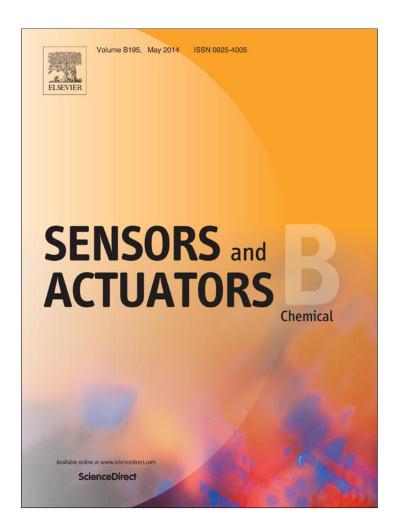
Provided for non-commercial research and education use. Not for reproduction, distribution or commercial use.



This article appeared in a journal published by Elsevier. The attached copy is furnished to the author for internal non-commercial research and education use, including for instruction at the authors institution and sharing with colleagues.

Other uses, including reproduction and distribution, or selling or licensing copies, or posting to personal, institutional or third party websites are prohibited.

In most cases authors are permitted to post their version of the article (e.g. in Word or Tex form) to their personal website or institutional repository. Authors requiring further information regarding Elsevier's archiving and manuscript policies are encouraged to visit:

http://www.elsevier.com/authorsrights

Author's personal copy

Sensors and Actuators B 195 (2014) 294-302



Contents lists available at ScienceDirect

Sensors and Actuators B: Chemical

journal homepage: www.elsevier.com/locate/snb



Screen-printed electrodes for electroanalytical sensing, of chromium VI in strong acid media



S.A. Miscoria a,b, C. Jacq c, T. Maeder c,*, R. Martín Negria,*

- a Instituto de Química Física de los Materiales, Medio Ambiente y Energía (INQUIMAE), Departamento de Química Inorgánica, Analítica y Química Física (DQIAyQF), Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Buenos Aires, Argentina
- ^b Departamento de Química, Facultad de Ciencias Naturales, Universidad Nacional de la Patagonia San Juan Bosco, Comodoro Rivadavia, Chubut, Argentina
- ^c Laboratoire de Production Microtechnique (LPM), École Polytechnique Fédérale de Lausanne (EPFL), CH-1015 Lausanne, Switzerland

ARTICLE INFO

Article history: Received 17 September 2013 Received in revised form 16 December 2013 Accepted 2 January 2014 Available online 11 January 2014

Keywords: Electrochemical sensors Hexavalent chromium Screen-printed electrodes Thick-film technology.

ABSTRACT

Simple, low-cost and acid-resistant carbon-based screen-printed electrodes (SPEs) addressed to detection of hexavalent chromium species, Cr(VI), in sulfuric acid at pH about 1, were prepared and characterized. Working and counter electrodes were prepared jointly on the same substrate in a single strip (workingcounter electrodes pair). The batch printing process allowed obtaining many working-counter electrode pairs in a unique step. The developed working electrodes are comprised of several layers deposited on an alumina substrate: (1) bottom silver conductor, (2) dense organic-graphite composite conductor, (3) active layer consisting of a porous organic-graphite composite which contains a Cr(III) ionophore for testing Cr(III) obtained after reducing Cr(VI), and (4) an insulating and protective dielectric. All materials except the bottom Ag conductor were made on an organic matrix based on a thermoplastic polymer, polyvinylbutyral (PVB). The amperometric determination of Cr(VI) species at pH 1 was performed over a wide concentration range (dynamic range 3 μ M-40 mM). The range for linear amperometric response is $3 \mu M-10 \text{ mM}$, with sensitivity about 0.08 mA mM^{-1} . The sensitivity is improved in comparison with previously developed sensors, while keeping a low limit of detection (LOD about 1 μ M). The response of the sensors are not interfered by the presence of Cr(III) in the solution, although the sensor can detect local formation of Cr(III) at the electrode surface after the reduction of Cr(VI) when diethylenetriaminepentaacetic acid (DTPA), a complexing agent for Cr(III) is incorporated into the pastes.

© 2014 Elsevier B.V. All rights reserved.

1. Introduction

Detection of chromium (VI) species in natural environments (referred generically as Cr(VI)) represents an issue of high relevance in environmental control because of the high toxicity and solubility of Cr(VI) compounds in comparison with chromium species in other oxidation states (Cr(III), Cr(II)) [1–9]. Cr(VI) species have mutagenic, genotoxic and carcinogenic effects on humans and animals, and its main sources are plating industries, leather tanning, wood preservation and electricity generation, which use large amounts of $K_2Cr_2O_7$ that are eliminated in acid effluents, including extreme pH values (≤ 1) [2,10–13]. Remediation

Abbreviations: C18E2, PEG-2 stearyl ether/steareth-2/Brij S2; DTPA, diethylenetriamine-pentaacetic acid; PVB, polyvinylbutyral; SPE, screen-printed electrodes: DMSO. dimethylsulfoxide.

E-mail addresses: thomas.maeder@epfl.ch (T. Maeder), rmn@qi.fcen.uba.ar (R. Martín Negri).

includes chemical treatments and bioremediation processes; e.g. the group at INQUIMAE have recently monitored the growth of the acidophilic bacteria *Acidithiobacillus thioxidans* (AT), used in Cr(VI) bioremediation by means of electronic noses coupled to bioreactors in acid media (pH \leq 1) [14]. AT is a chemolithotrophic bacteria used for biotechnological leaching and heavy metal bioremediation, grown in air-lift bioreactors using tiny bits of elemental sulfur as energy source, where the biofilms grow by oxidation of elemental sulfur to sulfate in a process that increases the acidity of the medium [15,16]. For instance, in the bioreactors implemented at INQUIMAE, pH decreases from 3 to 0 in few days during the phase of biofilm growth, depending of the conditions [14].

Hence, there is large interest for monitoring Cr(VI) at $pH \cong 0-1$ in sulfuric acid, for instance when using chemolithotrophic bacteria in bioremediation process used in the described bioreactors or when monitoring very acid effluents, such as those used in leather tanning and plating industries (which commonly use sulphuric acid media at $pH \cong 0-1$ also).

