distribution [290,291], without considering changes in land use that could also be driven by climate change. However, even in broad regional events, such as ENSO, the transmission clustered in some sites, so it is not synchronous in different areas close to each other [175].

Intermediate vegetation density at larger spatial scales could imply at smaller scales, forest remnants and wastelands that are intermingled with human settlements and ongoing changes in land use. Therefore, the positive association of the amount of phlebotomine collected with these intermediate vegetation indexes is related to environmental heterogeneity and edges [83,166,292]. In the same sense, the incidence of ACL at municipality levels was positively associated with the development of a human index instead of deforestation [293], but when residual and gallery forests were identified, the incidence was associated with the size of the deforested and urbanized areas, and thus, with the closeness of the edges to human dwellings [294,295].

## **Conclusion**

### **Focus Research in ACL Outbreaks**

The answers about the “Where”, “When”, “Why” of ACL transmission contribute to refine the questions to ask during the research of an active focus once an outbreak of ACL happens. To weigh each of the confluent factors that generate the eco-

epidemiological momentum of an ACL epidemic, allows evaluating the feasibility and sustainability of the alternative strategies for control.

The questions begin when the ACL event is suspected, or better if confirmed, before any eco-epidemiological field work is initiated. Any useful background information should be gathered to design a research protocol and to estimate the resources needed: potential vectors, reservoirs, and parasite species known in the area, their biology, bionomy, and time-space distribution at different scales from micro-habitat to village focus (i.e., vector endophily-endophagy, reservoir synanthropy), clinical and epidemiological antecedents, the currently reported situation, climatic and geographic data, local available human and logistic resources, and any recorded changes from environment to demography in the last decade. The experts in the local settings and the locals should be included in the team, or at least its perception of the causes of the outbreak event. Regarding the active or recent ACL human cases in this stage of the research, the clinical files should be enough as a source for preliminary information. It is better to avoid asking patients several times with the same topics until the in-depth epidemiological interview, otherwise the reiteration of the questions by agents of the health system tend to reiterative answers of the surveyed person based on preconceived concepts rather than actual antecedents. The clinical files also contribute to identifying the need of parasite characterization if there are differences with the previously reported clinical expression, and the complexity of the entomological studies if there are differences with the known pattern of transmission that require *a priori* a study for incrimination of new potential vectors.

In the field the focus research of the ACL outbreak involves defining the probable edges of transmission, the “Where” question, from the spatial distribution of cases and their autochthony contribution to the potential indoor transmission. The hypotheses that arise from the answers could then be specifically tested by proper controlled entomological protocols, indoor-outdoor collections, and sites nearby the domestic ones, but with better vector suitability. The in-depth interview to the ACL patient and his household members discriminated by gender and age implies to recollect domestic, recreational, and work-related activities before the case clinical expression appearance, period that varies according to the estimated intrinsic period of the parasite species and the date of the outbreak onset. To avoid biasing the answers, it is important to start with the site of perceived transmission before asking specific questions of places visited. The questionnaire should include even sites incidentally visited but with known environmental risk, and also the perception of the ACL case about the place where the transmission happened. These putative places perceived by the community as risky, although the actual risk of transmission at these sites may not have a biological bases, it should also be assessed by entomological captures. The results

of entomological captures should be shared afterward with the community to discuss the places with actual risk of transmission, mainly in the localities where all vector-borne diseases knowledge, perception and attitudes are based on mosquito-arbovirus information.

While the “When” question to discriminate ephemeral and transient of permanent edges contributes to design the optimal period for entomological collections at local levels, it is also asked to understand the timing of the eco-epidemiological momentum and its probability to happen again. Once the “Where” suggests probable areas of transmission, the “When” question can look for any changes or exceptional events in climate, environment (new edges), demography, practices, and activities that could be associated with increasing exposure. To draw the epidemic curve, the probable date of infection should be determined for each case, by computing a mobile average period according to the parasite’s estimated intrinsic cycle, and the recalled start date that clinical signs began. The anamnesis is required to identify and record which was the first manifestation recognized (redness, itching, bumps, ulcer, actual lesion, etc.), and if the date is too vague, to use before-after a milestone-date (Christmas, popular festivity, birthday, etc.). The lag between a trigger event and the outbreak goes from more than a year of delay as for extraordinary climate events, few months as the coming of electricity to town that changes the hours-site of domestic practices, to few days due to a religious assembly at the hour-place of risk that convenes the community. Furthermore, primarily in outbreaks in urban edges, we need to assess if the cases are sporadic or if the incidence has seasonal regularity, as well as if there are clusters with spatial recurrences.

Despite the active search of current and past ACL cases (ulcers and scars) based on the previous questions, and the review of historical clinical files for suspected, undiagnosed cases during the previous years, it is recommended to ask the patients about other cases to drive a “Snow-ball” search approach in the affected community, or even emigrants residing in the community during the transmission event. On the other hand, when the information provided in the interview starts to be redundant, the principle of saturation could be applied.

The “Why” question used to figure out how the “Where-when” changes in the area trigger the eco-epidemiological momentum involves the in-depth interview of the cases, but also other sources and key informants. Additionally, as the review of the literature above suggests, biological factors determine the possibility of an ACL outbreak, but the climate and mainly the social context actually modulates its probability. Therefore, despite the immediate, more visible “Why” determinants on individual lifestyles or health status, the structural ones, such as changes in the trends of economic global, regional and local economic markets, the correlated changes in land use and massive migration of people seeking economic opportunities or safety, compels the

ACL researchers to include a social determination assessment and to advocate for the right of health as a human right [296,297]. From the control standpoint, this public health attitude requires identification of the more vulnerable links of the transmission process that can be feasibly changed in short-, medium- and long-term interventions, and to involve the players within an agency to change them, including the legal responsibility of public and private “Constructor of edges” in prevention, mitigation and control ACL transmission. From a personal standpoint, the social responsibility of the medical entomologist as builder of the risk concepts and as a public health professional must be realized, that besides the immediate recommendations at focal time-space scales, we need to be aware on what Ulrich Beck in “World at risk” [298] and Zygmunt Bauman in “Wasted Lives: modernity and its outcasts” [299] told us about the use of ‘risk’ as a tool for social control, so to avoid claiming the persons for individual solutions to structural problems.

## References

1. WHO (2004) Scientific working group on Leishmaniasis Meeting report. 2-4 February 2004, TDR/SWG/04, Geneva, Switzerland.
2. Hotez PJ (2018) Human Parasitology and Parasitic Diseases: heading Towards 2050. *Adv Parasitol* 100: 29-38.
3. Alvar J, Vélez ID, Bern C, Herrero M, Desjeux P, et al. (2012) Leishmaniasis worldwide and global estimates of its incidence. *PLoS One* 7: e35671.
4. PAHO Leishmaniasis, informe epidemiológico de las Américas, N° 6, febrero 2018 (2018).
5. Rangel EF, Lainson R, Costa SM, Shaw JJ, Carvalho BM (2018) Sand fly vectors of American Cutaneous Leishmaniasis in Brazil. In: Rangel E.F & Shaw J.J. (editors). *Brazilian sand flies: Biology, taxonomy, medical importance and control*. Rio de Janeiro, Brazilian Ministry of Health. Oswaldo Cruz Foundation; 341-380.
6. Machado-Coelho GL, Assunção R, Mayrink W, Caiiffa WT (1999) American cutaneous leishmaniasis in Southeast Brazil: space-time clustering. *Int. J. Epidemiol* 28: 982-989.
7. Soares VB, Almeida AS, Sabroza PC, Vargas WP (2017) Epidemiological surveillance of tegumentary leishmaniasis: local territorial analysis. *Revista de Saúde Pública* 51: 51.
8. Susser M, Susser E (1996) Choosing a future for epidemiology: II. From black box to Chinese boxes and eco-epidemiology. *Am J Public Health* 86: 674-677.
9. Salomón OD, Mastrángelo AV, Santini MS, Liotta DJ, Yadón ZE (2016) La eco-epidemiología retrospectiva como herramienta aplicada a la vigilancia de la leishmaniasis en Misiones, Argentina, 1920-2014. *Revista Panamericana de Salud Pública* 40: 29-39.
10. Rojas CA (2001) An ecosystem approach to human health and the prevention of cutaneous leishmaniasis in Tumaco, Colombia. *Cadernos de Saúde Pública* 17 Suppl: 193-200.
11. Maia-Elkhoury AN, Yadón ZE, Saboyá Díaz MI, de Araújo Lucena FF, Castellanos LG, et al. (2016) Exploring Spatial and Temporal Distribution of Cutaneous Leishmaniasis in the Americas, 2001-2011. *PLoS Negl Trop Dis* 10: e0005086.



