

Neotropical Biodiversity



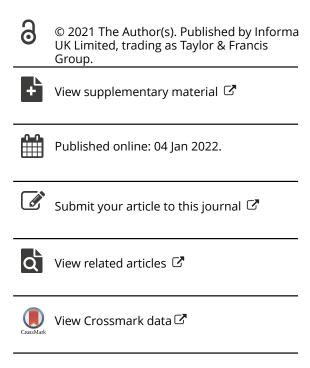
ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/tneo20

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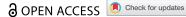
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To cite this article: G. Morera, S. Lupo, S. Alaniz & G. Robledo (2021) Diversity of the *Ganoderma* species in Uruguay, Neotropical Biodiversity, 7:1, 570-585, DOI: 10.1080/23766808.2021.1986329

To link to this article: https://doi.org/10.1080/23766808.2021.1986329









Diversity of the Ganoderma species in Uruguay

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ABSTRACT

Ganoderma is a cosmopolitan genus that includes a great diversity of species. Many of them have been historically described based only on morphological characteristics; however, due to their morphological plasticity, there is no complete understanding about their relationship and taxonomic status. Commonly applied names, particularly in the southern Neotropics, come from species of North Hemisphere distribution (e.g. G. lucidum, G. resinaceum and G. applanatum). The objective of the present work was to perform a survey of Ganoderma species thriving in Uruguay. We aimed to identify and characterize them through molecular, morphological and ecological analysis. The results confirm the presence of four reddish laccate species first registered for Uruguay (G. dorsale, G. platense, G. martinicense and G. mexicanum), and one non-laccate species (G. australe s.l.) composed of two clades. The species are morphologically differentiated mainly by its stipe, pilear surface, context, pores, basidiospores and cutis cells. Regarding the ecological data, the species present differences in substrate preferences. In addition, a taxonomic discussion regarding phylogenetic relationships and taxonomic status of Uruguayan Ganoderma species is presented.

ARTICLE HISTORY

Received 21 October 2020 Accepted 16 September 2021

KEYWORDS

ITS; mycogeography; phylogeny; systematics; South American polypores

Introduction

Ganoderma (P. Karst.) harbors at least 220 species, being the most diverse genus of Ganodermataceae [1–4]. However, due to their high diversity and phenotypic plasticity, the phylogenetic relationship and taxonomic status of many species remain unclear until now [5,6]. In the last 20 years, the use of molecular tools, mainly through the amplification and sequencing of Internal Transcribed Spacer (ITS), has been incorporated into systematic studies and into circumscription of Ganoderma species around the world. In this sense, through phylogenies based on molecular characters, the tendency in recent years has been to reinterpret the variations in characters of morphologically defined species, reinterpret the ecological relationships (relationship with hosts), determine their distribution and arrive at an understanding of the biogeographic processes that shape it [5,7–11].

Ganoderma species are morphologically characterized by the formation of sessile to stipitate basidiomata, with a glossy reddish laccate to opaque non-laccate cover, ellipsoid to ovoid double-walled basidiospores with truncated apex and endosporium with columnar ornamentations [12]. This cosmopolitan genus is comprised of parasitic and saprophytic species that decay the wood of plants from temperate and tropical areas around the world [6,13,14]. These fungi are described as white rot decayers that play a critical role in the dynamics of wood decomposition in tropical forests [15]. Moreover, they are the main cause of tree deterioration in public ornamental and commercial plantations [10,15-18].

In the last decades, some researchers have tried to elucidate the diversity of Ganoderma genus in the Neotropics, particularly Bazzalo and Wright [19], Gilbertson and Ryvarden [20], Gottlieb and Wright [21,22], Gottlieb et al. [7], Ryvarden [12,23], Torres-Torres and Dávalos [24], Torres-Torres et al. [25,26], and more recently Cabarroi-Hernández et al. [6] and Loyd et al. [11] for the northern limit of Neotropical distribution. Uruguay in particular harbors a great biodiversity due to its transitional condition and ecoregions diversity [27]. Some Ganoderma species have been historically recorded in Uruguay (Table 1), while Gazzano [28,29] and Martinez [30] have made more recent contributions to the diversity of the genus in Uruguay.

Some Uruguayan cited species as G. applanatum (Pers.) Pat. (= G. lipsiense (Batsch) G.F. Atk.), G. sessile Murrill, G. lucidum P. Karst. and G. resinaceum Boud were distributed out of the southern Neotropics [14,35,36], while others as G. lorenzianum (Kalchbr.)

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MS. Guillermo Morera. Main author of the work, he collected the specimens, analyzed the data and wrote the manuscript.Dra. Sandra Lupo Rizzo. Participated in field campaigns, molecular analysis and reviewed the manuscript. Dra. Sandra Alaniz Ferro. Participated in the revision and correction of the manuscript.Dr. Gerardo Robledo. Contributed in morphological and phylogenetic analyses, and reviewed the manuscript.

