

# Distinctness, phylogenetic relations and biogeography of intertidal mussels (*Brachidontes*, Mytilidae) from the south-western Atlantic

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*Rocky shore intertidal communities along the cold- and warm-temperate coasts of the south-western Atlantic are dominated by small mussels of the genus Brachidontes s.l. (Mytilidae), yet the status of species occurring in the region remains unresolved. Taxonomic studies have been based on shell morphology, but high phenotypic variability has led to much confusion. Based on mitochondrial and nuclear genes (COI, 28S rDNA and ITS1) from nine localities in Uruguay and Argentina we confirmed the occurrence of three species in the south-western Atlantic: Brachidontes darwinianus and B. rodriguezii in the warm-temperate and B. purpuratus in the cold-temperate sector. The latter two species coexist in the same beds along the transition zone (41–43°S). The phylogeny based on mitochondrial and nuclear genes, indicate an early divergence of B. purpuratus. At the intra-specific level, low genetic differentiation and absence of fossil record for B. purpuratus from the earlier Quaternary marine terraces of Patagonia likely result from a relatively recent (post-LGM) colonization originated from populations in the south-eastern Pacific. In the case of B. rodriguezii, by contrast, genetic intraspecific differentiation, a fossil record of phenotypically-related forms going back to the Late Miocene, and phylogenetic position in the COI-based phylogeny, prompts the hypothesis that this species is derived from a local stock with a long history of occurrence in the warm-temperate region of the south-western Atlantic. While intertidal mussel beds from the south-western Atlantic are ecologically similar in appearance, their assembly involves components clearly differentiated in terms of historical biogeography and phylogeny.*

**Keywords:** *Brachidontes*, mussel, COI barcoding, nuclear DNA, phylogeny, south-western Atlantic

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## INTRODUCTION

The ecological communities that characterize the rocky intertidal shores of South America are often dominated by dense, largely single-species stands of small mussels primarily belonging to the genus *Brachidontes* Swainson, 1840, *sensu lato* (s.l.) (Tanaka & Magalhães, 2002; Thiel & Ullrich, 2002; Silliman *et al.*, 2011). These *Brachidontes*-dominated beds are present along both coasts of South America, from south-east Brazil to Tierra del Fuego on the Atlantic side and from Tierra del Fuego to northern Chile on the Pacific side. The beds thus extend across biogeographic boundaries between the cold-temperate coasts of Patagonia (the so-called Magellanic Province; Balech & Ehrlich, 2008), and the warm temperate regions of the south-western Atlantic (Argentinian Province) and the south-east Pacific (Chile–Peru Province; Briggs & Bowen, 2012).

In the south-western Atlantic (the focal area of our study), there is evidence indicative of replacement among nominal species of *Brachidontes*, defined on the basis of shell

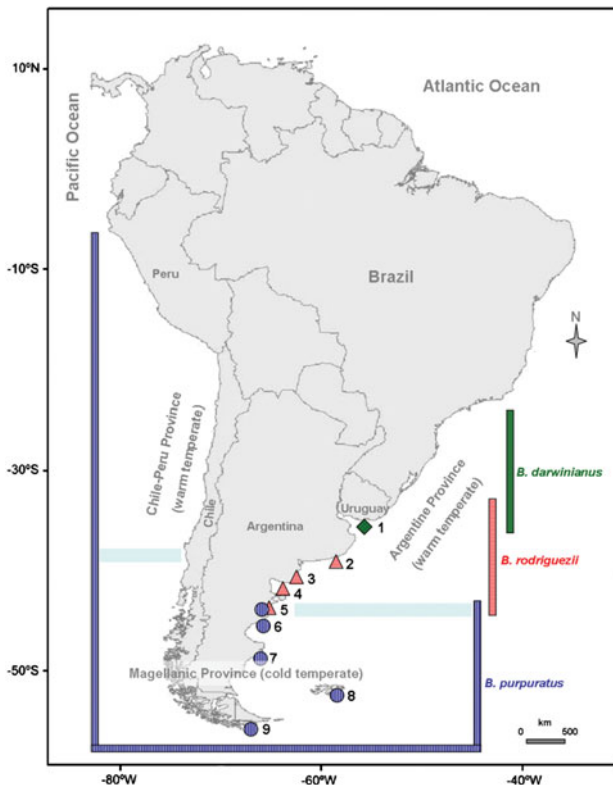
phenotypes, along environmental gradients (Figure 1). First, there is a gradual latitudinal replacement of *B. purpuratus* (Lamarck, 1819), a cold-temperate species, by *B. rodriguezii* (d'Orbigny, 1842), a warm-temperate species, between 43° and 41°S (northern Argentine Patagonia) (Scarabino, 1977). This transition appears to be tightly correlated with sea-surface temperature. Second, *B. rodriguezii* is replaced by the estuarine species *B. darwinianus* (d'Orbigny, 1842) along the northern coast of the La Plata River estuary, from east to west, presumably in relation to the salinity gradient (Scarabino *et al.*, 2006). The mussel beds are remarkable for their uniform appearance, particularly on exposed rocky shores, and in spite of shifts in the dominant species.

In the past, these and other species of *Brachidontes* have been separated solely on the basis of shell phenotypic characters. High phenotypic variability and homoplasy, however, have led to considerable confusion. For instance, recent molecular studies clarified the systematics and phylogeography of *Brachidontes* in Central and North America. Lee & O'Foighil (2004, 2005) found that what was generally considered a single species, *B. exustus* (Linnaeus, 1758), is actually a complex of five species in the Gulf of Mexico and the Caribbean Sea, plus two geminates on the Pacific coast of Central America. Opinions are contradictory in the case of the species from

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**Fig. 1.** Latitudinal range of distribution of *Brachidontes* s.l. species present in the south-western Atlantic (bars) and sampling sites for this study (symbols); (♦) *B. darwinianus*, (▲) *B. rodriguezii*, (●) *B. purpuratus*. Numbers indicate locations: 1, Punta Canario; 2, Santa Clara del Mar; 3, Bahía San Blas; 4, Bahía Rosas; 5, Puerto Madryn; 6, Caleta Carolina; 7, Puerto Deseado; 8, Surfer Bay; 9, Bahía Ensenada (see Table 1 for more information).

the south-western Atlantic. Aguirre *et al.* (2006a), based on the application of morphometric techniques to planar projections of shell outlines, concluded that the three species are indistinguishable; accordingly, *B. rodriguezii* and *B. darwinianus* would be junior synonyms of *B. purpuratus*. More recent morphological studies, however, highlighted characters purportedly allowing the differentiation of *B. rodriguezii* and *B. purpuratus* (van der Molen *et al.*, 2012; Adami *et al.*, 2013). Rios (1994), and other Brazilian authors after him, have considered *B. darwinianus* as a junior synonym of *B. exustus*, which as mentioned earlier is a complex of cryptic species. While many authors (e.g. Coan & Valentich-Scott, 2012) place *B. purpuratus* in the monotypic genus *Perumytilus* Olsson, 1961, others retain it in *Brachidontes* s.l. (e.g. Zelaya, 2009; Huber, 2010; Adami *et al.*, 2013). We prefer to follow the latter, at least until phylogenetic relations within *Brachidontes* s.l. are better understood.

Beyond the taxonomic conundrum, extreme viewpoints on the distinctness of these species implicitly support alternative ecophylogenetic hypotheses (Webb *et al.*, 2002; Mouquet *et al.*, 2012). If the three nominal species were closely related to each other, differentiated along environmental gradients (temperature, salinity), then physiognomic uniformity of intertidal mussel beds across biogeographic boundaries would reflect conservative design in the species dominating these beds. If, on the other hand, they belonged to separate clades, then the uniformity in appearance of these intertidal mussel beds from sub-antarctic to subtropical ecosystems, along both coasts of South America, would reflect

convergence in communities assembled with elements of different origins.

The objectives of our study were to clarify the status of the nominal species of *Brachidontes* from the south-western Atlantic. We hypothesized that *Brachidontes darwinianus*, *B. rodriguezii* and *B. purpuratus* are indeed different species and used molecular information (one mitochondrial and two nuclear markers), to investigate their phylogenetic relations, and to discuss their patterns of distribution in the light of current ideas on the biogeography of the region.

## The genus *Brachidontes* in the south-western Atlantic

The genus *Brachidontes* Swainson, 1840, s.l. (including *Hormomya* Mörch, *Austromytilus* Laseron, and *Perumytilus* Olsson, 1961) includes between 25 and 35 valid species (depending on accepted synonymies) distributed world-wide along warm and warm-temperate coastlines. Only one species, *B. purpuratus*, extends into cold-temperate waters (Figure 1). This species is also remarkable because its geographic range spans two entire biogeographic provinces located in different regions as defined by water temperature: the cold-temperate Magellanic Province (around southern South America), and the warm-temperate Chile–Peru Province (Zelaya, 2009). In the south-western Atlantic, *B. purpuratus* ranges northward to the transition zone between the cold and warm-temperate regions (41–43°S), corresponding, respectively, to the Magellanic and Argentine biogeographic provinces. The fact that *B. purpuratus* ranges into the warm-temperate region in the south-east Pacific but not in the south-western Atlantic is somewhat puzzling, but may be a consequence of density-dependent and founder effects (Waters *et al.*, 2012) and the pre-existence in the warm-temperate region of the south-western Atlantic of a close competitor, i.e. *Brachidontes rodriguezii* (see below). As mentioned earlier, two other species have been consistently recorded in the south-western Atlantic. *Brachidontes rodriguezii*, originally described on the basis of specimens collected by Alcide d’Orbigny (1842, 1846) in Bahía San Blas (northern Argentine Patagonia, ~40°26’S), ranges from Punta Ninfas (42°58’S) to Rio Grande (southern Brazil, 32°10’S) and is thus confined to the Argentine province. *Brachidontes darwinianus* was described on the basis of specimens collected in Rio de Janeiro (~22°56’S, Brazil), Maldonado (~34°55’S, Uruguay) and Bahía Rosas (= Ensenada de Ros, 41°09’S, northern Argentine Patagonia). This species has often been considered a synonym of the nominal species *B. exustus* (Linnaeus, 1758) (Rios, 1994), which is in fact a complex of cryptic species (Lee & Ó Foighil, 2004, 2005).