12. Salomón OD, Feliciangeli MD, Quintana MG, Afonso MM, Rangel EF (2015) *Lutzomyia longipalpis* urbanisation and control. Memórias do Instituto Oswaldo Cruz 110: 831-846.
13. Hashiguchi Y, Arias O, Maciel D, Mansur J, Furuya M, et al. (1991) Cutaneous leishmaniasis in south-eastern Paraguay: a study of an endemic area at Limoy. Trans R Soc Trop Med Hyg 85: 592-594.
14. Corte AÂ, Nozawa MR, Ferreira Md, Pignatti MG, Rangel O, et al. (1996) Aspectos eco-epidemiológicos da leishmaniose tegumentar americana no Município de Campinas. Cadernos de Saúde Pública 12: 465-472.
15. Feliciangeli MD, Rabinovich J (1998) Abundance of *Lutzomyia ovallesi* but not *Lu. gomezi* (Diptera: Psychodidae) correlated with cutaneous leishmaniasis incidence in north-central Venezuela. Med Vet Entomol 12:121-131.
16. González R, Devera R, Madrid C, Zghayer S (2000) Evaluación de un brote de leishmaniasis tegumentaria americana en una comunidad rural del Estado Bolívar, Venezuela. Revista da Sociedade Brasileira de Medicina Tropical 33:31-37.
17. Azulay RD, Queiroz CM, Ferreira AM, de Oliveira-Neto MP (2001) An interesting outbreak of leishmaniasis in a family in the state of Rio de Janeiro. Int J Dermatol 40: 79-80.
18. Salomón, OD, Sosa Estani S, Canini L, Córdoba Lanús E (2001) Leishmaniasis tegumentaria en un área con niveles epidémicos de transmisión, Salta, Argentina, 1998. Medicina (Buenos Aires) 61: 284-290.
19. Salomón OD, Sosa Estani S, Monzani AS, Studer C (2001) Brote epidémico de leishmaniasis tegumentaria en Puerto Esperanza, provincia de Misiones, 1998. Medicina (Buenos Aires) 61: 385-390.
20. Salomon OD, Zaidenberg M, Burgos R, Heredia VI, Caropresi SL (2001) American cutaneous leishmaniasis outbreak, Tartagal city, province of Salta, Argentina, 1993. Cadernos de Saúde Pública 43:105-108.
21. Salomón OD, Quintana MG, Flores I, Andina AM, Molina S, et al. (2006) Phlebotominae sand flies associated with a tegumentary leishmaniasis outbreak, Tucumán Province, Argentina. Revista da Sociedade Brasileira de Medicina Tropical 39: 341-346.
22. Salomón OD, Quintana MG, Zaidenberg M (2008) Urban distribution of Phlebotominae in a cutaneous leishmaniasis focus, Argentina Memórias do Instituto Oswaldo Cruz 103: 282-287.
23. Garcia AL, Tellez T, Parrado R, Rojas E, Bermudez H, et al. (2007) Epidemiological monitoring of American tegumentary leishmaniasis: molecular characterization of a peridomestic transmission cycle in the Amazonian lowlands of Bolivia. Trans R Soc Trop Med Hyg 101:1208-1213.
24. Nunes WdS, Araújo SR, Calheiros CM (2010) Epidemiological profile of leishmaniasis at a reference service in the state of Alagoas, Brazil, from January 2000 to September 2008. Braz J Infect Dis 14: 342-345.
25. Ramos WR, Medeiros JF, Julião GR, Ríos-Velásquez CM, Marialva EF, et al. (2014) Anthropoc effects on sand fly (Diptera: Psychodidae) abundance and diversity in an Amazonian rural settlement, Brazil. Acta Trop 139: 44-52.
26. Bonfante-Garrido R, Barroeta S, Mejía de Alejos MA, Meléndez E, Arredondo C, et al. (1987) Leishmaniasis tegumentaria urbana en Barquisimeto, Venezuela. Bull Pan Am Health Organ 21: 149-155.
27. Salomón OD, Sosa-Estani S, Ramos K, Orellano PW, Sanguesa G, et al. (2006) Tegumentary leishmaniasis outbreak in Bella Vista City, Corrientes, Argentina during 2003. Memórias do Instituto Oswaldo Cruz 101: 767-774.
28. Salomon OD, Bogado De Pascual M, Molinari ML, Verri V (2001) Study of a cutaneous leishmaniasis outbreak in General Vedia, Province of Chaco, 1996. Cadernos de Saúde Pública 43: 99-104.
29. Cortés LA, Fernández JJ (2008) Especies de *Lutzomyia* en un foco urbano de leishmaniasis visceral y cutánea en El Carmen de Bolívar, Bolívar, Colombia. Biomédica 28: 433-440.
30. Teles CB, Santos AP, Freitas RA, Oliveira AF, Ogawa GM, et al. (2016) Phlebotomine sandfly (Diptera: Psychodidae) diversity and their *Leishmania* DNA in a hot spot of American Cutaneous Leishmaniasis human cases along the Brazilian border with Peru and Bolivia. Memórias do Instituto Oswaldo Cruz 111: 423-432.
31. Souza WJ, Sabroza PC, Santos CS, de Sousa E, Henrique MF, et al. (1992) Montenegro skin tests for American cutaneous leishmaniasis carried out on school children in Rio de Janeiro, Brazil: an indicator of transmission risk. Acta Trop 52: 111-119.
32. Souza NA, Silva JB, Godoy RE, Souza FJ, Andrade-Coelho CA, et al. (2015) Studies on Phlebotominae (Diptera: Psychodidae) in the campus FIOCRUZ mata Atlântica, Jacarepaguá, in the City of Rio de Janeiro, Brazil. Revista da Sociedade Brasileira de Medicina Tropical 48: 26-32.
33. Pedrosa FdA, Ximenes RA (2009) Sociodemographic and environmental risk factors for American Cutaneous Leishmaniasis (ACL) in the State of Alagoas, Brazil. Am J Trop Med Hyg 81: 195-201.
34. Carvalho GM, Gontijo CM, Falcão AL, Andrade Filho JD (2010) Study of phlebotomine sand flies (Diptera: Psychodidae) collected in a *Leishmania*-endemic area of the metropolitan region of Belo Horizonte, Brazil. J Med Entomol 47: 972-976.
35. Silva AF, Latorre MdR, Galati EA (2010) Fatores relacionados à ocorrência de leishmaniose tegumentar no Vale do Ribeira. Revista da Sociedade Brasileira de Medicina Tropical 43: 46-51.
36. Moschin JC, Ovallos FG, Sei IA, Galati EA (2013) Ecological aspects of phlebotomine fauna (Diptera, Psychodidae) of Serra da Cantareira, Greater São Paulo Metropolitan region, state of São Paulo, Brazil. Revista Brasileira de Epidemiologia 16: 190-201.
37. Nascimento BW, Saraiva L, Neto RG, Meira PC, Sanguinette CdC, et al. (2013) Study of sand flies (Diptera: Psychodidae) in visceral and cutaneous leishmaniasis areas in the central-western state of Minas Gerais, Brazil. Acta Trop 125: 262-268.
38. de Souza CF, Quaresma PF, Andrade Filho JD, Bevilacqua PD (2014) Phlebotomine fauna in the urban area of Timóteo, State of Minas Gerais, Brazil. Acta Trop 134: 72-79.
39. Ferreira JVS, Vasconcelos dos Santos T, Santos EM, Gorayeb IS (2014) Phlebotomine sand flies (Diptera: Psychodidae) in forest fragments of Belém metropolitan area, Pará State, Brazil, with considerations on vectors of American cutaneous leishmaniasis agents. Rev Panamazonica Saude 5: 29-35.
40. Carneiro FRO, Amin GA, Cruz LBP, Daher BA (2018) Urban American cutaneous leishmaniasis. Anais Brasileiros de Dermatologia 93: 156-158.

41. Thies SF, Bronzoni RVM, Michalsky EM, Santos ESD, Silva DJFD, et al. (2018) Aspects on the ecology of phlebotomine sand flies and natural infection by *Leishmania hertigi* in the Southeastern Amazon Basin of Brazil. Acta Trop 177: 37-43.
42. Trüeb I, Portela RD, Franke CR, Carneiro IO, Ribeiro Jr GJ, et al. (2018) *Trypanosoma cruzi* and *Leishmania* sp. infection in wildlife from urban rainforest fragments in Northeast. J Wildlife Dis 54: 76-84.
43. Teodoro U, Kühl JB, Rodrigues M, dos Santos ES, dos Santos DR, et al. (1998) Flebotomíneos coletados em matas remanescentes e abrigos de animais silvestres de zoológico no perímetro urbano de Maringá, sul do Brasil. Estudo preliminar. Revista da Sociedade Brasileira de Medicina Tropical 31: 517-522.
44. Bernardes Filho F, Bonatto DC, Martins G, Maier LdM, Nery JA, et al. (2014) Occurrence of two autochthonous cases of American cutaneous leishmaniasis in the neighborhood of Caju, city of Rio de Janeiro, Brazil. Anais Brasileiros de Dermatologia 89: 848-850.
45. Sánchez Uzcátegui YdV, Vasconcelos dos Santos T, Silveira FT, Ramos PKS, Póvoa MM (2019) Phlebotomines (Diptera: Psychodidae) from a urban park of Belém, Pará State, northern Brazil and potential implications in the transmission of American Cutaneous Leishmaniasis. J Med Entomol doi: 10.1093/jme/tjz153. (Sep 24. Epub ahead of print)
46. Kawa H, Sabroza PC (2002) Espacialização da leishmaniose tegumentar na cidade do Rio de Janeiro. Cadernos de Saúde Pública 18: 853-865.
47. Leonardo FS, Rebêlo JM (2004) A periurbanização de *Lutzomyia whitmani* em área de foco de leishmaniose cutânea, no Estado do Maranhão, Brasil. Revista da Sociedade Brasileira de Medicina Tropical 37: 282-284.
48. Aparicio C, Bitencourt MD (2004) Modelagem espacial de zonas de risco da leishmaniose tegumentar americana. Revista de Saúde Pública 38: 511-516.
49. Monteiro WM, Neitzke HC, Silveira TG, Lonardoni VM, Teodoro U, et al. (2009) Pólos de produção de leishmaniose tegumentar americana no norte do Estado do Paraná, Brasil. Cadernos de Saúde Pública 25: 1083-1092.
50. Kawa H, Sabroza PC, Oliveira RM, Barcellos C (2010) A produção do lugar de transmissão da leishmaniose tegumentar: o caso da Localidade Pau da Fome na cidade do Rio de Janeiro, Brasil. Cadernos de Saúde Pública 26: 1495-1507.
51. Passos VM, Falcão AL, Marzochi MC, Gontijo CM, Dias ES, Barbosa-Santos EG, Guerra HL, et al. (1993) Epidemiological aspects of American cutaneous leishmaniasis in a periurban area of the metropolitan region of Belo Horizonte, Minas Gerais, Brazil. Memórias do Instituto Oswaldo Cruz 88: 103-110.
52. Oliveira CC, Lacerda HG, Martins DR, Barbosa JD, Monteiro GR, et al. (2004) Changing epidemiology of American Cutaneous Leishmaniasis (ACL) in Brazil: a disease of the urban-rural interface. Acta Trop 90: 155-162.
53. Dutari LC, Loaiza JR (2014). American Cutaneous Leishmaniasis in Panama: a historical review of entomological studies on anthropophilic *Lutzomyia* sand fly species. Parasit Vectors 7: 218.
54. Bustamante MC, Pereira MJ, Schubach AO, da Fonseca AH (2009) Epidemiological profile of cutaneous leishmaniasis in an endemic region in the State of Rio de Janeiro, Brazil. Revista Brasileira de Parasitologia Veterinária 18: 34-40.
55. Passos VM, Falcão AL, Katz N (1990) Urban American cutaneous leishmaniasis in the Metropolitan Region of Belo Horizonte, Minas Gerais State, Brazil. Memórias do Instituto Oswaldo Cruz 85: 243-244.
56. Bejarano EE, Uribe S, Rojas W, Velez ID (2002) Phlebotomine sand flies (Diptera: Psychodidae) associated with the appearance of urban Leishmaniasis in the city of Sincelejo, Colombia. Memórias do Instituto Oswaldo Cruz 97: 645-647.
57. Agudelo LA, Uribe L, Sierra D, Ruiz F, Velez ID (2002) Presence of American cutaneous Leishmaniasis vectors surrounding the city of Medellín, Colombia. Memórias do Instituto Oswaldo Cruz 97: 641-642.
58. Gil JF, Nasser JR, Cajal SP, Juarez M, Acosta N, et al. (2010) Urban transmission of American cutaneous leishmaniasis in Argentina: spatial analysis study. Am J Trop Med Hyg 82: 433-440.
59. Ballart C, Vidal G, Picado A, Cortez MR, Torrico F, et al. (2016) Intra-domiciliary and peridomiciliary captures of sand flies (Diptera: Psychodidae) in the leishmaniasis endemic area of Chapare province, tropic of Cochabamba, Bolivia. Acta Trop 154: 121-124
60. Carvalho GM, De Vasconcelos FB, Da Silva DG, Botelho HA, Filho JD (2011) Diversity of phlebotomine sand flies (Diptera: Psychodidae) in Ibitipoca State Park, Minas Gerais, Brazil. J Med Entomol 48: 764-769.
61. Carvalho BM, Maximo M, Costa WA, de Santana AL, da Costa SM, et al. (2013) Leishmaniasis transmission in an ecotourism area: potential vectors in Ilha Grande, Rio de Janeiro State, Brazil. Parasit Vectors 6: 325.
62. Brilhante AF, Nunes VL, Kohatsu KA, Galati EA, Rocca ME, Ishikawa EA (2015) Natural infection of phlebotomines (Diptera: Psychodidae) by *Leishmania (Leishmania) amazonensis* in an area of ecotourism in Central-Western Brazil. J Venom Anim Toxins Incl Trop Dis 21: 39.
63. Lana RS, Michalsky EM, Fortes-Dias CL, França-Silva JC, Lara-Silva FdO, et al. (2015) Phlebotomine sand fly fauna and *leishmania* infection in the vicinity of the Serra do Cipó National Park, a natural Brazilian heritage site. Biomed Res Int 2015: 385493.
64. Pereira Filho AA, Bandeira MdC, Fonteles RS, Moraes JL, Lopes CR, et al. (2015) An ecological study of sand flies (Diptera: Psychodidae) in the vicinity of Lençóis Maranhenses National Park, Maranhão, Brazil. Parasit Vectors 8: 442.
65. Saldaña A, Chaves LF, Rigg CA, Wald C, Smucker JE, et al. (2013) Clinical cutaneous leishmaniasis rates are associated with household *Lutzomyia gomezi*, *Lu. panamensis*, and *Lu. trapidoi* abundance in Trinidad de Las Minas, western Panama. Am J Trop Med Hyg 88: 572-574.
66. Benício E, Cordeiro M, Monteiro H, Saboia Moura MA, Oliveira C, et al. (2015) Sustained Presence of Cutaneous Leishmaniasis in Urban Manaus, the Largest Human Settlement in the Amazon. Am J Trop Med Hyg 93: 1208-1213
67. Salomón OD, Acardi SA, Liotta DJ, Fernández MS, Lestani E, et al. (2009) Epidemiological aspects of cutaneous leishmaniasis in the Ig-uazú falls area of Argentina. Acta Trop 109: 5-11.
68. Salomón OD, Orellano PW, Quintana MG, Pérez S, Sosa Estani S, et al. (2006) Transmisión de la leishmaniasis tegumentaria en Argentina Medicina (Buenos Aires) 66: 211-219.