Supplemental data for this article can be accessed here.

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Pat., G. nitens (Fr.) Pat. lack of phylogenetic studies and their taxonomic status is unclear. Until now, there is no complete understanding on the systematic of the Ganoderma genus in Uruguay and its species diversity remains unknown.

The objective of the present work was to perform an updated survey of the Ganoderma species thriving in Uruguay and characterize them through molecular, morphological and ecological analysis. We also aimed to discuss the taxonomic status of the species of Uruguay and contrast with previous reports. We hypothesized that the Ganoderma specimens of Uruguay correspond to native species that have not been previously reported for the country. Those species are different from those registered to date (with names of species described from the Northern Hemisphere).

Materials and methods

Fungal specimens, basidiomata description and host characterization

Fresh Ganoderma basidiomata were collected from indigenous and urban ecosystems of Uruguay, during 2017 and 2018 field expeditions. Geographic location, host species and substrate condition (living tree, dead trunk, roots and stump) data were taken at the collection site following Urcelay and Robledo [37]. For fresh basidiomata, small pieces were aseptically taken from the context, placed into 2% malt extract agar (MEA) and incubated in darkness at 25°C. Pure cultures and basidiomata were deposited in MVHC. In addition, specimens from national herbaria (MVHC and MVM) were examined. Herbarium acronyms follow Thiers [38] (continuously updated, http://sweetgum.nybg.org/).

For morphological analyses and basidiomata identification, macroscopic and microscopic observations were made on basidiomata following the terminology and methodology according to authors [6,7,19,22-26]. Macroscopic features of basidiomata were analyzed and measured, particularly: pileus dimension, colour, texture, shape and appearance of surface, margin and stipe. The colour, presence of melanoid deposits and texture of context were also inspected and pores were described and measured (pores/mm). Microscopic features (basidiospores, chlamydospores in context, generative and somatic hyphae and cuticular cells) were analyzed with an optical microscope by mounting small sections of basidiomata with 5% KOH or Melzer's Reagent (to test for dextrinoid or amyloid reaction). In particular, for the analysis of the hyphal system, sections of basidiomata were treated for 24-48 h in 3% NaOH at 50-60°C [39]. Thirty basidiospores were measured from each specimen (length and width) and values were expressed as rank of mean values. Width was measured in the widest part of the spore and the length considered from the base to the truncated apex of the basidiospore. Then, Q ratio was calculated as the relation: length/width. Measures of cuticular cells were made from the middle part of the basidiomes.

Host relationships of each Ganoderma species in Uruguay were characterized by host range and native/exotic status. The preference of substrate condition was analyzed through the relative frequency of each Ganoderma species in each substrate condition (LT = stem of living tree, DT = dead trunks, S = stumps and R = soil, arising from roots of living or dead trees) following Urcelay and Robledo [37]. Then, substrate preference was determined by transforming this relative frequency into a percentage.

DNA extraction, PCR and sequencing

DNA extractions from pure cultures were performed, using the CTAB protocol of Doyle and Doyle [40] with modifications [41]. The PCR reaction of the ITS region (including ITS1, 5.8s and ITS2) was performed. The following PCR primers were alternatively used: ITS1/ITS4 [42], ITS1-F/ITS4-B [43], and ITS4/ITS5 [42]. The PCR mixture was prepared in a 25 µl final volume, with 2 µl of genomic DNA solution (10 ng), 16 µl of mQ water, 0.25 µl of Tag polymerase (1 U), 0.5 µl of each primer (10 mM), 0.7 µl of 50 mM MgCl2, 2.5 µl of dntps (2.5 mM) and 2.5 µl (10X) of buffer. PCR reactions were performed in a MultiGene Optimax thermocycler (Labnet International Inc) with the cycling conditions as follows: 3 min at 94°C, followed by 35 cycles, each

Table 1. Ganoderma species recorded in Uruguay.

Taxon	Author
G. australe (Fr.) Pat.	Felippone [6667]
G. applanatum (Pers.) Pat. (= G. lipsiense (Batsch) G.F. Atk.)	Felippone [66], Herter [31], Gazzano [28,29,32] Martínez [30]
G. fornicatum (Fr.) Pat.	Felippone [66]
G. lorenzianum (Kalchbr.) Pat.	Patouillard [65], Felippone [66]
G. lucidum (Curtis) P. Karst.	Spegazzini [33], Felippone [66], Wright & Blumenfeld [34], Gazzano [28,29,32], Martínez [30]
G. nitens (Fr.) Pat.	Patouillard [65], Felippone [66]
G. resinaceum Boud.	Felippone [66], Gazzano [32], Martínez [30]
G. sessile Murrill	Gazzano [29]

Table 2. Ganoderma species, specimens, location, Gen Bank accession numbers for ITS sequences and reference source.