Besides these three species, others have been reported for the region of focal interest. *Brachidontes solisianus* (d’Orbigny, 1842), placed under the genus *Mytilaster* Monterosato by some authors (Scarabino, 2003; Huber, 2010, followed by WoRMS), was originally described on the basis of specimens collected in the rocky intertidal of Rio de Janeiro (Brazil) and Maldonado (Uruguay). While never recorded again for the Uruguayan littoral, it has been reported to occur along the coasts of Brazil (Rios, 1994). The status of this species requires clarification, but given its reported distribution range this is beyond the scope of the present study. *Brachidontes blakeanus* Melvill & Standen (1914) was

described based on sublittoral specimens collected in Roy Cove (Malvinas/Falkland Islands), and later reported for Patagonia in some ecological studies (Rios *et al.*, 2003; Kelaher *et al.*, 2007). Kelaher *et al.* (2007) also reported *B. granulatus* (Hanley, 1843) for the rocky intertidal zone of Argentine Patagonia. Otherwise, its known range of geographic distribution is restricted to the warm-temperate Chile–Peru Province.

## MATERIALS AND METHODS

### Specimens

Specimens attributable to the nominal species *Brachidontes darwinianus*, *B. rodriguezii* and *B. purpuratus* were collected from nine localities in the south-western Atlantic, ranging from Montevideo (Uruguay) to the Beagle Channel (Tierra del Fuego) and the Malvinas/Falkland Islands (Table 1), and preserved in 95% ethanol. The sampling sites of Punta Canario (34°55'S 56°09'W, Uruguay, Figure 1) and Bahía San Blas (40°32'S 62°15'W, Argentina, Figure 1) are close to type localities of *B. darwinianus* and *B. rodriguezii*, respectively. In order to support assignment of specimens used in our genetic study to nominal species based on phenotypic shell characters we obtained high quality pictures of all the type materials of *B. darwinianus*, *B. rodriguezii* (Figure 2) and *B. solisianus*, deposited in the British Museum of Natural History, and of *B. blakeanus*, deposited in the collections of The Manchester Museum. The type material of *B. purpuratus* appears to have been lost (Dr Guido Pastorino, personal communication). Separation of *B. rodriguezii* and *B. purpuratus* was further supported with results from recent morphological and morphometric studies (van der Molen *et al.*, 2012; Adami *et al.*, 2013). Specimens selected for sequencing included juveniles of *B. rodriguezii* from Bahía Rosas (41°01'S 64°06'W, Figure 1), superficially resembling *B. darwinianus* (e.g. Figure 2K), as well as worn-out or atypical specimens from the region where the ranges of *B. purpuratus* and *B. rodriguezii* overlap, obvious candidates to be misclassified. We also examined extensive collections covering the geographic range of our study and kept at the Museo Nacional de Historia Natural (Montevideo, Uruguay), the Museo Argentino de Ciencias Naturales (Buenos Aires

Argentina), and the Museo de Ciencias Naturales de La Plata (Argentina). Some of those collections were also used in related studies focused on taxonomic and ecological aspects (Adami *et al.*, 2013).

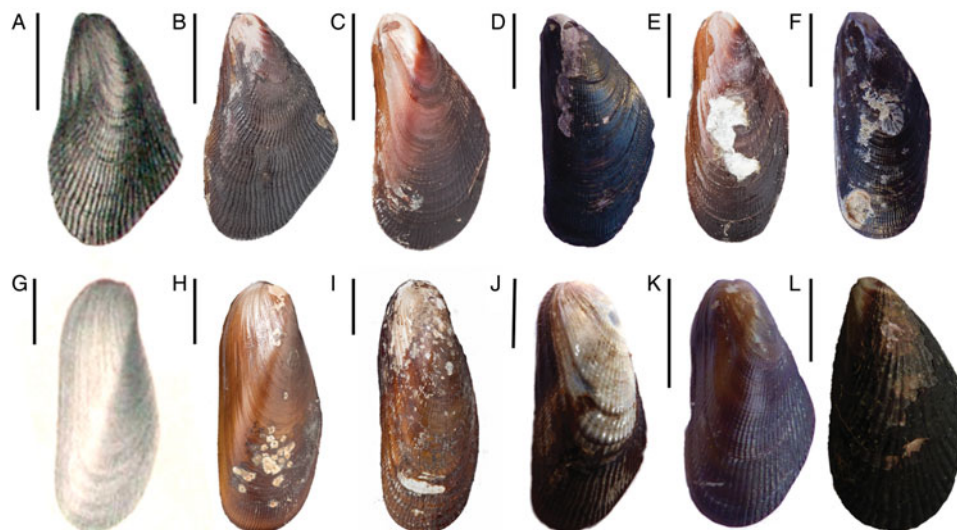
### DNA extraction, amplification and alignment

Total DNA was isolated from the posterior adductor muscle using the phenol-chloroform protocol (modified from Sambrook *et al.*, 1989). Three target gene fragments were amplified and directly sequenced from the study taxa. A 560 nucleotide (nt) portion of mt cytochrome c oxidase subunit I (COI) was amplified via Folmer *et al.*'s (1994) primers, LCO1490 and HCO2198, and another primer pair designed for this study nested within the Folmer's primers, position 46 and 650, CO1aF, 5'AAT GTT TGG TAT ATG AAG 3'/CO1aR, 5' ATC TCC GCC TCC TAT WGG ATC 3'. In addition, two discrete segments of the nuclear ribosomal gene cluster were sequenced. A 709 nt (aligned length) fragment of the large nuclear subunit (28S) rDNA, encompassing domain 2 and part of domain 3, was characterized using primers D23F and D6R (Park & Ó Foighil, 2000). The nuclear ribosomal first internal spacer (ITS1, 563 nt aligned length) was characterized using primers annealing to flanking regions of the 18S and the 5.8S (White *et al.*, 1996).

When possible we sequenced ten specimens per locality for the mitochondrial gene COI and two per locality for the nuclear genes 28S rDNA and ITS1, obtaining a total of 124 sequences (Table 1). To amplify the genes we used Tsg polymerase (Bio Basic Inc., Canada). The protocol used included an initial denaturing temperature of 95°C for 5 min, followed by 30 cycles of 95°C for 45 s, an annealing temperature of 45°C for 1 min for the COI and 52°C for the 28S rDNA and ITS1, 72°C for 1 min, and a final extension at 72°C for 10 min. After extraction and amplification the DNA was visualized by UV transillumination in 1% agarose gels stained with green gel (BIOTUM). Extractions and amplifications of DNA samples were performed in the Gene Probe Laboratory of Dalhousie University (Nova Scotia, Canada) and in the Laboratory of Molecular Biology (CENPAT, Argentina), while the purification of PCR products and sequencing of both strands of DNA were carried out mostly by Macrogen Inc. (Maryland, USA); the remainder was performed at the CENPAT laboratory using the same primers in the

**Table 1.** Sampling locations, corresponding minimum and maximum monthly mean seawater temperature (SST, min–max), and number of samples sequenced per gene. AR, Argentina; UY, Uruguay.

Nominal species	Locality	Latitude S	Longitude W	SST (°C) min–max	Sequences		
					COI	28S rDNA	ITS1
<i>Brachidontes darwinianus</i>	1 Punta Canario (Montevideo, UY)	34° 55'	56° 09'	10.5–23.6	8	2	1
<i>Brachidontes rodriguezii</i>	2 Santa Clara del Mar (AR)	37° 50'	57° 30'	9.3–21.3	6	2	2
	3 Bahía San Blas (AR)	40° 32'	62° 15'	7.5–22.6	8	5	2
	4 Bahía Rosas (AR)	41° 01'	64° 06'	8.7–19.7	9	2	1
	5 Puerto Madryn (AR)	42° 46'	65° 00'	8.7–17.9	8	2	2
	6 Caleta Carolina (AR)	44° 47'	65° 43'	7.3–16.6	9	7	2
<i>Brachidontes purpuratus</i>	7 Puerto Deseado (AR)	47° 44'	65° 53'	3.8–14.4	8	1	2
	8 Surfer Bay, Malvinas/Falkland I.	51° 41'	57° 46'	2.4–8.0	10	2	2
	9 Bahía Ensenada (Beagle Channel, Tierra del Fuego, AR)	54° 49'	68° 15'	2.6–10.8	8	1	2
Total					78	28	18



**Fig. 2.** Selected specimens of *Brachidontes s.l.* sequenced in this study and type specimens. (A–F): *B. darwinianus*; (A, B) syntype illustrated by d'Orbigny (1842): original illustration (A) and presumably the same specimen conserved at the British Museum (B); (C, E) syntypes, (D, F) sequenced specimens from Punta Canario, (Montevideo, Uruguay). (G–K) *B. rodriguezii*; (G, H) lectotype (H), presumably corresponding to the specimen illustrated by d'Orbigny (1842) (G); (I) paralectotype (J, K) sequenced specimens collected at the type locality (Bahia San Blas). (L) *B. purpuratus*, sequenced specimen from Puerto Deseado. Scale bar: 1 cm.

amplification. DNA sequence data were edited in CodonCode Aligner v.2.0.4. and aligned using default parameters with Clustal W (Thompson *et al.*, 1994) in Bioedit (Hall, 1999), and then adjusted manually where necessary. No gap or stop codon was found in the sequences. A total of 124 sequences of a mitochondrial (COI) and two nuclear (28S rDNA and ITS1) markers of the nominal species *B. darwinianus*, *B. rodriguezii* and *B. purpuratus* were obtained. All DNA sequences were deposited in GenBank under the Accession Numbers KC844362–KC844484.

