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Targeting DNA Damage Response Kinases in Cancer Therapy

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SUMMARY

Cancer cells die when their decimated DNA damage response (DDR) unsuccessfully handles DNA damage. This notion has been successfully exploited when targeting PARP (poly ADP-ribose polymerase) in homologous recombination-deficient cells. With the greater understanding of DDR achieved in the last decade, new cancer therapy targets within the DDR network have been identified. Intriguingly, many of the molecules that have advanced into clinical trials are inhibitors of DDR kinases. This special issue is devoted to discussing the mechanism of cell killing and the level of success that such inhibitors reached in pre-clinical and clinical settings.

INTRODUCTION

The DNA Damage response is a signaling network that senses DNA damage and promotes both the adaptation of DNA replication and mitosis to changes in the quality of DNA templates and the activation of DNA repair mechanisms [1]. Because classic chemotherapeutic agents increase the load of DNA damage, it has been suggested that targeting the DDR of cancer cells may enhance their killing. Over the last decades, numerous screenings were performed to identify the effect of eliminating single DDR components on cancer cell killing. Breast cancer gene 1 (BRCA1) and 2 (BRCA2)- deficient tumors models were particularly suitable for this type of study as only the tumors cells, but not the healthy cells of the patients, are entirely devoided from BRCA expression. In 2005 two groups simultaneously reported that BRCA1 and BRCA2 breast cancers cell models are selectively sensitive to PARP inhibitors [2, 3]. Taking upon a notion initially used by developmental biologists, such a synergic killing was defined as "synthetic lethality". Today, three PARP inhibitors have already been approved by the FDA for treating ovarian, primary peritoneal and fallopian tube cancer: olaparib (Lynparza™, AstraZeneca, Cambridge UK), rucaparib (Rubraca®, Clovis Oncology, Boulder, CO, USA), and niraparib (Zejula™, Tesaro Inc., Waltham, MA, USA). Remarkably, although PARP inhibitors were initially developed as treatments that should be used in precision medicine on specific genetic backgrounds only, clinical trials in ovarian cancers demonstrated that they are effective independently of the BRCA status, perhaps in a manner that is

related to the high rate of BRCAness (Homologous recombination -HR deficiency in the context of BRCA wt status) of ovarian tumors [4-6].

The DDR network is not limited to PARP-dependent signaling and other synthetic lethal combinations have been reported [7]. Some DDR proteins (for example, the tumor suppressor p53) are frequently mutated in tumors [8]; some others, such as the Ataxia telangiectasia mutated (ATR) and checkpoint protein 1 kinase (Chk1), are not mutated in tumors[9]. Different strategies were designed to target either mutant p53 and ATR or Chk1 in vitro and ATR and Chk1 inhibitors have reached clinical trials.[10] ATR and Chk1 kinases are expressed both in healthy and cancer cells and their function is crucial to promote DNA replication when the DNA damage load increases [11]. While ATR and Chk1 are essential molecules in healthy and cancer cells, their selective effect on cancer cells has been ascribed to their increased replication stress [12]. In patients, such inhibitors were tested in monotherapy schemes and in combination with classical chemotoxins already validated in patients. Comprehensive reviews covering the progress in ATR and Chk1 targeting and the use of their inhibitors in the clinic have recently been published, for example [13]. Hence this special issue focuses on other kinases that have received less attention than ATR and Chk1.

SPECIAL ISSUE TOPICS

In this special issue, outstanding reviews written by experts in the field present different key DDR components from the perspective of their relevance as targets for cancer treatment. All reviews first provide a comprehensive introductory overview of the role of each kinase in the DDR; then discussing the potential of the inhibitors that have been designed so far; highlighting the progress of the best candidates into clinical trials.

In this issue, the review entitled "Clinical potential of ATM inhibitors" written by Martin F. Lavin and Abrey J. Yeo, the contribution of the ataxia telangiectasia mutated (ATM) kinase to the cell response to double-strand breaks (DSBs) accumulation is discussed.[14] Similarly to ATR and Chk1, ATM is relevant both for normal and cancer cell survival. Moreover, this review also presents other ATM functions, such as protection against oxidative and nutrient stress, which may also be relevant to the homeostasis of healthy and malignant cells [15-19]. Moreover, the authors also remind us that ATM was initially described as a kinase which loss causes a cancer-prone syndrome, Ataxia Telangiectasia.[20] Thereafter, it may seem puzzling to foresee a therapeutic advantage in eliminating a protein that, when lost, propels cancer genesis. The critical observation is that highly proliferating cells deficient in ATM expression are exquisitely

sensitive to a variety of DNA damaging agents.[21] The authors thoroughly developed this notion, focusing on the relevance of ATM activity both for the nuclear and mitochondrial function. Another subject covered in this review are the 10 ATM inhibitors developed so far, focusing on their effectiveness after transient exposure of cells to treatment, tolerance in mouse models, and capacity to enhance classical chemotherapy [22]. The advantages found when selectively targeting these types of small molecules to cancer tissues and when combining them with other DDR inhibitors such as PARPs are also discussed. The ongoing clinical trials for solid tumors using ATM inhibitors (AZD0156, KU-60019, AZD1390) are also described.