In all the mentioned cases, after dissolution of $K_2Cr_2O_7$ at pH <2, chromium remains as Cr(VI), the main species at those pH

^{*} Corresponding authors.

values being HCrO₄⁻ and H₂CrO₄ [13]. In the industrial effluents, before remediation, the concentrations of these species are between 0.1 and 10 mM. These concentrations are higher than the limit recommended by the World Health Organization (WHO) for drinking water, 1 µM [8], thus effluents require pretreatments and exhaustive monitoring (see also reference [2]). Since the pioneer works of Urone [17,18] the more common technique for quantifying Cr(VI) in solution is spectrophotometric based on the formation of the colored complex between chromium and 1,5diphenylcarbazide (DPC), performed at pH 1. Atomic absorption spectrometry and inductively coupled plasma mass spectrometry are the latest designs [19–26]. In general, previous treatment of samples involving preconcentration and separation schemes is required, such as chromatography, precipitation, extraction, etc. Additionally, turbidity of the samples, difficulties of handling, the unfeasibility to measure on time and the cost of operation and spectroscopic devices for in situ determinations, represent significant drawbacks.

Alternatively, electrochemical techniques are always interesting alternatives as they offer a rapid detection of the analyte in different matrices because they do not require prior separation and can be used in the presence of turbidity. Adsorptive stripping voltammetric methods have been reported. Jorge et al. [27] presented an adsorptive stripping voltammetric protocol coupled with a rotating disc bismuth film electrode obtaining very low limit of quantification (LOQ), in the range on nM, but at pH 4.6, acetate buffer. Lin et al. [28] described an adsorptive stripping voltammetric protocol at a bismuth-coated glassy-carbon electrode for trace measurements of chromium (VI) for pH between 4.5 and 7.5. Grabarczyk [29] developed a procedure based on a quantitative extraction method to extract total Cr(VI) (soluble and insoluble forms) from a solid sample, detecting Cr(VI) in the extract by a catalytic adsorptive stripping voltammetric technique for pH between 5 and 6

On the other hand, concerning solid-state sensors coupled to amperometric or voltammetric detection, Bergamini et al. [30] and Sanchez-Moreno et al. [31] reported detection of Cr(VI) at pH 2 and 3. Fiol et al. [32] prepared vegetable waste-based sensors with selectivity for Cr(VI) and Hg(II) for pH 2-3 with LOD about 2 mM and sensitivity of -61 mV/decade. In these works the experiments were performed at $pH \ge 2$. Other authors developed relatively more sophisticated modified electrodes. For instance, Svancara et al. [33] reported Cr(VI) sensor working at pH between 1 and 3 based on synergistic pre-concentration of the chromate anion at a carbon paste electrode modified in situ with quarternary ammonium salts. Hallam et al. [34] described screen printed electrodes (SPE) with platforms based on graphite for Cr(VI) sensors, working at pH 1 obtaining a limit of detection (LOD) about 0.4 µM, a limit of quantification (LOQ) of 2 μM, but in short range for linear response (up to $20 \,\mu\text{M}$). Welch et al. [13] found that Cr(VI) in pH 1 solutions can undergo chemically irreversible reduction on the surface of diamond electrodes, doped with gold, glassy carbon and boron with an LOD of $4 \mu M$.

Thus, there is continuous interest to develop low-cost, serial fabricated and robust sensors, for detection and quantification of Cr(VI) species in pH < 1

Screen-printed electrodes (SPE) represent a highly interesting option when considering technological aspects related to mass production, including improvements in reproducibility and replicability, low costs and size reduction. Coupling these aspects with the general advantages of electroanalytical detection appears attractive for environmental monitoring of acid effluents containing Cr(VI). High sensitivity, excellent limits of detection, no requirements concerning large sample volume and possibilities for analyzing turbid media are relevant characteristics of SPE sensors.

However development and application of fully-integrated screen-printed electrodes (for instance integrating working, counter and reference electrodes) for Cr(VI) detection in acid media (pH \approx 1) had not been achieved yet. Actually, application of screen printed working electrodes for Cr(VI) detection and quantification has been reported in very few cases and at higher pH values [29–31,35]. The challenges lie in the formulation of screen-printable inks compatible with processing at moderate temperatures, for obtaining acid-resistant electrodes that allow working at pH \leq 1, with an active surface tailored to achieve selective response to Cr(VI) under these conditions. Ideally, the three electrodes (working, counter and reference) should be integrated into a single device.

Screen-printed thick-film sensors, as previously developed [36–38], provide an ideal platform for realizing such an SPE. Hence, the aims of the present work, originated in a preliminary conference paper [39], are to present the steps of the fabrication process and the quality parameters of the obtained sensors. The SPE graphite electrodes for Cr(VI) quantification were fabricated by integrating working and counter electrode into a unique strip. These systems proved very highly robust in acid media, with the possibility of recording amperometric signals at two different potentials.

2. Material and methods

2.1. Reagents

 $K_2Cr_2O_7$ was from Anedra and H_2SO_4 from Cicarelli (Argentina, analytical grade). Millipore-MilliQ water (ρ = 18 $M\Omega$ cm) was used for preparing the solutions. All chemicals were reagent grade and used without further purification. The silver ink (Ag, type 9912K) was from ESL, ElectroScience Laboratories, USA. Polyvinylbutyral (PVB), acetyltributylcitrate (ATBC), cetanol, steareth-2 (C18E2) and diethylenetriamine-pentaacetic acid (DTPA) were from Sigma–Aldrich. Synthetic graphite powder was KS4 from TIMCAL (ovaloid shape, \leq 4 μ m). Detailed information on the materials used for the SPEs is given in Supplement S1.

2.2. Apparatus

Amperometric measurements and cyclic voltammetry experiments were performed with a Potentiostat/Galvanostat Teq3 Analyzer (Argentina). The electrodes were inserted into an electrochemical cell (BAS, Model MF 1084) through its Teflon cover. The reference was a Ag/AgCl, 3 M NaCl (BAS, Model RE-5B) electrode and all the potentials are referred to it. The set up is similar to those described in previous work [40,41]. Scanning electron microscope (SEM) images were obtained with a field Emission Scanning Electron Microscope, Zeiss Supra 40 Genimi.