Species/Voucher/Culture reference	Locality	ITS GenBank accession number	References
Ganoderma adspersum CBS351.74	Belgium	X78742/X78763	Moncalvo et al. [13]
GaTO00	Italy	AM906057	Guglielmo et al. [44]
Sanoderma annulare	,		•
CTC 16803	Brazil	JQ520160	GenBank
anoderma applanatum		10500444	
ATCC44053	Japan	JQ520161	GenBank
anoderma australe s.l.	Harranov	MN101560	This work
NVHC 5620 NVHC 5564	Uruguay Uruguay	MN191569 MN191568	This work
NVHC 5564 NVHC 5582	Uruguay	MN191570	This work
NVHC 5659	Uruguay	MN191554	This work
NVHC 5660	Uruguay	MN191553	This work
NVHC 5668	Uruguay	MN191555	This work
NVHC 5601	Uruguay	MN191556	This work
IVHC 5605	Uruguay	MN191557	This work
IVHC 5661	Uruguay	MN191558	This work
1VHC 5646	Uruguay	MN191552	This work
IVHC 5640	Uruguay 	MN191551	This work
IVHC 5641	Uruguay 	MN191550	This work
IVHC 5680	Uruguay	MN191547	This work
IVHC 5647 IVHC 5587	Uruguay Uruguay	MN191549 MN191548	This work This work
IVHC 5568	Uruguay Uruguay	MN191548 MN191559	This work
IVHC 5566 IVHC 5645	Uruguay	MN191560	This work
IVHC 5697	Uruguay	MN191561	This work
IVHC 5717	Uruguay	MN191572	This work
IVHC 5711	Uruguay	MN191562	This work
IVHC 5722.2	Uruguay	MN191563	This work
IVHC 5724	Uruguay	MN191565	This work
IVHC 5725	Uruguay	MN191564	This work
IVHC 5746	Uruguay	MN191573	This work
IVHC 5688	Uruguay	MN191566	This work
IVHC 5708	Uruguay	MN191567	This work
IVHC 5579	Uruguay 	MN191546	This work
IVHC 5732	Uruguay	MN191545	This work
I C 7177 705	Uruguay Taiwan	MN191571 X78750.1	This work Moncalvo and Buchanan [5]
anoderma concinnum	Talwall	X/8/30.1	Moricalvo and Buchanan [3]
obledo 3192 (FCOS)	Bolivia	MN077522	Costa-Rezende et al. [53]
obledo 3235 (FCOS)	Bolivia	MN077523	Costa-Rezende et al. [53]
anoderma dorsale	Donvia	1111077323	costa nezeride et di. [55]
NVHC 5588	Uruguay	MN191582	This work
IVHC 5701	Uruguay	MN191581	This work
IVHC 5653	Uruguay	MN191578	This work
IVHC 5648	Uruguay	MN191579	This work
IVHC 5654	Uruguay	MN191580	This work
anoderma ecuadoriense		W1400504	6
SL799	Ecuador	KU128524	Crous et al. [45]
MC126	Ecuador	KU128525	Crous et al. [45]
oly-2-4 anoderma gibbosum	Ecuador	KU128526	Crous et al. [45]
anoaerma gibbosum SD34	China	EU273513	GenBank
anoderma lipsiense	Cinia	L02/3313	Gelibalik
AFC2424	Argentina	AF169977/8	Gottlieb et al. [7]
anoderma lobatum	5		
AFC2411	Argentina	AF169989/90	Gottlieb et al. [7]
BS222.48	USA	X78740/X78761	Moncalvo et al. [13]
anoderma lucidum			
AFC2419	Argentina	AF170007/AF170008	Gottlieb et al. [7]
AFC2493	Argentina	AF170009/A1470010	Gottlieb et al. [7]
MNUT1	USA	MG654070	Loyd et al. [11]
175217	England	KJ143911	Zhou et al. [46]
YV 33217 T	Norway	Z37096/ Z37073	Moncalvo et al. [13]
BS 270.81	France	Z37049/Z37099	Moncalvo et al. [13]
ai 2272	Sweden	JQ781851	Cao et al. [14]
ai11593 CBC 37043	Finland	JQ781852	Cao et al. [14]
CRC 37043 anoderma martinicense	China	EU021460	Wang et al. [9]
anoaerma martinicense IVHC 5635	Uruguay	MN191574	This work
1VHC 5583	Uruguay	MN191574 MN191575	This work
IVHC 55637	Uruguay	MN191373 MN191577	This work
1VHC 5684	Uruguay	MN191577 MN191576	This work
46TX	USA	MG654185	Loyd et al. [11]
Mart08_55 T	Martinique	KF963256	GenBank
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anoderma mexicanum			

Table 2. (Continued).