69. Condino ML, Galati EA, Holcman MM, Salum MR, Silva DC, et al. (2008) Leishmaniose tegumentar americana no Litoral Norte Paulista, período 1993 a 2005. *Revista da Sociedade Brasileira de Medicina Tropical* 41: 635-641.
70. Silva NS, Muniz VD (2009) Epidemiologia da leishmaniose tegumentar americana no Estado do Acre, Amazônia brasileira. *Cadernos de Saúde Pública* 25:1325-1336.
71. Silva APOD, Miranda DEO, Santos MAB, Guerra NR, Marques SR, et al. (2017) Phlebotomines in an area endemic for American cutaneous leishmaniasis in northeastern coast of Brazil. *Revista Brasileira de Parasitologia Veterinária* 26: 280-284.
72. Campbell-Lendrum D, Dujardin JP, Martinez E, Feliciangeli MD, Perez JE, et al. (2001) Domestic and peridomestic transmission of American cutaneous leishmaniasis: changing epidemiological patterns present new control opportunities. *Memórias do Instituto Oswaldo Cruz* 96: 159-162.
73. Brasil - Ministry of Health. Secretary of Surveillance in Health. Department of Surveillance in Transmissible Diseases. Guide to surveillance of tegumentary leishmaniasis (in Portuguese). 2nd ed. Brasília: Ministério da Saúde press (2017)
74. Organización Panamericana de la Salud. Manual de procedimientos para vigilancia y control de las leishmaniasis en las Américas. Washington, D.C.: OPS (2019)
75. Firey W (1946) Ecological Considerations in Planning for Rurban Fringes. *American Sociological Review* 11: 411-423.
76. Vilela ML, de Pita-Pereira D, Azevedo CG, Godoy RE, Britto C, et al. (2013) The phlebotomine fauna (Diptera: Psychodidae) of Guarai, state of Tocantins, with an emphasis on the putative vectors of American cutaneous leishmaniasis in rural settlement and periurban areas. *Memórias do Instituto Oswaldo Cruz* 108: 578-585.
77. Rodríguez EM, Díaz F, Pérez MV (2013) Spatio-temporal clustering of American Cutaneous Leishmaniasis in a rural municipality of Venezuela. *Epidemics* 5:11-19.
78. Teodoro U, La Salvia Filho V, de Lima EM, Spinosa RP, Barbosa OC, et al. (1993) Observações sobre o comportamento de flebotomíneos em ecótopos florestais e extraflorestais, em área endêmica de leishmaniose tegumentar americana, no norte do Estado do Paraná, sul do Brasil *Revista de Saúde Pública* 27: 242-249.
79. Travi BL, Adler GH, Lozano M, Cadena H, Montoya-Lerma J (2002) Impact of habitat degradation on phlebotominae (Diptera: Psychodidae) of tropical dry forests in Northern Colombia. *J Med Entomol* 39: 451-456.
80. Zeilhofer P, Kummer OP, Santos ES, Ribeiro AL, Missawa NA (2008) Spatial modelling of *Lutzomyia (Nyssomyia) whitmani* s.l. (Antunes & Coutinho, 1939) (Diptera: Psychodidae: Phlebotominae) habitat suitability in the state of Mato Grosso, Brazil. *Memórias do Instituto Oswaldo Cruz* 103: 653-660.
81. Cerino DA, Teodoro U, Silveira TG (2009) Sand flies (Diptera: Psychodidae) in the urban area of the municipality of Cianorte, Paraná State, Brazil. *Neotrop Entomol* 38: 853-858.
82. Brandão-Filho SP, Donalisio MR, da Silva FJ, Valença HF, Costa PL, et al. (2011) Spatial and temporal patterns of occurrence of *Lutzomyia* sand fly species in an endemic area for cutaneous leishmaniasis in the Atlantic Forest region of northeast Brazil. *J Vector Ecol* 36 Suppl 1: S71-76.
83. Valderrama A, Tavares MG, Andrade Filho JD (2011) Anthropogenic influence on the distribution, abundance and diversity of sandfly species (Diptera: Phlebotominae: Psychodidae), vectors of cutaneous leishmaniasis in Panama. *Memórias do Instituto Oswaldo Cruz* 106: 1024-1031
84. Donalisio MR, Peterson AT, Costa PL, da Silva FJ, Valença HF, et al. (2012) Microspatial distributional patterns of vectors of cutaneous leishmaniasis in pernambuco, northeastern Brazil. *J Trop Med* 2012: 642910.
85. Campos AM, Matavelli R, Santos CL, Moraes LS, Rebêlo JM (2013) Ecology of phlebotomines (Diptera: Psychodidae) in a transitional area between the Amazon and the Cerrado in the State of Maranhão, Brazil. *J Med Entomol* 50: 52-58.
86. Ferreira, AL, Falqueto A, Grimaldi Jr G, Peixoto AA, Pinto IdS (2013) Ecological and epidemiological aspects of the sand fly (Diptera, Psychodidae) fauna of the National Monument of Pontões Capixabas, State of Espírito Santo, Southeastern Brazil. *J Med Entomol* 50: 1215-1223.
87. Aguiar GM, de Azevedo AC, Medeiros WM, Alves JR, Rendeiro V (2014) Aspects of the ecology of phlebotomines (Diptera: Psychodidae: Phlebotominae) in an area of cutaneous leishmaniasis occurrence, municipality of Angra dos Reis, coast of Rio de Janeiro State, Brazil. *Revista do Instituto de Medicina Tropical de São Paulo* 56: 143-149.
88. Rêgo FD, Shimabukuro PHF, Quaresma PF, Coelho IR, Tonelli GB, et al. (2014) Ecological aspects of the Phlebotominae fauna (Diptera: Psychodidae) in the Xakriabá Indigenous Reserve, Brazil. *Parasit Vectors* 7: 220.
89. Saraiva L, Silva Reis A, Marteleto Nunes Rugani J, Sampaio Pereira AA, Rêgo FD, et al. (2015) Survey of sand flies (Diptera: Psychodidae) in an environmentally protected area in Brazil. *PLoS One* 10: e0134845.
90. Vieira VR, Azevedo AC, Alves JR, Guimarães AE, Aguiar GM (2015) Ecological Aspects of Phlebotomine Sand Flies (Diptera, Psychodidae, Phlebotominae) in Areas of American Cutaneous Leishmaniasis, in the Municipality of Paraty, Rio de Janeiro, Brazil. I-Index of Abundance by Location and Type of Capture. *J Med Entomol* 52: 886-895.
91. Chagas ECDS, Silva AS, Fé NF, Ferreira LS, Sampaio VS, et al. (2018) Composition of sand fly fauna (Diptera: Psychodidae) and detection of *Leishmania* DNA (Kinetoplastida: Trypanosomatidae) in different ecotopes from a rural settlement in the central Amazon, Brazil. *Parasit Vectors* 11: 180.
92. Resadore F, Júnior AMP, de Paulo PMF, Gil LHS, de Souza Rodrigues MM, et al. (2019) Composition and Vertical Stratification of Phlebotomine Sand Fly Fauna and the Molecular Detection of *Leishmania* in Forested Areas in Rondônia State Municipalities, Western Amazon, Brazil. *Vector Borne Zoonotic Dis* 19: 347-357
93. Szelag EA, Rosa JR, Quintana MG, Salomon OD (2018) Temporal distribution of, and effect of anthropic modifications on, phlebotomine populations in the Chaco Bioregion, Argentina. *Med Vet Entomol* 32: 206-215.
94. Vasconcelos Dos Santos T, Prévot G, Ginouvès M, Duarte R, Silveira FT, et al. (2018) Ecological aspects of Phlebotomines (Diptera: Psychodidae) and the transmission of American cutaneous leishmaniasis agents in an Amazonian/ Guianan bordering area. *Parasit Vectors* 11: 612.