2.3. Electrode preparation

2.3.1. Fabrication of the electrodes

Fig. 1A shows a layout of the screen-printed electrodes (Fig. 1A) and the ink depositions steps (Fig. 1B) (more details are given in Supplements S2 (cross section) and S3 (substrate)). The electrodes, which have a convenient elongated shape, were batch fabricated on 96% alumina substrates (Al₂O₃, standard thick-film grade) by screen printing and thermal processing of thick-film inks [42]. The complete materials stack is given in Table 1. Dense (to completely cover the Ag layer) and porous (to allow intimate contact with the solution) carbon pastes were subsequently deposited on the working electrode, Except the first layer (thick-film silver, Ag), which was fired at 10 min peak dwell at 850 °C in a belt furnace, thermal processing of the other layers simply involved evaporating the solvent, with a typical cycle of 10 minutes at 120 °C in a ventilated

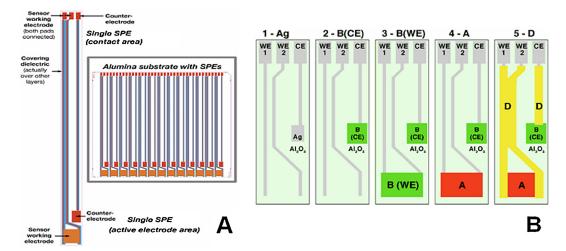


Fig. 1. (A) Layout of SPEs, single bar and as complete substrate for batch printing on alumina (inset). Layers according to Table 1; conductors in contact with solution: dense polymer–KS4 composite (both working electrode and counter electrode), and porous polymer–KS4–DTPA (working only). (B) Ink deposition steps: (1) Ag conductor; (2, 3) dense polymer–graphite (B) for counter (CE) and working (WE) electrode; (4) porous, polymer–graphite–ionophore active layer; (5) protective dielectric.

Table 1Thick-film compositions used in the fabrication of the SPEs based on PVB binder (Fig. 1).

Layer	Composition (see Supplement S2)
Ag conductor tracks and contact pads Dense carbon layer (KS4≈33 vol.%)	ESL 9912K PVB_ATBC_KS4
Porous active carbon layer (KS4 \approx 70 vol.%)	PVB-ATBC-KS4-DTPA
Protective dielectric	PVB-cetanol-C18E2/ATBC

oven. The nominal (geometric) area of the working electrode was $4.0\,\text{mm}\times6.0\,\text{mm}.$

The working electrode was fabricated with two pads in order to measure the conductivity of the graphite paste layer after the whole process when testing different pastes based on different polymers (later, those pads were short circuited). Because of the aim of this first work of the joint groups was to obtain the appropriated paste and to define the whole fabrication protocol in order to produce series of sensors with good quality parameters, the system presented here integrates only the working and counter electrode, but not the reference electrode, since to fabricate series of fully integrated systems (with the three electrodes) requires to set additional conditions, which are being adjusted at present.

2.3.2. Formulation of the screen-printing inks

Except the first silver (Ag) conductor layer, all inks were formulated and prepared in-house (Table 1) as screen-printing pastes, based on polyvinylbutyral (PVB) plasticized by acetyltributylcitrate

(ATBC), dissolved in appropriate high-boiling solvents (see Supplement S1). These starting mixtures serve both as screen-printing vehicles and, after evaporation of the solvent, as organic matrices. For electrode materials, synthetic graphite powder KS4 from TIM-CAL was used as the conductive phase, with the degree of porosity being set by changing the ratio between graphite and matrix (PVB+ATBC). PVB was chosen over more standard ethylcellulose (EC) because its carbon backbone is not sensitive to acid hydrolysis (as it is for EC[43]; see also Supplement S4). Additionally, diethylenetriamine-pentaacetic acid (DTPA), a Cr(III) ionophore [42,45] was incorporated into the vehicle of the active sensing electrode ink by first dissolving it (15% mass) into dimethylsulfoxide (DMSO) and subsequently adding this resulting solution. The presented results correspond to samples with 4.7% (w/w) DTPA in the ink, although samples with DTPA varying from 2.0% (w/w) to 8.5% (w/w) were tested. Increased DTPA concentration corresponded to decreased binder and plasticizer content, with KS4 (conductive filler) content somewhat increased to maintain printable rheology (see Table S1 in supplement). These changes led to small variations in the sensor response which do not change the main conclusion and results presented here. The protective dielectric layer was prepared by incorporating cetanol and C18E2 (steareth-2) into the base formulation. Upon drying and cooling, these waxes crystallize out of the PVB + ATBC matrix to yield a tough, hot-melt chemically resistant layer. For comparison purposes, samples with two layers (dense carbon and dielectric) based on an epoxy matrix instead of PVB were also fabricated (Supplement S5).

Fig. 2 shows a photograph of two sensors (101.6 mm \times 7 mm) along a cm ruler.

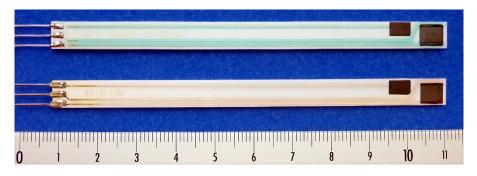


Fig. 2. Photograph of two sensors ($101.6 \text{ mm} \times 7.0 \text{ mm}$, ruler in cm).

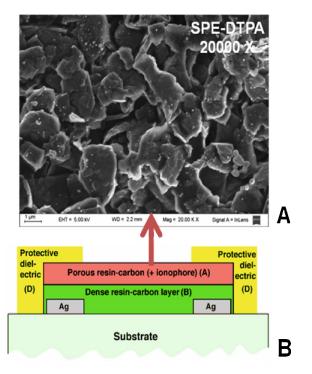


Fig. 3. (A) Scanning electronic microscopy image of PVB-DTPA-based system by magnification $20,000\times$. (B) For WE; (5) insulating dielectric (D). (B) Cross section of the active electrode (expanded vertical scale). The counter electrode is similar, but without the porous active layer.

Fig. 3A shows a SEM image of the graphite screen printed electrochemical sensor where a well-defined electrode surface is observed from randomly orientated graphite particles bound together with the inert polymeric binder. This layer sits on the dense carbon conductor (Fig. 3).

2.4. Electroanalytical measurements

Cyclic voltammetries and amperometric experiments were performed in a three electrodes electrochemical cell at $(25\pm1)^{\circ}$ C. Solutions were under permanent agitation provided by a magnetic stirrer during the amperometric determinations. The support electrolyte was sulfuric acid (H_2SO_4). The amperometric measurements

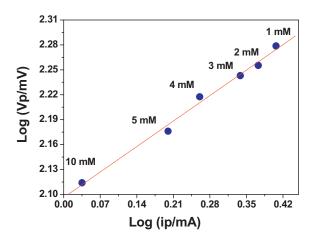


Fig. 5. $\log(V_p/mA)$ vs $\log(I_p/mV)$ (taken from Fig. 4 for the peak at positive voltages (vs. Ag/AgCl) for different concentration of $K_2Cr_2O_7$ in H_2SO_4 , pH 1.