Species/Voucher/Culture reference	Locality	ITS GenBank accession number	References	
(AL D.Jarvio 143	Mexico	MK531823	Cabarroi-Hernández et al. [6]	
AFC2580 anoderma multipileum	Brazil	AH008108	Gottlieb et al. [7]	
oanoderma mutupiieum OAI 9447	China	KJ143914	Zhou et al. [46]	
Ganoderma multiplicatum				
SPC9	Brazil	KU569553	Bolaños et al. [60]	
CC8	Brazil	KU569515	Bolaños et al. [60]	
JRM 83346 Ganoderma oerstedii	Brazil	JX310823	Bolaños et al. [60]	
ATCC 52409	Argentina	Z37058/Z37083	Moncalvo et al. [13]	
ATCC 52410	Argentina	X78739/X78760	Moncalvo et al. [13]	
ATCC 52411	Argentina	Z37059/Z37084	Moncalvo et al. [13]	
Ganoderma parvulum		11/24224		
JRM 83344	Brazil Brazil	JX310819	Correia de Lima Júnior et al. [57	
JRM 2948 Ganoderma platense	Drazii	JX310821	Correia de Lima Júnior et al. [57	
MVHC 5586	Uruguay	MN191585	This work	
MVHC 5565	Uruguay	MN191584	This work	
MVHC 5721	Uruguay	MN191591	This work	
MVHC 5732.2	Uruguay	MN191592	This work	
MVHC 5686	Uruguay	MN191593	This work	
MVHC 5692 MVHC 5687.2	Uruguay	MN191596 MN191597	This work This work	
WVHC 5687.2 WVHC 5687	Uruguay Uruguay	MN191597 MN191594	This work	
WVHC 5687 WVHC 5690	Uruguay	MN191595	This work	
NC 5332	Uruguay	MN191587	This work	
NC 7187	Uruguay	MN191588	This work	
NC 5104	Uruguay	MN191589	This work	
MVHC 5694	Uruguay	MN191590	This work	
MVHC 5691	Uruguay	MN191586	This work	
BAFC384 Ganoderma polychromum	Argentina	AH008109	Gottlieb et al. [7]	
330OR	USA	MG654196	Loyd et al. [11]	
3J280CA	USA	MG910492	Loyd et al. [11]	
Ganoderma resinaceum				
DP2	Italy	AM906060	Guglielmo et al. [44]	
CIRM BRFM 753	France	FJ805250	GenBank	
HMAS86599	England	AY884177	Wang et al. [9]	
BR 4150	France	KJ143915	Zhou et al. [46]	
GR-101 CBS 22036	India The Netherlands	GU451246 JQ520201	Mohanty et al. [63] Park et al. [60]	
BCRC 36147	The Netherlands	KJ143916	Zhou et al. [46]	
Ganoderma sessile	The Hellerianas	.0.1.02.10	2.100 00 01 01	
111TX	USA	MG654306	Loyd et al. [11]	
103SC	USA	MG654304	Loyd et al. [11]	
113FL	USA	MG654307	Loyd et al. [11]	
228DC	USA	MG654319	Loyd et al. [11]	
117TX BAFC2373	USA Argentina	MG654309 AH008111	Loyd et al. [11] Gottlieb et al. [7]	
Ganoderma sp.	Aigentina	AHOOTTI	dottileb et al. [7]	
MUCL27886	India	AF255190	Moncalvo and Buchanan [5]	
_XT.8	Vietnam	AF255188	Moncalvo and Buchanan [5]	
PKB96/330	Japan	AF255105	Moncalvo and Buchanan [5]	
ΓAI-05	Taiwan	AF255193/4	Moncalvo and Buchanan [5]	
CP331	Papua New Guinea	AF255125	Moncalvo and Buchanan [5]	
M97/31	USA China	AF255098	Moncalvo and Buchanan [5]	
M98/132 M98/2	China South Africa	AF255115 AF255149	Moncalvo and Buchanan [5] Moncalvo and Buchanan [5]	
ME-GAN-24	USA	AF255149 AF255131/2	Moncalvo and Buchanan [5]	
RV-PR10	Puerto Rico	AF255133	Moncalvo et al. [13]	
MCR.132	Costa Rica	AF255138	Moncalvo and Buchanan [5]	
MUCL40406	Ecuador	AF255139	Moncalvo and Buchanan [5]	
MUCL40324	French Guiana	AF255141	Moncalvo and Buchanan [5]	
NIAST824	South Korea	AF255114	Moncalvo and Buchanan [5]	
HMAS60686	China	AF255191/2	Moncalvo and Buchanan [5]	
BAFC2531 BAFC2449	Chile Argentina	AF255176 ΔF255187	Moncalvo and Buchanan [5] Moncalvo and Buchanan [5]	
SAFC2449 Ganoderma subamboinense var. laevisporum	Argentina	AF255187	INIONICATAO ANA DACUANAN [2]	
ATCC 52419	Argentina	X78736	Gottlieb et al. [7]	
ATCC 52419 ATCC 52420	Argentina	JQ520205	Park et al. [60]	
	USA	MG654373	Loyd et al. [11]	
JMNFL 100				
JMNFL 100 Ganoderma tornatum				
Ganoderma tornatum BAFC2764	Argentina	AF169993/4	Gottlieb et al. [7]	
Ganoderma tornatum BAFC2764 BAFC2582	Brazil	AF169985/6	Gottlieb et al. [7]	
Ganoderma tornatum BAFC2764				