95. McDermott EG, Mullens BA (2018) The Dark Side of Light Traps. J Med Entomol 55: 251-261.
96. Colla-Jacques FE, Casanova C, Prado AP (2010) Study of sand fly fauna in an endemic area of American cutaneous leishmaniasis and canine visceral leishmaniasis in the municipality of Espírito Santo do Pinhal, São Paulo, Brazil. Memórias do Instituto Oswaldo Cruz 105: 208-215.
97. Figueira EA, Silva G, Chagas EC, Shimabukuro PH (2013) Phlebotomine sandflies (Diptera: Psychodidae) from Lábrea, state of Amazonas, Brazil, with a description of *Evandromyia (Aldamyia) apurinan* Shimabukuro, Figueira & Silva, sp. nov. Memórias do Instituto Oswaldo Cruz 108: 280-287.
98. Casanova C, Costa AI, Natal D (2005) Dispersal pattern of the sand fly *Lutzomyia neivai* (Diptera: Psychodidae) in a cutaneous leishmaniasis endemic rural area in Southeastern Brazil. Memórias do Instituto Oswaldo Cruz 100: 719-724.
99. Caja Rivera R, Barradas I (2018) Vector Preference Annihilates Backward Bifurcation and Reduces Endemicity. Bull. Math. Biol.
100. Quintana MG, Salomón OD, de Grosso MS (2010) Distribution of phlebotomine sand flies (Diptera: Psychodidae) in a primary forest-crop interface, Salta, Argentina. J Med Entomol 47:1003-1010.
101. Weigle KA, Saravia NG, de Dávalos M, Moreno LH, D'Alessandro A (1986) *Leishmania braziliensis* from the Pacific coast region of Colombia: foci of transmission, clinical spectrum and isoenzyme phenotypes. Am J Trop Med Hyg 35: 722-731.
102. Fonseca Eda S, D'Andrea LA, Taniguchi HH, Hiramoto RM, Tolezano JE, et al. (2014) Spatial epidemiology of American cutaneous leishmaniasis in a municipality of west São Paulo State, Brazil. J Vector Borne Dis 51: 271-275.
103. Haydon DT, Cleaveland S, Taylor LH, Laurenson MK (2002) Identifying reservoirs of infection: a conceptual and practical challenge. Emerg Infect Dis 8: 1468-1473.
104. Quaresma PF, Rêgo FD, Botelho HA, da Silva SR, Moura Júnior AJ, et al. (2011) Wild, synanthropic and domestic hosts of *Leishmania* in an endemic area of cutaneous leishmaniasis in Minas Gerais State, Brazil. Trans R Soc Trop Med Hyg 105: 579-585.
105. Lima BS, Dantas-Torres F, de Carvalho MR, Marinho-Junior JF, de Almeida Brito ME, et al. (2013) Small mammals as hosts of *Leishmania* spp. in a highly endemic area for zoonotic leishmaniasis in Northeastern Brazil. Trans R Soc Trop Med Hyg 107: 592-597.
106. Andrade MS, Courtenay O, Brito ME, Carvalho FG, Carvalho AW, et al. (2015) Infectiousness of Sylvatic and Synanthropic Small Rodents Implicates a Multi-host Reservoir of *Leishmania (Viannia) braziliensis*. PLoS Negl Trop Dis 9: e0004137.
107. Miranda C, Marques CC, Massa JL (1998) Sensoriamento remoto orbital como recurso para análise da ocorrência da leishmaniose tegumentar americana em localidade urbana da região Sudeste do Brasil. Revista de Saúde Pública. 32: 455-463.
108. Guerra JA, Barbosa MD, Loureiro AC, Coelho CP, Rosa GG, et al. (2007) Leishmaniose tegumentar americana em crianças: aspectos epidemiológicos de casos atendidos em Manaus, Amazonas, Brasil. Cadernos de Saúde Pública 23: 2215-2223.
109. Nasser JT, Donalísio MR, Vasconcelos CH (2009) Distribuição espacial dos casos de leishmaniose tegumentar americana no município de Campinas, Estado de São Paulo, no período de 1992 a 2003. Rev Soc Bras Med Trop 42: 309-314.
110. Cruz CF, Cruz MF, Galati EA (2013) Sandflies (Diptera: Psychodidae) in rural and urban environments in an endemic area of cutaneous leishmaniasis in southern Brazil. Memórias do Instituto Oswaldo Cruz 108: 303-311.
111. Chanampa MDM, Gleiser RM, Hoyos CL, Copa GN, Mangudo C, et al. (2018) Vegetation Cover and Microspatial Distribution of Sand Flies (Diptera: Psychodidae) in an Endemic Locality for Cutaneous Leishmaniasis in Northern Argentina. J Med Entomol 55: 1431-1439.
112. Thies SF, Bronzoni RV, Espinosa MM, Souza CO, Ribeiro AL, et al. (2016) Frequency and diversity of phlebotomine sand flies (Diptera: Psychodidae) in Sinop, State of Mato Grosso, Brazil. Revista da Sociedade Brasileira de Medicina Tropical 49: 544-552.
113. Teles CB, Medeiros JF, Santos AP, Freitas LA, Katsuragawa TH, et al. (2015) Molecular characterization of American Cutaneous Leishmaniasis in the tri-border area of Assis Brasil, Acre State, Brazil. Revista do Instituto de Medicina Tropical de São Paulo 57: 343-347.
114. Thomaz-Soccol V, Gonçalves AL, Piechnik CA, Baggio RA, Boeger WA, et al. (2018) Hidden danger: Unexpected scenario in the vector-parasite dynamics of leishmaniasis in the Brazil side of triple border (Argentina, Brazil and Paraguay). PLoS Negl Trop Dis 12: e0006336.
115. Santini MS, Fernández MS, Cavia R, Salomón OD (2018) Co-occurrence and seasonal and environmental distributions of the sandflies *Lutzomyia longipalpis* and *Nyssomyia whitmani* in the city of Puerto Iguazú, northeastern Argentina. Med Vet Entomol 32: 197-205.
116. O'Neal SE, Guimarães LH, Machado PR, Alcântara L, Morgan DJ, et al. (2007) Influence of helminth infections on the clinical course of and immune response to *Leishmania braziliensis* cutaneous leishmaniasis. J Infect Dis 195: 142-148.
117. Braga LdS, Navasconi TR, Leatte EP, Skraba CM, Silveira TG, et al. (2015) Presence of anti-*Leishmania (Viannia) braziliensis* antibodies in blood donors in the West-Central region of the State of Paraná, Brazil. Revista da Sociedade Brasileira de Medicina Tropical 48: 622-625.
118. Azeredo-Coutinho RB, Pimentel MI, Zanini GM, Madeira MF, Cataldo JI, et al. (2016) Intestinal helminth coinfection is associated with mucosal lesions and poor response to therapy in American tegumentary leishmaniasis. Acta Trop 154: 42-49.
119. Castellucci L, Cheng LH, Araújo C, Guimarães LH, Lessa H, et al. (2005) Familial aggregation of mucosal leishmaniasis in northeast Brazil. Am J Trop Med Hyg 73: 69-73.
120. Ampuero J, Urdaneta M, Macêdo VdO (2005) Factores de riesgo para la transmisión de leishmaniasis cutánea en niños de 0 a 5 años en un área endémica de *Leishmania (Viannia) braziliensis*. Cadernos de Saúde Pública 21: 161-170.
121. Bustamante M, Diaz M, Espinoza J, Parrado R, Reithinger R, et al. (2012) Sand fly fauna in Chapare, Bolivia: an endemic focus of *Leishmania (Viannia) braziliensis*. J Med Entomol 49: 1159-1162.
122. Alexander B, Jaramillo C, Usma MC, Quesada BL, Cadena H, et al. (1995) An attempt to control Phlebotomine sand flies (Diptera: Psychodidae) by residual spraying with deltamethrin in a Colombian village. Memórias do Instituto Oswaldo Cruz 90: 421-424.
123. Ferro C, Marín D, Góngora R, Carrasquilla MC, Trujillo JE, et al. (2011) Phlebotomine vector ecology in the domestic transmission of American cutaneous leishmaniasis in Chaparral, Colombia. Am J Trop Med Hyg 85: 847-856.