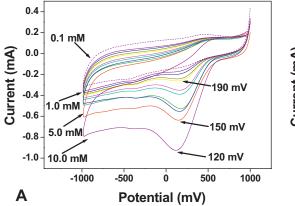
were carried out by applying the desired potential and allowing the transient current to decay to a plateau. Afterwards Cr(VI) was added and the generated current monitored after stabilization.

3. Results and discussion

Two types of measurements were performed: (1) cyclic voltammetries varying scan rates, (2) amperometries at fixed voltages by successively adding concentrated aliquots of $K_2Cr_2O_7$ in H_2SO_4 , adjusted to obtain the desired concentration.

3.1. Voltammograms

Fig. 4 shows cyclic voltammograms (scan rate of $50\,\text{mV}\,\text{s}^{-1}$) for a PVB-SPE containing 4.7% (w/w) of DTPA in oxygenated solutions of H_2SO_4 pH: 1.0. The scans were performed from +1000 mV to $-1000\,\text{mV}$ and returning finally to the initial potential. Fig. 4A illustrates the irreversibility of the electrode process and the presence of shoulders with maxima at about +150 mV and $-370\,\text{mV}$ (although the exact position of both peaks has a slight dependence on $\text{K}_2\text{Cr}_2\text{O}_7$ concentration). Fig. 4B shows a zoom for potentials between $\pm 600\,\text{mV}$ and concentrations lower than 1.0 mM in steps of 0.1 mM (only the reduction window is shown for a better visualization).



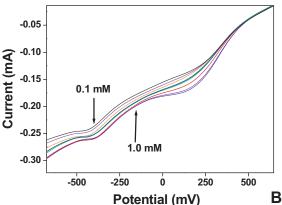


Fig. 4. Cyclic voltammograms for Cr(VI) (A): at different concentrations, from 1.0 mM to 5.0 mM with 0.5 mM breaks (the curves for 0.1 mM and 10 mM are also included). The scans were performed from 1000 mV to -1000 mV to return finally to the initial potential. (B): zoom for potentials between (-600,600) mV and concentrations lower than 1.0 mM in steps of 0.1 mM. Peaks at -370 and +175 mV are observed. Scan rate: 50 mV/s. Supporting electrolyte: H₂SO₄ pH: 1.0. Reference electrode: Ag/AgCl. Counter electrode: graphite.

Fig. 5 shows a logarithmic relationship between the peak voltage, $V_{\rm p}$, and peak current, $I_{\rm p}$, for different concentrations of $\rm K_2Cr_2O_7$ (and for the positive voltages). The value of $V_{\rm p}$ remains almost constant (+150 mV) for concentrations lower than 1 mM. Nevertheless, $V_{\rm p}$ deceases for higher concentrations, observing a linear relation relationship between $V_{\rm p}$ and $I_{\rm p}$ for concentrations between 1 and 10 mM. In the case of a reversible process, commonly used electrochemical models predict that $V_{\rm p}$ must be independent of the analyte concentration, but those models do not apply for irreversible systems. Thus, the decrease of $V_{\rm p}$ when increasing Cr(VI) concentration (at a fixed scan rate) seems to be related to the irreversible behavior of the electrodes (a complete understanding of the $V_{\rm p}$ vs. $I_{\rm p}$ relationship is beyond the scope of the present work).

3.2. Scan rate experiments

Scan rate studies were performed in the absence of stirring. It was observed that the position of the reduction peak shifts towards more negative values with increasing the scan rate, in agreement with the irreversible behavior shown in Fig. 4A. The dependence of the current peak with the scan rate is given in Fig. 6 for +150 mV (Fig. 6A) and $-370 \, \text{mV}$ (Fig. 6B). The scans were performed from +1000 mV to $-1000 \, \text{mV}$ and returning finally to the initial potential. The linear relationship between current peak (at +150 and $-370 \, \text{mV}$) and the square root of the scan rate indicates that signals are the resultant of a diffusion-controlled process.

Summarizing the scan-rate experiments, it is concluded that the response of the sensor is associated to irreversible processes that are kinetically controlled by diffusion of the analyte towards the surface and not by its adsorption or the consequent reduction.

3.3. Origin of the analyzed peaks

The origin of the peaks at +150 mV and -370 mV is now discussed. The first peak correspond to a broad band that begins at about + 450 mV (depending on the Cr(VI) concentration) when decreasing the voltage from +1000 mV. This band is reasonable assigned to reduction of Cr(VI) to lower oxidation states, since current decreases and there are not higher oxidation states available. The presence of DTPA is irrelevant in the observation of the band centered at +150 mV since the apparition of that band was verified (and peak) when using working electrodes prepared without DTPA (the experimental conditions remaining the same). Thus, as already reported in other cases [13,34], no need for Cr(VI) ionophore is required: the surface provides the conditions for the reduction. In agreement, the band centered at +150 mV do not appears when only Cr(III), but not Cr(VI), is present $(CrCl_3 \cdot nH_2O)$ was used). Therefore, we conclude that the surface donates the electrons for Cr(VI) reduction for voltages lower than approximately +475 mV (the voltages are always referred to Ag/AgCl reference electrode)

On the other hand, the band centered at $-370\,\text{mV}$ is assigned to the subsequent reduction of Cr(III) formed at the surface after reducing Cr(VI). This is strongly suggested by observing that the peak at -370 mV only appears when using pastes containing DTPA, a well-known ionophore for selective determination of Cr(III) [44,45]. The formation of a Cr(III)–DTPA complex has been proposed as participating in the reduction of Cr(III) where DTPA acts as chelating agent. After forming the complex on the surface, its reduction on the electrode surface causes the observed reduction peak. It is worth to remark that this peak has much lower intensity than the peak at +150 mV, in agreement with the hypothesis that the signal at -370 mV is originated in a secondary process. The relative low intensity at -370 mV is related also to the fact that the fraction of DTPA available for chelating is restricted by two factors: a) at pH<1 DTPA appears mainly protonated (pKa 1.8) thus deactivating its chelating ability; b) most of DTPA molecules are inside the

paste but not exposed to the electrode surface. As a matter of fact, no clear peaks were observed in the voltamogramms of $CrCl_3 \cdot nH_2O$ solutions.

Therefore, the hypothesis for describing the behavior of the electrodes presented in this work when introducing $K_2Cr_2O_7$ is summarized as follows: the primary process is reduction of Cr(VI) to Cr(III) (and probably to lower states also). The reduction of Cr(VI) is provided by the surface (except the voltage is higher than +450 mV) without need of any ionophore. Then a surface Cr(III)–DTPA complex is formed whose reduction is observed at lower voltages, i.e. $-370\,\text{mV}$. The amperometric analysis fixing the voltage at $-380\,\text{mV}$ can be also used for monitoring Cr(VI) since the primary process that allows the peak at $-380\,\text{mV}$ to be observed is Cr(VI) reduction by the surface.