Also in this issue, the manuscript entitled "Targeting DNA-PK in cancer", Giovanna Damia reviews the contribution of the DNA-dependent protein kinase (DNA-PK) to DDR [23]. Similarly to ATM, DNA-PK has a primary role in the repair of DSBs. DNA-PK has a crucial role in non-homologous end joining (NHEJ), a DSBs repair pathway that is highly error-prone but nevertheless essential outside the S and G2 phases [24]. Similarly to ATM, DNA-PK has other DNA repair-independent functions, such as regulating the transcriptional machinery, the function of various transcriptional factors, and histones [25, 26]. The role of DNA-PK in protecting telomeres, promoting immune diversity, and generating innate immune response when dealing with foreign pathogens is presented [27-29]. The association between the dysregulation of DNA-PK expression and cancer and metastasis risk is also discussed [30]. Given its role in NHEJ, it has been hypothesized and demonstrated that acute downregulation of DNA-PK causes hypersensitivity to γ irradiation. Likewise, encouraging results were obtained when combining DNA-PK inhibitors with genotoxins and γ irradiation in pre-clinical models [31]. Ongoing and completed clinical trials are summarized, with the discussion focused on the unfortunate toxicity resulting from the systemic delivery of DNA-PK inhibitors. Strategies that could alleviate such toxicity, for example, identifying genetic backgrounds that are exquisitely sensitive to DNA-PK inhibitors, are debated as well [32-34].

In the manuscript entitled "Wee1 kinase limits CDK activities to safeguard DNA replication and mitotic entry ", which is part of this issue, Camille Elbæk, Valdemaras Petrosius and Claus Sørensen discuss the central role of Wee1 in the control of the signals that shut down the S phase [35]. While working in coordination, Wee1 and Chk1 have different roles. Wee1 phosphorylates and regulates the activity of CDKs, controlling the timing of S phase finalization. Wee1 can delay mitosis onset, but it also functions within S phase. Wee1 loss causes an intra-S-phase replication catastrophe, characterized by increased origin firing, reduction in the nucleotide pool, accumulation of forks with augmented regions of single-stranded DNA, and dysregulated nucleases which pulverize chromosomes [36-38]. The interaction of Chk1

and Wee1 functions is discussed, focusing on the nature of the synthetic lethality between their inhibitors. The challenges faced by researchers during the design of specific Wee1 inhibitors are also debated [39]. Moreover, the review also discusses novel approaches to the design of inhibitors such as the use of Proteolysis-Targeting Chimeras, which may boost the therapeutic potential of Wee1 inhibitors due to their specificity [40]. The evolution of clinical strategies involving Wee1 inhibitors is also put into the spotlight, and current clinical trials focused on the combination and sequential therapies that circumvent toxicity are presented [41-45].

In the review entitled "Therapeutic opportunities for PLK1 inhibitors: Spotlight on BRCA1-deficiency and triple-negative breast cancers" which is part of this special issue, Gastón Soria and coworkers concentrate their analysis on the DDR events regulated by the master polo-like mitotic kinase (PLK1) [46]. Antimitotic agents or mitotic kinases have received much attention as a target of cancer treatment; [47] however, a high toxicity of PLK1 inhibitors has been demonstrated in the clinic. Such a disappointing outcome might have to do with the crucial role of PLK1 in the survival of both healthy and cancer cells. In fact, the authors highlight the extremely sporadic frequency of PLK1 mutations in human cancers and the striking lethality observed after PLK1 ablation in more than 700 cell lines. The authors also remind us that, even in the most successful clinical trials, the dosage limitations imposed by the toxicity issues have allowed only moderate disease stabilization in a limited number of patients (for example, [48-50]). Such a modest success has demonstrated that the only way to profit from these FDA-approved inhibitors is to find a genetic background that is particularly sensitive to PLK1 inhibition. The authors propose two potential niches for PLK1 inhibitors. The tumor cell killing capacity of PLK1 inhibitors is augmented in triple-negative breast cancers (TNBCs) and BRCA1 deficient cancers [51-54]. The authors emphasize that a retrospective analysis of human breast tumors demonstrated that BRCA1 deficient tumors had augmented levels of PLK1. In this review, they include a similar analysis demonstrating a similar scenario for TNBCs. An increase reliance of these types of cancers on PLK1 function may decrease the PLK1 inhibitor doses, reducing the toxicity issues observed in the healthy cells of patients.