124. Oliveira-Neto MP, Pirmez C, Rangel E, Schubach A, Grimaldi Júnior G (1988) An outbreak of american cutaneous leishmaniasis (*Leishmania braziliensis braziliensis*) in a periurban area of Rio de Janeiro city, Brazil: clinical and epidemiological studies. Memórias do Instituto Oswaldo Cruz 83: 427-435.
125. Sosa-Estani S, Segura EL, Gomez A, Salomón OD, Peralta M, et al. (2001) Leishmaniose cutânea no Norte da Argentina. Fatores de risco identificados num estudo caso-coorte em três municípios de Salta. Revista da Sociedade Brasileira de Medicina Tropical 34: 511-517.
126. Meneses CR, de Azevedo AC, da Costa SM, Costa WA, Rangel EF (2002) Ecology of American cutaneous leishmaniasis in the state of Rio de Janeiro, Brazil. J Vector Ecol 27: 207-214.
127. Yadon ZE, Rodrigues LC, Davies CR, Quigley MA (2003) Indoor and peridomestic transmission of American cutaneous leishmaniasis in northwestern Argentina: a retrospective case-control study. Am J Trop Med Hyg 68: 519-526.
128. Membrive NA, Rodrigues G, Gualda KP, Bernal MV, Oliveira DM, et al. (2012) Environmental and animal characteristics as factors associated with American cutaneous leishmaniasis in rural locations with presence of dogs, Brazil. PLoS One 7: e47050.
129. Vieira VP, Ferreira AL, Biral dos Santos C, Leite GR, Ferreira GE, et al. (2012) Peridomestic breeding sites of phlebotomine sand flies (Diptera: Psychodidae) in an endemic area of American cutaneous leishmaniasis in southeastern Brazil. Am. J. Trop. Med. Hyg. 87: 1089-1093.
130. Guerra JAO, Vale Barbosa Guerra MdG, Vasconcelos ZS, da Silva Freitas N, Rodrigues Fonseca F, et al. (2019) Socioenvironmental aspects of the Purus Region - Brazilian Amazon: Why relate them to the occurrence of American Tegumentary Leishmaniasis. PLoS One 14: e0211785.
131. Silva J, Queiroz A, Moura I, Sousa RS, Guimarães LH, et al. (2017) Dynamics of American tegumentary leishmaniasis in a highly endemic region for *Leishmania (Viannia) braziliensis* infection in northeast Brazil. PLoS Negl Trop Dis 11: e0006015.
132. Grangeiro Júnior CRP, Pimentel JVC, Teixeira Júnior AG, Jesus AF, Galvão TCF, et al. (2018) American cutaneous leishmaniasis in a northeast Brazilian city: clinical and epidemiological features. Revista da Sociedade Brasileira de Medicina Tropical 51: 837-842.
133. McIlwee BE, Weis SE, Hosler GA (2018) Incidence of Endemic Human Cutaneous Leishmaniasis in the United States. JAMA Dermatol. 154: 1032-1039.
134. Desjeux P, Mollinedo S, Le Pont F, Paredes A, Ugarte G (1987) Cutaneous leishmaniasis in Bolivia. A study of 185 human cases from Alto Beni (La Paz Department). Isolation and isoenzyme characterization of 26 strains of *Leishmania braziliensis braziliensis*. Trans R Soc Trop Med Hyg 81: 742-746.
135. Andrade-Narvaez F, Canto-Lara SB, Del Rosario Garcia-Miss M (2009) Leishmaniasis entomological field studies: ethical issues. Dev World Bioeth 9: 157-160.
136. Felinto de Brito ME, Andrade MS, de Almeida EL, Medeiros AC, Werkhäuser RP, et al. (2012) Occupationally acquired american cutaneous leishmaniasis. Case Rep Dermatol Med 2012: 279517.
137. Krolewiecki AJ, Gil JF, Quipildor M, Cajal SP, Pravia C, et al. (2013) Restricted outbreak of American tegumentary leishmaniasis with high microfocal transmission. Am J Trop Med Hyg 88: 578-582.
138. Martin-Blondel G, Iriart X, El Baidouri F, Simon S, Mills D, et al. (2015) Outbreak of *Leishmania braziliensis* Cutaneous Leishmaniasis, Saül, French Guiana. Emerg Infect Dis 21: 892-894.
139. Soares L, Abad-Franch F, Ferraz G (2014) Epidemiology of cutaneous leishmaniasis in central Amazonia: a comparison of sex-biased incidence among rural settlers and field biologists. Trop Med Int Health 19: 988-995.
140. Zlotogorski A, Gilead L, Jonas F, Horev L, Klaus SN (1998) South American cutaneous leishmaniasis: report of ten cases in Israeli travelers. J Eur Acad Dermatol Venereol 11: 32-36.
141. Maguire GP, Bastian I, Arianayagam S, Bryceson A, Currie BJ (1998) New World cutaneous leishmaniasis imported into Australia. Pathology 30: 73-76.
142. Lawn SD, Whetham J, Chiodini PL, Kanagalingam J, Watson J, et al. (2004) New world mucosal and cutaneous leishmaniasis: an emerging health problem among British travellers. QJM 97: 781-788.
143. Schwartz E, Hatz C, Blum J (2006) New world cutaneous leishmaniasis in travellers. Lancet Infect Dis 6: 342-349.
144. Sanchez JL, Diniega BM, Small JW, Miller RN, Andujar JM, et al. (1992) Epidemiologic investigation of an outbreak of cutaneous leishmaniasis in defined geographic focus of transmission. Am J Trop Med Hyg 47: 47-54.
145. Hepburn NC, Tidman MJ, Hunter JA (1993) Cutaneous leishmaniasis in British troops from Belize. Br J Dermatol 128: 63-68.
146. Banzet S (2000) Leishmaniose cutanee chez les militaires en operation en Guyane Francaise, Revue du Corps de Santé Colonial 60: 297-302.
147. Berger F, Romary P, Brachet D, Rapp C, Imbert P, et al. (2006) Épidémie de leishmaniose cutanée chez des militaires de retour de mission en Guyane. Revue d'épidémiologie et de santé publique 54: 213-221.
148. Andrade MS, Brito ME, Silva ST, Ishikawa E, Carvalho SM, et al. (2009) Novo surto de leishmaniose tegumentar americana em área de treinamento militar na Zona da Mata norte do Estado de Pernambuco. Revista da Sociedade Brasileira de Medicina Tropical 42: 594-596.
149. Bailey MS (2011) Cutaneous leishmaniasis in British troops following jungle training in Belize. Travel Med Infect Dis 9: 253-254.
150. van Thiel PP, Zeegelaar JE, van Gool T, Faber WR, Kager PA (2011) Cutaneous leishmaniasis in three Dutch military cohorts following jungle training in Belize. Travel Med Infect Dis 9: 153-160.
151. Gomes LH, Albuquerque MI, Rocha LC, Pinheiro FG, Franco AM (2013) Diversity and distribution of sandflies (Diptera: Psychodidae: Phlebotominae) in a military area in the state of Amazonas, Brazil. Memórias do Instituto Oswaldo Cruz 108: 651-656
152. Andrade MS, Valença HF, da Silva AL, Almeida FdA, Almeida EL, et al. (2005) Sandfly fauna in a military training area endemic for American tegumentary leishmaniasis in the Atlantic Rain Forest region of Pernambuco, Brazil. Cadernos de Saúde Pública 21: 1761-1767.
153. Andrade MS, Brito ME, Silva ST, Lima BS, Almeida EL, et al. (2005) Leishmaniose tegumentar americana causada por *Leishmania (Viannia) braziliensis*, em área de treinamento militar na Zona da Mata de Pernambuco, Revista da Sociedade Brasileira de Medicina Tropical 38: 229-233.

154. Oré M, Sáenz E, Cabrera R, Sanchez JF, De Los Santos MB, et al. (2015) Outbreak of Cutaneous Leishmaniasis in Peruvian Military Personnel Undertaking Training Activities in the Amazon Basin, 2010. Am J Trop Med Hyg 93: 340-346.
155. Dantas-Torres F, Sales KG, Miranda DE, da Silva FJ, Figueredo LA, et al. (2017) Sand fly population dynamics and cutaneous leishmaniasis among soldiers in an Atlantic forest remnant in northeastern Brazil. PLoS Negl Trop Dis 11: e0005406.
156. Vélez ID, Carrillo LM, López L, Rodríguez E, Robledo SM (2012) An epidemic outbreak of canine cutaneous leishmaniasis in Colombia caused by *Leishmania braziliensis* and *Leishmania panamensis*. Am J Trop Med Hyg 86: 807-811.
157. Mubayi A, Paredes M, Ospina JA (2018) A Comparative Assessment of Epidemiologically Different Cutaneous Leishmaniasis Outbreaks in Madrid, Spain and Tolima, Colombia: An Estimation of the Reproduction Number via a Mathematical Model. Trop Med Infect Dis 3: 43.
158. Patino LH, Mendez C, Rodriguez O, Romero Y, Velandia D, et al. (2017) Spatial distribution, *Leishmania* species and clinical traits of Cutaneous Leishmaniasis cases in the Colombian army. PLoS Negl Trop Dis 11: e0005876
159. Ready PD, Lainson R, Shaw JJ (1983) Leishmaniasis in Brazil: XX. Prevalence of "Enzootic rodent leishmaniasis" (*Leishmania mexicana amazonensis*), and apparent absence of "Pian bois" (*Le. braziliensis guyanensis*), in plantations of introduced tree species and in other non-climax forests in eastern Amazônia. Trans R Soc Trop Med Hyg 77: 775-785.
160. Wong L, Netto EM, Wiese K, França F, Cuba CC, et al. (1986) Unusual prevalence of *Leishmania braziliensis braziliensis* in four families. Revista da Sociedade Brasileira de Medicina Tropical 19: 195-196.
161. Costa JM, Osaho NK, Vale KC, Lago EL, França F, et al. (1986) Ocorrência familiar da Leishmaniose Tegumentar americana em uma região endêmica, Corte de Pedra, Bahia. Revista da Sociedade Brasileira de Medicina Tropical 19: 197-198.
162. Brilhante AF, Dorval ME, Galati EA, da Rocha HC, Cristaldo G, et al. (2015) Phlebotomine fauna (diptera: psychodidae) in an area of fishing tourism in Central-Western Brazil. Revista do Instituto de Medicina Tropical de São Paulo 57: 233-238.
163. Carvalho BM, Dos Santos TV, da R Barata I, Lima JAN, Silveira FT, Vale MM, et al. (2018) Entomological surveys of *Lutzomyia flaviscutellata* and other vectors of cutaneous leishmaniasis in municipalities with records of *Leishmania amazonensis* within the Bragança region of Pará State, Brazil. J Vector Ecol 43:168-178.
164. Jones TC, Johnson Jr WD, Barretto AC, Lago E, Badaro R, et al. (1987) Epidemiology of American cutaneous leishmaniasis due to *Leishmania braziliensis braziliensis*. J Infect Dis 156: 73-83.
165. García Bustos, MF, González-Prieto G, Ramos F, Mora MC, Hashiguchi Y, et al. (2016) Clinical and epidemiological features of leishmaniasis in northwestern-Argentina through a retrospective analysis of recent cases. Acta Trop 154: 125-132.
166. Temponi AOD, Brito MG, Ferraz ML, Diniz SA, Silva MX, et al. (2018) Ocorrência de casos de leishmaniose tegumentar americana: uma análise multivariada dos circuitos espaciais de produção, Minas Gerais, Brasil, 2007 a 2011. Cadernos de Saúde Pública 34: e00165716.
167. Gomez EA, Kato H, Torres-Romero EX, Velez LN, Villegas NV, et al. (2018) Leishmaniasis caused by *Leishmania (Viannia) guyanensis* in north-central Pacific region of Ecuador: A clinico-epidemiological feature. Acta Trop 185: 204-211.
168. Chaves LF (2009) Climate and recruitment limitation of hosts: the dynamics of American cutaneous leishmaniasis seen through semi-mechanistic seasonal models. Ann Trop Med Parasitol 103: 221-234.
169. Salomón OD, Wilson ML, Munstermann LE, Travi BL (2004) Spatial and temporal patterns of phlebotomine sand flies (Diptera: Psychodidae) in a cutaneous leishmaniasis focus in northern Argentina. J Med Entomol 41: 33-39.
170. Lewnard JA, Jirmanus L, Júnior NN, Machado PR, Glesby MJ, et al. (2014) Forecasting temporal dynamics of cutaneous leishmaniasis in Northeast Brazil. PLoS Negl Trop Dis 8: e3283.
171. Chaves LF, Pascual M (2006) Climate cycles and forecasts of cutaneous leishmaniasis, a nonstationary vector-borne disease. PLoS Med 3: e295.
172. Cardenas R, Sandoval CM, Rodríguez-Morales AJ, Franco-Paredes C (2006) Impact of climate variability in the occurrence of leishmaniasis in northeastern Colombia. Am J Trop Med Hyg 75: 273-277.
173. Chaves LF, Calzada JE, Valderrama A, Saldaña A (2014) Cutaneous leishmaniasis and sand fly fluctuations are associated with El Niño in Panamá. PLoS Negl Trop Dis 8: e3210.
174. Ferreira de Souza RA, Andreoli RV, Toshie Kayano M, Lima Carvalho A (2015) American cutaneous leishmaniasis cases in the metropolitan region of Manaus, Brazil: association with climate variables over time. Geospat Health 10: 314.
175. Yamada K, Valderrama A, Gottdenker N, Cerezo L, Minakawa N, et al. (2016) Macroecological patterns of American Cutaneous Leishmaniasis transmission across the health areas of Panamá (1980-2012). Parasite Epidemiol Control 1: 42-55.
176. Simon S, Nacher M, Carme B, Basurko C, Roger A, et al. (2017) Cutaneous leishmaniasis in French Guiana: revising epidemiology with PCR-RFLP. Trop Med Health 45: 5.
177. Azevedo AC, Souza NA, Meneses CR, Costa WA, Costa SM, et al. (2002) Ecology of sand flies (Diptera: psychodidae: phlebotominae) in the north of the state of Mato Grosso, Brazil. Memórias do Instituto Oswaldo Cruz 97: 459-464.
178. Herrero A, Christensen HA, Beumer RJ (1976) Epidemiological patterns of cutaneous leishmaniasis in Panama. II. Incidental occurrence of cases in non-endemic settlements. Ann Trop Med Parasitol 70: 67-71.
179. Barry MA, Koshelev MV, Sun GS, Grekin SJ, Stager CE, et al. (2014) Cutaneous leishmaniasis in Cuban immigrants to Texas who traveled through the Darién Jungle, Panama. Am J Trop Med Hyg 91: 345-347.
180. Cannella AP, Nguyen BM, Piggott CD, Lee RA, Vinetz JM, et al. (2011) A cluster of cutaneous leishmaniasis associated with human smuggling. Am J Trop Med Hyg 84: 847-850.
181. Low-A-Chee RM, Rose P, Ridley DS (1983) An outbreak of cutaneous leishmaniasis in Guyana: epidemiology, clinical and laboratory aspects. Ann Trop Med Parasitol 77: 255-260.
182. Weigle KA, Santrich C, Martinez F, Valderrama L, Saravia NG (1993) Epidemiology of cutaneous leishmaniasis in Colombia: environmental and behavioral risk factors for infection, clinical manifestations, and pathogenicity. J Infect Dis 168: 709-714.
183. Bermúdez H, Torrico F, Rojas E, Balderrama F, Le Ray D, et al. (1993) Leishmaniasis in the lowlands of Bolivia, prevalence of the disease in two groups of localities with different settlement ages in Carrasco Tropical, Cochabamba. Archives de l'Institut Pasteur de Tunis 70: 443-453.