Some mytilids have a unique form of mtDNA inheritance known as 'doubly uniparental inheritance' (DUI) (Zouros, 1992; Skibinski *et al.*, 1994). A maternally inherited mitochondrial genome is present in the eggs and the somatic tissues of female and male individuals, whereas a different paternally inherited mitochondrial genome appears in the male germ line (Rawson & Hilbish, 1995). The paternal mtDNA is preferentially replicated, particularly in the gonad (Skibinski *et al.*, 1994), although there are some exceptions (Garrido-Ramos *et al.*, 1998; Kyriakou *et al.*, 2010). This phenomenon has been found in some species of *Brachidontes*, but not in others (Lee & Ó Foighil, 2004; Terranova *et al.*, 2007). Following Lee & Ó Foighil (2005) we expect that by targeting DNA from posterior adductor muscle tissue, possible problems associated with heteroplasmy were minimized. Infiltration of the muscle tissue by germ line tissue is unlikely, and so we expected it to be dominated by maternally transmitted mitochondria, irrespective of the gender of the individual mussel sampled.

### Within-nominal species genetic structure

Genetic variation of mitochondrial DNA (COI) of *B. darwinianus*, *B. rodriguezii* and *B. purpuratus* was estimated using the number of polymorphic sites (S), number of haplotypes (H), haplotype diversity (Hd), average number of nucleotide differences (k) and nucleotide diversity (Pi) in DnaSP version 5 (Librado & Rosas, 2009). To determine whether

the individuals were in mutation–genetic drift equilibrium of mitochondrial COI sequences a test of Tajima's D was performed in DnaSP. Levels of among-population genetic differentiation were estimated by pairwise  $F_{ST}$  (mtDNA) in Arlequin version 3.5.1.2 (Excoffier & Lischer, 2010) and were visualized constructing a principal component analysis (PCA) implemented in Infostat 2011 (Di Rienzo *et al.*, 2011). Intralineage divergences were calculated using MEGA 4 (Tamura *et al.*, 2007). Maximum-parsimony COI haplotype networks were constructed individually for each nominal species using the median joining algorithm (Bandelt *et al.*, 1999) with default parameters in Network 4.6.1 software (Polzin & Daneschmand, 2003).

### Among-nominal species genetic structure

To calculate interlineage differences (mean genetic distance) between groups of taxa we used MEGA 4 and the method selected was p-distance. To determine the partition that maximizes the differences among groups we performed an analysis of molecular variation (AMOVA; Excoffier *et al.*, 1992) implemented in Arlequin. The AMOVA was performed based on a distance matrix of pairwise differences and the significance was estimated using 10,000 iterations. The pattern of geographic structure was visualized with GenGIS v1.08 (Parks *et al.*, 2009), a bioinformatics application that provides a graphical interface for the merging of information on molecular diversity (DNA sequences) with the geographic location from which the sequences were collected.

### Phylogenetic analysis

Two global phylogenies of *Brachidontes s.l.* were constructed based on COI and 28S rDNA sequences obtained by us and retrieved from the GenBank data base (Table 2); *Ischadium recurvum* and *Geukensia granosissima* were used as outgroups; the same species were also used as outgroups by Lee & Ó Foighil (2005). The phylogeny of *Brachidontes s.l.* could not

**Table 2.** Sequences of COI and 28S rDNA from *Brachidontes* species and other mytilids from other studies used in the phylogenetic analyses, with indication of locations of origin, GenBank Accession codes, and references.

Taxa	Locality	GenBank ID		Reference
		COI	28S rDNA	
<i>Brachidontes semistriatus</i>	Kwazulu-Natal, South Africa	–	AY825099.1	Lee & Ó Foighil, 2005
<i>Brachidontes mutabilis</i>	Japan	–	AB103124.1	Hashimoto & Matsumoto, unpublished data
<i>Brachidontes</i> sp. 1	Darwin Harbor, Australia	–	AY825081.1	Lee & Ó Foighil, 2005
<i>Brachidontes</i> sp. 2	Darwin Harbor, Australia	–	AY825080.1	Lee & Ó Foighil, 2005
<i>Brachidontes semilaevis</i>	Chumical, Panama	AY825117.1	AY825089.1	Lee & Ó Foighil, 2005
<i>Brachidontes adamsianus</i>	Pto.Vallarta/Isla Jicarón, Panama	AY825170.1	AY825091.1–100.1	Lee & Ó Foighil, 2005
<i>Brachidontes exustus I</i>	Veracruz, Mexico	AY825216.1	AY825104.1	Lee & Ó Foighil, 2005
<i>Brachidontes exustus II</i>	Panacea, Florida, USA	AY621900.1	AY621999.1	Lee & Ó Foighil, 2005
<i>Brachidontes exustus III</i>	Boca Chica Key, Florida, USA	–	AY621993.1	Lee & Ó Foighil, 2005
<i>Brachidontes variabilis</i>	Hong Kong	DQ836021.1–20.1	AY825101.1	Lee & Ó Foighil, 2005
<i>Brachidontes modiolus</i>	Key Biscayne, Florida, USA	AY621916.1	AY622002.1	Lee & Ó Foighil, 2005
<i>Brachidontes puniceus</i>	Cape Verde	HM999789.1	–	Cunha <i>et al.</i> , unpublished data
<i>Brachidontes</i> sp. 1	Palau Islands	AB465560.1	–	Goto <i>et al.</i> , 2011
<i>Brachidontes</i> sp. 2	Palau Islands	AB465574.1	–	Goto <i>et al.</i> , 2011
<i>Brachidontes</i> sp. 3	Palau Islands	AB465568.1	–	Goto <i>et al.</i> , 2011
<i>Brachidontes pharaonis</i>	Indian Ocean	DQ836013.1	–	Terranova <i>et al.</i> , 2007
<i>Geukensia granosissima</i>	Marco, Florida, USA	AY621927.1	AY622006.1	Lee & Ó Foighil, 2005
<i>Ischadium recurvum</i>	Panacea, Florida, USA	AY621933.1	AY622009.1	Lee & Ó Foighil, 2005

be resolved using as outgroups members of more distantly related mytilid subfamilies (e.g. *Perna*, *Mytilus*, *Mytella*). To evaluate character congruence among COI and 28S rDNA datasets a partition-homogeneity test (Farris *et al.*, 1995) was performed (100 random replications) using PAUP\*4.0b10 (Swofford, 2003); the phylogeny of the ITS1 gene was not included due to the low number of sequences available in comparison with the other two genes. The two datasets were not significantly incongruent ( $P = 0.08$ ) and were analysed as a combined dataset. The same general tree topology was obtained with the combined data sets and with the COI data alone.

Bayesian analyses were performed on each dataset using Mr Bayes v3.1.2 (Ronquist & Huelsenbeck, 2003) under the best-fit substitution model (HKY + G + I for COI and HKY + G for 28S rDNA datasets) determined by Bayesian information criteria (BIC) as implemented in jModelTest (Guindon & Gascuel, 2003; Posada, 2008). Model parameters were treated as unknown and were estimated for each analysis. To produce one 50% majority rule consensus tree for each analysis, random starting trees were used. Each Bayesian analysis comprised four chains which were sampled every 10,000 generations for a total of 10,000,000 generations and the burn-in used was 25%. Maximum likelihood (ML) analyses were conducted with PAUP 4.0 (Swofford, 1998) using the heuristic search option. Nodal support was estimated through bootstrap analysis (Felsenstein, 1985) using 1000 replications with 10 random additions per each bootstrap replicate. The editing of the trees was carried out in Dendroscope 2.7.4. (Huson *et al.*, 2007).