3.4. Quality parameters of the amperometric signal at ± 140 mV

The sensitivity, limit of detection, robustness, response times, etc., of the sensors for the detection and quantification of Cr(VI) was determined by amperometric experiments instead of voltammograms, observing the effect of consecutive addition of analyte increments on the detected current at fixed voltage. This is illustrated in the following paragraphs (calibration plots obtained from voltammograms are much noiser and presents excellent linearity only below 1 mM).

Fig. 7A and C shows amperometric recordings obtained from an experiment performed by successive additions of $K_2Cr_2O_7$ in H_2SO_4 pH: 1.0, fixing the voltage of the working electrode at +140 mV (10 mV lower than the value of V_p , +150 mV, in the considered concentration range).

The rise time of the signal after injection is 2-3 s for all concentrations. The time required for amperometric signal stabilization is dependent on the $K_2Cr_2O_7$ concentration: it takes 5-10 s for concentrations below 1 mM, but can take 3-5 min for concentrations between 6 and 40 mM.

The momentary dips in current observed for some additions in Fig. 7A and C are sometimes referred as "injection noise". The origin of that noise is the difficulties for injecting the analyte solution in the electrochemical cell, particularly when its dimensions are relatively small (volume of the solution, 30–50 mL, approximately). It has a purely mechanical origin and does not disturb the amperometric response, which is stabilized after few seconds (injection is always performed under stirring).

Reproducibility test on a given electrode, different freshly made electrodes in the same batch (12 electrodes) and inter-batches were performed. The typical relative standard deviation (RSD) value in all cases was about 3–5% (depending of the measurement conditions, indicated in text). As an example, we made 10 calibration plots in the range 0.010–10 mM using the same electrode, obtaining a RSD value of 5.2% (the same electrode was used 10 times). In other series we made calibration plots using 10 different electrodes from the same batch (each electrode was used only one time), obtaining RSD 2.7%. When using electrodes from different batches RSD about 3% were obtained.

Calibration plots are shown in Fig. 7B and D. The figures are shown only for a narrow concentration window. A sensitive cathodic signal with fast response time is observed. The sensitivity of the sensor for the amperometric determination at +140 mV, S_{140} , is around 0.08 mA mM^{-1} .

The presented calibration plots begin at $5 \,\mu M$ (0.005 mM in Fig. 7B). However, we verified that can be extended up to $3 \,\mu M$ of $K_2Cr_2O_7$. The difference between the amperometric signal recorded at $3 \,\mu M$ and the signal presented in H_2SO_2 at pH 1 in the absence of $K_2Cr_2O_7$, is three times higher than the electronic noise observed in the absence of $K_2Cr_2O_7$. Thus, the estimated limit of quantification, LOQ, is $3 \,\mu M$. The observed limit of detection, LOD, is

S.A. Miscoria et al. / Sensors and Actuators B 195 (2014) 294–302

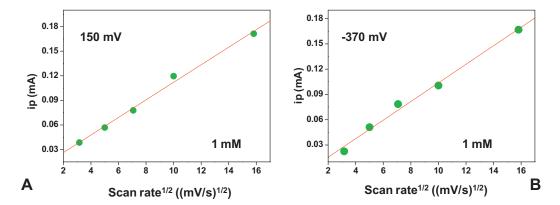


Fig. 6. Reduction peak currents (A): at 150 mV, (B): at -370 mV versus its scan rate square root, obtained from the cyclic voltammograms for 1 mM of Cr(VI). The scans were carried out from 1000 mV to -1000 mV to return finally to the initial potential. Supporting electrolyte: H_2SO_4 pH: 1.0. Reference electrode: Ag/AgCl. Counter electrode: graphite.

about 1.0–1.5 μ M, where the difference of time-averaged signals is slightly higher than the noise. These values of LOQ and LOD may have a 3–5% of variation with the particular considered sensor and the considered batch.

3.5. Quality parameters of the amperometric signal at -380 mV

Fig. 8A and C also shows amperometric recordings and (B and D) calibration plots obtained from the same experiment exposed in Fig. 7 but now at $-380\,\mathrm{mV}$ working potential. Similarly, a fast cathodic signal is obtained at the electrode. Fig. 8C shows a broader concentration range. The sensitivity of the electrode is around $0.1\,\mathrm{mA\,mM^{-1}}$.

Fig. 9 illustrates the behavior of the sensor showing an amperometric recording obtained from experiments performed by

increasing concentration of $K_2Cr_2O_7$ in a extended concentration range (10 μ M–40 mM).

The response is quite linear, although sensitivity slightly decreases at concentrations above 20 mM. The rise time of the signal is about 2–3 s in the whole concentration range. Stabilization of the signal may take 3–5 min after injection, depending of the analyte concentration, as discussed in the previous section. The sensitivity of the electrode is clearly enough to unambiguously detect concentration steps of 1–2 μM of Cr(VI), with a limit of detection (LOD) of 1 μM .

3.6. Stability, reproducibility and potentiality for environmental applications

The short-term stability of the electrodes was evaluated by analyzing the sensitivity of ten calibrations performed using the same

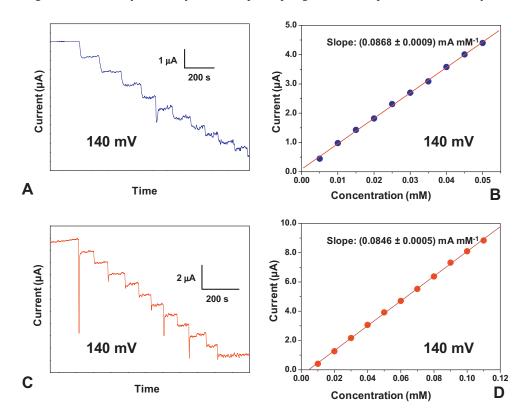


Fig. 7. Amperometric recordings (A and C) and calibration plots (B and D) obtained from experiments performed by successive additions of 0.005 mM (A and B) and 0.1 mM (C and D) Cr(VI), at +140 mV. Supporting electrolyte: H₂SO₄ pH: 1.0. Reference electrode: Ag/AgCl. Counter electrode: graphite.