184. Coimbra Júnior CE, Santos RV, do Valle AC (1996) Cutaneous leishmaniasis in Tupi-Mondé Amerindians from the Brazilian Amazonia. *Acta Trop* 61: 201-211.
185. Rebollar-Téllez EA, Ramírez-Fraire A, Andrade-Narvaez FJ (1996) A two years study on vectors of cutaneous Leishmaniasis. Evidence for sylvatic transmission cycle in the state of Campeche, Mexico. *Memórias do Instituto Oswaldo Cruz* 91: 555-560.
186. Rawlins SC, Tiwari T, Chadee DD, Validum L, Alexander H, et al. (2001) American cutaneous leishmaniasis in Guyana, South America. *Ann Trop Med Parasitol* 95: 245-251.
187. Rotureau B, Joubert M, Clyti E, Djossou F, Carme B (2006) Leishmaniasis among gold miners, French Guiana. *Emerg. Infect. Dis* 12:1169-1170.
188. Fouque F, Gaborit P, Issaly J, Carinci T, Gantier JC, et al. (2007) Phlebotomine sand flies (Diptera: Psychodidae) associated with changing patterns in the transmission of the human cutaneous leishmaniasis in French Guiana. *Memórias do Instituto Oswaldo Cruz* 102: 35-40.
189. Rotureau B, Couppié P, Nacher M, Dedet JP, Carme B (2007) Les leishmanioses cutanées en Guyane française. *Bulletin de la Société de pathologie exotique* 100: 251-251.
190. Kent AD, Dos Santos TV, Gangadin A, Samjhawan A, Mans DR, et al. (2013) Studies on the sand fly fauna (Diptera: Psychodidae) in high-transmission areas of cutaneous leishmaniasis in the Republic of Suriname. *Parasit Vectors* 6: 318.
191. Thies SF, Ribeiro AL, Michalsky É, Miyazaki RD, Fortes-Dias CL, et al. (2013) Phlebotomine sandfly fauna and natural *Leishmania* infection rates in a rural area of Cerrado (tropical savannah) in Nova Mutum, State of Mato Grosso in Brazil. *Revista da Sociedade Brasileira de Medicina Tropical* 46: 293-298.
192. Gosch CS, Marques CP, Resende BS, Souza JDS, Rocha RADS, et al. (2017) American tegumentary leishmaniasis: epidemiological and molecular characterization of prevalent *Leishmania* species in the State of Tocantins, Brazil, 2011-2015. *Revista do Instituto de Medicina Tropical de São Paulo* 59: e91.
193. Detoni MB, Lima DM, Silva TPD, Machado LF, Tomiotto-Pellissier F, et al. (2019) Temporal and spatial distribution of American tegumentary leishmaniasis in north Paraná: 2010-2015 *Revista da Sociedade Brasileira de Medicina Tropical* 52: e20180119.
194. Mastrángelo AV, Salomón OD (2009) Trabajo forestal y Leishmaniasis Cutánea: Un análisis social centrado en el riesgo para el N de Misiones (Argentina). *Talleres. ULA-Inst Experimental JW Torrealba, Mérida, Venezuela* 12: 60-68.
195. Mastrángelo AV, Salomón OD (2010) Contribución de la antropología a la comprensión ecoepidemiológica de la Leishmaniasis Tegumentaria Americana en las "2000 hectáreas", Puerto Iguazú, Misiones, Argentina. *Revista Argentina de Salud Pública* 1: 6-13.
196. Eid D, Guzman-Rivero M, Rojas E, Goicolea I, Hurtig AK, et al. (2018) Risk factors for cutaneous leishmaniasis in the rainforest of Bolivia: a cross-sectional study. *Trop Med Health* 46: 9.
197. Davies CR, Llanos-Cuentas EA, Campos P, Monge J, Villaseca P, et al. (1997) Cutaneous leishmaniasis in the Peruvian Andes: risk factors identified from a village cohort study. *Am J Trop Med Hyg* 56: 85-95.
198. Costa JM, Balby IT, Rocha EJ, da Silva AR, Rebêlo JM, et al. (1998) Estudo comparativo da leishmaniose tegumentar americana em crianças e adolescentes procedentes das áreas endêmicas de Buriticupu (Maranhão) e Corte de Pedra (Bahia), Brasil. *Revista da Sociedade Brasileira de Medicina Tropical* 31: 279-288.
199. Lockard RD, Wilson ME, Rodríguez NE (2019) Sex-Related Differences in Immune Response and Symptomatic Manifestations to Infection with *Leishmania* Species. *J. Immunol. Res.* 2019: 4103819.
200. Torres Espejo JM, Le Pon F, Mouchet J, Desjeux P, Richard A (1989) Epidemiologie de la Leishmaniose tegumentaire en Bolivie. 1. Description des zones d'étude et fréquence de la maladie. *Annales de la Société Belge de Médecine Tropicale* 69: 307-312.
201. França F, Lago EL, Tada S, Costa JM, Vale K, et al. (1991) An outbreak of human *Leishmania (Viannia) braziliensis* infection. *Memórias do Instituto Oswaldo Cruz* 86: 169-174.
202. Davies CR, Reithinger R, Campbell-Lendrum D, Feliciangeli D, Borges R, et al. (2000) The epidemiology and control of leishmaniasis in Andean countries. *Cadernos de Saúde Pública* 16: 925-950.
203. Alexander B, Oliveria EB, Haigh E, Almeida LL (2002) Transmission of *Leishmania* in coffee plantations of Minas Gerais, Brazil. *Memórias do Instituto Oswaldo Cruz* 97: 627-630.
204. Ovallos FG, Silva YR, Fernandez, N. Gutierrez R, Galati EA, et al. (2013) The sandfly fauna, anthropophily and the seasonal activities of *Pintomyia spinicrassa* (Diptera: Psychodidae: Phlebotominae) in a focus of cutaneous leishmaniasis in northeastern Colombia. *Memórias do Instituto Oswaldo Cruz* 108: 297- 302.
205. Carrada Figueroa GdC, Leal Ascencio VJ, Jiménez Sastré A, López Álvarez J (2014) Transmission of cutaneous leishmaniasis associated with cacao (*Theobroma cacao*) plantations in Tabasco. *Gaceta Médica de México* 150: 499-508.
206. Alexander B, Agudelo LA, Navarro JF, Ruiz JF, Molina J, et al. (2009) Relationship between coffee cultivation practices in Colombia and exposure to infection with *Leishmania*. *Trans R Soc Trop Med Hyg* 103:1263-1268.
207. Ocampo CB, Ferro MC, Cadena H, Gongora R, Pérez M, et al. (2012) Environmental factors associated with American cutaneous leishmaniasis in a new Andean focus in Colombia. *Trop Med Int Health* 17: 1309-1317.
208. Muñoz G, Davies CR. (2006) *Leishmania panamensis* transmission in the domestic environment: the results of a prospective epidemiological survey in Santander, Colombia. *Biomedica* 26 Suppl 1: 131-144.
209. Barros GC, Sessa PA, de Mattos EA, Carias VR, Mayrink W, et al. (1985) Foco de Leishmaniose Tegumentar Americana nos municípios de Viana Ee Cariacica, Estado do Espírito Santo, Brasil. *Revista de Saúde Pública* 19: 146-153.
210. da Silva O, de Sousa ME, dos Santos FA, da Silava P, Gazin P (2000) La leishmaniose tégumentaire américaine dans la région sucrière du Pernambouc, Nord-Est du Brésil. *Santé: Cahiers d'Etudes et de Recherches Francophones* 10: 123-126.
211. Herrero A, Christensen HA (1976) Epidemiological patterns of cutaneous leishmaniasis in Panama. I. Epidemics among small groups of settlers. *Ann Trop Med Parasitol* 70: 59-65.
212. Gontijo CM, da Silva ES, de Fuccio MB, de Sousa MC, Pacheco RS, et al. (2002) Epidemiological studies of an outbreak of cutaneous leishmaniasis in the Rio Jequitinhonha Valley, Minas Gerais, Brazil. *Acta Trop* 81: 143-150.
213. Dedet JP, Pradinaud R, Gay F (1989) Epidemiological aspects of human cutaneous leishmaniasis in French Guiana. *Trans R Soc Trop Med Hyg* 83: 616-620.
214. Soccol VT, de Castro EA, Schnell e Schühli G, de Carvalho Y, Marques E, et al. (2009) A new focus of cutaneous leishmaniasis in the central area of Paraná State, southern Brazil. *Acta Trop* 111: 308-315.