Table 2. (Continued).

Species/Voucher/Culture reference	Locality	ITS GenBank accession number	References
PLM684	USA	MG654369	Loyd et al. [11]
UMNFL160	USA	MG654364	Loyd et al. [11]
Ganoderma zonatum			·
FL-02	USA	KJ143921	Zhou et al. [46]
179NC	USA	MG654417	Loyd et al. [11]
UMNFL105	USA	MG654408	Loyd et al. [11]
UMNSC4	USA	MG654415	Loyd et al. [11]
123FL	USA	MG654416	Loyd et al. [11]
BAFC2374	Argentina	AH008110	Gottlieb and Wright [22]
Cristataspora coffeata	-		3
FLOR 50933	Brazil	KU315204	Costa-Rezende et al. [53]
Foraminispora rugosa			
HUEFS_DHCR560	Brazil	MF409963	Costa-Rezende et al. [53]
FLOR 52191	Brazil	KU315200	Costa-Rezende et al. [4]

consisting of 60 s at 94°C, 45 s at 50°C, 60 s at 72°C, and a final extension step at 72°C for 5 min. PCR products were verified by electrophoresis in 1.0% agarose gels in TBE buffer, stained with EZ vision®One (Amresco®) and visualized under UV light transillumination. GeneRuler DNA Ladder Mix marker (Thermo) was used as molecular size marker. PCR products were purified and sequenced by Macrogen (Seoul, Korea). Sequences were submitted to GenBank (Table 2).

Phylogenetic analyses

The sequences obtained were manually edited (visual inspection of sequences and chromatograms, resolution of conflicts and pair the extremes) with Bioedit V.7.0.5.3 [47] and incorporated into alignments with sequences of specimens from other parts of the world obtained from GenBank (Table 2). Multiple alignment was made using ProbCons 1.12 from the CIPRES Science Gateway [48]. Subsequently, the best evolutionary model for each region (ITS1, 5.8S and ITS2) was estimated using the Corrected Akaike Informational Criteria (AICc), implemented by the jModelTest2 v.1.6 software [49]. The phylogenetic analysis was conducted in two independent ways: Bayesian Inference (BI) and Maximum Likelihood (ML), performed with MrBayes 3.2.7 [50] and RAxML 8.2.12 [51], respectively, in CIPRES Science Gateway [52]. Cristataspora coffeata (Murrill) Robledo, Rezende and de Madrignac Bonzi (FLOR 50933) and Foraminispora rugosa (Berk.) Costa-Rezende, Drechsler-Santos and Robledo (FLOR 52191 and HUEFS DHCR560) were used as outgroup [4,53]. For the BI, two independent runs were performed, starting with random trees, with four independent and simultaneous chains, 10,000,000 MCMC generations, and maintaining 1 tree every 1000 generations. Burn in discarded values was indicated as 0.25. The estimated models for each partition were incorporated, as indicated below (see Results). The average standard deviation of split frequencies was limited to

below 0.01. Convergence of the Markov chains to a stationary distribution was visually inspected using the Tracer v.1. 7. 1 program [54]. A GTRGAMMA nucleotide model and 1000 bootstrap iterations were indicated for the ML analysis. The rest of the parameter values were set by default. Since the topologies of trees obtained in each analysis were convergent, only the consensus BI tree is shown with values of Bayesian posterior probability (BPP) and ML bootstrap (ML) separated by cross bars (BPP/BS). A clade was considered strongly supported if it showed a 0.95 BPP and/or 80% BS [55].

Results

Fungal specimens, basidiomata description and host characterization

A total of 163 Ganoderma specimens from collections generated in this work (n = 90), MVHC (n = 57) and MVM (n = 16) were morphologically and ecologically analyzed (Tables 3 and 4). Ganoderma australe was the most commonly collected species, with 101 specimens. The 62 remaining specimens belong to four reddish laccate species: G. mexicanum (n = 3), G. martinicense (n = 10), G. platense (n = 22) and G. dorsale (n = 27).