In this issue, the review entitled "Aurora kinases and DNA damage response" by Hoi Tang Ma and Randy Y.C. Poon, the mitotic aurora kinases A (AURKA) and B (AURKB) are discussed [55]. Their contribution to mitosis progression, cell differentiation events, stem cell maintenance, chromatin remodeling, and telomeres preservation is presented. The review thoroughly discusses the role of AURKA and AURKB (AURKs) in guarding against chromosome segregation mistakes [56-58]. DDR dysregulation after AURKs overexpression is also discussed, highlighting that AURKs augmentation is frequently observed in solid

tumors [59-61]. Alterations in the function of AURKs causes the transmission of DNA damage to the subsequent G1, impacting on the activation of the tumor suppressor p53 in G1, which in turn downregulates AURKs [62]. Likewise, AURKs dysregulation modulates HR, NHEJ, and PARP activation, and in turn, DNA repair pathways downregulate excess AURKs activation [63, 64]. Such observation suggested clinical benefits after AURK inhibition. Aurora kinase inhibitors, specific for either AURKA and AURKB, are currently available and were successfully combined with DNA damaging agents in Phase II clinical trials designed to treat neuroblastoma and acute myeloid leukemia [65, 66]. However, the authors remind us that AURKA and AURKB inhibitors per se trigger DDR [67, 68], and AURKA and AURKB are turned off by DNA damage. Hence sequential rather than combined therapies may be a better choice, which should be tested shortly.

The review entitled "Targeting AKT/PKB to improve treatment outcomes for solid tumors" from this issue is authored by Mahmoud Toulany and coworkers [69]. They focus the manuscript on the current understanding of the role of protein kinase B, also known as AKT, from the perspective of its interaction with the DDR response. Most known AKT functions are related to its activation at the cellular membrane by receptor tyrosine kinases (RTK) and other proteins [70-72]. However, AKT can also be activated in the nucleus by DSBs associated kinases, ATM and DNA-PK, and perhaps other kinases to promote the DNA damage tolerance event (TLS) and the DSB repair pathway (HR and NHEJ) [73-75]. They also devote a section to discuss the dysregulation of AKT and its relevance for cancer genesis as AKT hyperactivation is frequent and is considered a marker of poor prognosis (for example, [76]). The fact that conventional chemo- and radio-therapy induce the activation of the nuclear fraction of AKT is also discussed, exploring as well its link with the blockage of apoptosis and the augmentation of resistance events in different cancer models [77, 78]. Specific AKT inhibition and compounds that block AKT activity, such as perifosine and nelfinavir [79, 80] are also discussed in this review. The clinical trials involving AKT inhibitors, the most successful combinations found so far, and the challenges associated with toxicity and resistance events are also analyzed.

CONCLUSIONS:

The fast proliferation of cancer cells and their incapacity to arrest when facing increased levels of DNA damage has encouraged the development of therapeutic agents that increase the load of DNA damage. The limited success of those treatments has prompted the exploration of DDR as a therapeutic target, which may reduce the cancer cell capacity to complete DNA replication in the face of DNA damage. Several DDR kinases were successfully targeted by specific inhibitors; and those compounds have been tested

with a certain degree of success in clinical trials. Those results also indicate options which should be explored: 1) identifying genetic backgrounds which are more sensitive to the treatment, 2) identifying treatment combinations, possibly involving immune checkpoint therapy, which may be more effective in cancers when compared to highly proliferating normal cells, 3) attempting selective delivery of those agents to tumor cells. It is highly likely that inhibitors of kinases or other DDR factors will follow PARP inhibitors' successful path in the years to come.

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References

- [1] P.G. Pilie, C. Tang, G.B. Mills, T.A. Yap, State-of-the-art strategies for targeting the DNA damage response in cancer, *Nature reviews. Clinical oncology*, 16 (2019) 81-104.
- [2] H.E. Bryant, N. Schultz, H.D. Thomas, K.M. Parker, D. Flower, E. Lopez, S. Kyle, M. Meuth, N.J. Curtin, T. Helleday, Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase, *Nature*, 434 (2005) 913-917.
- [3] H. Farmer, N. McCabe, C.J. Lord, A.N. Tutt, D.A. Johnson, T.B. Richardson, M. Santarosa, K.J. Dillon, I. Hickson, C. Knights, N.M. Martin, S.P. Jackson, G.C. Smith, A. Ashworth, Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy, *Nature*, 434 (2005) 917-921.
- [4] C.J. Lord, A. Ashworth, PARP inhibitors: Synthetic lethality in the clinic, *Science*, 355 (2017) 1152-1158.
- [5] M.R. Mirza, B.J. Monk, J. Herrstedt, A.M. Oza, S. Mahner, A. Redondo, M. Fabbro, J.A. Ledermann, D. Lorusso, I. Vergote, N.E. Ben-Baruch, C. Marth, R. Madry, R.D. Christensen, J.S. Berek, A. Dorum, A.V. Tinker, A. du Bois, A. Gonzalez-Martin, P. Follana, B. Benigno, P. Rosenberg, L. Gilbert, B.J. Rimel, J. Buscema, J.P. Balser, S. Agarwal, U.A. Matulonis, E.-O.N. Investigators, Niraparib Maintenance Therapy in Platinum-Sensitive, Recurrent Ovarian Cancer, *The New England journal of medicine*, 375 (2016) 2154-2164.
- [6] D. Killock, Gynaecological cancer: PARP inhibition - moving beyond BRCA-mutated disease, *Nature reviews. Clinical oncology*, 14 (2017) 71.
- [7] A. Huang, L.A. Garraway, A. Ashworth, B. Weber, Synthetic lethality as an engine for cancer drug target discovery, *Nature reviews. Drug discovery*, 19 (2020) 23-38.
- [8] V.J.N. Bykov, S.E. Eriksson, J. Bianchi, K.G. Wiman, Targeting mutant p53 for efficient cancer therapy, *Nature reviews. Cancer*, 18 (2018) 89-102.
- [9] S. Rundle, A. Bradbury, Y. Drew, N.J. Curtin, Targeting the ATR-CHK1 Axis in Cancer Therapy, *Cancers*, 9 (2017).
- [10] D. Nazareth, M.J. Jones, B. Gabrielli, Everything in Moderation: Lessons Learned by Exploiting Moderate Replication Stress in Cancer, *Cancers*, 11 (2019).
- [11] M.A. Gonzalez Besteiro, V. Gottifredi, The fork and the kinase: a DNA replication tale from a CHK1 perspective, *Mutation research. Reviews in mutation research*, 763 (2015) 168-180.

