



Food Additives & Contaminants: Part B Surveillance

ISSN: 1939-3210 (Print) 1939-3229 (Online) Journal homepage: <http://www.tandfonline.com/loi/tfab20>

Fumonisin occurrence in wheat-based products from Argentina

Eugenia Cendoya, Maria J. Nichea, María P. Monge, Michael Sulyok, Stella M. Chiacchiera & María L. Ramirez

To cite this article: Eugenia Cendoya, Maria J. Nichea, María P. Monge, Michael Sulyok, Stella M. Chiacchiera & María L. Ramirez (2018): Fumonisin occurrence in wheat-based products from Argentina, Food Additives & Contaminants: Part B, DOI: [10.1080/19393210.2018.1520308](https://doi.org/10.1080/19393210.2018.1520308)

To link to this article: <https://doi.org/10.1080/19393210.2018.1520308>



Published online: 03 Oct 2018.



Submit your article to this journal [↗](#)



View Crossmark data [↗](#)

ARTICLE



Fumonisin occurrence in wheat-based products from Argentina

Eugenia Cendoya^{a,d}, María J. Nichea^{a,d}, María P. Monge^{b,d}, Michael Sulyok^c, Stella M. Chiacchiera^{b,d} and María L. Ramirez^{a,d}

^aDepartamento de Microbiología e Inmunología, Facultad de Ciencias Exactas Fco-Qcas y Naturales, Universidad Nacional de Río Cuarto, Río Cuarto, Córdoba, Argentina; ^bDepartamento de Química, Facultad de Ciencias Exactas Fco-Qcas y Naturales, Universidad Nacional de Río Cuarto, Río Cuarto, Córdoba, Argentina; ^cCenter for Analytical Chemistry, Department for Agrobiotechnology (IFA-Tulln), University of Natural Resources and Life Sciences, (BOKU), Vienna, Tulln, Austria; ^dConsejo Nacional de Investigaciones Científicas y Técnicas (CONICET)

ABSTRACT

In Argentina, wheat is the most consumed cereal by the human population. Since fumonisins occurrence in wheat grains and wheat-based products have been reported worldwide, a survey was conducted in order to determine fumonisin contamination in 91 wheat-based products (white wheat flour samples, wheat flour used at bakery products and whole-wheat flour samples) collected from different retail stores of Río Cuarto city in Argentina using HPLC-MS/MS. Sixty-seven samples (74%) showed contamination by fumonisins. From these samples, 16 showed fumonisin levels between LOD and LOQ (between 0.01 to 0.05 ng/g), while fumonisins (FB₁ + FB₂) in quantifiable samples ranged from 0.05 ng/g to 18.9 ng/g. Although FB₁ was more prevalent, FB₂ was found in higher levels than FB₁. Overall, fumonisin prevalence was high, but concentrations were far below EU or USA limits set for maize and maize-based products.

ARTICLE HISTORY

Received 23 June 2018
Accepted 1 September 2018

KEYWORDS

Fumonisin; wheat-based products; HPLC-MS/MS; Argentina; quantitative analysis

Introduction

Wheat is the most important source of calories as well as protein for humans. World wheat production in 2016 was 749.5 million tons (FAO 2017). Wheat is the most important cereal used for human consumption in Argentina. This cereal is ground into flour and semolina for consumption, being the basic ingredients for bread and other bakery products, pasta, breakfast cereals, cookies, and cupcakes (Chandrika and Shahidi 2006). Human consumption of manufactured wheat products, either semolina (*Triticum turgidum* L. var. durum) or bread (*T. aestivum*) in Argentina is much greater than products made from other cereals (Pacin et al. 2012). Wheat flour per capita consumption varies widely by region worldwide. In Argentina its consumption per capita was estimated at 84 kg/habitant/year during 2017 (FAIM 2017).

Cereals like wheat are commonly colonised by *Fusarium* species and can be contaminated with mycotoxins, which are defined as second metabolites produced by fungi that could have a major impact on health, welfare and productivity. Among all *Fusarium* species that could be present in wheat grains and wheat-based products, several fumonisin (FB) producing *Fusarium* species could be isolated. Although several *Fusarium* species are able to produce FBs, the two

most important ones are *Fusarium verticillioides* and *F. proliferatum*, both associated with maize, but also isolated from wheat (Palacios et al. 2011; Chehri et al. 2010; Busman et al. 2012; Amato et al. 2015; Guo et al. 2016). Thus, FBs are usually present in maize and maize-based products, but also, in wheat and wheat-based products (Cendoya et al. 2018a).