Morphologically assigned species are presented in Figures 1 and 2. Ganoderma species of Uruguay were primarily differentiated by their pilear surface into two groups: reddish laccate and non-laccate (Table 3 and Figure 1).

The first group is composed of two species with great and robust basidiomes (G. martinicense and G. platense) and two species with smaller basidiomes (G. dorsale and G. mexicanum).

Ganoderma martinicense is characterized by a large, commonly substipitate basidiomata with a tuberculous concentric zonated pilear surface and conspicuous melanoid deposits in the homogeneous context. Microscopically, it is characterized by the presence of a distinct smooth basidiospores ornamentation, formed by free pillars and cutis cells with weak

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				Contex	ext		Pores		Basi	Basidiospores	Chlamydospores	oores	J	Cutis cells	
Taxon	Stipe	Surface	Margin	Color	Resinous/ melanoid deposits	Color	Shape	Pores/ mm	Size (µm)	Ornamentation) Š	Size (µm)	Shape	Melzer reaction Size (µm)	Size (µm)
ale M1	Rarely present, mostly sessile	Tubercular, rugose, concentric zonate, brownish to mate	Brownish, flat to slightly lobulated	Homogeneous, chocolate	Always present	Whitish to brownish	Circular to irregular		ļ'	Thick, free to sub free pillars		10-13 1 × 9- 11	Branches of terminal skeletal hyphae embedded in melanoid substance	N	reaction 70 × × × × 3 - 5 5
6. australe M2	Rarely present, mostly sessile	Concentric zonate, brownish to greyish	Brownish, flat to slightly lobulated	Homogeneous, chocolate	Always present	Whitish to brownish	Circular to irregular	9	6.5- 10.1 × 4.2- 6.1	Thick, free to sub free pillars	walled. In context, very 11 × 9 rare, slightly ellipsoid, double- walled.	6 ×	Branches of terminal skeletal hyphae embedded in melanoid	Ž <u>.</u> ⊑	ull 50- reaction 70 X 4- 4-
G. dorsale	Always present, lateral, vertical	Rough, semi-concentric zonation, wine purplish to reddish	Reddish, flat to acute	Not fully homogeneous, first light brown becoming dark brown near the	Always present	Whitish to brownish	Circular to irregular	3-5 8		Distinct, thick pillars, sometimes with anatomosed	In context, rare, 5–13 × late or 4– intercalary, 7.5 double- walled.	,5-13 × 4- 7.5	Substance Cylindrical to clavate, usually with apical protuberances (up to 2)	Substance Indirical to Strongly Clavate, usually amyloid protuberances (up to 2)	lly 20– //loid 39 X 3– 7.5
G. martinicense Substipitate to sessile	Substipitate to sessile	Tuberculous, rugose, concentric colored, first purplish, then reddish and yellowish toward the margin	Whitish, Iobulated	Not fully Not fully homogeneous, alternate dark and light brown	Always present	Whitish to yellowish	irregular to	9	10- 12.9 × 5.7- 7.7	appearance Distinct, free pillars	In culture, terminal or intercalary, double walled, variably smooth, sometimes with appendages, yellowish. In context, terminal or intercalary.	12.5 × 2.5 × 8- 9-12 5-8	Spheroid pedunculated to clavate, occasionally with apical or basal branches	Weakly Steed amyloid amyloid steed amyloid s	y 30- Noid 60 X 5- 5- 15

Table 3. (Continued).

				Context	ext		Pores		Bas	Basidiospores	Chlamydospores	ores	Ū	Cutis cells
					Resinous/									
					melanoid			Pores/	Size			Size		Melzer
Taxon	Stipe	Surface	Margin	Color	deposits	Color	Shape	mm	(mm)	Ornamentation Description	Description	(mm)	Shape	reaction Size (μm)
6. mexicanum	Present,		Whitish,	Mostly light	Present,	Whitish to Circular to 4–6	Circular to		× 6-2	$7-9 \times$ Inconspicuous,	ln context,	9-15.5	, 9–15.5 Cylindrical to	Strongly 30 ×
	lateral,	concentric zonation,	acute	colored. Slightly	inconspicuous brownish	brownish	irregular		2–6	mostly free	frequent,	× 7-	slightly	amyloid 4–
	horizontal	shiny, bordered to		heterogeneous,						pillars	final or	11	clavate,	9
		reddish		being darker							intercalary,		generally	
				near the tubes							globose to		without	
											slightly		protuberances	es
											ellipsoid,			
											-əlqnop			
											walled.			
6. platense	Always	Rough to variably striated, Whitish,		Not fully		>		3-4	7.5–9	Inconspicuous,		8.3-	Cylindrical to	Strongly 30–
	sessile	semi-orbicular	lobulated	homogeneous,	inconspicuous	brownish			× 4-	\times 4– free pillars	infrequent,		slightly	amyloid 50
		zonation, shiny, wine	to acute	first light brown					6.2		terminal or		clavate,	×
		purplish on board		becoming dark							intercalary,	5.5-	sometimes	4-
				brown near the							brownish.		with knots	or 9
				tubes									apical	
													constrictions	S