- [12] P. Gralewska, A. Gajek, A. Marczak, A. Rogalska, Participation of the ATR/CHK1 pathway in replicative stress targeted therapy of high-grade ovarian cancer, *Journal of hematology & oncology*, 13 (2020) 39.
- [13] L. Carrassa, G. Damia, DNA damage response inhibitors: Mechanisms and potential applications in cancer therapy, *Cancer treatment reviews*, 60 (2017) 139-151.
- [14] M.F. Lavin, A.J. Yeo, Clinical potential of ATM inhibitors, *Mutation research*, 821 (2020) 111695.
- [15] M.F. Lavin, Ataxia-telangiectasia: from a rare disorder to a paradigm for cell signalling and cancer, *Nature reviews. Molecular cell biology*, 9 (2008) 759-769.
- [16] D. Watters, P. Kedar, K. Spring, J. Bjorkman, P. Chen, M. Gatei, G. Birrell, B. Garrone, P. Srinivasa, D.I. Crane, M.F. Lavin, Localization of a portion of extranuclear ATM to peroxisomes, *The Journal of biological chemistry*, 274 (1999) 34277-34282.
- [17] J. Zhang, D.N. Tripathi, J. Jing, A. Alexander, J. Kim, R.T. Powell, R. Dere, J. Tait-Mulder, J.H. Lee, T.T. Paull, R.K. Pandita, V.K. Charaka, T.K. Pandita, M.B. Kastan, C.L. Walker, ATM functions at the peroxisome to induce pexophagy in response to ROS, *Nature cell biology*, 17 (2015) 1259-1269.
- [18] D.S. Lim, D.G. Kirsch, C.E. Canman, J.H. Ahn, Y. Ziv, L.S. Newman, R.B. Darnell, Y. Shiloh, M.B. Kastan, ATM binds to beta-adaptin in cytoplasmic vesicles, *Proceedings of the National Academy of Sciences of the United States of America*, 95 (1998) 10146-10151.
- [19] A. Morita, K. Tanimoto, T. Murakami, T. Morinaga, Y. Hosoi, Mitochondria are required for ATM activation by extranuclear oxidative stress in cultured human hepatoblastoma cell line Hep G2 cells, *Biochemical and biophysical research communications*, 443 (2014) 1286-1290.
- [20] Y. Shiloh, The cerebellar degeneration in ataxia-telangiectasia: A case for genome instability, *DNA repair*, 95 (2020) 102950.
- [21] I. Corcoles-Saez, K. Dong, R.S. Cha, Versatility of the Mec1(ATM/ATR) signaling network in mediating resistance to replication, genotoxic, and proteotoxic stresses, *Current genetics*, 65 (2019) 657-661.
- [22] M.H. Jin, D.Y. Oh, ATM in DNA repair in cancer, *Pharmacology & therapeutics*, 203 (2019) 107391.
- [23] G. Damia, Targeting DNA-PK in cancer, *Mutation research*, 821 (2020) 111692.
- [24] N.R. Pannunzio, G. Watanabe, M.R. Lieber, Nonhomologous DNA end-joining for repair of DNA double-strand breaks, *The Journal of biological chemistry*, 293 (2018) 10512-10523.
- [25] J.F. Goodwin, K.E. Knudsen, Beyond DNA repair: DNA-PK function in cancer, *Cancer discovery*, 4 (2014) 1126-1139.
- [26] M. Bustin, F. Catez, J.H. Lim, The dynamics of histone H1 function in chromatin, *Molecular cell*, 17 (2005) 617-620.
- [27] V.C. George, S.A. Ansari, V.S. Chelakkot, A.L. Chelakkot, C. Chelakkot, V. Menon, W. Ramadan, K.R. Ethiraj, R. El-Awady, T. Mantso, M. Mitsiogianni, M.I. Panagiotidis, G. Dellaire, H.P. Vasantha Rupasinghe, DNA-dependent protein kinase: Epigenetic alterations and the role in genomic stability of cancer, *Mutation research*, 780 (2019) 92-105.
- [28] R. Hill, P.W. Lee, The DNA-dependent protein kinase (DNA-PK): More than just a case of making ends meet?, *Cell cycle*, 9 (2010) 3460-3469.
- [29] B.J. Ferguson, D.S. Mansur, N.E. Peters, H. Ren, G.L. Smith, DNA-PK is a DNA sensor for IRF-3-dependent innate immunity, *eLife*, 1 (2012) e00047.
- [30] C. Schwartz, O. Rohr, C. Wallet, Targeting the DNA-PK complex: Its rationale use in cancer and HIV-1 infection, *Biochemical pharmacology*, 160 (2019) 80-91.
- [31] C.R. Timme, B.H. Rath, J.W. O'Neill, K. Camphausen, P.J. Tofilon, The DNA-PK Inhibitor VX-984 Enhances the Radiosensitivity of Glioblastoma Cells Grown In Vitro and as Orthotopic Xenografts, *Molecular cancer therapeutics*, 17 (2018) 1207-1216.
- [32] N. Albarakati, T.M. Abdel-Fatah, R. Doherty, R. Russell, D. Agarwal, P. Moseley, C. Perry, A. Arora, N. Alsubhi, C. Seedhouse, E.A. Rakha, A. Green, G. Ball, S. Chan, C. Caldas, I.O. Ellis, S. Madhusudan, Targeting BRCA1-BER deficient breast cancer by ATM or DNA-PKcs blockade either alone or in combination with cisplatin for personalized therapy, *Molecular oncology*, 9 (2015) 204-217.