In terms of occurrence and toxicity Fumonisin B₁ (FB₁) is the most significant, followed by Fumonisin B₂ (FB₂), which differs structurally from FB₁ in the number and placement of hydroxyl groups on the molecule's hydrocarbon (Voss et al. 2007). FB₁ can cause leukoencephalomalacia in horses (Marasas et al. 1988), pulmonary oedema syndrome and hydrothorax in pigs (Haschek et al. 1992) and nephrotoxic, hepatotoxic and hepatocarcinogenic activities in rats (Wan Norhasima et al. 2009). In human populations that consume FB-contaminated maize, this mycotoxin has been epidemiologically associated with oesophageal cancer (Marasas 2001) and neural tube defects (Missmer et al. 2006). Moreover, recent studies suggest that FB exposure could be related with stunting in children (Kimanya et al. 2010; Shirima et al. 2015). Due to the toxicological effect in animals and humans, the International Agency for Research on Cancer (IARC) designated FB₁ in Group 2B as "possible

carcinogenic to humans" (IARC 2002) and the Joint FAO/WHO Expert Committee on Food Additives (JECFA) has determined a provisionally maximum tolerable daily intake (PMTDI) of 2 µg/kg body weight per day for FB₁, FB₂ and fumonisin B₃ (FB₃) alone or in combination (WHO 2001). The European Union in 2007 (EC N°1126/2007) established maximum limits for FBs in cereals and cereal-based foods for human consumption, but not specifically for wheat or wheat-based foods.

In Argentina wheat contamination with FBs has been reported (Palacios et al. 2011; Cendoya et al. 2014), but there are no studies about contamination with FBs in wheat-based products yet. Thus, due to the importance of wheat in the diet of the Argentinean population and because FB intake occurs mainly via the intake of wheat and wheat-products (Bakker et al. 2003), the aim of this survey was to determine the natural occurrence of FBs in wheat-based products used for human consumption marketed in this country.

Materials and methods

Sample collection

Ninety-one wheat-based products for human consumption were randomly collected from local retail stores in Rio Cuarto (Cordoba, Argentina) in order to investigate presence of FBs. Forty-six and 45 samples were collected during 2015 and 2016, respectively. These consisted of white wheat flour (76), wheat flour used at bakery products (11) and whole-wheat flour (4). To reduce the samples to analytical portions, three subsamples (200 g) were drawn, from the top, middle and bottom layers of each individual package (1 kg). Then, these three subsamples were pooled, mixed thoroughly and quartered. Opposite quarters were rejected and the remainder re-mixed thoroughly to obtain a final reduced sample of 200 g for analysis. All reduced samples were kept in ziplock plastic bags at -20 °C prior to analysis, of which a 25-g test portion was taken for FB extraction.

Extraction

FB extraction was performed according to the method originally reported by Shephard et al. (1990) and modified by Doko et al. (1995). Twenty-five grams of each sample (25 g) were shaken with 50 mL of methanol: water (3:1) for 30 min and filtered through Whatman N° 4 filter paper (Whatman Int. Ltd, Maidstone, Kent, United Kingdom). Ten millilitres of the filtered extract were applied to a Bond-Elut strong anion-exchange

(SAX) cartridge (Agilent Technologies Inc., Santa Clara, CA, USA) fitted to a Supelco solid phase extraction (SPE) manifold (Supelco, Bellefonte, PA, USA), previously conditioned by the successive passage of methanol (5 mL) and methanol:water (3:1, 5 mL), while the flow rate was maintained below 2 mL/min. After that, the cartridge was washed with methanol:water (3:1, 8 mL) followed by methanol (3 mL) and finally the FBs were eluted with 0.5% acetic acid in methanol (14 mL). The eluates were evaporated to dryness at 40 °C under a moderate stream of nitrogen and stored dry at 4 °C until HPLC MS/MS analysis.

HPLC-MS/MS analysis

Analysis was performed with a Waters 2695 module equipped with an auto sampler and interfaced to a Micromass®-Quattro Ultima™ Platinum tandem quadrupole mass spectrometer with electrospray ionisation (ESI) in positive mode, after separation on a XBridge™ C18 column (3.5 µm, 2.1 x 150 mm) connected to a XBridge BEH C18 Sentry Guard Cartridge (130 Å, 3.5 µm, 2.1 x 10 mm), all from Waters (Milford, MA, USA). The mobile phase consisted of a gradient of aqueous 1% formic acid (solvent A) and methanol with 1% formic acid (solvent B) according to Cendoya et al. (2018b). The flow rate was 0.2 mL/min. Column temperature was kept at 20 °C. The nitrogen flow was adjusted to 109 and 726 L/h for cone and desolvation gases, respectively. Blank matrix extracts were investigated to confirm the specificity of the method. Data acquisition and processing were performed using Mass Lynx V.4.1, Waters INC software (Milford, MA, USA). Nebulisation and desolvation temperatures were 150 and 200 °C, respectively. The capillary voltage was 3.00 kV. Multiple-reaction monitoring (MRM) was used for FB determination. The precursor peak [M + H]⁺ and two product peaks monitored to accomplish both quantification and qualification criteria, as well as retention times and detector settings, are given in Table 1. The most abundant transition, 722 > 352, was used for the quantification of FB₁, while 706 > 336 was used for FB₂.