## RESULTS

### Screening of reference collections

Specimens used for sequencing were assigned to three nominal species differentiated on the basis of shell phenotypes: *Brachidontes darwinianus* (Figure 2A–F), *B. rodriguezii*

(Figure 2G–K) and *B. purpuratus* (Figure 2L). Specimens of *B. rodriguezii* collected at the type locality (Bahia San Blas) (Figure 2J, K) share the phenotypic diagnostic characters of the species, defined by Adami *et al.* (2013) observable in the type series (BMNH 1854-12-4-809, 6 specimens consisting of paired valves, two of them shown in Figure 2H, I). The specimen presumably used by d'Orbigny (1842) for illustration (Figure 2G; BMNH1854.12.4.809/1) is easily singled out in the series (Figure 2H), and is the one selected as lectotype by Aguirre (1994). Shell ribs are relatively thin as compared to other specimens (Figure 2I, J), but diagnostic characters (Adami *et al.*, 2013) are easily observable. The location of origin of specimens of *B. darwinianus* deposited in the British Museum (BMNH 1854-12-4-799 (six paired shells), -800 (four specimens) and -801 (five paired plus three single shells)) is not clearly identified (three locations were indicated by d'Orbigny, 1842), but specimens collected in Punta Canario (Montevideo, close to Maldonado, one of d'Orbigny's locations) fall within the range of phenotypic variation of the syntype series (compare Figure 2C, E with Figure 2D, F). We were unable to confirm the unlikely presence of this species in northern Argentine Patagonia. Directed collections made in Bahia Rosas (= Ensenada de Ros, one of the three locations indicated by d'Orbigny) yielded small specimens of *B. rodriguezii* (Figure 2K) with an outline that resembles the subtriangular shell outline of the specimen of *B. darwinianus* selected by d'Orbigny for illustration (Figure 2A, B). We did not include *B. solisianus* in this study because its occurrence in the region of interest remains unconfirmed. Based on phenotypic characters, the type series (BMNH 1854-12-4-797, three paired and three single shells) is difficult to separate from eroded specimens of *B. rodriguezii*. Re-examination of the holotype of *B. blakeanus*, deposited in The Manchester Museum (Catalogue # EE.7674, Figure S1) revealed that it does not correspond to a mytilid, but rather belongs in the genus *Philobrya* Carpenter (family Philobryidae) (see Supplementary Material). Kelaher *et al.* (2007) reported *B. granulatus* for

the intetidal zone of Argentine Patagonia, where its occurrence is considered unlikely. No voucher specimens from that study appear to exist (Professor J.C. Castilla, personal communication). Extensive studies conducted in the same region and habitats failed to confirm its presence (Liuzzi & López-Gappa, 2008).

### Within-nominal species genetic structure

Genetic diversity indices such as the number of polymorphic sites (*S*), number of haplotypes (*h*), haplotype diversity (*Hd*), nucleotide differences (*k*) and nucleotide diversity (*Pi*) were variable among the three nominal species (Table 3). *Brachidontes purpuratus* showed higher diversity values than *B. darwinianus* and *B. rodriguezii*. Haplotype diversity was similar among *B. darwinianus* and *B. rodriguezii*. Tajima's *D* test was negative for all nominal species but was significant only for *B. purpuratus*, suggesting population expansion for this species but not for the others.

Pairwise  $F_{ST}$  values within *B. rodriguezii* indicate that the samples from Santa Clara del Mar differ from the other localities (0.32–0.60) (Figure 3). On the other hand, pairwise  $F_{ST}$  values within *B. purpuratus* indicate a lack of genetic differentiation among localities (Figure 3). Pairwise  $F_{ST}$  values were not estimated for *B. darwinianus* because all haplotypes were obtained from a single location. Individual median-joining network analyses resulted in star-like genealogies for *B. darwinianus* and *B. rodriguezii* (Figure 4A, B) and in a more expanded genealogy for *B. purpuratus* (Figure 4C). No haplotype is shared among the three nominal species. In the network analysis for *B. darwinianus*, notwithstanding the fact that all samples for this species come from a single locality, haplotype 1 has the highest frequency and many connections (Figure 4A), while in the analysis for *B. rodriguezii* (Figure 4B) haplotype 4 is the most frequent and shows a wide geographical distribution. *Brachidontes rodriguezii* from Santa Clara del Mar do not share haplotypes with the other localities to the south, a result consistent with the strong genetic differentiation revealed by the pairwise  $F_{ST}$  analysis mentioned earlier. *Brachidontes purpuratus* showed a complex genealogy (Figure 4C) with high levels of genetic similarity among populations. These results support the low level of genetic differentiation among localities indicated by the pairwise  $F_{ST}$  analysis.

### Among-nominal species genetic structure

The pairwise  $F_{ST}$  among localities of mtCOI point out a significant genetic differentiation among the three nominal species (0.95–0.99; Figure 3). In addition, the intralocus

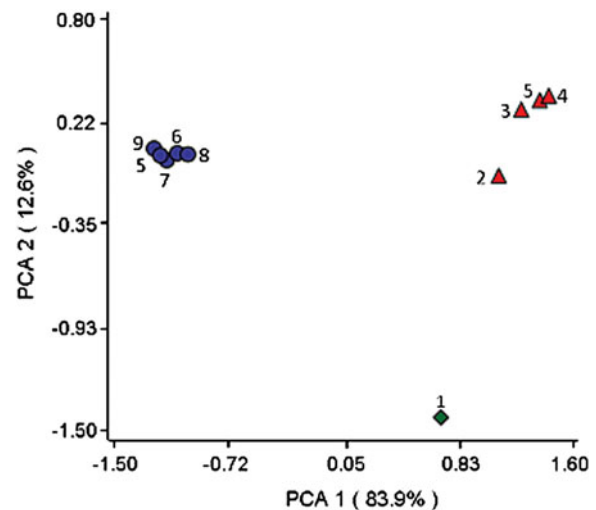


Fig. 3. Principal component analysis (PCA) of  $F_{ST}$  obtained from the COI mitochondrial gene. (◆) *B. darwinianus*, (▲) *B. rodriguezii*, (●) *B. purpuratus*. Numbers indicate locations: 1, Punta Canario; 2, Santa Clara del Mar; 3, Bahia San Blas; 4, Bahia Rosas; 5, Puerto Madryn; 6, Caleta Carolina; 7, Puerto Deseado; 8, Surfer Bay; 9, Bahia Ensenada (see Table 1 for more information).

divergences within *B. darwinianus*, *B. rodriguezii* and *B. purpuratus* ranged between 0.1% and 1.1% ( $SE = 0-0.3$ ), whereas the interlineage sequence differences ranged from 20.0% to 21.0% (Table 4). The hypothesis of panmixia, tested with the AMOVA with all localities of the three nominal species as one-gene pool, was rejected ( $P < 0.05$ ). We found that variation among phenotypically defined groups ('darwinianus', 'rodriguezii' and 'purpuratus'; Table 5) explain 96.61%, 99.79%, and 99.87% of the genetic variance in COI, 28S rDNA and ITS1, respectively. Strong genetic differentiation is consistent with the genetic divergence detected by the three molecular markers observed in the phylograms (Figure 5A–C). In all the recovered trees, *B. purpuratus* is sister to the well supported clade formed by *B. rodriguezii* and *B. darwinianus*.

### Phylogenetic analysis

Phylogenies of *Brachidontes s.l.* constructed from the nuclear COI and 28S rDNA datasets are shown in Figure 6. Both ML and Bayesian analyses reveal two well-supported nuclear/mitochondrial clades sister to *Geukensia/Ischadium*. One clade was formed by the specimens of *B. purpuratus* (Bayesian posterior probabilities 0.99/ML bootstrap 100 and 1.00/100, from both mitochondrial and nuclear genes trees) and the other clade includes all the other species of *Brachidontes s.l.* analysed. In the latter clade, the specimens of *B. rodriguezii* form a strongly supported clade (Bayesian posterior probabilities 1.00/ML bootstrap 99 and 1.00/100, from both mitochondrial and nuclear genes trees), separated from the clade formed by the specimens of *B. darwinianus* (Bayesian posterior probabilities 1.00/ML bootstrap 100 and 0.98/100, from both mitochondrial and nuclear genes trees). For *B. darwinianus*, the topologies obtained from the COI mitochondrial gene were generally consistent with those from the 28S rDNA nuclear gene, although some differences were observed. In the mitochondrial tree, *B. rodriguezii* is sister to a well supported clade containing a polytomy

Table 3. Genetic diversity indices for *Brachidontes s.l.* species based on mtDNA (COI) sequences. *n*, number of specimens analysed; *h*, number of haplotypes; *S*, number of polymorphic sites; *Hd*, haplotype diversity; *k*, average number of nucleotide differences; *Pi*, nucleotide diversity. Number in bold has  $P$ -value  $< 0.05$ .

Nominal species	<i>n</i>	<i>h</i>	<i>S</i>	<i>Hd</i>	<i>k</i>	<i>Pi</i>	Tajima's <i>D</i>
<i>B. darwinianus</i>	8 <sup>a</sup>	4	3	0.64	0.75	0.001	−1.45
<i>B. rodriguezii</i>	31	10	15	0.69	2.21	0.004	−1.37
<i>B. purpuratus</i>	39	30	41	0.98	4.54	0.010	−1.88

a, all haplotypes derived from a single location.