- [33] F. Dietlein, L. Thelen, M. Jokic, R.D. Jachimowicz, L. Ivan, G. Knittel, U. Leeser, J. van Oers, W. Edelmann, L.C. Heukamp, H.C. Reinhardt, A functional cancer genomics screen identifies a druggable synthetic lethal interaction between MSH3 and PRKDC, *Cancer discovery*, 4 (2014) 592-605.
- [34] H. Jiang, H.C. Reinhardt, J. Bartkova, J. Tommiska, C. Blomqvist, H. Nevanlinna, J. Bartek, M.B. Yaffe, M.T. Hemann, The combined status of ATM and p53 link tumor development with therapeutic response, *Genes & development*, 23 (2009) 1895-1909.
- [35] C.R. Elbaek, V. Petrosius, C.S. Sorensen, WEE1 kinase limits CDK activities to safeguard DNA replication and mitotic entry, *Mutation research*, 819-820 (2020) 111694.
- [36] H. Beck, V. Nahse-Kumpf, M.S. Larsen, K.A. O'Hanlon, S. Patzke, C. Holmberg, J. Mejlvang, A. Groth, O. Nielsen, R.G. Syljuasen, C.S. Sorensen, Cyclin-dependent kinase suppression by WEE1 kinase protects the genome through control of replication initiation and nucleotide consumption, *Molecular and cellular biology*, 32 (2012) 4226-4236.
- [37] L.I. Toledo, M. Altmeyer, M.B. Rask, C. Lukas, D.H. Larsen, L.K. Povlsen, S. Bekker-Jensen, N. Mailand, J. Bartek, J. Lukas, ATR prohibits replication catastrophe by preventing global exhaustion of RPA, *Cell*, 155 (2013) 1088-1103.
- [38] H. Duda, M. Arter, J. Gloggnitzer, F. Teloni, P. Wild, M.G. Blanco, M. Altmeyer, J. Matos, A Mechanism for Controlled Breakage of Under-replicated Chromosomes during Mitosis, *Developmental cell*, 40 (2017) 421-422.
- [39] A.F. Serpico, G. D'Alterio, C. Vetrei, R. Della Monica, L. Nardella, R. Visconti, D. Grieco, Wee1 Rather Than Plk1 Is Inhibited by AZD1775 at Therapeutically Relevant Concentrations, *Cancers*, 11 (2019).
- [40] Z. Li, B.J. Pinch, C.M. Olson, K.A. Donovan, R.P. Nowak, C.E. Mills, D.A. Scott, Z.M. Doctor, N.A. Eleuteri, M. Chung, P.K. Sorger, E.S. Fischer, N.S. Gray, Development and Characterization of a Wee1 Kinase Degradable, *Cell chemical biology*, 27 (2020) 57-65 e59.
- [41] K.C. Cuneo, M.A. Morgan, V. Sahai, M.J. Schipper, L.A. Parsels, J.D. Parsels, T. Devasia, M. Al-Hawaray, C.S. Cho, H. Nathan, J. Maybaum, M.M. Zalupski, T.S. Lawrence, Dose Escalation Trial of the Wee1 Inhibitor Adavosertib (AZD1775) in Combination With Gemcitabine and Radiation for Patients With Locally Advanced Pancreatic Cancer, *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 37 (2019) 2643-2650.
- [42] C.J. Matheson, S. Venkataraman, V. Amani, P.S. Harris, D.S. Backos, A.M. Donson, M.F. Wempe, N.K. Foreman, R. Vibhakkar, P. Reigan, A WEE1 Inhibitor Analog of AZD1775 Maintains Synergy with Cisplatin and Demonstrates Reduced Single-Agent Cytotoxicity in Medulloblastoma Cells, *ACS chemical biology*, 11 (2016) 921-930.
- [43] A.B. Bukhari, C.W. Lewis, J.J. Pearce, D. Luong, G.K. Chan, A.M. Gamper, Inhibiting Wee1 and ATR kinases produces tumor-selective synthetic lethality and suppresses metastasis, *The Journal of clinical investigation*, 129 (2019) 1329-1344.
- [44] S. Hauge, C. Naucke, G. Hasvold, M. Joel, G.E. Rodland, P. Juzenas, T. Stokke, R.G. Syljuasen, Combined inhibition of Wee1 and Chk1 gives synergistic DNA damage in S-phase due to distinct regulation of CDK activity and CDC45 loading, *Oncotarget*, 8 (2017) 10966-10979.
- [45] Y. Fang, D.J. McGrail, C. Sun, M. Labrie, X. Chen, D. Zhang, Z. Ju, C.P. Vellano, Y. Lu, Y. Li, K.J. Jeong, Z. Ding, J. Liang, S.W. Wang, H. Dai, S. Lee, N. Sahni, I. Mercado-Urbe, T.B. Kim, K. Chen, S.Y. Lin, G. Peng, S.N. Westin, J. Liu, M.J. O'Connor, T.A. Yap, G.B. Mills, Sequential Therapy with PARP and WEE1 Inhibitors Minimizes Toxicity while Maintaining Efficacy, *Cancer cell*, 35 (2019) 851-867 e857.
- [46] I.A. Garcia, C. Garro, E. Fernandez, G. Soria, Therapeutic opportunities for PLK1 inhibitors: Spotlight on BRCA1-deficiency and triple negative breast cancers, *Mutation research*, 821 (2020) 111693.
- [47] C. Dominguez-Brauer, K.L. Thu, J.M. Mason, H. Blaser, M.R. Bray, T.W. Mak, Targeting Mitosis in Cancer: Emerging Strategies, *Molecular cell*, 60 (2015) 524-536.
- [48] E. Pujade-Lauraine, F. Selle, B. Weber, I.L. Ray-Coquard, I. Vergote, J. Sufliarsky, J.M. Del Campo, A. Lortholary, A. Lesoin, P. Follana, G. Freyer, B. Pardo, L. Vidal, B. Tholander, L. Gladieff, M. Sassi, P. Garin-