Table 1. MS parameters used to investigate fumonisins by HPLC-MS/MS.

Mycotoxin	Precursorion (m/z)	t _R	Productions (m/z)	CV (V)	CE (V)
FB ₁	722	13.5	334	101	38
			352		35
FB ₂	706.3	15.5	318.5	106	31
			336.3		49

Precursor ion [M + H]⁺;
t_R, retention time (min.);
CV, cone voltage;
CE, collision energy.

Aliquots of 45 μL of sample extracts were injected into the HPLC unit. Four points of identification were used to identify FB_1 , i.e. retention time, the precursor ion $[\text{M} + \text{H}]^+$ and two product ions (m/z 334 and 352). FB_1 and FB_2 stock solutions in acetonitrile: water (1:1) were used (Biopure, Tull, Austria). A calibration curve was obtained by injecting 10 μL of mixed ($\text{FB}_1 + \text{FB}_2$) standard solutions of 0.5, 1.0 and 2.0 $\mu\text{g}/\text{mL}$. Good linearity with a correlation coefficient higher than 0.996 was obtained for the calibration range. The limit of detection (LOD) for FB_1 and FB_2 was 0.01 ng/g , calculated as $S/N = 3$ and the limit of quantification (LOQ) with $S/N = 5$ was 0.05 ng/g . Relative within day and between-day standard deviations (RSD) were 6.5%.

A recovery experiment was performed in triplicate by spiking 25 g of FB -free wheat flour with FB_1 and FB_2 at levels of 10, 100 and 200 ng/g . Spiked samples were left overnight at room temperature to allow solvent evaporation prior to extraction. Mean FB_1 and FB_2 recoveries were 99.9% and 93.6%, respectively.

Results and discussion

FB mean values, as well as maximum amounts of FB_1 and FB_2 found in the present study and the incidence percentage of positive samples are shown in Table 2. Figure 1 resumes FB content in total wheat-based products analysed. Natural FB contamination was present in 74% of all analysed wheat-based samples (67/91). From these positive samples, 16 showed trace levels, i.e. levels between LOD and LOQ (13 white wheat flour samples, 1 wheat flour sample used at bakery and 2 whole-wheat flour samples). Total FB amounts ($\text{FB}_1 + \text{FB}_2$) reached 18.9

ng/g , with a mean level of 1.54 ng/g . FB_1 was present in 64 out of 91 positive samples, while FB_2 was present in 52 samples. Most samples were contaminated with both FB_1 and FB_2 (55%), while 14 samples only contained FB_1 and 2 samples only FB_2 . Although the incidence percentage of FB_1 was higher than of FB_2 , amounts of FB_2 were higher than those of FB_1 (high ratio FB_2/FB_1) when these mycotoxins were found together. Moreover, the highest amount of FB found in the present study was 17.5 ng/g of FB_2 . Maximum amounts of total FB s were found in white wheat flour samples as well as in wheat flour samples used for bakery products. In both types of samples, FB_2 content was at least 8 fold higher than FB_1 . Regarding whole-wheat flour samples, FB content was lower than in other analysed samples, being FB_1 content higher than FB_2 .

Since FB s have become important due to their toxicity, the willingness to study their incidence has been

Table 2. Incidence rate and concentration of fumonisins in wheat-based samples collected from retail stores in Argentina.

Sample types	Mycotoxin	$N_{\text{pos}}/N_{\text{tot}}$ ^a	Incidence (%)	Maximum (ng/g)	Mean ^b (ng/g)
Total samples	FB_1	64/91	70	2.10	0.32
	FB_2	52/91	57	17.51	1.24
	$\text{FB}_1 + \text{FB}_2$	67/91	74	18.94	1.54
White wheat flour	FB_1	52/76	68	2.10	0.28
	FB_2	40/76	53	17.52	0.92
	$\text{FB}_1 + \text{FB}_2$	52/76	68	18.94	1.19
Wheat flour used at bakery	FB_1	9/11	82	2.00	0.66
	FB_2	11/11	100	16.13	3.80
	$\text{FB}_1 + \text{FB}_2$	9/11	82	18.14	4.33
Whole-wheat flour	FB_1	3/4	75	1.10	0.28
	FB_2	2/4	50	0.72	0.19
	$\text{FB}_1 + \text{FB}_2$	3/4	75	1.82	0.47

^aNumber of positive samples/number of total samples.