215. Tedesqui VL, Calleja GN, Parra R, Pabón JP, Bóia MN, et al. (2012) Active surveillance of American tegumentary leishmaniasis in endemic areas in rural Bolivia. *Revista da Sociedade Brasileira de Medicina Tropical* 45: 30-34.
216. Ferro C, López M, Fuya P, Lugo L, Cordovez JM, et al. (2015) Spatial Distribution of Sand Fly Vectors and Eco-Epidemiology of Cutaneous Leishmaniasis Transmission in Colombia. *PLoS One* 10: e0139391.
217. Alcasis A, Abel L, David C, Torrez ME, Flandre P, et al. (1997) Risk factors for onset of cutaneous and mucocutaneous leishmaniasis in Bolivia. *Am J Trop Med Hyg* 57: 79-84.
218. Monteiro WM, Neitzke HC, Lonardon MV, Silveira TG, Ferreira ME, et al. (2008) Distribuição geográfica e características epidemiológicas da leishmaniose tegumentar americana em áreas de colonização antiga do Estado do Paraná, Sul do Brasil. *Cadernos de Saúde Pública* 24:1291-1303.
219. Sousa AQ, Pearson R (2009) Drought, smallpox, and emergence of *Leishmania braziliensis* in northeastern Brazil. *Emerg Infect Dis* 15: 916-921.
220. Desjeux P (2001) The increase in risk factors for leishmaniasis worldwide. *Trans R Soc Trop Med Hyg* 95: 239-243.
221. de Castro EA, Luz E, Telles FQ, Pandey A, Biseto A, et al. (2005) Eco-epidemiological survey of *Leishmania (Viannia) braziliensis* American cutaneous and mucocutaneous leishmaniasis in Ribeira Valley River, Paraná State, Brazil. *Acta Trop* 93: 141-149.
222. Monteiro WM, Neitzke-Abreu HC, Ferreira ME, Melo GC, Barbosa Md, et al. (2009) Mobilidade populacional e produção da leishmaniose tegumentar americana no Estado do Paraná, sul do Brasil. *Revista da Sociedade Brasileira de Medicina Tropical* 42: 509-514.
223. García AL, Parrado R, Rojas E, Delgado R, Dujardin JC, et al. (2009) Leishmaniasis in Bolivia: comprehensive review and current status. *Am J Trop Med Hyg* 80: 704-711.
224. Vilela ML, Azevedo CG, Carvalho BM, Rangel EF (2011) Phlebotomine fauna (Diptera: Psychodidae) and putative vectors of leishmaniasis in impacted area by hydroelectric plant, State of Tocantins, Brazil. *PLoS One* 6: e27721.
225. Gonçalves Neto VS, Barros Filho AK, Santos AM, Prazeres MP, Bezerril AC, et al. (2013) An analysis of the spatiotemporal distribution of American cutaneous leishmaniasis in counties located along road and railway corridors in the State of Maranhão, Brazil. *Revista da Sociedade Brasileira de Medicina Tropical* 46: 322-328.
226. Furtado NV, Galardo AK, Galardo CD, Firmino VC, Vasconcelos Dos Santos T (2016) Phlebotomines (Diptera: Psychodidae) in a Hydroelectric System Affected Area from Northern Amazonian Brazil: Further Insights into the Effects of Environmental Changes on Vector Ecology. *J Trop Med* 2016: 9819723.
227. Godoy RE, de Santana AL, Graser C, Rangel EF, Vilela ML (2017) Aspects on the Ecology of Phlebotomine Sand Flies (Diptera: Psychodidae) From Guarai, State of Tocantins, Brazil, Endemic Area for American Cutaneous Leishmaniasis. *J Med Entomol* 54: 229-235.
228. Zorrilla V, De Los Santos MB, Espada L, Santos RDP, Fernandez R, et al. (2017) Distribution and identification of sand flies naturally infected with *Leishmania* from the Southeastern Peruvian Amazon. *PLoS Negl Trop Dis* 11: e000602
229. Berry I, Berrang-Ford L (2016) Leishmaniasis, conflict, and political terror: a spatio-temporal analysis. *Soc Sci Med* 167:140-149.
230. Brilhante AF, Melchior LAK, Nunes VLB, Cardoso CO, Galati EAB (2017) Epidemiological aspects of American Cutaneous Leishmaniasis (ACL) in an endemic area of forest extractivist culture in western Brazilian Amazonia. *Revista do Instituto de Medicina Tropical de São Paulo* 59: e12.
231. Melchior LAK, Brilhante AF, Chiaravalloti-Neto F (2017) Spatial and temporal distribution of American cutaneous leishmaniasis in Acre state, Brazil. *Infect Dis Poverty* 6: 99.
232. Valderrama-Ardila C, Alexander N, Ferro C, Cadena H, Marín D, et al. (2010) Environmental risk factors for the incidence of American cutaneous leishmaniasis in a sub-Andean zone of Colombia (Chaparral, Tolima). *Am J Trop Med Hyg* 82: 243-250.
233. Chaves LF, Cohen JM, Pascual M, Wilson ML (2008) Social exclusion modifies climate and deforestation impacts on a vector-borne disease. *PLoS Negl Trop Dis* 2: e176.
234. Armijos RX, Weigel MM, Izurieta R, Racines J, Zurita C, et al. (1997) The epidemiology of cutaneous leishmaniasis in subtropical Ecuador. *Trop Med Int Health* 2: 140-152.
235. Rodríguez-Morales AJ, Pascual-González Y, Benítez JA, López-Zambrano MA, Harter-Griep R, et al. (2010) Asociación entre la incidencia de leishmaniosis cutánea y el índice de desarrollo humano y sus componentes en cuatro estados endémicos de Venezuela *Revista Peruana de Medicina Experimental y Salud Pública* 27:22-30.
236. Medina-Morales DA, Machado-Duque ME, Machado-Alba JE (2017) Epidemiology of Cutaneous Leishmaniasis in a Colombian Municipality. *Am J Trop Med Hyg* 97: 1503-1507.
237. Melo HA, Rossoni DF, Teodoro U (2018) Effect of vegetation on cutaneous leishmaniasis in Paraná, Brazil. *Memórias do Instituto Oswaldo Cruz* 113: e170505.
238. Galvão EL, Pedras MJ, Cota GF, Rabello A, Simões TC (2019) How cutaneous leishmaniasis and treatment impacts in the patients' lives: A cross-sectional study. *PLoS One* 14: e0211374.
239. Araujo AR, Portela NC, Feitosa AP, Silva OA, Ximenes RA, et al. (2016) Risk factors associated with American Cutaneous Leishmaniasis in an endemic area of Brazil. *Revista do Instituto de Medicina Tropical de São Paulo* 58: 86.
240. Hertig M, Fairchild GB (1954) The control of Phlebotomus in Peru with DDT. *Am J Trop Med Hyg* 28: 207-230.
241. Nery-Guimarães F, Bustamante FM (1954) A aplicação domiciliar de DDT como base da profilaxia das leishmanioses. Estudo de um foco de leishmaniose muco-cutâneo cinco anos depois de aspersão periódica com aquele inseticida. *Revista brasileira de malariologia e doenças tropicais* 6: 127-130.
242. Falcão AL, Falcão AR, Pinto CT, Gontijo CMF, Falqueto A (1991) Effect of deltamethrin spraying on the sandfly populations in a focus of American cutaneous leishmaniasis. *Memórias do Instituto Oswaldo Cruz* 86: 399-404.
243. Davies CR, Llanos-Cuentas A, Canales J, Leon E, Alvarez E, et al. (1994) The fall and rise of Andean cutaneous leishmaniasis: transient impact of the DDT campaign in Peru. *Trans R Soc Trop Med Hyg* 88: 389-393.
244. Gomes AdC, Neves VL (1998) Estratégia e perspectivas de controle da leishmaniose tegumentar no Estado de São Paulo. *Revista da Sociedade Brasileira de Medicina Tropical* 31: 553-558.