Table 4. Number of specimens, distribution and host characterization (species, native/exotic status and substrate preferences) of Ganoderma species collected and studied in this work.

				Host characterization	
Taxon	Total specimens	Distribution (Departments)	Native	Exotic	Substrate preferences
G. australe M1	13	Lavalleja, Montevideo, Maldonado, Paysandú, Rivera, Rocha, Florida, Colonia, Flores	Scutia buxifolia, Myrsine laetevirens, Alophyllus edullis, Lithraea sp.	Acacia melanoxylon, Gleditsia triachanthos, Tilia sp., Eucalyptus sp., Quercus sp., Salix sp., Grevillea sp.	LT (45%), DT (33%) and S (22%)
G. australe M2	88	Canelones, Lavalleja, Maldonado, Montevideo, Rivera, Rocha, Tacuarembó, Treinta y Tres, Florida, Cerro Largo, Río Negro, Artigas, Colonia, San José	Alophillus edulis, Lithraea sp., Myrcianthes sp., Scutia buxifolia, Myrsine laetevirens	Acacia longifolia, Acacia melanoxylon, Acacia sp., Eucalyptus globulus, Robinia pseudoacacia, Prunus persica, Pinus pseudostrobus, Quercus suber, Salix sp.	DT (46%), LT (39%), S (14%) and R (1%)
G. dorsale	27	Montevideo, Rocha, Treinta y Tres, Canelones, Rivera	Alophyllus edulis, Lithraea sp., Ruprechtia salicifolia, Scutia buxifolia, Celtis sp., Sebastiana sp.	Quercus suber	R (58%), LT (37%) and S (5%)
G. martinicense	10	Canelones, Montevideo, Paysandú, Rocha, Artigas	Tipuana tipu	Acacia sp.	R (87%) and S (13%)
G. mexicanum	3	Rocha, Paysandú	Unidentified wood	-	DT (100%)
G. platense	22	Canelones, Montevideo, Maldonado	-	Acacia dealbata, Fraxinus sp., Platanus acerifolia, Salix sp., Quercus sp., Araucaria sp.	LT (85%), DT (8%) S (7%)

LT = stem of living tree, DT = dead trunks, S = stumps, R = soil arising from roots of living or dead trees.

reaction and spheroid-pedunculated shape. On the other hand, G. platense basidiomata are smaller, semiorbicular zonate, homogeneous colored and always sessile. Its context is also different, presenting extremely inconspicuous melanoid lines and microscopically characterized by particular cutis elements with apical constrictions (Figure 2).

Ganoderma mexicanum produces stylized, flabelliform, laterally and horizontally stipitate basidiomata characterized by an almost light-colored context and basidiospores with fine ornamentation, whereas G. dorsale produces stylized, spatuliform, shellshaped and almost laterally and vertically stipitate basidiomata with no homogeneous distinctive context. Its basidiospores present notorious rough ornamentation and almost cylindrical cutis cells with a strong amyloid reaction (Figure 2).

Specimens with non-laccate surfaces are morphologically very similar, discernible however by their pilear surface, which is distinctly tubercular in G. australe M1 and distinctly zonate in Ganoderma australe M2.

Herbaria specimens showed a morphology consistent with the species recorded in this study. In that sense, G. lucidum is a name previously used for specimens corresponding to the G. dorsale, G. martinicense, G. platense and G. mexicanum species. The name G. resinaceum was previously used to name specimens corresponding to G. platense and G. martinicense. The names G. lipsiense, G. applanatum and G. marmoratum were used to refer to specimens of G. australe in the broad sense (from now on sensu lato or s.l.). A morphological key for Ganoderma species found in Uruguay is presented below.

Host relationships of each Ganoderma species in Uruguay and presence in distinct departments are presented in Table 4. Ganoderma martinicense and G. australe were found alternatively in several departments growing on native or exotic trees. On the other hand, G. platense was only recorded in the southeast (Montevideo, Canelones and Maldonado departments), growing preferentially on exotic trees, G. mexicanum was only found on dead stems of native forest, and G. dorsale, almost exclusively on native trees. Regarding their substrate preferences, G. mexicanum and G. australe M2 were found preferentially on dead stem wood (100% and 46%, respectively); G. martinicense and G. dorsale, preferentially on roots (87% and 58%, respectively); and G. australe M1 and G. platense, preferentially on live stems (45% and 85%, respectively).

