

# Draft Genome Sequence of *Geobacillus* sp. Isolate T6, a Thermophilic Bacterium Collected from a Thermal Spring in Argentina

Elio M. Ortiz,<sup>a,c</sup> Marcelo F. Berretta,<sup>a,c</sup> Laura E. Navas,<sup>a,c</sup> Graciela B. Benintende,<sup>a</sup>  Ariel F. Amadio,<sup>b,c</sup> Rubén O. Zandomeni<sup>a,c</sup>

Instituto de Microbiología y Zootecnia Agrícola, Instituto Nacional de Tecnología Agropecuaria (INTA), Buenos Aires, Argentina<sup>a</sup>; Estación Experimental Agropecuaria Rafaela, Instituto Nacional de Tecnología Agropecuaria (INTA), Santa Fe, Argentina<sup>b</sup>; Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Buenos Aires, Argentina<sup>c</sup>

A.F.A. and R.O.Z. contributed equally to and directed different aspects of the work.

***Geobacillus* sp. isolate T6 was collected from a thermal spring in Salta, Argentina. The draft genome sequence (3,767,773 bp) of this isolate is represented by one major scaffold of 3.46 Mbp, a second one of 207 kbp, and 20 scaffolds of <13 kbp. The assembled sequences revealed 3,919 protein-coding genes.**

Received 29 May 2015 Accepted 15 June 2015 Published 16 July 2015

**Citation** Ortiz EM, Berretta MF, Navas LE, Benintende GB, Amadio AF, Zandomeni RO. 2015. Draft genome sequence of *Geobacillus* sp. isolate T6, a thermophilic bacterium collected from a thermal spring in Argentina. *Genome Announc* 3(4):e00743-15. doi:10.1128/genomeA.00743-15.

**Copyright** © 2015 Ortiz et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Laura E. Navas, [navas.laura@inta.gob.ar](mailto:navas.laura@inta.gob.ar).

Some members of the genus *Geobacillus* are thermophilic Gram-positive endospore-forming bacteria, usually isolated from high-temperature environments. Thermophilic strains of this genus have attracted interest as sources of thermostable enzymes with potential biotechnological applications, including proteases, lipases, and glycoside-hydrolases (1). *Geobacillus* sp. isolate T6 was obtained from a hot water spring in Rosario de la Frontera, Salta, in the northwest of Argentina.

The draft genome sequence was generated using a combined approach between the Roche 454 GS-FLX and Illumina MiSeq platforms (MWG Eurofins), producing unpaired and paired-end reads, respectively. A total of 218,128 single reads with an average length of 350 bp were produced by Roche 454, resulting in 16-fold coverage of the genome. Furthermore, 3,682,898 paired-end plus 3,587,489 singleton reads with an average of 126 bp were obtained using a long jumping distance library with an insert size of 8 kbp from an Illumina MiSeq 2 × 150-bp run, reaching 239-fold coverage. *De novo* hybrid assembly of the reads was performed using Velvet version 1.2.10 (2) with a *k*-mer size of 99. The assembly consists of one major scaffold of 3,468,726 bp (containing 211 contigs), a second scaffold of 207,976 bp, and 20 minor scaffolds of >1 kbp. The order and orientation of the contigs into scaffolds were assessed using Mega BLAST (3) to align the resultant assembly and the genome of *Geobacillus kaustophilus* HTA426, and were visualized using the Artemis comparison tool (4).

The draft genome sequence of *Geobacillus* sp. isolate T6 consists of 3,767,773 bp with an average G + C content of 53%. It was subjected to automated annotation using the RAST server version 2.0 (5) and the NCBI Prokaryotic Genome Annotation Pipeline (PGAP). The tRNA and rRNA genes were predicted by tRNAscan-SE version 1.21 (6) and RNAmmer version 1.2 (7), respectively.

In total, 4,011 genes were identified, including only one copy of 16S rRNA, 13 copies of 23S/5S rRNA, and 78 tRNA genes. RAST predicted 3,919 protein-coding genes, 47% of which were as-

signed to 452 subsystems. Phylogenetic analysis of the 16S rRNA gene confirmed the affiliation of *Geobacillus* sp. isolate T6 to the genus *Geobacillus*. *Geobacillus kaustophilus* HTA426 (score 531), *Geobacillus thermodenitrificans* NG80-2 (score 495), and *Geobacillus* sp. Y412MC61 (score 495) are the closest neighbor genomes based on RAST analysis.

A group of 878 enzyme-coding genes was appointed as potentially useful in several industries according to their predicted activity, including agriculture, environment, biosensor, biotechnology, medicine, biofuel, food, and other industries. Cloning and expression strategies are under way to confirm the functionality of some of these enzymes. The main goals of our work in gaining genome-based knowledge of thermophilic organisms are to characterize their thermostable enzymes and unravel the metabolic pathways adapted to extreme living conditions.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [LDNZ000000000](https://www.ncbi.nlm.nih.gov/nuccore/LDNZ000000000). The version described in this paper is the first version, LDNZ01000000.

## ACKNOWLEDGMENTS

We thank Irma Fuxan for the technical support in the genomic DNA isolation and the shipping of the sample for sequencing.

This work was supported by project INTA PNAIyAV-1130032.

## REFERENCES

- Niehaus F, Bertoldo C, Kähler M, Antranikian G. 1999. Extremophiles as a source of novel enzymes for industrial application. *Appl Microbiol Biotechnol* 51:711–729. <http://dx.doi.org/10.1007/s002530051456>.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <http://dx.doi.org/10.1186/1471-2105-10-421>.
- Carver TJ, Rutherford KM, Berriman M, Rajandream MA, Barrell BG,

- Parkhill J. 2005. ACT: the Artemis comparison tool. *Bioinformatics* 21: 3422–3423. <http://dx.doi.org/10.1093/bioinformatics/bti553>.
5. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
  6. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25: 955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
  7. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.