245. Alexander B, Maroli M (2003) Control of phlebotomine sandflies. *Med Vet Entomol* 17: 1-18.
246. Nunes MP, Jackson JM, Carvalh RW, Furtado NJ, Coutinho SG (1991) Serological survey for canine cutaneous and visceral leishmaniasis in areas at risk for transmission in Rio de Janeiro where prophylactic measures had been adopted. *Memórias do Instituto Oswaldo Cruz* 86: 411-417.
247. Lima LC, Marzochi MC, Sobroza PC, de Souza MA (1988) Observações sobre a leishmaniose tegumentar, cinco anos após profilaxia. *Revista de Saúde Pública* 22: 73-77.
248. Christensen HA, de Vasquez AM (1982) The tree-buttress biotope: a pathobiocenose of *Leishmania braziliensis*. *Am J Trop Med Hyg* 31: 243-251.
249. Ready PD, Arias JR, Freitas RA (1985) A pilot study to control *Lutzomyia umbratilis* (Diptera: Psychodidae), the major vector of *Leishmania braziliensis guyanensis*, in a peri-urban rainforest of Manaus, Amazonas State, Brazil. *Memórias do Instituto Oswaldo Cruz* 80: 27-36.
250. Olalla HR, Velez LN, Kato H, Hashiguchi K, Caceres AG, et al. (2015) An analysis of reported cases of leishmaniasis in the southern Ecuadorian Amazon region, 1986-2012. *Acta Trop* 146: 119-126.
251. Chaniotis BN, Parsons RE, Harlan HJ, Correa MA (1982) A pilot study to control Phlebotomine sand flies (Diptera: Psychodidae) in a Neotropical rain forest. *J Med Entomol* 19: 1-5.
252. Perich MJ, Hoch AL, Rizzo N, Rowton ED (1995) Insecticide barrier spraying for the control of sand fly vectors of cutaneous leishmaniasis in Rural Guatemala. *Am J Trop Med Hyg* 52: 485-488.
253. Feliciangeli MD, Mazzarri MB, Campbell-Lendrum D, Maroli M, Maingon R (2003) Cutaneous leishmaniasis vector control perspectives using lambda-cyhalothrin residual house spraying in El Ingenio, Miranda State, Venezuela. *Trans R Soc Trop Med Hyg* 97: 641-646.
254. Le Pont F, Padilla JM, Desjeux P, Richard A, Mouchet J (1989) Impact de pulverisations de deltamethrine dans un foyer de leishmaniose de Bolivie. *Annales de la Societe belge de medecine tropicale* 69: 223-232.
255. Chaves LF, Calzada JE, Rigg C, Valderrama A, Gottedenker N, et al. (2013) Leishmaniasis sand fly vector density reduction is less marked in destitute housing after insecticide thermal fogging. *Parasit Vectors* 6: 164.
256. Calzada JE, Saldaña A, Rigg C, Valderrama A, Romero L, et al. (2013) Changes in Phlebotomine sand fly species composition following insecticide thermal fogging in a rural setting of western Panama. *PLoS One* 8.
257. Davies CR, Llanos-Cuentas EA, Campos P, Monge J, Leon E, et al. (2000) Spraying houses in the Peruvian Andes with lambda-cyhalothrin protects residents against cutaneous leishmaniasis. *Trans R Soc Trop Med Hyg* 94: 631-636.
258. Reithinger R, Davies CR, Cadena H, Alexander B (1997) Evaluation of the fungus *Beauveria bassiana* as a potential biological control agent against phlebotomine sand flies in Colombian coffee plantations. *J Invertebr Pathol* 70: 131-135.
259. González U, Pinart M, Sinclair D, Firooz A, Enk C, et al. (2015) Vector and reservoir control for preventing leishmaniasis. *Cochrane Database Syst Rev* 2015.
260. Le Pont F, Pajot FX (1981) La leishmaniose en Guyane française 2. Modalités de la transmission dans un village forestier: Cacao. *Cahiers ORSTOM. Série entomologie médicale et parasitologie* 19: 223-231.
261. Esterre P, Chippaux JP, Lefait JF, Dedet JP (1986) Evaluation d'un programme de lutte contre la leishmaniose cutanée dans un village forestier de Guyane française. *Bull World Health Organ* 64: 559-565.
262. Reinhold-Castro KR, Fenelon VC, Rossi RM, Brito JE, Freitas JS, et al. (2013) Impact of control measures and dynamics of sand flies in southern Brazil. *J Vector Ecol* 38: 63-68.
263. Reinhold-Castro KR, Scodro RB, Dias-Sversutti AdC, Neitzke HC, Rossi RM, et al. (2005) Avaliação de medidas de controle de flebotomíneos. *Revista da Sociedade Brasileira de Medicina Tropical* 41: 269-276.
264. Teodoro U, dos Santos DR, dos Santos AR, Oliveira Od, dos Santos ES, et al. (2006) Avaliação de medidas de controle de flebotomíneos no Município de Lobato, Estado do Paraná, Sul do Brasil. *Cadernos de Saúde Pública* 22: 451-455.
265. Gouveia C, de Oliveira RM, Zwetsch A, Motta-Silva D, Carvalho BM, et al. (2012) Integrated Tools for American Cutaneous Leishmaniasis Surveillance and Control: Intervention in an Endemic Area in Rio de Janeiro, RJ, Brazil. *Interdiscip Perspect Infect Dis* 2012.
266. Orellano PW, Vazquez N, Salomon OD (2013) Cost-effectiveness of prevention strategies for American tegumentary leishmaniasis in Argentina. *Cadernos de Saúde Pública* 29: 2459-2472.
267. Schreck CE, Kline DL, Chaniotis BN, Wilkinson N, McGovern TP, et al. (1982) Evaluation of personal protection methods against phlebotomine sand flies including vectors of leishmaniasis in Panama. *Am J Trop Med Hyg* 31: 1046-1053.
268. Alexander B, Cadena H, Usma MC, Rojas CA (1995) Laboratory and field evaluations of a repellent soap containing diethyl toluamide (DEET) and permethrin against phlebotomine sand flies (Diptera: Psychodidae) in Valle del Cauca, Colombia. *Am J Trop Med Hyg* 52: 169-173.
269. Soto J, Medina F, Dember N, Berman J (1995) Efficacy of permethrin-impregnated uniforms in the prevention of malaria and leishmaniasis in Colombian soldiers. *Clin Infect Dis* 21: 599-602.
270. Lightburn E, Meynard JB, Morand JJ, Garnotel E, Kraemer P, et al. (2002) Surveillance épidémiologique de la leishmaniose cutanée en Guyane. *Résumé des données militaires recueillies sur 10 ans. Médecine tropicale: revue du corps de sante colonial* 62: 545-553.
271. Dorval ME, Alves TP, Cristaldo G, Rocha HC, Alves MA, et al. (2010) Sand fly captures with Disney traps in area of occurrence of *Leishmania (Leishmania) amazonensis* in the state of Mato Grosso do Sul, mid-western Brazil. *Revista da Sociedade Brasileira de Medicina Tropical* 43: 491-495.
272. Dedet JP, Esterre P, Pradinaud R (1987) Individual clothing prophylaxis of cutaneous leishmaniasis in the Amazonian area. *Trans R Soc Trop Med Hyg* 81: 748.
273. Reithinger R, Dujardin JC, Louzir H, Pirmez C, Alexander B, Brooker S (2007) Cutaneous leishmaniasis. *Lancet Infect Dis* 7:581-596.
274. Cabrera OL, Santamaría E, Pardo RH (2018) Experimental hut to study the indoor behaviour and effects of insecticide-treated bednets on phlebotomine sand flies (Diptera: Psychodidae). *Memórias do Instituto Oswaldo Cruz* 113.

275. Acosta MM, Santini MS, Pérez AA, Salomón OD (2017) Evaluation of efficacy of impregnated curtains in experimental hen houses as a phlebotomine control tool in northeast Argentina. *Med Vet Entomol* 31:161-166.
276. Feliciangeli MD, Wheeler A, Towson H, Ward R, Maignon R (1995) Sandfly control trial with deltamethrin impregnated curtains in el Ingenio, Miranda state Venezuela. *Boletín de la Dirección de Malaria-riología y Saneamiento Ambiental* 35: 127-135.
277. Kroeger A, Avila EV, Morison L (2002) Insecticide impregnated curtains to control domestic transmission of cutaneous leishmaniasis in Venezuela: cluster randomised trial. *BMJ* 325: 810-813.
278. Alexander BU, Cadena H, Quesada BL, Solarte Y, Rosa W, et al. (1995) Evaluation of deltamethrin-impregnated bed nets and curtains against phlebotomine sandflies in Valle del Cauca, Colombia. *Med Vet Entomol* 9: 279-283.
279. Rojas CA, Weigle KA, Tovar R, Morales AL, Alexander B (2006) A multifaceted intervention to prevent American cutaneous leishmaniasis in Colombia: results of a group-randomized trial. *Biomédica* 26: 152-166.
280. Trujillo AV, Reina AEG, Orjuela AG, Suárez EP, Palomares JE, et al. (2013) Seasonal variation and natural infection of *Lutzomyia antunesi* (Diptera: Psychodidae: Phlebotominae), an endemic species in the Orinoquia region of Colombia. *Memórias do Instituto Oswaldo Cruz* 108: 463-469.
281. Rigg CA, Calzada JE, Saldaña A, Perea M, Chaves LF, et al. (2019) *Leishmania* spp. Infection Rate and Feeding Patterns of Sand Flies (Diptera: Psychodidae) from a Hyperendemic Cutaneous Leishmaniasis Community in Panamá. *Am J Trop Med Hyg* 100: 798-807.
282. Gutiérrez JD, Martínez-Vega R, Ramoni-Perazzi J, Diaz-Quijano FA, Gutiérrez R, et al. (2017) Environmental and socio-economic determinants associated with the occurrence of cutaneous leishmaniasis in the northeast of Colombia. *Trans R Soc Trop Med Hyg* 111: 564-571.
283. Peterson AT, Shaw J (2003) *Lutzomyia* vectors for cutaneous leishmaniasis in Southern Brazil: ecological niche models, predicted geographic distributions, and climate change effects. *Int J Parasitol* 33: 919-931.
284. Karagiannis-Voules DA, Scholte RG, Guimarães LH, Utzinger J, Vou-natsou P (2013) Bayesian geostatistical modeling of leishmaniasis incidence in Brazil. *PLoS Negl Trop Dis* 7.
285. Pérez-Flórez M, Ocampo CB, Valderrama-Ardila C, Alexander N (2016) Spatial modeling of cutaneous leishmaniasis in the Andean region of Colombia. *Memórias do Instituto Oswaldo Cruz*.
286. Purse BV, Masante D, Golding N, Pigott D, Day JC, et al. (2017) How will climate change pathways and mitigation options alter incidence of vector-borne diseases? A framework for leishmaniasis in South and Meso-America. *PLoS One* 12.
287. Pupo Nogueira Neto J, Basso G, Cipoli AP, El Kadre L (1998) American cutaneous leishmaniasis in the State of São Paulo, Brazil—epidemiology in transformation. *Ann. Agric. Environ. Med.* 5:1- 5.
288. Stolf HO, Marques SA, Marques ME, Yoshida EL, Dillon NL (1993) Surto de leishmaniose tegumentar americana em Itaporanga, São Paulo (Brasil). *Revista do Instituto de Medicina Tropical de São Paulo* 35: 437-442.
289. McIntyre S, Rangel EF, Ready PD, Carvalho BM (2017) Species-specific ecological niche modelling predicts different range contractions for *Lutzomyia intermedia* and a related vector of *Leishmania braziliensis* following climate change in South America. *Parasite Vectors* 10: 157.
290. Carvalho BM, Rangel EF, Ready PD, Vale MM (2015) Ecological Niche Modelling Predicts Southward Expansion of *Lutzomyia (Nyssomyia) flaviscutellata* (Diptera: Psychodidae: Phlebotominae), Vector of *Leishmania (Leishmania) amazonensis* in South America, under Climate Change. *PLoS One* 10.
291. da Costa SM, Cordeiro JLP, Rangel EF (2018) Environmental suitability for *Lutzomyia (Nyssomyia) whitmani* (Diptera: Psychodidae: Phlebotominae) and the occurrence of American cutaneous leishmaniasis in Brazil. *Parasit Vectors* 11: 155.
292. Shimabukuro PH, da Silva TR, Ribeiro FO, Baton LA, Galati EA (2010) Geographical distribution of American cutaneous leishmaniasis and its phlebotomine vectors (Diptera: Psychodidae) in the state of São Paulo, Brazil. *Parasit Vectors* 3: 121.
293. Rodrigues MGA, de Brito Sousa JDB, Dias ALB, Monteiro WM, Sampaio VS (2019) The role of deforestation on American Cutaneous Leishmaniasis incidence: spatial-temporal distribution, environmental and socioeconomic factors associated in the Brazilian Amazon. *Trop Med Int Health* 24: 348-355.
294. Camargo-Neves VL, Gomes AdC, Antunes JL (2002) Correlação da presença de espécies de flebotomíneos (Diptera: Psychodidae) com registros de casos da leishmaniose tegumentar americana no Estado de São Paulo, Brasil. *Revista da Sociedade Brasileira de Medicina Tropical* 35: 299-306.
295. Arraes SM, Veit RT, Bernal VM, Becker TC, Nanni MR (2008) Leishmaniose tegumentar americana em municípios da região noroeste do estado do Paraná: utilização de sensoriamento remoto para análise do tipo de vegetação e os locais de ocorrência da doença. *Revista da Sociedade Brasileira de Medicina Tropical* 41: 642-647.
296. Shaw J (2007) The leishmaniasoses—survival and expansion in a changing world. A mini-review. *Memórias do Instituto Oswaldo Cruz* 102: 541-547.
297. Beyrer C, Villar JC, Suwanvanichkij V, Singh S, Baral SD, et al. (2007) Neglected diseases, civil conflicts, and the right to health. *Lancet* 370: 619-627.
298. Beck U (2015) *World at risk*. (13<sup>th</sup> edition) Polity Press, Cambridge, UK and Malden, MA. 269.
299. Bauman Z (2015) *Wasted Lives: modernity and its outcasts*. (18<sup>th</sup> edition), Polity Press. Cambridge, UK. 140.