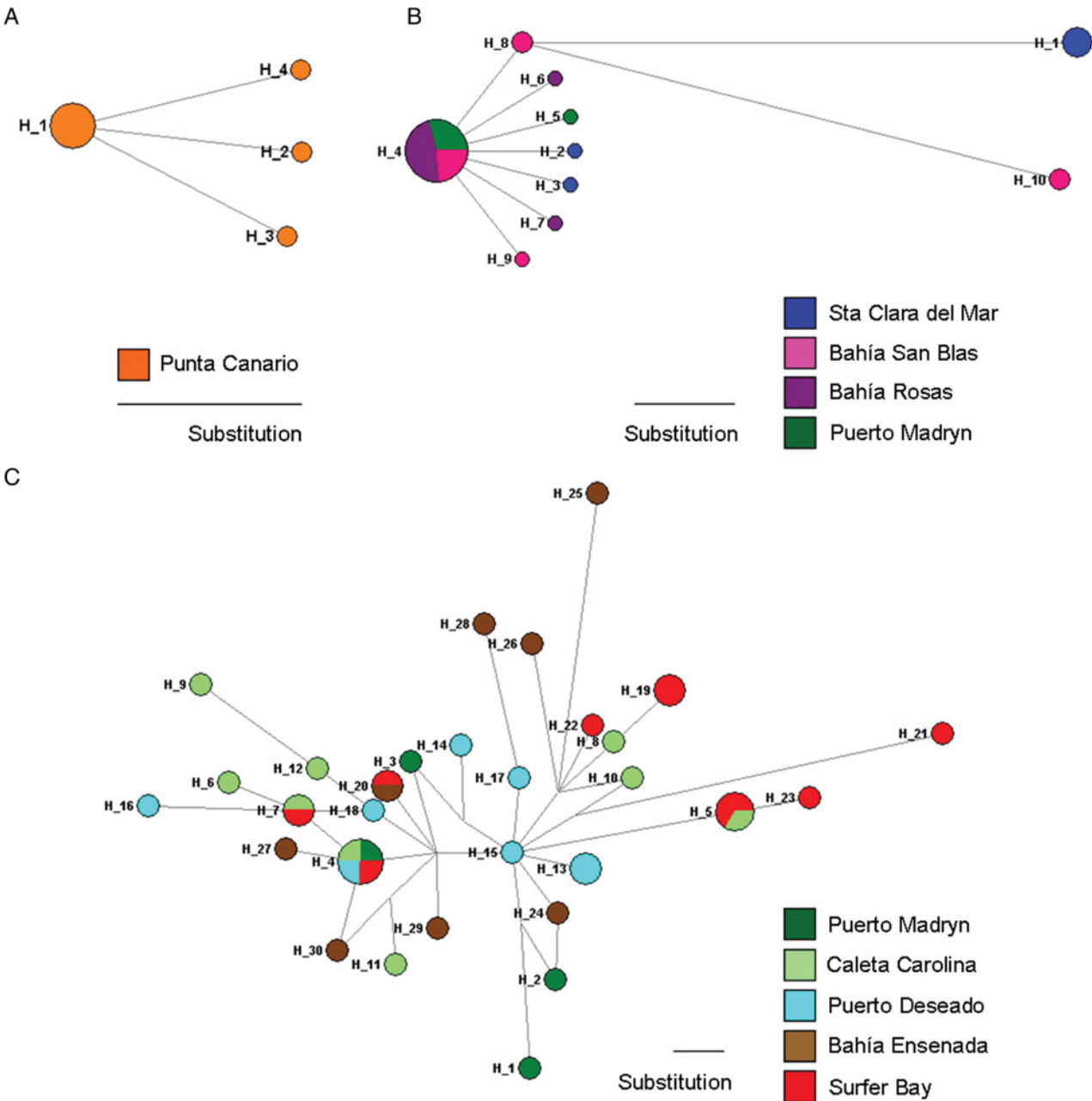


Fig. 4. Median joining haplotype networks based on COI mitochondrial gene. (A) *Brachidontes darwinianus*; (B) *B. rodriguezii*; (C) *B. purpuratus*. Each haplotype is represented by a circle whose size is proportional to its frequency; colours indicate locality of origin. Latitude and longitude information for each locality indicated on Table 1.

which includes *B. darwinianus* and the other species of *Brachidontes* studied (Figure 6A); while in the nuclear gene tree, *B. darwinianus* is basal to the remainder species (Figure 6B).

Table 4. Uncorrected genetic distances among the three species of *Brachidontes s.l.* present in the south-western Atlantic, based on the gene COI. Between groups mean distance below the diagonal; *P* values above the diagonal.

	<i>B. darwinianus</i>	<i>B. rodriguezii</i>	<i>B. purpuratus</i>
<i>B. darwinianus</i>	–	0.016	0.016
<i>B. rodriguezii</i>	0.21	–	0.017
<i>B. purpuratus</i>	0.20	0.20	–

DISCUSSION

Status of *Brachidontes* species from the south-western Atlantic

Mitochondrial and nuclear DNA sequences revealed three different lineages of *Brachidontes s.l.* in the south-western Atlantic. These units satisfy the phylogenetic species concept (Nixon & Wheeler, 1990) and correspond to *B. darwinianus*, *B. rodriguezii* and *B. purpuratus*. The AMOVA for all three genes, the pairwise  $F_{ST}$  and the genetic distance analyses for COI consistently indicate a high level of genetic divergence among the three species. A variety of metazoan species exhibit what has come to be known as a barcoding gap in

**Table 5.** Molecular variation analysis (AMOVA) of the genes COI, 28S rDNA and ITS1 for the three species of *Brachidontes* present in the south-western Atlantic.

Gene	Source of variation	df	Fixation indices	% of variation
COI	<i>darwinianus</i> vs <i>rodriguezii</i> vs <i>purpuratus</i>	2	$F_{CT} : 0.97^*$	96.61
	Among populations within groups	7	$F_{ST} : 0.97^*$	0.50
	Within populations	68	$F_{SC} : 0.15^*$	2.89
28S rDNA	<i>darwinianus</i> vs <i>rodriguezii</i> vs <i>purpuratus</i>	2	$F_{CT} : 0.99^*$	99.79
	Among populations within groups	7	$F_{ST} : 1.00^*$	0.21
	Within populations	18	$F_{SC} : 1.00^*$	0.00
ITS1	<i>darwinianus</i> vs <i>rodriguezii</i> vs <i>purpuratus</i>	2	$F_{CT} : 0.99^*$	99.87
	Among populations within groups	7	$F_{ST} : 0.99^*$	-0.01
	Within populations			

\*,  $P < 0.05$ .

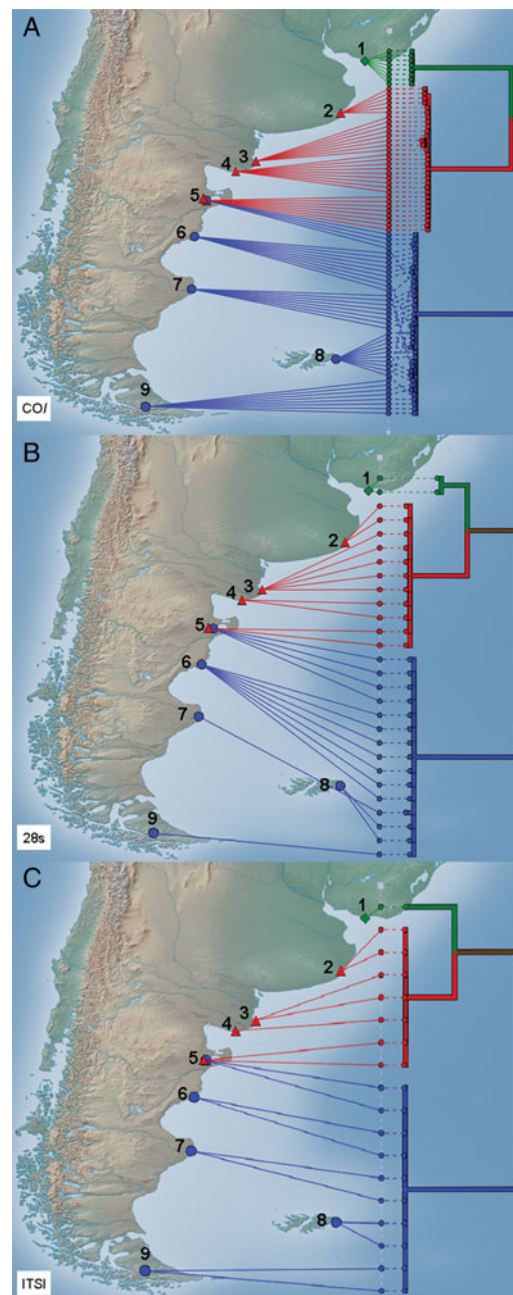
the COI region of the mitochondrial DNA whereby intraspecific divergences are typically  $< 3\%$ , while interspecific divergences range between 10% and 25% (Hebert *et al.*, 2003; Bucklin *et al.*, 2011). The estimated genetic distances among the three *Brachidontes* groups in our study (20–21%) suggest that these groups are indeed three different species. Hypotheses postulating that *B. darwinianus* and *B. rodriguezii* are junior synonyms of *B. purpuratus* (Aguirre *et al.*, 2006a, 2008), and that *B. darwinianus* is a junior synonym of *B. exustus* (Rios, 1994), are thus rejected. This is consistent with results on chromosomal mapping of several genes in *B. purpuratus* and *B. rodriguezii* (Pérez-García *et al.*, 2010a, b).

In the south-western Atlantic *B. purpuratus* is confined to the cold-temperate region (Magellanic Biogeographic Province), while *B. darwinianus* and *B. rodriguezii* occur in the warm-temperate region (Argentine Biogeographic Province) (Figure 1). Our results confirm that *B. rodriguezii* and *B. purpuratus* coexist in the same beds along the transition zone ( $41-43^\circ\text{S}$ ), represented in our study by the collections from Puerto Madryn. The presence of *B. darwinianus* in the Montevideo area (La Plata River estuary) is consistent with previous studies indicating that this is an estuarine species (Avelar & Narchi, 1983; Scarabino *et al.*, 2006).

The status of other species reported for the region is questionable. *Brachidontes solisianus* needs to be evaluated with materials from south-eastern Brazil, north of our area of focal interest. *Brachidontes blakeanus*, originally described from the Falkland/Malvinas Islands is not even a mytilid, and must be transferred to the genus *Philobrya* (Philobryidae) (see Supplementary Material). Records of *B. blakeanus* and *B. granulatus* from Argentine Patagonia may correspond to other species, most likely *B. purpuratus* or juveniles of other mytilids.