- Chesa, S. Nazabadioko, K. Marzin, K. Pilz, F. Joly, Volasertib Versus Chemotherapy in Platinum-Resistant or -Refractory Ovarian Cancer: A Randomized Phase II Groupe des Investigateurs Nationaux pour l'Etude des Cancers de l'Ovaire Study, *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 34 (2016) 706-713.
- [49] D. Olmos, D. Barker, R. Sharma, A.T. Brunetto, T.A. Yap, A.B. Taegtmeyer, J. Barriuso, H. Medani, Y.Y. Degenhardt, A.J. Allred, D.A. Smith, S.C. Murray, T.A. Lampkin, M.M. Dar, R. Wilson, J.S. de Bono, S.P. Blagden, Phase I study of GSK461364, a specific and competitive Polo-like kinase 1 inhibitor, in patients with advanced solid malignancies, *Clinical cancer research : an official journal of the American Association for Cancer Research*, 17 (2011) 3420-3430.
- [50] H. Dohner, M. Lubbert, W. Fiedler, L. Fouillard, A. Haaland, J.M. Brandwein, S. Lepretre, O. Reman, P. Turlure, O.G. Ottmann, C. Muller-Tidow, A. Kramer, E. Raffoux, K. Dohner, R.F. Schlenk, F. Voss, T. Taube, H. Fritsch, J. Maertens, Randomized, phase 2 trial of low-dose cytarabine with or without volasertib in AML patients not suitable for induction therapy, *Blood*, 124 (2014) 1426-1433.
- [51] V. Maire, F. Nemati, M. Richardson, A. Vincent-Salomon, B. Tesson, G. Rigai, E. Gravier, B. Marty-Prouvost, L. De Koning, G. Lang, D. Gentien, A. Dumont, E. Barillot, E. Marangoni, D. Decaudin, S. Roman-Roman, A. Pierre, F. Cruzalegui, S. Depil, G.C. Tucker, T. Dubois, Polo-like kinase 1: a potential therapeutic option in combination with conventional chemotherapy for the management of patients with triple-negative breast cancer, *Cancer research*, 73 (2013) 813-823.
- [52] A. Ueda, K. Oikawa, K. Fujita, A. Ishikawa, E. Sato, T. Ishikawa, M. Kuroda, K. Kanekura, Therapeutic potential of PLK1 inhibition in triple-negative breast cancer, *Laboratory investigation; a journal of technical methods and pathology*, 99 (2019) 1275-1286.
- [53] J. Zou, K. Rezvani, H. Wang, K.S. Lee, D. Zhang, BRCA1 downregulates the kinase activity of Polo-like kinase 1 in response to replication stress, *Cell cycle*, 12 (2013) 2255-2265.
- [54] S. Carbajosa, M.F. Pansa, N.S. Paviolo, A.M. Castellaro, D.L. Andino, A.D. Nigra, I.A. Garcia, A.C. Racca, L. Rodriguez-Berdini, V. Angiolini, L. Guantay, F. Villafanez, M.B. Federico, M.C. Rodriguez-Baili, B.L. Caputto, G. Drewes, K.P. Madauss, I. Gloger, E. Fernandez, G.A. Gil, J.L. Bocco, V. Gottifredi, G. Soria, Polo-like Kinase 1 Inhibition as a Therapeutic Approach to Selectively Target BRCA1-Deficient Cancer Cells by Synthetic Lethality Induction, *Clinical cancer research : an official journal of the American Association for Cancer Research*, 25 (2019) 4049-4062.
- [55] H.T. Ma, R.Y.C. Poon, Aurora kinases and DNA damage response, *Mutation research*, 821 (2020) 111716.
- [56] S.F. Bakhoum, L. Kabeche, J.P. Murnane, B.I. Zaki, D.A. Compton, DNA-damage response during mitosis induces whole-chromosome missegregation, *Cancer discovery*, 4 (2014) 1281-1289.
- [57] E. Petsalaki, G. Zachos, Building bridges between chromosomes: novel insights into the abscission checkpoint, *Cellular and molecular life sciences : CMLS*, 76 (2019) 4291-4307.
- [58] L. Kabeche, H.D. Nguyen, R. Buisson, L. Zou, A mitosis-specific and R loop-driven ATR pathway promotes faithful chromosome segregation, *Science*, 359 (2018) 108-114.
- [59] M.M. Tanner, S. Grenman, A. Koul, O. Johannsson, P. Meltzer, T. Pejovic, A. Borg, J.J. Isola, Frequent amplification of chromosomal region 20q12-q13 in ovarian cancer, *Clinical cancer research : an official journal of the American Association for Cancer Research*, 6 (2000) 1833-1839.
- [60] P. Meraldi, R. Honda, E.A. Nigg, Aurora-A overexpression reveals tetraploidization as a major route to centrosome amplification in p53^{-/-} cells, *The EMBO journal*, 21 (2002) 483-492.
- [61] A. Gonzalez-Loyola, G. Fernandez-Miranda, M. Trakala, D. Partida, K. Samejima, H. Ogawa, M. Canamero, A. de Martino, A. Martinez-Ramirez, G. de Carcer, I. Perez de Castro, W.C. Earnshaw, M. Malumbres, Aurora B Overexpression Causes Aneuploidy and p21Cip1 Repression during Tumor Development, *Molecular and cellular biology*, 35 (2015) 3566-3578.
- [62] S.S. Chen, P.C. Chang, Y.W. Cheng, F.M. Tang, Y.S. Lin, Suppression of the STK15 oncogenic activity requires a transactivation-independent p53 function, *The EMBO journal*, 21 (2002) 4491-4499.