^bMean of all samples.

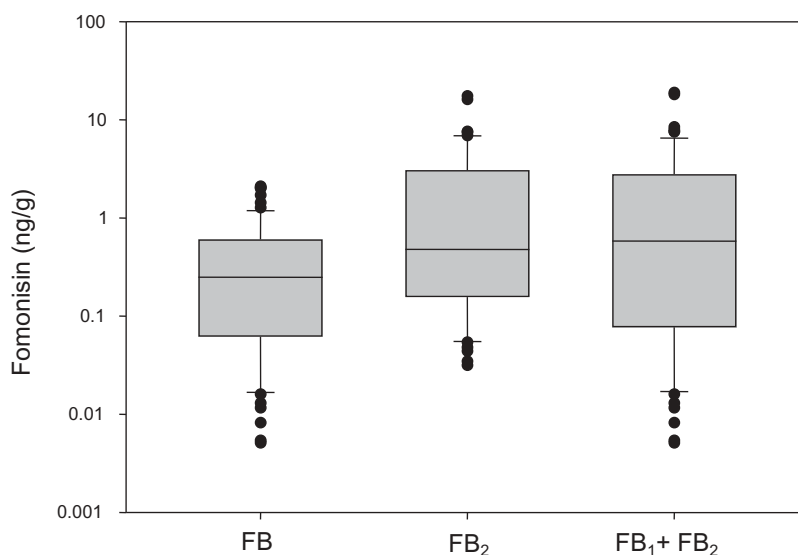


Figure 1. Box plot of FB_1 , FB_2 , and $\text{FB}_1 + \text{FB}_2$ content in wheat-based products marketed in Argentina.

increasing over the years. Together with the development of more sophisticated analytical techniques that allow the detection of small amounts of mycotoxins in different foods, this raised multiple studies related to their incidence in different foods. Thus, the occurrence of FBs in wheat and wheat-based products has been reported in many countries worldwide (Cendoya et al. 2018a).

The results presented above are in agreement with similar surveys performed with HPLC-MS/MS for FB detection and quantitation in different wheat-based samples. In most studies the incidence and concentration of FBs found were low. In China, Liu et al. (2012) detected total FBs (FB₁ + FB₂ + FB₃) in 7% of 30 wheat flour samples, with a mean level of 0.1 ng/g. In the same way, also in China Li et al. (2015) detected FB₁ incidence of 6.2% in 369 wheat flour samples with levels ranging from 0.3 to 34.6 ng/g. In the same country Sun et al. (2011) analysed 16 wheat flour samples and observed 81% of FB₁ incidence with a mean level of 200 ng/g. Similar results were found in other wheat-based products for human consumption. In France, Sirot et al. (2013) analysed FB₁ and FB₂ occurrence in 14 bread and dried bread products and 8 sweet or savoury biscuits and bars, detecting mean levels of 1.4 and 0.3 ng/g and 35 and 75 ng/g for FB₁/FB₂, respectively. Also in France, Rubert et al. (2013) studied FB₁ and FB₂ incidence in 188 wheat-based products. These authors observed 27 positive samples (FB₁ and FB₂ contamination) with levels ranging from 20.2 to 125.8 ng/g and from 21.7 to 101.1 ng/g for FB₁ and FB₂, respectively. Cirillo et al. (2003a and 2003b) performed two different surveys in Italy in order to analyse FBs occurrence in different wheat-based products, obtaining similar results regarding incidence percentages and quantities. These authors also observed a higher incidence of FB₂ than FB₁ in both studies for all analysed samples and in some cases higher amounts of FB₂ than FB₁. Although those results are in agreement with the results obtained in the present study, they are different to most results reported in the literature, where commonly FB₁ is the most frequent FB found in different foods and also commonly higher amounts of FB₁ than FB₂ or FB₃ are found. In Tunisia, 2 wheat-based products showed FB₁ contamination with levels ranging from 88.3 to 184 ng/g (Serrano et al. 2012). In Canada, Roscoe et al. (2008) studied FB incidence in 166 breakfast cereals (maize-, oat-, wheat- and rice-based cereals, as well as mixed-grain cereals) samples collected from the Canadian retail market. As a result total FBs (FB₁ + FB₂) were found in 17% of wheat-based samples with a mean level of 3 ng/g and in one buckwheat cereal sample 5 ng/g. Although FB levels found in that study