### Within-species genetic structure and phylogeographic implications

The low genetic differentiation among populations of *B. purpuratus*, coupled with the negative sign for Tajima's D, suggest that this species experienced a recent population expansion in



**Fig. 5.** Association of the phylogenetic relationship among *Brachidontes* species and their geographic distribution in the region of interest based on (A) mtDNA COI, (B) nuclear 28S rDNA and (C) nuclear ITS1. (◆) *B. darwinianus*, (▲) *B. rodriguezii*, (●) *B. purpuratus*. Numbers indicate locations: 1, Punta Canario; 2, Santa Clara del Mar; 3, Bahía San Blas; 4, Bahía Rosas; 5, Puerto Madryn; 6, Caleta Carolina; 7, Puerto Deseado; 8, Surfer Bay; 9, Bahía Ensenada (see Table 1 for more information).

the south-western Atlantic. Similar patterns have been documented for the notothenioid fish *Eleginops maclovinus* (Ceballos *et al.*, 2012) and the limpet *Nacella magellanica* (de Aranzamendi *et al.*, 2011), both endemic to the Magellanic Province. The cold-temperate portion of the south-western Atlantic, most of which was never glaciated, must be considered in the context of Quaternary glaciation episodes (Rabassa *et al.*, 2011; Fraser *et al.*, 2012). At the time of the Last Glacial Maximum (LGM, ca. 23–25 cal. ka BP) the coasts of Chile south of approximately  $43^\circ\text{S}$  were

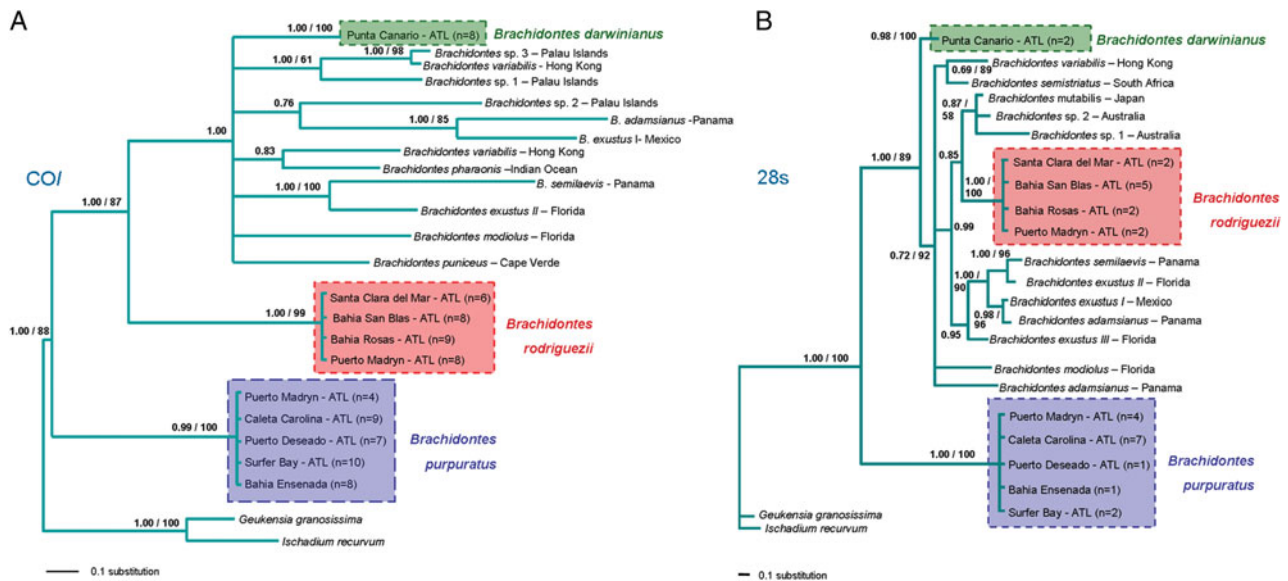


Fig. 6. Bayesian trees of *Brachidontes* s.l. based on (A) the mitochondrial gene COI and (B) the nuclear gene 28S rDNA, including species sequenced by us and in previous studies and using *Ischadium recurvum* and *Geukensia granosissima* (Mytilinae) as outgroups. Numbers above the branches represent the Bayesian posterior probabilities/maximum likelihood bootstrap values (>60 only) for the supported nodes. Numbers in parentheses following sampling locations indicate multiple individuals sharing the same genotype.

covered by ice (Ruzzante *et al.*, 2008, their figure 1). At the same time the coastline of the south-west Atlantic was located 200–400 km east of its current position depending on latitude (Ponce *et al.*, 2011), and was presumably dominated by depositional environments. As the coastline moved westward following a rise in sea level during post-LGM millennia, availability of hard substrates and sheltered habitats was very limited, in contrast to the convoluted coastline and rocky shores (sheltered and exposed) from the south-east Pacific. It is likely that hard substrates in the south-western Atlantic, dominated by abrasion platforms of friable sedimentary rocks, were colonized by immigrants from the south-east Pacific, where diversification of invertebrates associated with hard substrates was likely facilitated by habitat diversity (González-Wevar *et al.*, 2011) and glaciation-related fragmentation of coastscapes (Fraser *et al.*, 2012). Low genetic differentiation of *Nacella magellanica* and *B. purpuratus* in the south-western Atlantic could be explained by relatively recent (post-LGM) colonization originating from populations in the south-eastern Pacific. *Brachidontes purpuratus* is well represented in Early Pleistocene–Holocene terraces of Argentine Patagonia (Aguirre *et al.*, 2006b), but no related fossils are recorded from earlier periods, including the rich Late Miocene deposits of northern Patagonia (Madrin Formation; del Río & Martínez, 1998), a region of overlap with *B. rodriguezii* during Holocene and Recent times. Testing the hypothesis of a post-LGM range expansion of *B. purpuratus* will require further research on haplotypic diversity along the south-eastern Pacific, which is beyond the scope of the present study.

*Brachidontes purpuratus* is the only species of *Brachidontes* s.l. whose range extends into cold-temperate waters, encompassing two entire biogeographic provinces: Magellanic (cold-temperate) and Chile–Peru (warm-temperate). A broad transition zone between these two provinces extends approximately between 32° and 42° (Fernández *et al.*, 2000). We doubt that the low genetic differentiation observed in

the south-western Atlantic will be mirrored in the south-east Pacific once sequences become available from that region. In fact, Pérez *et al.* (2008) observed significant genetic heterogeneity (using microsatellite markers) between populations from central Chile (Talca, 32°26'S, warm-temperate region) and Argentine Patagonia (Puerto Lobos, 41°42'S, cold-temperate region), the latter close to the northern range limit of the species in the south-west Atlantic. Genetic diversity related to the biogeographic divide of the south-east Pacific has been reported for other groups of marine organisms (Fraser *et al.*, 2010; Macaya & Zuccarello, 2010).

In contrast to the low level of genetic differentiation observed in *B. purpuratus*, pairwise  $F_{ST}$  values in *B. rodriguezii* indicate that the samples from the northernmost location where this species was sampled in our study, Santa Clara del Mar, differ from the other localities to the south. Sequenced specimens from Santa Clara del Mar do not share haplotypes with other localities, a result consistent with the strong genetic differentiation revealed by the pairwise  $F_{ST}$  analysis mentioned earlier. Between Santa Clara del Mar and the other locations to the south considered in this study (Bahia San Blas, Bahia Rosas, Puerto Madryn) there are hundreds of kilometres of coastline consisting of exposed sandy beaches and mud flats (SEGEMAR, 2000), habitats not occupied by *Brachidontes* species. This scenario created a substantial barrier to dispersal by means of pelagic larvae. The three locations from northern Patagonia, on the contrary, are connected by a continuum of intertidal hard substrate habitats (SEGEMAR, 2000). Fossil shells of *B. rodriguezii*, presumably of Pleistocene age, have been recovered from cores obtained at 68 m depth off of Mar del Plata (Richards & Craig, 1963), near to our sampling sites, confirming a long presence of the species in that region during the Quaternary. Fossils of *Brachidontes* from the Late Miocene (Madrin and Paraná Formation; del Río & Martínez, 1998) are phenotypically close enough to *B. rodriguezii* to have been classified as a subspecies, *B. r. lepida* (Philippi, 1893). Genetic intraspecific

differentiation, a long history of occurrence in the region, and the phylogenetic position of *B. rodriguezii* within the main *Brachidontes* clade in the COI-based phylogeny (discussed later) prompt the hypothesis that *B. rodriguezii* is derived from a local stock, with a history of presence in the warm-temperate waters of the south-western Atlantic traceable to the Late Miocene. Evaluation of this hypothesis will require the concatenation of new molecular, palaeontological and morphometric data.

### Phylogenetic relations of *Brachidontes s.l.*

The Mytilidae have been classified into a variable number of subfamilies (see Huber, 2010, for an overview). Several studies (Distel, 2000; Matsumoto, 2003; Owada & Hoeksema, 2011) have contributed substantially to the clarification of the relationships among mytilid genera based on genetic data. Huber (2010) summarized available information and subdivided the Mytilidae into ten subfamilies. *Brachidontes s.l.* (including *Hormomya* Mörch, *Austromytilus* Laseron, and *Perumytilus* Olsson) was placed in a revived Brachidontinae Nordsieck, together with *Mytilaster* Monterosato and the modioliform *Geukensia* van de Poel (after genetic results presented by Distel, 2000).

Our results strongly support an early divergence of *B. purpuratus* from the other species of *Brachidontes s.l.*. Studies involving other species of *Brachidontes* and related genera will be required before the taxonomy of the group is settled.