- [63] T. Sourisseau, D. Maniotis, A. McCarthy, C. Tang, C.J. Lord, A. Ashworth, S. Linardopoulos, Aurora-A expressing tumour cells are deficient for homology-directed DNA double strand-break repair and sensitive to PARP inhibition, *EMBO molecular medicine*, 2 (2010) 130-142.
- [64] T.V. Do, J. Hirst, S. Hyter, K.F. Roby, A.K. Godwin, Aurora A kinase regulates non-homologous end-joining and poly(ADP-ribose) polymerase function in ovarian carcinoma cells, *Oncotarget*, 8 (2017) 50376-50392.
- [65] S.G. DuBois, Y.P. Mosse, E. Fox, R.A. Kudgus, J.M. Reid, R. McGovern, S. Groshen, R. Bagatell, J.M. Maris, C.J. Twist, K. Goldsmith, M.M. Granger, B. Weiss, J.R. Park, M.E. Macy, S.L. Cohn, G. Yanik, L.M. Wagner, R. Hawkins, J. Courtier, H. Lai, F. Goodarzian, H. Shimada, N. Boucher, S. Czarnecki, C. Luo, D. Tsao-Wei, K.K. Matthay, A. Marachelian, Phase II Trial of Alisertib in Combination with Irinotecan and Temozolomide for Patients with Relapsed or Refractory Neuroblastoma, *Clinical cancer research : an official journal of the American Association for Cancer Research*, 24 (2018) 6142-6149.
- [66] A.M. Brunner, T.M. Blonquist, D.J. DeAngelo, M. McMasters, G. Fell, N.M. Hermance, E.S. Winer, R.C. Lindsley, G.S. Hobbs, P.C. Amrein, H.R. Hock, D.P. Steensma, J.S. Garcia, M.R. Luskin, R.M. Stone, K.K. Ballen, J. Rosenblatt, D. Avigan, M.R. Nahas, L.M. Mendez, S.L. McAfee, J.A. Moran, M. Bergeron, J. Foster, C. Bertoli, A.L. Manning, K.L. McGregor, K.M. Fishman, F.C. Kuo, M.T. Baltay, M. Macrae, M. Burke, T. Behnan, M.C. Wey, T.T. Som, A.Y. Ramos, J. Rae, J. Lombardi Story, N. Nelson, E. Logan, C. Connolly, D.S. Neuberg, Y.B. Chen, T.A. Graubert, A.T. Fathi, Alisertib plus induction chemotherapy in previously untreated patients with high-risk, acute myeloid leukaemia: a single-arm, phase 2 trial, *The Lancet. Haematology*, 7 (2020) e122-e133.
- [67] Y. Liu, O.E. Hawkins, Y. Su, A.E. Vilgelm, T. Sobolik, Y.M. Thu, S. Kantrow, R.C. Splittgerber, S. Short, K.I. Amiri, J.A. Ecsedy, J.A. Sosman, M.C. Kelley, A. Richmond, Targeting aurora kinases limits tumour growth through DNA damage-mediated senescence and blockade of NF-kappaB impairs this drug-induced senescence, *EMBO molecular medicine*, 5 (2013) 149-166.
- [68] M.R. Dreier, A.Z. Grabovich, J.D. Katusin, W.R. Taylor, Short and long-term tumor cell responses to Aurora kinase inhibitors, *Experimental cell research*, 315 (2009) 1085-1099.
- [69] M. Iida, P.M. Harari, D.L. Wheeler, M. Toulany, Targeting AKT/PKB to improve treatment outcomes for solid tumors, *Mutation research*, 819-820 (2020) 111690.