were low, the authors concluded that the risk of human exposure to these mycotoxins is present. Just 2 studies showed high amounts of FBs when wheat-based samples were analysed by HPLC-MS/MS. Mashinini and Dutton (2006) analysed 210 wheat-based products from South Africa and observed FB₁ occurrence in four samples, but with FB levels ranging from 1000 to 30,000 ng/g. In Spain Rubert et al. (2013) studied FB₁ and FB₂ incidence in 35 organic Gofio wheat samples and observed both 77% incidence of FBs (FB₁ + FB₂) and high amounts.

On the other hand there are studies that report higher amounts of FBs in wheat-based products than those observed in the present study, but the method used for detection and quantification was ELISA. In Iran Roohi et al. (2012) studied total FB (FB₁ + FB₂ + FB₃) occurrence in different types of wheat flour by ELISA and observed percentages of incidence higher than 80% and FB levels that reached 4500 ng/g. A worldwide survey was conducted by Rodrigues and Naehrer (2012), who analysed mycotoxins by ELISA and HPLC in 7049 samples between 2009 and 2011 from America (North and South), Europe, Oceania and Asia. Regarding FB presence in wheat and wheat bran samples no positive samples out of 7 analysed samples were found in North America, while 2 out of 40 samples were contaminated with FBs (average: 1407 ng/g) in South America. No FBs were detected in a single sample analysed from Northern Europe, while, respectively, 33% and 30% of wheat and wheat bran samples from Central and Southern Europe were contaminated with this mycotoxin (mean levels: 268 and 386 ng/g, respectively). In North Asia, South-East Asia and Oceania 11, 5 and 12% of wheat-based samples were contaminated with FBs, respectively, with mean levels of 371, 172 and 269 ng/g, respectively.

It is remarkable that in the present study most wheat-based samples were contaminated with FBs at levels comparable to those found in the studies described above when the analytical method used was the same: HPLC-MS/MS. This is an important point as other techniques such as ELISA demonstrated to be able to produce false positive results in wheat and wheat-based samples (Shephard et al. 2005). These authors reviewed several reports on FBs in wheat and concluded that careful evaluation of the analytical method and possible source of contamination, as well as confirmation by an appropriate and valid analytical method is required in order to prevent the report of false positive results.

The high incidence of FBs in wheat-based samples found in the present study is important as it has been demonstrated by other studies, using biomarkers

(Szabo-Fodor et al. 2016), that in global areas where maize is contaminated with FBs, the local population is exposed to a higher risk when compared to consider only the individual mycotoxin. Therefore, in those regions where FB-contaminated wheat and wheat-based products consumption is high, the same exposure risk could occur. Bakker et al. (2003) estimated FB₁ intake of the Dutch population by comparing FB₁ concentrations in different food products with consumption rates of those products and reported wheat to be the main contributor (73%) to the total FB₁ intake. Thus, although FB₁ concentration in maize was higher than that in wheat, it appeared that wheat and not maize was the main source of FB intake in the Netherlands.

Similar situations were reported in other countries without specific FB legislation, where a common staple food was reported to be contaminated with FBs. In some occasions at low contamination incidence/levels, in which case most authors concluded that daily intake of low doses of mycotoxin-contaminated food stuff over a period of time could lead to chronic mycotoxicosis. For example, Garcia et al. (2016) studied FBs presence in dry soybean samples sold for human consumption in Brazil and found that 10% of all samples were contaminated with FBs, with levels up to 1495 ng/g. Therefore, the authors emphasised the need for frequent monitoring of FBs in soybeans. Sorghum is a main staple cereal in some districts of Ethiopia and its consumption per day per person is high. Therefore, Taye et al. (2016) studied AFB₁ and FBs presence in sorghum grains they and found all 90 sorghum samples to be contaminated with *Aspergillus* and *Fusarium* species, as well as with AFB₁ and FBs. Aflatoxin B₁ was determined at levels ranging from < LOD to 33.10 µg/kg and total FB levels varied between 907 and 2041 µg /kg. This leads to the conclusion their survey could serve as a starting point to create awareness on toxicogenic fungi and associated mycotoxins in the studied area of the country. Lui et al. (2017) analysed FBs in maize kernel samples from 8 provinces of China and found that 28 samples exhibited higher levels than set by the Food and Drug Administration and 12 samples exhibited higher levels as set by the European Commission. Although the average exposure to FBs which they calculated to be 0.12 µg body weight/day, was within the provisional maximum tolerable daily intake as set by the Joint FAO/WHO Expert Committee on Food Additives.