In contrast to *B. purpuratus*, *B. darwinianus* and *B. rodriguezii* are nested within a clade that includes all the other *Brachidontes* species for which sequences of our target genes (nuclear and mitochondrial) are available. Discordance between phylogenies based on different genes regarding the position of *B. rodriguezii* within the clade is not uncommon in phylogenetic contexts. Quite often, this occurrence has been attributed to introgression or hybridization, but can also be explained by differences in mutational rates between marker types, sensitivity to effective population size, incomplete lineage sorting, or mode of inheritance (Ballard & Whitlock, 2004). One potential generic cause of the partial incongruence between the nuclear and mitochondrial DNA-based trees is a weak phylogenetic signal, which may result in poor phylogenetic resolution or inaccurate gene trees as an artefact of phylogenetic reconstruction (Funk & Omland, 2003). Further studies including more species of the *Brachidontes s.l.* and related genera, covering also other regions of the world ocean are needed to resolve the full phylogeny of the group.

While their intrinsic and extrinsic traits (*sensu* Webb *et al.*, 2002) overlap, the small mussels (*Brachidontes* spp. *s.l.*) that dominate the appearance of intertidal communities along rocky shores of the south-western Atlantic belong to different lineages. Determining whether this reflects evolutionary convergence or conservation of ancestral traits will require a better resolution of mytilid phylogeny.

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### Supplementary materials and methods

The Supplementary material referred to in this paper can be found online at [journals.cambridge.org/mbi](http://journals.cambridge.org/mbi).

### REFERENCES

- Adami M.L., Pastorino G. and Orensanz J.M. (2013) Phenotypic differentiation of ecologically-significant *Brachidontes* species co-occurring in intertidal mussel beds from the southwestern Atlantic. *Malacologia* 56.
- Aguirre M.L. (1994) Type specimens of Quaternary marine bivalves from Argentina. *Ameghiniana* 31, 347–374.
- Aguirre M.L., Perez S.I. and Negro-Sirch Y. (2006a) Morphological variability of *Brachidontes* Swainson (Bivalvia, Mytilidae) in the marine Quaternary of Argentina (SW Atlantic). *Palaeogeography, Palaeoclimatology, Palaeoecology* 239, 100–125.
- Aguirre M.L., Richiano S. and Negro-Sirch Y. (2006b) Palaeoenvironments and palaeoclimates of the Quaternary molluscan faunas from the coastal area of Bahia Vera-Camarones (Chubut, Patagonia). *Palaeogeography, Palaeoclimatology, Palaeoecology* 229, 251–286.
- Aguirre M.L., Hlebszevitsch-Savalsky J.C. and Dellatorre F. (2008) Late Cenozoic invertebrate paleontology of Patagonia and Tierra del Fuego, with emphasis on molluscs. In Rabassa J. (ed.) *The Late Cenozoic of Patagonia and Tierra del Fuego*. Amsterdam: Elsevier, pp. 285–324.
- Avelar W.E.P. and Narchi W. (1983) Behavioral aspects of *Brachidontes darwinianus darwinianus* (Orbigny, 1846) and *Brachidontes solisianus* (Orbigny, 1846) (Bivalvia, Mytilidae) in response to a salinity gradient. *Iheringia (Zool)* 63, 125–132.
- Balech E. and Ehrlich M.D. (2008) Esquema Biogeográfico del Mar Argentino. *Revista de Investigación y Desarrollo Pesquero* 19, 45–75.
- Ballard J.W.O. and Whitlock M.C. (2004) The incomplete natural history of mitochondria. *Molecular Ecology* 13, 729–744.
- Bandelt H.J., Forster P.I. and Röhl A. (1999) Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution* 16, 37–48.

- Briggs J.C. and Bowen B.W. (2012) A realignment of marine biogeographic provinces with particular reference to fish distributions. *Journal of Biogeography* 39, 12–30.
- Bucklin A., Steinke D. and Blanco-Bercial L. (2011) DNA Barcoding of marine metazoa. *Annual Review of Marine Science* 3, 471–508.
- Ceballos S.G., Lessa E.P., Victorio M.F. and Fernández D.A. (2012) Phylogeography of the sub-Antarctic notothenioid fish *Eleginops maclovinus*: evidence of population expansion. *Marine Biology* 159, 499–505.
- Coan E.V. and Valentich-Scott P. (2012) *Bivalve seashells of tropical West America. Marine bivalve mollusks from Baja, California to Northern Peru*. Santa Barbara, CA: Santa Barbara Museum.
- de Aranzamendi M.C., Bastida R. and Gardenal C.N. (2011) Different evolutionary histories in two sympatric limpets of the genus *Nacella* (Patellogastropoda) in the South-western Atlantic coast. *Marine Biology* 158, 2405–2418.
- del Río C. and Martínez S.A. (1998) Moluscos marinos miocenos de la Argentina y del Uruguay. *Monografías de la Academia Nacional de Ciencias Exactas, Físicas y Naturales* 15, 95 pp.
- Di Rienzo J.A., Casanoves F., Balzarini M.G., Gonzalez L., Tablada M. and Robledo C.W. (2011) *InfoStat*. Grupo InfoStat FCA, Universidad Nacional de Córdoba, Argentina.
- Distel D.L. (2000) Phylogenetic relationships among Mytilidae (Bivalvia): 18S rRNA data suggest convergence in mytilid body plans. *Molecular Phylogenetics and Evolution* 15, 25–33.
- d'Orbigny A. (1842) *Mollusques. Voyage dans l'Amérique Méridionale* 5. Paris: P. Bertrand, pp. 83–85.
- d'Orbigny A. (1846) *Mollusques. Voyage dans l'Amérique Méridionale* 5. Paris: P. Bertrand, pp. 489–758.
- Excoffier L. and Lischer H.E.L. (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology and Resources* 10, 564–567.
- Excoffier L., Smouse P.E. and Quattro J.M. (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* 131, 479–491.
- Farris J.S., Källersjö M., Kluge A.G. and Bult C. (1995) Testing significance of congruence. *Cladistics* 10, 315–319.
- Felsenstein J. (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39, 783–791.
- Fernández M., Jaramillo E., Marquet P.A., Moreno C.A., Navarrete S.A., Ojeda F.P., Valdovinos C.R. and Vásquez J.A. (2000) Diversity, dynamics and biogeography of Chilean benthic nearshore ecosystems: an overview and guidelines for conservation. *Revista Chilena de Historia Natural* 73, 797–830.
- Folmer O., Black M., Hoeh W., Lutz R. and Vrijenhoek R. (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3, 294–299.
- Fraser C.I., Thiel M., Spencer H.G. and Waters J.M. (2010) Contemporary habitat discontinuity and historic glacial ice drive genetic divergence in Chilean kelp. *Evolutionary Biology* 10, 203.
- Fraser C.I., Nikula R., Ruzzante D.E. and Waters J.M. (2012) Poleward bound: biological impacts of Southern Hemisphere glaciation. *Trends in Ecology and Evolution* 27, 462–471.
- Funk D.J. and Omland K.E. (2003) Species-level paraphyly and polyphyly: frequency, causes, and consequences, with insights from animal mitochondrial DNA. *Annual Review of Ecology, Evolution and Systematics* 34, 397–423.
- Garrido-Ramos M.A., Stewart D.T., Sutherland B.W. and Zouros E. (1998) The distribution of male-transmitted and female-transmitted mitochondrial DNA types in somatic tissues of blue mussels: implications for the operation of doubly uniparental inheritance of mitochondrial DNA. *Genome* 41, 818–824.
- González-Wevar C.A., Nakano T., Cañete J.I. and Poulin E. (2011) Concerted genetic, morphological and ecological diversification in *Nacella* limpets in the Magellanic Province. *Molecular Ecology* 20, 1936–1951.
- Goto T.V., Tamate H.B. and Hanzawa N. (2011) Phylogenetic characterization of three morphs of mussels (Bivalvia, Mytilidae) inhabiting isolated marine environments in Palau Islands. *Zoological Science* 28, 568–579.
- Guindon S. and Gascuel O. (2003) A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Systematic Biology* 52, 696–704.
- Hall T.A. (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41, 95–98.
- Hebert P.D.N., Cywinska A., Ball S.L. and deWaard J.R. (2003) Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London, Series B—Biological Sciences* 270, 313–322.
- Huber M. (2010) *Compendium of bivalves. A full-color guide to 3300 of the world's marine bivalves. A status on Bivalvia after 250 years of research*. Hackenheim: ConchBooks.
- Huson D.H., Richter D.C., Rausch C., DeZulian T., Franz M. and Rupp R. (2007) Dendroscope: an interactive viewer for large phylogenetic trees. *Bioinformatics* 8, 460.
- Kelahr B.P., Castilla J.C., Prado L., York P., Schwindt E. and Bortolus A. (2007) Spatial variation in molluscan assemblages from coralline turfs of Argentinean Patagonia. *Journal of Molluscan Studies* 73, 139–146.
- Kyriakou E., Zouros E. and Rodakis G.C. (2010) The atypical presence of the paternal mitochondrial DNA in somatic tissues of male and female individuals of the blue mussel species *Mytilus galloprovincialis*. *BMC Research Notes* 3, 222.
- Lee T. and Ó Foighil D. (2004) Hidden Floridian biodiversity: mitochondrial and nuclear gene trees reveal four cryptic species within the scorched mussel, *Brachidontes exustus*, species complex. *Molecular Ecology* 13, 3527–3542.
- Lee T. and Ó Foighil D. (2005) Placing the floridian marine genetic disjunction into regional evolutionary context using the scorched mussel, *Brachidontes exustus*, species complex. *Evolution* 59, 39–58.
- Librado P. and Rozas J. (2009) DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25: 1451–1452.
- Liuzzi M.G. and López Gappa J. (2008) Macrofaunal assemblages associated with coralline turf: species turnover and changes in structure at different spatial scales. *Marine Ecology Progress Series* 363, 147–156.
- Macaya E.C. and Zuccarello G.C. (2010) Genetic structure of the giant kelp *Macrocystis pyrifera* along the southeastern Pacific. *Marine Ecology Progress Series* 420, 103–112.
- Matsumoto M. (2003) Phylogenetic analysis of the subclass Pteriomorpha (Bivalvia) from mtDNA COI sequences. *Molecular Phylogenetics and Evolution* 27, 429–440.
- Melville J.C. and Standen R. (1914) Notes on Mollusca collected in the North-west Falklands by Mr Rupert Vallentin FLS, with descriptions of six new species. *Annual Magazine of Natural History* 13, 110–137.
- Mouquet N., Devictor V., Meynard C.N., Munoz F., Bersier L.F., Chave J., Couteron P., Dalecky A., Fontaine C., Gravel D., Hardy O.J., Jabot F., Lavergne S., Leibold M., Mouillot D., Münkemüller T.,

