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**SARS-CoV-2 virus and liver expression of host receptors: Putative mechanisms of liver involvement in COVID-19**

**Short running title:** COVID-19 and liver

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*To the Editor,*

Zhang et al. showed that COVID-19 affected patients' present liver biochemistry abnormalities, including elevation of aminotransferases, gamma-glutamyl transferase, and alkaline phosphatase <sup>1</sup>.

Hence, several possible clinical scenarios in the setting of liver diseases have been postulated.

First, patients with chronic liver disease may be more vulnerable to the severe clinical consequences of COVID-19, including oxygen desaturation and hypoxemia due to severe pneumonia or the cytokine storm <sup>1,2</sup>. Second, liver biochemistry abnormalities are the consequence of drug toxicity.

There is a third potential but poorly explored clinical scenario, which is the possibility that the novel 2019 coronavirus, also known as SARS-CoV-2, may directly or indirectly cause liver injury. In fact, SARS-CoV2 viral load in the stool, which has been detected in about 48% of patients even in stool collected after respiratory samples tested negative <sup>3</sup>, is likely to be associated with portal venous viremia.

We assessed the gene expression levels of SARS-CoV2-interacting host receptors in the liver tissue and their distribution across cell types according to single-cell transcriptomic experiments retrieved from the Single Cell Portal. We focused on angiotensin-converting enzyme 2 (*ACE2*), transmembrane serine protease 2 (*TMPRSS2*), and paired basic amino acid cleaving enzyme (*FURIN*) gene expression levels. Our analysis shows that the three human host receptors are expressed in the liver tissue; however, expression levels extensively vary across cell types. *ACE2* presents the highest expression levels in cholangiocytes, followed by hepatocytes (**Figure 1C**). *TMPRSS2* is expressed in cholangiocytes, hepatocytes, periportal liver sinusoidal endothelial cells, erythroid cells, and in a much lesser extent in non-inflammatory macrophages and alpha-beta T cells (**Figure 1D**). *FURIN* shows expression levels across all cell types, from hepatocytes to all populations of liver resident cells (**Figure 1E**).

Together, these findings support the possibility that SARS-CoV-2 may cause direct liver injury by viral cytopathic effect (directly by lysis and/or by inducing necrotic/apoptotic effect/s).

Furthermore, the expression pattern in cell clusters associated with numerous active immune pathways, for example, inflammatory macrophages, natural killer cells, plasma cells, mature B cells, and cells of the liver endothelial microenvironment, opens the possibility of SARS-CoV-2 - immune-mediated liver damage.

Not surprisingly, reports from the past 2003-SARS (severe acute respiratory syndrome) epidemic showed not only liver impairment in up to 60% of the patients but also confirmed the presence of

SARS-coronavirus by RT-PCR in liver biopsies presenting mild to moderate lobular inflammation and apoptosis <sup>4</sup>.

In conclusion, to understand the pathogenesis of SARS-CoV-2 –related liver disease, additional research must be guaranteed, including the search for evidence of viral replication in hepatocytes and liver histology characterization.

**Author contributions:** SS: study concept and design; data acquisition; data analysis and interpretation; manuscript drafting; securing funding. CJP: study concept and design; data acquisition; data analysis and interpretation; manuscript drafting; securing funding.

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## Figure 1

### Liver gene expression profiling across cell types of host receptors implicated in SARS-CoV-2 infection

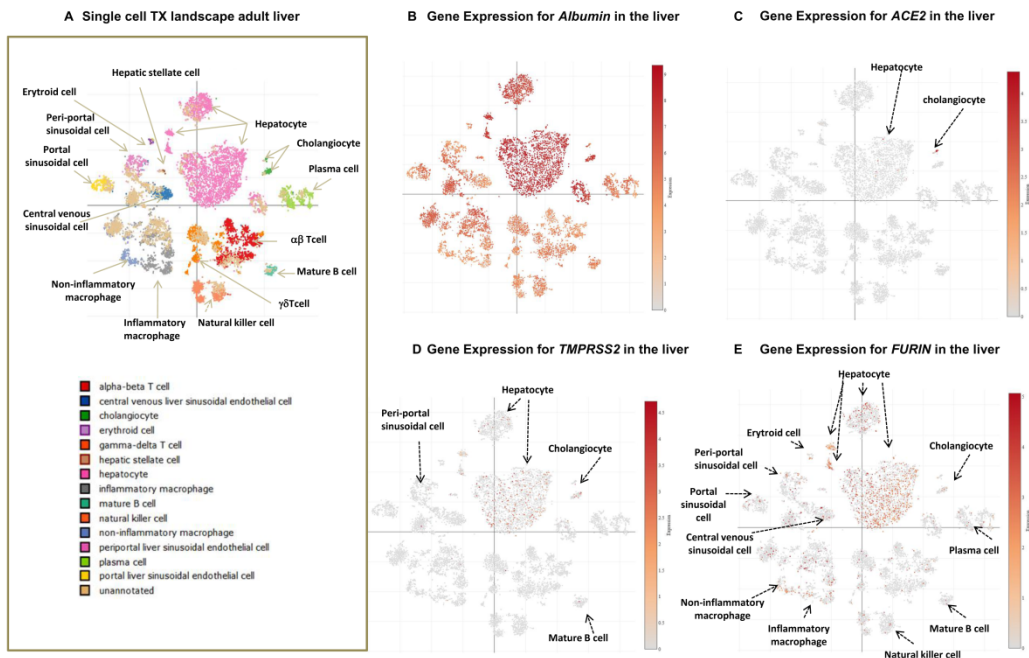
Profiling of gene expression was retrieved from the Single Cell Portal available at [https://singlecell.broadinstitute.org/single\\_cell](https://singlecell.broadinstitute.org/single_cell). The analysis was focused on the adult liver dataset from the Human Cell Atlas March 2020 Release, a collection of 23 human single-cell datasets.

The human liver cellular landscape analysis by single cell RNA-seq is based on the study of MacParland et al. <sup>5</sup>. Human liver tissue was obtained from livers procured from deceased donors deemed acceptable for liver transplantation.

**A.** Annotation of liver whole transcriptome involved 15 clusters, including hepatocytes, alpha-beta T cells, central liver sinusoidal endothelial cells, cholangiocytes, erythroid cells, gamma-delta T cells, hepatic stellate cells, inflammatory macrophage, mature B cells, natural killer cells, non-inflammatory macrophages, periportal liver sinusoidal endothelial cells, plasma cells, portal liver sinusoidal endothelial cells, and unannotated cells (**A**).

**B.** To illustrate the pattern and magnitude of differential gene expression levels at different cells in the liver, we assessed the pattern of gene expression of albumin (*ALB*)- the most abundant protein in human blood that is highly expressed in the liver.

**C-E.** Exploration of *ACE2*, *TMPRSS*, and *FURIN* expression in the liver.



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