Another important item to consider is the fact that FB exposure determined by routine analysis could lead to underestimate the actual FB intake by consumers, generally estimated at 40%, as bio-accessibility is not taken into consideration (Szabo-Fodor et al. 2016).

Conventional analytical methods fail to take into account the mycotoxin content bound to the food matrix, which is bio-accessible at consumption as it can be absorbed from the gastrointestinal tract (Versantvoort et al. 2005). Thus, mycotoxin burden in cereals could be significantly higher if the “modified” forms of mycotoxins are considered as well. In agreement with that, Dall’Asta and Battilani (2016) concluded that modified FB should be included in food monitoring plans in order to have an overview of the possible contribution to human exposure. Moreover, in the present study we did not consider co-occurrence of FBs with other mycotoxins like trichothecenes and zearalenone, which are known to occur in wheat and wheat-based products as well, raising the question of possible interactions, synergistic or antagonistic actions in the manifestation of toxicity (Stanciu et al. 2015). Simultaneous exposure to more than only one mycotoxin is of increasing concern and requires more study.

Conclusion

From the findings as established in this survey can be concluded that more studies of FB presence in wheat and wheat-based products as well as co-occurrence with other mycotoxins are needed in order to determine the actual FB intake of the population of Argentina. Also these studies could result in regulatory limits for cereals for human consumption, as nowadays legislation for FB levels in wheat and wheat-based products does not exist in Argentina. Thus, in the near future we are planning to perform further analyses, including more wheat-based products intended for human consumption, especially those consumed by infants and babies, as well as determination of FBs bound to the food matrix and co-occurrence with other mycotoxins, especially deoxynivalenol. These considerations are in agreement with the views of the Joint FAO/WHO Expert Committee on Food Additives, who expressed the need to analyse FB content in food samples using appropriate analytical methods. Moreover, this Committee highlighted the need to perform studies on bound mycotoxins as well (JECFA 2017).

Acknowledgements

Nichea, M.J. and Cendoya, E. are fellows of CONICET and Ramirez, M.L., Chiacchiera, S., Zacchetti, V. and Monge, M.P. are members of the Research Career of CONICET.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by the Secretaría de Ciencia y Técnica, Universidad Nacional de Río Cuarto (SECyT-UNRC) and CONICET (the National Scientific and Technical Research Council of Argentina).