- Pavoine S., Prinzing A., Rodrigues A.S.L., Rohr R.P., Thébault E. and Thuiller W. (2012) Ecophylogenetics: advances and perspectives. *Biological Reviews* 87, 769–785.
- Nixon K.C. and Wheeler Q.D. (1990) An amplification of the phylogenetic species concept. *Cladistics* 621, 1–223.
- Olsson A.A. (1961) *Mollusks of the tropical eastern Pacific. Panamic-Pacific Pelecypoda*. Ithaca, NY: Paleontological Research Institution.
- Owada M. and Hoeksema B.W. (2011) Molecular phylogeny and shell microstructure of *Fungiacava eilatensis* Goreau et al. 1968, boring into mushroom corals (Scleractinia: Fungiidae), in relation to other mussels (Bivalvia: Mytilidae). *Contributions to Zoology* 80, 169–178.
- Park J. and Ó Foighil D. (2000) Sphaeriid and corbiculid clams represent separate heterodont bivalve radiations into freshwater environments. *Molecular Phylogenetics and Evolution* 14, 75–88.
- Parks D.H., Porter M., Churcher S., Wang S., Blouin C., Whalley J., Brooks S. and Beiko R.G. (2009) GenGIS: a geospatial information system for genomic data. *Genome Research* 19, 1896–1904.
- Pérez M., Guíñez R., Llanova A., Toro J.E., Astorga M. and Presa P. (2008) Development of microsatellite markers for the ecosystem bioengineer mussel *Perumytilus purpuratus* and cross-priming testing in six Mytilinae genera. *Molecular Ecology Resources* 8, 449–451.
- Pérez-García C., Cambeiro J.M., Morán P. and Pasantes J.J. (2010a) Chromosomal mapping of rDNAs, core histone genes and telomeric sequences in *Perumytilus purpuratus* (Bivalvia: Mytilidae). *Journal of Experimental Marine Biology and Ecology* 395, 199–205.
- Pérez-García C., Guerra-Varela J., Morán P. and Pasantes J.J. (2010b) Chromosomal mapping of rRNA genes, core histone genes and telomeric sequences in *Brachidontes puniceus* and *Brachidontes rodriguezi* (Bivalvia, Mytilidae). *BMC Genetics* 11, 1–8.
- Polzin T. and Daneschmand S.V. (2003) On Steiner trees and minimum spanning trees in hypergraphs. *Operations Research Letters* 31, 12–20.
- Ponce J.F., Rabassa J., Coronato A. and Borromei A.M. (2011) Palaeogeographical evolution of the Atlantic coast of Pampa and Patagonia from the last glacial maximum to the Middle Holocene. *Biological Journal of the Linnean Society* 103, 363–379.
- Posada D. (2008) jModelTest: Phylogenetic Model Averaging. *Molecular Biology and Evolution* 25, 1253–1256.
- Rabassa J., Coronato A. and Martínez O. (2011) Late Cenozoic glaciations in Patagonia and Tierra del Fuego: an updated review. *Biological Journal of the Linnean Society* 103, 316–335.
- Rawson P.D. and Hilbish T.J. (1995) Evolutionary relationships among the male and female mitochondrial DNA lineages in the *Mytilus edulis* species complex. *Molecular Biology and Evolution* 12, 893–901.
- Richards H.G. and Craig J.R. (1963) Pleistocene sedimentation and fauna of the Argentine shelf. II. Pleistocene mollusks from the continental shelf of Argentina. *Proceedings of the Academy of Natural Sciences of Philadelphia* 115, 113–152.
- Ríos C., Mutschke E. and Morrison E. (2003) Benthic sublittoral biodiversity in the Strait of Magellan, Chile. *Revista de Biología Marina y Oceanografía* 38, 1–12.
- Rios E.C. (1994) *Seashells of Brazil*. Rio Grande do Sul: Museu Oceanográfico da FURG.
- Ronquist F. and Huelsenbeck J.P. (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. *BMC Bioinformatics* 19, 1572–1574.
- Ruzzante D.E., Walde S.J., Gosse J.C., Cussac V.E., Habit E., Zemlak T.S. and Adams E.D.M. (2008) Climate control on ancestral population dynamics: insight from Patagonian fish phylogeography. *Molecular Ecology* 17, 2234–2244.
- Sambrook J., Fritsch E.F. and Maniatis T. (1989) *Molecular cloning. A laboratory manual*. New York: Cold Spring Harbor Laboratory Press.
- Scarabino F. (2003) Lista sistemática de los Bivalvia marinos y estuarinos vivientes de Uruguay. *Comunicaciones de la Sociedad Malacológica del Uruguay* 8, 229–259.
- Scarabino F., Zaffaroni J.C., Clavijo C., Carranza A. and Nin M. (2006) Bivalvos marinos y estuarinos de la costa uruguaya: faunística, distribución, taxonomía y conservación. In Menafrá R., Rodríguez-Gallego L., Scarabino F. and Conde D. (eds) *Bases para la Conservación y el Manejo de la Costa Uruguaya*. Montevideo: Vida Silvestre Publicaciones, pp. 157–170.
- Scarabino V. (1977) Moluscos del Golfo San Matías (Prov. De Río Negro, Rep. Argentina). *Comunicaciones de la Sociedad Malacológica del Uruguay* 4, 177–286.
- SEGEMAR (Servicio Geológico Minero Argentino) (2000) *Mapa geológico de la República Argentina, escala 1:2.500.000*. Buenos Aires: Instituto de Geología y Recursos Naturales.
- Silliman B.R., Bertness M.D., Altieri A.H., Griffin J.N., Bazterrica M.C., Hidalgo F.J., Crain C.M. and Reyna M.V. (2011) Whole-community facilitation regulates biodiversity on Patagonian rocky shores. *PLoS ONE* 6, e24502. doi:24510.21371/journal.pone.0024502
- Skibinski D.O.F., Gallagher C. and Beynon C.M. (1994) Mitochondrial DNA inheritance. *Nature* 368, 817–818.
- Swofford D.L. (1998) *PAUP\*. Phylogenetic analysis using Parsimony (\*and other methods)*. Sunderland, MA: Sinauer Associates.
- Tamura K., Dudley J., Nei M. and Kumar S. (2007) MEGA 4: Molecular Evolutionary Genetics Analysis. *Molecular Biology and Evolution* 24, 1596–1599.
- Tanaka M.O. and Magalhães C.A. (2002) Edge effects and succession dynamics in *Brachidontes* mussel beds. *Marine Ecology Progress Series* 237, 151–158.
- Terranova M.S., Lo Brutto S., Arculeo M. and Mitton J.B. (2007) A mitochondrial phylogeography of *Brachidontes variabilis* (Bivalvia: Mytilidae) reveals three cryptic species. *Journal of Zoological Systematics and Evolutionary Research* 45, 289–298.
- Thiel M. and Ullrich N. (2002) Hard rock versus soft bottom: the fauna associated with intertidal mussel beds on hard bottoms along the coast of Chile, and considerations on the functional role of mussel beds. *Helgolander Marine Research* 56, 21–30.
- Thompson J.D., Higgins D.G. and Gibson T.J. (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research* 22, 4673–4680.
- van der Molen S., Márquez F., Idaszkin Y.L. and Adami M. (2012) Use of shell-shape to discriminate between *Brachidontes rodriguezii* and *Brachidontes purpuratus* species (Mytilidae) in the transition zone of their distributions (south-western Atlantic). *Journal of the Marine Biological Association of the United Kingdom*, doi: <http://dx.doi.org/10.1017/S0025315412001221>
- Waters J.M., Fraser C.I. and Hewitt G.M. (2012) Founder takes all: density-dependent processes structure biodiversity. *Trends in Ecology and Evolution* 28, 78–85.
- Webb C.O., Ackerly D.D., McPeck M.A. and Donoghue M.J. (2002) Phylogenies and community ecology. *Annual Review of Ecology, Evolution, and Systematics* 33, 475–505.

**White L.R., McPherson B.A. and Stauffer J.R.** (1996) Molecular genetic identification tools for the unionids of French Creek, Pennsylvania. *Malacologia* 38, 181–202.

**WoRMS (World Registry of Marine Species)** (2013) Available at: <http://www.marinespecies.org> (accessed 25 March 2013).

**Zelaya D.G.** (2009) Bivalvia. In Häussermann V. and Försterra G. (eds) *Marine benthic fauna of Chilean Patagonia*. Puerto Montt: Nature in Focus, pp. 426–454.

and

**Zouros E., Freeman K.R., Ball A.O. and Pogson G.H.** (1992) Direct evidence for extensive paternal mitochondrial DNA inheritance in the marine mussel *Mytilus*. *Nature* 359, 412–414.

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