S.A. Miscoria et al. / Sensors and Actuators B 195 (2014) 294-302

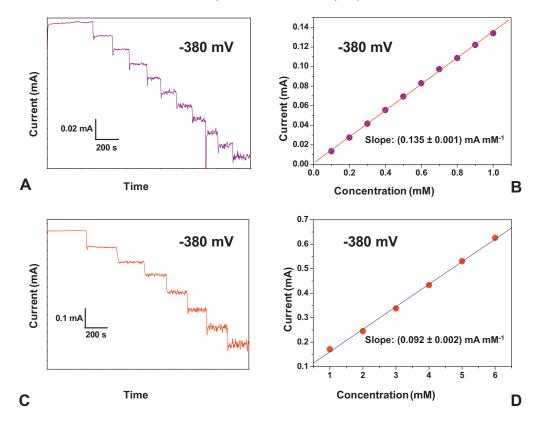


Fig. 8. Amperometric recordings (A and C) and calibration plot (B and D) obtained from experiments performed by successive additions of Cr(VI), at -380 mV. Supporting electrolyte: H₂SO₄ pH: 1.0. Reference electrode: Ag/AgCl. Counter electrode: graphite.

sensor. The resulting RSD of 5.2% demonstrates the robustness of the electrode even though they are disposables. The effect of using different surfaces was also checked by evaluating the sensitivity of ten corresponding calibration plots, resulting in an RSD of 2.7%. Concerning long-term stability, after 6 months of storage in air, at room temperature and in darkness, the response of the electrode remained at around 100% of the original one.

Concerning the effect of oxygen in the media, it has been reported [46] that the oxygen reduction in H_2SO_4 starts at potentials more negative than $-500\,\mathrm{mV}$ for carbon platforms. Therefore, in the case of the present device, the oxidation of the solvent starts at more negative potentials and thus does not interfere with the studied signals (in fact oxygenated and deoxygenated solutions present the same profile). Therefore, nitrogen purge is not required for the developed sensors, which is a very important result concerning to possible *in situ* determinations for environmental monitoring, where nitrogen sparging is not possible or impractical.

The results with epoxy-based dense conductor show similar, but lower and noisier response (see Supplement S5). As the active electrode material was the same, this poorer performance was attributed to the degradation of the electrode by excessive high-temperature exposure during hardening of the dielectric (150 °C 2 h), with loss of plasticizer or DTPA being the most likely mechanisms. This also illustrates the need for further study of the impact of the electrode formulation (resin, plasticizer, type of carbon, ionophore) on the electrochemical behavior.

3.7. Comparison with other sensors and methods for Cr(VI) detection

A table for comparison of the performance between sensors developed by different authors is presented (Table 2), indicating range of pH, LOD, LOQ, concentration range for linear response and sensitivity. When comparing with other sensors reported by

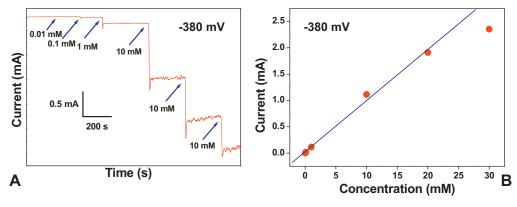


Fig. 9. Amperometric recordings (A) and calibration plot (B) obtained from experiments performed by successive additions of different Cr(VI) concentration, at -380 mV. Supporting electrolyte: H₂SO₄ pH: 1.0. Reference electrode: Ag/AgCl. Counter electrode: graphite.

Table 2Comparison of quality parameters between Cr(VI) sensors developed by different authors.

Authors/reference	Electrode/surface	pH of work or range	LOD	LOQ	Concentration range for linear response	Sensitivity
Present work	Graphite-SPE	pH 1	1 μΜ	3 μΜ	3 μM-10 mM	0.08 mA/mM
Bergamini et al. [30]	Modified-SPE	pH 2	0.05 μΜ	$0.1 \mu\text{M}$	0.1 μM-0.15 mM	1.1 mA/mM
Sanchez-Moreno et al. [31]	Graphite-epoxy	pH 3	0.6 μΜ	1 μΜ	1 μM–10 mM	60 mV/dec (potentiometry)
Fiol et al. [32]	Graphite powder, epoxy resin	pH 2-3	$2\mu M$	$20\mu M$	20 μM-3 mM	61 mV/dec (potentiometry)
Svancara et al. [33]	Modified-CPE	pH 1	0.05 μΜ	0.5 μΜ	$0.550\mu\text{M}$	(stripping preconcentration)
Hallam et al. [34]	SPE-graphite	pH 1	$0.4 \mu M$	$2 \mu M$	2-20 μΜ	0.03 mA/mM
Domímguez-Renedo et al. [35]	Modified-SPE	pH 3-6	0.4 μΜ	0. 4 μΜ	0.4-30 μM	0.02 mA/mM (differential pulsed voltammetry)
Welch et al. [13]	Gold electrode	pH 1-3	$4 \mu M$	0.1 mM	0.1- 1.5 mM	0.05 mA/mM
Neto et al. [27]	Bismuth film electrode	pH 4.6	0.3 nM	1nM	-	9.53 μAnM ⁻¹ (stripping preconcentration)
Lin et al. [28]	Bismuth film electrode	рН 6	0.3 nM	1 nM	5–50nM	0.07 μAnM ⁻¹ (stripping preconcentration)

authors which use amperometric or potentiometric detection, it can be observed the very good limit of detection, extended range for linear response, and high sensitivity for working at pH 1 of the presented sensors. For instance, in comparison with the Cr(VI) sensor developed by Hallam et al. [34], the sensors presented here have twice the sensitivity, a much larger dynamic range (two orders of magnitude) with a similar limit of detection. On the other hand, when using selective adsorbents, masking agents or formation of complexes, the LOD can be in the range of nM in the case of electrochemical methods also (e.g. for stripping voltammetry, see Table 2). In the case of spectroscopic methods, including Atomic Absorption Spectroscopy (AAS) and ICP-tandems, the LODs can range from nM to μM depending on the preconcentration technique used. If no preconcentration technique is used, the LODs are typically about 0.1–1 μM .

4. Conclusions

The present work shows the excellent quality parameters for Cr(VI) detection using screen printed sensors in extremely low pH values. The most relevant characteristics of the sensing platforms proposed here are the remarkable sensitivities, the extended dynamic range, and the low detection limit. Remarkably, the sensor do not display signal when a Cr(III) salt is dissolved at pH 1 (thus, it can discriminate C(VI) from Cr(III)), but is sensitive to local formation of Cr(III) at the surface when Cr(VI) is reduced on the electrode. The developed sensors are very efficient for using in bioremediation processes control since they have an excellent performance at low pH values. Oxygen removal is not required, which is an essential issue when working in the field. The combination of such an excellent analytical performance with the enormous possibilities offered by simple, low-cost disposable screen-printed electrodes are highly promising for addressing the requirements of environmental chemical analysis.