References

- Amato B, Pfohl K, Tonti S, Nipoti P, Dastjerdi R, Pisi A. 2015. *Fusarium proliferatum* and fumonisin B₁ co-occur with *Fusarium* species causing Fusarium head blight in durum wheat in Italy. *J Appl Bot Food Qual*. 88:228–233.
- Bakker MI, Speijers GJA, Paulsch WE. 2003. Risk assessment of fumonisin B₁ in the Netherlands. Laboratory for Food and Residue Analysis, National Institute for Public Health and the Environment (RIVM). RIVM report 310301001/2003. Available from: P.O. Box 1 (3720 BA Bilthoven, the Netherlands).
- Busman M, Desjardins AE, Proctor RH. 2012. Analysis of fumonisin contamination and the presence of *Fusarium* in wheat with kernel black point disease in the United States. *Food Addit Contam Part A*. 29:1092–1100.
- Cendoya E, Chiotta ML, Zachetti V, Chulze SN, Ramirez ML. 2018a. Fumonisin and fumonisin-producing *Fusarium* occurrence in wheat and wheat by products: A review. *J Cereal Sci*. 80:158–166.
- Cendoya E, Monge MP, Chiacchiera S, Farnochi MC, Ramirez ML. 2018b. Influence of water activity and temperature on growth and fumonisin production by *Fusarium proliferatum* strains on irradiated wheat grains. *Int J Food Microbiol*. 266:158–166. ISSN 0168-1605.
- Cendoya E, Monge MP, Palacios S, Chiacchiera S, Torres A, Farnochi MC, Ramirez ML. 2014. Fumonisin occurrence in naturally contaminated wheat grain harvested in Argentina. *Food Control*. 37:56–61.
- Chandrika ML, Shahidi F. 2006. Importance of insoluble-bound phenolics to antioxidant properties of wheat. *J Agric Food Chem*. 54:1256–1264.
- Chehri K, Jahromi ST, Reddy KR, Abbasi S, Salleh B. 2010. Occurrence of *Fusarium spp.* and fumonisins in stored wheat grains marketed in Iran. *Toxins*. 2:2816–2823.
- Cirillo T, Ritieni A, Galvano F, Amodio Cocchieri R. 2003a. Natural co-occurrence of deoxynivalenol and fumonisins B₁ and B₂ in Italian marketed foodstuff. *Food Addit Contam*. 20:566–571.
- Cirillo T, Ritieni A, Visone M, Amodio Cocchieri R. 2003b. Evaluation of conventional and organic Italian foodstuffs for deoxynivalenol and fumonisins B₁ and B₂. *J Agric Food Chem*. 51:8128–8131.
- Dall'Asta C, Battilani P. 2016. Fumonisin and their modified forms, a matter of concern in future scenario? *World Mycotoxin J*. 9:727–739.
- Doko MB, Rapior S, Visconti A, Schjoth JE. 1995. Incidence and levels of fumonisin contamination in maize genotypes grown in Europe and Africa. *J Agric Food Chem*. 43:429–434.
- [FAIM] Federacion Argentina de la Industria Molinera. 2017. Molienda de trigo pan y candel a nivel nacional. Produccion de harina y consumo per capita. [Common and durum wheat national grinding. Wheat flour production and per capital consumption]. Spanish. [accessed March 2018]. <https://www.faim.org.ar/Nacional.aspx>.
- [FAO] Food and Agriculture Organization of the United Nations. 2017. Faostat. [accessed 2018 Mar]. <http://www.fao.org/faostat/en/#data/QC>.
- Garcia LP, Savi GD, Santos K, Scussel VM. 2016. Fumonisin and fungi in dry soybeans (*Glycine Max L.*) for human consumption. *Food Addit Contam Part B*. 9:79–84.
- Guo Z, Pfohl K, Karlovsky P, Dehne HW, Altincicek B. 2016. Fumonisin B₁ and beauvericin accumulation in wheat kernels after seed-borne infection with *Fusarium proliferatum*. *Agric Food Sci*. 25:138–145.
- Haschek WM, Motelin G, Ness DK, Harlin KS, Hall WF, Vesonder RF. 1992. Characterization of fumonisin toxicity in orally and intravenously dosed swine. *Mycopathologia*. 117:83–96.
- [IARC] International Agency for Research on Cancer. 2002. Fumonisin B₁. In: IARC, editor. IARC monograph on the evaluation of carcinogenic risk to human, some traditional herbal medicines, somemycotoxins, naphthalene and styrene. Vol. 82. Geneve. IARC Press. p. e301–e366.
- [JECFA] Joint FAO/WHO Expert Committee on Food Additives. 2017. Evaluation of certain contaminants in food 83rd report of the joint FAO/WHO expert committee on food additives. WHO Technical Report Series, No. 1002.
- Kimanya ME, De Meulenaer B, Roberfroid D, Lachat C, Kolsteren P. 2010. Fumonisin exposure through maize in complementary foods is inversely associated with linear growth of infants in Tanzania. *Mol Nutr Food Res*. 54:1659–1667.
- Li F, Jiang D, Zheng F, Chen J, Li W. 2015. Fumonisin B₁, B₂ and B₃ in corn products, wheat flour and corn oil marketed in Shandong province of China. *Food Addit Contam Part B*. 8:169–174.
- Liu Y, Jiang Y, Li R, Pang M, Liu Y, Dong J. 2017. Natural occurrence of fumonisins B₁ and B₂ in maize from eight provinces of China in 2014. *Food Addit Contam Part B*. 10:113–117.
- Liu YP, Yang LX, Yang NJ, Dong B, Cao LL, Wang K, Yang LX. 2012. Occurrence of fumonisins and aflatoxins in cereals from markets of Hebei province of China. *Food Addit Contam Part B*. 5:208–211.
- Marasas WF. 2001. Discovery and occurrence of the fumonisins: a historical perspective. *Environ Health Perspect*. 2:239–243.
- Marasas WF, Kellerman TS, Gelderblom WC, Coetzer JA, Thiel PG, Van der Lugt JJ. 1988. Leukoencephalomalacia in a horse induced by fumonisin B₁ isolated from *Fusarium moniliforme*. *Onderstepoort J Vet Res*. 55:197–203.
- Mashini K, Dutton MF. 2006. The incidence of fungi and mycotoxins in South Africa wheat and wheat-based products. *J Environ Sci Health Part B*. 41:285–296.
- Missmer SA, Suarez L, Felkner M, Wang E, Merrill AH, Rothman KJ. 2006. Exposure to fumonisins and the occurrence of neural tube defects along the Texas-Mexico border. *Environ Health Perspect*. 114:237–241.
- Pacin A, Ciancio Bovier E, Canoa G, Taglieri D, Hernandez Pezzani C. 2012. Effect of the bread making process on wheat flour contaminated by deoxynivalenol and exposure estimate. *Food Control*. 21:492–495.
- Palacios SA, Ramirez ML, Cabrera Zalazar M, Farnochi MC, Zappacosta D, Chiacchiera SM, Reynoso MM, Chulze SN,