- [70] T. Regad, Targeting RTK Signaling Pathways in Cancer, *Cancers*, 7 (2015) 1758-1784.
- [71] E. Rozengurt, Mitogenic signaling pathways induced by G protein-coupled receptors, *Journal of cellular physiology*, 213 (2007) 589-602.
- [72] M. Delcommenne, C. Tan, V. Gray, L. Rue, J. Woodgett, S. Dedhar, Phosphoinositide-3-OH kinase-dependent regulation of glycogen synthase kinase 3 and protein kinase B/AKT by the integrin-linked kinase, *Proceedings of the National Academy of Sciences of the United States of America*, 95 (1998) 11211-11216.
- [73] K. Szymonowicz, S. Oeck, N.M. Malewicz, V. Jendrosseck, New Insights into Protein Kinase B/Akt Signaling: Role of Localized Akt Activation and Compartment-Specific Target Proteins for the Cellular Radiation Response, *Cancers*, 10 (2018).
- [74] F. Villafanez, I.A. Garcia, S. Carbajosa, M.F. Pansa, S. Mansilla, M.C. Llorens, V. Angiolini, L. Guantay, H. Jacobs, K.P. Madauss, I. Gloger, V. Gottifredi, J.L. Bocco, G. Soria, AKT inhibition impairs PCNA ubiquitylation and triggers synthetic lethality in homologous recombination-deficient cells submitted to replication stress, *Oncogene*, 38 (2019) 4310-4324.
- [75] M. Toulany, M. Iida, S. Keinath, F.F. Iyi, K. Mueck, B. Fehrenbacher, W.Y. Mansour, M. Schaller, D.L. Wheeler, H.P. Rodemann, Dual targeting of PI3K and MEK enhances the radiation response of K-RAS mutated non-small cell lung cancer, *Oncotarget*, 7 (2016) 43746-43761.
- [76] J. Tsurutani, J. Fukuoka, H. Tsurutani, J.H. Shih, S.M. Hewitt, W.D. Travis, J. Jen, P.A. Dennis, Evaluation of two phosphorylation sites improves the prognostic significance of Akt activation in non-small-cell lung

cancer tumors, *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 24 (2006) 306-314.

[77] J.C. Hahne, A. Honig, S.R. Meyer, S. Gambaryan, U. Walter, J. Wischhusen, S.F. Haussler, S.E. Segerer, N. Fujita, J. Dietl, J.B. Engel, Downregulation of AKT reverses platinum resistance of human ovarian cancers in vitro, *Oncology reports*, 28 (2012) 2023-2028.

[78] K. Gohr, A. Hamacher, L.H. Engelke, M.U. Kassack, Inhibition of PI3K/Akt/mTOR overcomes cisplatin resistance in the triple negative breast cancer cell line HCC38, *BMC cancer*, 17 (2017) 711.

[79] A.K. Gupta, G.J. Cerniglia, R. Mick, W.G. McKenna, R.J. Muschel, HIV protease inhibitors block Akt signaling and radiosensitize tumor cells both in vitro and in vivo, *Cancer research*, 65 (2005) 8256-8265.

[80] J.J. Gills, P.A. Dennis, Perifosine: update on a novel Akt inhibitor, *Current oncology reports*, 11 (2009) 102-110.