Acknowledgements

Financial support was received from the University of Buenos Aires (UBACyT 2012–2015, 20020110100098), Ministry of Science, Technology and Innovations of Argentina (PICT 2010-2829 and PICT 2011-0377) and from the CODEV Seed Money Programme of the *École Polytechnique Fédérale de Lausanne* (EPFL), Switzerland. The authors thank Mr. Matthias Garcin, EPFL, for help in fabricating the samples, the Center of Advanced Microscopy (CMA), School

of Sciences, University of Buenos Aires, for obtaining the SEM images, and are also grateful to TIMCAL for kindly donating the KS4 graphite powder.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at http://dx.doi.org/10.1016/j.snb.2014.01.013.

References

- Q. Wu, Y. Qu, X. Li, D. Wang, Chromium exhibits adverse effects at environmental relevant concentrations in chronic toxicity assay system of nematode *Caenorhabditis elegans*, Chemosphere 87 (2012) 1281–1287.
- [2] A.K. Shanker, B. Venkateswarlu, Chromium: environmental pollution, health effects and mode of action, in: Encyclopedia of Environmental Health, 2011, pp. 650–659.
- [3] Z.-H. Li, P. Li, T. Randak, Evaluating the toxicity of environmental concentrations of waterborne chromium (VI) to a model teleost, *Oncorhynchus mykiss*: a comparative study of in vivo and in vitro, Comp. Biochem. Physiol. Part C: Toxicol. Pharmacol. 153 (2011) 402–407.
- [4] E. Kiliç, R. Puig, G. Baquero, J. Font, S. Çolak, D. Gürler, Environmental optimization of chromium recovery from tannery sludge using a life cycle assessment approach, J. Hazard. Mater. 192 (2011) 393–401.
- [5] J.B. Vincent, The bioinorganic chemistry of chromium (III), Polyhedron 20 (2001) 1–26.
- [6] J. Kotás, Z. Stasicka, Chromium occurrence in the environment and methods of its speciation, Environ. Pollut. 107 (2000) 263–283.
- [7] M. Gagneten, J.C. Paggi, Effects of heavy metal contamination (Cr, Cu, Pb, Cd) and eutrophication on zooplankton in the Lower Basin of the Salado River (Argentina). Water Air Soil Pollut. (2009) 317–334.
- [8] Guidance for Drinking Water Quality, Recommendations, vol. 1, second ed., WHO, Geneva, 1993, pp. 45–46.
- [9] G. Hanrahan, D.G. Patil, J. Wang, Electrochemical sensors for environmental monitoring: design, development and applications, J. Environ. Monit. 6 (2004) 657–664
- [10] F.J. Alguacil, M. Alonso, F. Lopez, A. Lopez-Delgado, Uphill permeation of Cr(VI) using Hostarex A327 as ionophore by membrane-solvent extraction processing, Chemosphere 72 (2008) 684–689.
- [11] F.J. Alguacil, A.G. Coedo, M.T. Dorado, A.M. Sastre, Uphill permeation of chromium (VI) using Cyanex 921 as ionophore across an immobilized liquid membrane, Hydrometallurgy 61 (2001) 13–19.
- [12] M. Tuzen, M. Soylak, Multiwalled carbon nanotubes for speciation of chromium in environmental samples, J. Hazard. Mater. 147 (2007) 219–225.
- [13] C.M. Welch, O. Nekrassova, R.G. Compton, Reduction of hexavalent chromium at solid electrodes in acidic media: reaction mechanism and analytical applications, Talanta 65 (2005) 74–80.
- [14] P.E. Rosi, S.A. Miscoria, D.L. Bernik, R.M. Negri, Customized design of electronic noses placed on top of air-lift bioreactors for in situ monitoring the off-gas patterns, Bioprocess Biosyst. Eng. 35 (2012) 835–842.
- [15] K. Bredberg, H.T. Karlsson, O. Holst, Reduction of vanadium(V) with Acidithiobacillus ferrooxidans and Acidithiobacillus thiooxidans, Bioresour. Technol. 92 (2004) 93–96.