- Torres AM. 2011. Occurrence of *Fusarium spp* and fumonisin in durum wheat grains. *J Agric Food Chem*. 59:12264–12269.
- Rodrigues I, Naehrer K. 2012. A three-year survey on the worldwide occurrence of mycotoxins in feedstuffs and feed. *Toxins*. 4:663–675.
- Roohi S, Gholampour Azizi I, Hashemi M. 2012. Fumonisin contamination based on flour quality used in bakeries and confectioneries in Qaemshahr (city of the Northern Iran). *Afr J Microbiol Res*. 6:1815–1818.
- Roscoe V, Lombaert GA, Huzel V, Neumann G, Melietio J, Kitchen D, Kotello S, Krakalovich T, Trelka R, Scott PM. 2008. Mycotoxins in breakfast cereals from the Canadian retail market: A 3-year survey. *Food Addit Contam Part A*. 25:347–355.
- Rubert J, Soriano JM, Manes J, Soler C. 2013. Occurrence of fumonisins in organic and conventional cereal-based products commercialized in France, Germany and Spain. *Food Chem Toxicol*. 56:387–391.
- Serrano AB, Font G, Ruiz MJ, Ferrer E. 2012. Co-occurrence and risk assessment of mycotoxins in food and diet from Mediterranean area. *Food Chem*. 135:423–429.
- Shephard GS, Sydenham EW, Thiel PG, Gelderblom WCA. 1990. Quantitative determination of fumonisins B₁ and B₂ by high performance liquid chromatography with fluorescence detection. *J Liq Chromatogr*. 13:2077–2087.
- Shephard GS, van der Westhuizen L, Gatyeni PM, Katerere DR, Marasas WF. 2005. Do fumonisin mycotoxins occur in wheat? *J Agric Food Chem*. 53:9293–9296.
- Shirima CP, Kimanya ME, Routledge MN, Srey C, Kinabo JL, Humpf HU. 2015. A prospective study of growth and biomarkers of exposure to aflatoxin and fumonisin during early childhood in Tanzania. *Environ Health Perspect*. 123:173–178.
- Sirot V, Freymy JM, Leblanc JC. 2013. Dietary exposure to mycotoxins and health risk assessment in the second French total diet study. *Food Chem Toxicol*. 52:1–11.
- Stanciu O, Banc R, Cozma A, Filip L, Miere D, Manes J. 2015. Occurrence of *Fusarium* mycotoxins in wheat from Europe- a review. *Acta Univ Cibiniensis Ser E Food Technol*. 19:35–60.
- Sun G, Wang S, Hu X, Su J, Zhang Y, Xie Y. 2011. Co-contamination of aflatoxin B₁ and fumonisin B₁ in food and human dietary exposure in three areas of China. *Food Addit Contam: Part A*. 28:461–470.
- Szabo-Fodor J, Bors I, Szabo A, Kovacs M. 2016. Comparison of the amount of bioaccessible fumonisin B₁ and B₂ in maize and rice inoculated with *Fusarium verticillioides* (MRC 826) and determined by in vitro digestion-preliminary results. *Mycotoxin Res*. 32:173–178.
- Taye W, Ayalew A, Chala A, Dejene M. 2016. Aflatoxin B₁ and total fumonisin contamination and their producing fungi in fresh and stored sorghum grain in East Hararghe, Ethiopia. *Food Addit Contam Part B*. 9:237–245.
- Versantvoort CH, Oomen AG, Van de Kamp E, Rempelberg CJ, Sips AJ. 2005. Applicability of an in vitro digestion model in assessing the bioaccessibility of mycotoxins from food. *Food Chem Toxicol*. 43:31–40.
- Voss KA, Smith GW, Haschek WM. 2007. Fumonisin: toxicokinetics, mechanism of action and toxicity. *Anim Feed Sci Technol*. 137:299–325.
- Wan Norhasima WM, Abdulmir AS, Abu Bakar F, Son R, Norhafniza A. 2009. The health and toxic adverse effects of *Fusarium* fungal mycotoxin, fumonisins, on human population. *Am J Infect Dis*. 4:273–281.
- [WHO] World Health Organization. 2001. Safety evaluation of certain mycotoxins in food (WHO food additives series 47). In: International Programme on Chemical Safety. Geneva: World Health Organization; p. 103–279.