- [16] R.M. Gargarello, D. Di Gregorio, H. Huck, J. Fernandez Niello, G. Curutchet, Reduction of uranium(VI) by Acidithiobacillus thiooxidans and Acidithiobacillus ferrooxidans, Hydrometallurgy 104 (2010) 529–532.
- [17] R.T. Pflaum, L.C. Howick, The chromium-diphenylcarbazide reaction, J. Am. Chem. Soc. 78 (19) (1956) 4862–4866.
- [18] P.F. Urone, Stability of colorimetric reagent for chromium, s-diphenylcarbazide, in various solvents, Anal. Chem. 27 (8) (1955) 1354–1355.
- [19] Y. Inoue, T. Sakai, H. Kumaguai, Simultaneous determination of Cr(III) and Cr(VI) by ion chromatography with inductively coupled plasma mass spectrometry, J. Chromatogr. A 706 (1995) 127–136.
- [20] P. Liang, T.Q. Shi, H.B. Lu, Z.C. Jiang, B. Hu, Adsorption of Cr(VI) and speciation of Cr(VI) and Cr(III) in aqueous solutions using chemically modified chitosan, Spectrochim. Acta B 58 (2003) 1709–1714.
- [21] P. Wu, H. Chen, G. Cheng, X. Hou, Exploring surface chemistry of nano-TiO(2) for automated speciation analysis of Cr(III) and Cr(VI) in drinking water using flow injection and ET-AAS detection, J. Anal. At. Spectrom. 24 (2009) 1098–1104.
- [22] M.T. Siles Cordero, E. Vereda Alonso, A. Garcia de Torres, J.M. Cano Pavon, Development of a new system for the speciation of chromium in natural waters and human urine samples by combining ion exchange and ETA-AAS, J. Anal. At. Spectrom. 19 (2004) 398–403.
- [23] Y.L. Chang, S.J. Jiang, Determination of chromium species in water samples by liquid chromatography-inductively coupled plasma-dynamic reaction cellmass spectrometry, J. Anal. Atom. Spectrom. 16 (2001) 858.
- [24] V. Gómez, M.P. Callao, Chromium determination and speciation since 2000, Trends Anal. Chem. 25 (2006) 1006–1015.
- [25] M.S. Tehrani, P.A. Azar, S.W. Husain, F. Shafaei, Dispersive liquid-liquid microextraction of Cr(VI) in water and hair samples by electrothermal atomic absorption spectrometry, Asian J. Chem. 22 (2010) 6302–6310.
- [26] C. Diniz Pereira, J.G. Techy, E. Moreira Ganzarollib, S.P. Quináia, Chromium fractionation and speciation in natural waters, J. Environ. Monit. 14 (2012) 1559–1564.
- [27] E.O. Jorge, M.M. Rocha, I.T.E. Fonseca, M.M.M. Neto, Studies on the stripping voltammetric determination and speciation of chromium at a rotating-disc bismuth film electrode, Talanta 81 (2010) 556–564.
- [28] L. Lin, N.S. Lawrence, S. Thongngamdeea, J. Wang, Y. Lin, Catalytic adsorptive stripping determination of trace chromium (VI) at the bismuth film electrode, Talanta 65 (2005) 144–148.
- [29] M. Grabarczyk, Speciation analysis of chromium by adsorptive stripping voltammetry in tap and river water samples, Electroanalysis 20 (2008) 2217–2222.
- [30] M.F. Bergamini, D.P. dos Santos, M.V.B. Zanoni, Development of a voltammetric sensor for chromium(VI) determination in wastewater sample, Sens. Actuators B 123 (2007) 902–907.
- [31] R.A. Sánchez-Moreno, M.J. Gismera, M.T. Sevilla, J.R. Procopio, Evaluation of solid-state platforms for chromium (VI) potentiometric sensor development, Anal. Bioanal. Chem. 397 (2010) 331–338.
- [32] N. Fiol, F. de la Torre, P. Demeyere, A. Florido, I. Villaescusa, Vegetable waste-based sensors for metal ion determination, Sens. Actuators B 122 (2007) 187–194.
- [33] I. Svancara, P. Foret, K. Vytras, A study on the determination of chromium as chromate at a carbon paste electrode modified with surfactants, Talanta 64 (2004) 844–852.
- [34] P.M. Hallam, D.K. Kampouris, R.O. Kadara, N. Jenkinson, C.E. Banks, Graphite screen printed electrodes for the electrochemical sensing of chromium(VI), Analyst 135 (2010) 1947–1951.
- [35] O. Domínguez-Renedo, L. Ruiz-Espelt, N. García-Astorgano, M.J. Arcos-Martínez, Electrochemical determination of chromium(VI) using metallic nanoparticle-modified carbon screen-printed electrodes, Talanta 76 (2008) 854-858.

- [36] N. Serra, T. Maeder, P. Ryser, Piezoresistive effect in epoxy-graphite composites, Sens. Actuators A 186 (2012) 198–202.
- [37] B. Jiang, P. Muralt, P. Heeb, A.J. Santis-Alvarez, M. Nabavi, D. Poulikakos, P. Niedermann, T. Maeder, A micro heater platform with fluid channels for testing micro-solid oxide fuel cell components, Sens. Actuators B 175 (2012) 218–224.
- [38] F. Gruet, F. Vecchio, C. Affolderbach, Y. Pétremand, N.F. de Rooij, T. Maeder, G. Mileti, A miniature frequency-stabilized VCSEL system emitting at 795 nm based on LTCC modules, Opt. Lasers Eng. 51 (2013) 1023–1027.
- [39] T. Maeder, S. Miscoria, C. Jacq, P. Ryser, R.M. Negri, Screen-printed electrochemical chromium (VI) sensing electrodes for effluent bioremediation monitoring, Proc. Eng. 47 (2012) 1303–1306.
- [40] G.A. Rivas, S.A. Miscoria, J. Desbrières, G.D. Barrera, New biosensing platforms based on the layer-by-layer self-assembling of polyelectrolytes on Nafion/carbon nanotubes-coated glassy carbon electrodes, Talanta 71 (2007) 270-275.
- [41] S.A. Miscoria, J. Desbrieres, G.D. Barrera, P. Labbè, G.A. Rivas, Glucose biosensor based on the layer-by-layer self-assembling of glucose oxidase and chitosan derivatives on a thiolated gold surface, Anal. Chim. Acta 578 (2006) 137-144.
- [42] K. Pitt, Handbook of Thick Film Technology, 2nd ed., Electrochemical Publications, Isle of Man, 2005.
- [43] Y.V. Moissev, N.A. Khalturinskii, G.E. Zaikov, The mechanism of the acid-catalysed hydrolysis of polysaccharides, Carbohydr. Res. 51 (1976) 39–54.
- [44] R. Bucci, A.D. Magrì, A.L. Magrì, A. Napoli, On the reaction of iron(III) with chromium(III)-DTPA chelating agent, Polyhedron 19 (2000) 2421–2425.
- [45] Y. Li, H. Xue, Determination of Cr(III) and Cr(VI) species in natural waters by catalytic cathodic stripping voltammetry, Anal. Chim. Acta 448 (2001) 121–134.
- [46] M.D. Rubianes, G.A. Rivas, Carbon nanotubes paste electrode, Electrochem. Commun. 5 (2003) 689–694.

Biographies

Silvia Alejandra Miscoria graduated at Universidad Nacional de la Patagonia "San Juan Bosco", Argentina, as Biochemist and continued a PhD in Chemistry working in biosensors designs. Since 2010 is carrying out a post-doc in the Instituto de Quimica Fisica de Materiales, Ambiente y Energía (INQUIMAE), Universidad de Buenos Aires in chemistry sensors designs.

Caroline Jacq graduated with a Master thesis in materials science from the Institut Supe'rieur des Mate'riaux du Mans (ISMANS), and is working as a researcher at the Ecole Polytechnique Fe'de'rale de Lausanne (EPFL) since 2001. Her main research interests concern the development and study of novel lead-free thick-film materials systems and their application to medical devices.

Thomas Maeder graduated at the Ecole Polytechnique Fédérale de Lausanne (EPFL) in materials science, and continued with a PhD in piezoelectric thin films, then a post-doc at IBM Rüschlikon in single-crystal conductive oxides. He now heads the thick-film technology group at the EPFL, where current areas of interest are thick-film and LTCC technology for advanced sensor, packaging, biomedical and harsh-environment applications.

R. Martín Negri graduated in physics at the University of Buenos Aires (UBA) where made a PhD in chemistry. He is member of the research career of CONICET (Argentina) and Professor at School of Science-UBA, heading a group in the field of materials science and sensors development.