Phylogenetic analyses

A total of 53 new sequences were generated from Uruguayan specimens. The dataset alignment resulted in 140 DNA sequences comprising 639 bp. The best evolutionary models for each partition were as follows: K80 + G (ITS1), JC (5.8 S), K80 + G (ITS 2). The partition scheme was K80 + G (ITS1) with -lnL = 1373.0169, and equal base frequencies as follows: A = 0.25, C = 0.25, G = 0.25, T = 0.25, JC (5.8 S) with -lnL = 300.8947 with equal base frequencies, and K80 + G (ITS 2) with 1498.7406 and equal base frequencies. The Bayesian Inference consensus tree is presented in Figure 3.

A total of 19 supported clades were recovered through a phylogenetic analyses (Figure 3). Sequences corresponding to Uruguayan specimens

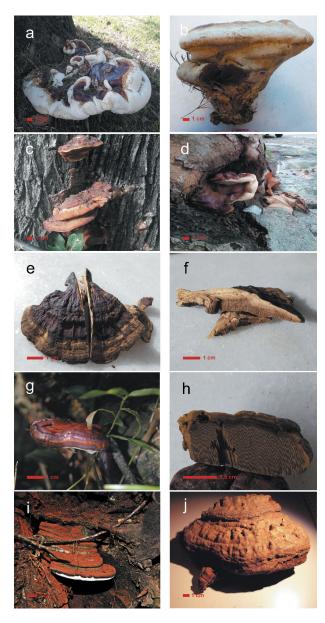


Figure 1. Basidiomata of Ganoderma species from Uruguay. G. martinicense: (a) (MVHC 5583) and (b) (MVHC 5635), G. platense: (c) (MVHC 5586) and (d) (MVHC 5687), G. mexicanum (MVHC 5652): (e) and (f), G. dorsale: (g) (MVHC 5648) and (h) (MVHC 5655), G. australe M2 (MVHC 5587): (i) G. australe M1 (MVHC 5582): (j).

were distributed in six terminal clades representing the following species (BPP/BS): Ganoderma platense Speg. (1/98), Ganoderma dorsale (Lloyd) Torrend (1/100), martinicense Welti & Courtec. (0.98/80),G. mexicanum Pat. (1/100) and Ganoderma australe s.l. composed of two terminal clades: one of them termed clade 1 (0.97/73), composed of specimens of G. australe M1 and the other one termed clade 2 (0.91/68) and composed of specimens of G. australe M2. The remaining clades represent next species: G. multipileum Hou. (0.96/97), G. multiplicatum (Mont.) Pat. (1/90), lucidum" sensu authors [22] (0.76/94),G. tuberculosum Murrill (1/100), G. concinnum Ryvarden (1/100), G. lucidum (1/100), G. resinaceum

(0.9/59), G. polychromum Murrill (1/96), G. sessile (0.99/82), G. zonatum Murrill (1/100), G. ecuadoriense W.A. Salazar, C.W. Barnes & Ordoñez (1/100) and two unnamed taxa of the G. australe/applanatum complex from the Neotropic and Asia (1/89) and the Northern Hemisphere (1/88).

Key to Ganoderma species from Uruguay

Reddish to wine-purplish pileus surface, glossy to opaque
1'. Greyish brown pileus surface, matte, never reddish <i>G. australe</i> s.l. 5
Usually robust basidiomata, mostly sessile to occasionally substipitate
2'. Stylized basidiomata, slender, laterally to eccentrically stipitate
3. Pores 4–6/mm, basidiospores with thick endosporic pillars <i>G. martinicense</i>
3'. Pores 3–4/mm, basidiospores with thin and tiny endosporic pillars
Vertically stipitate basidiomata, brownish context and basidiospores with prominent endosporic pillars
4'. Horizontally stipitate basidiomata, cream colored context and basidiospores with tiny endosporio pillar
5. Conspicuosly, tuberculous pileus surface, sometimes presenting concentric zones
5'. Concentrically zonated to slightly tubercular

Discussion and integrative taxonomy

The taxonomic status of Ganoderma species in Uruguay was evaluated through ITS-based phylogenetic analyses in combination with morphological and ecological data. The topology recovered in our phylogenetic analyses is congruent with previous works [4-6]. Three main clades were recovered. One of them is composed of Ganoderma species with reddish laccate basidiomata, stipe and pilear surface, including traditional G. lucidum and G. resinaceum complexes (0.99/67). The second one is composed of almost-sessile Ganoderma species with non-laccate pilear surface, including G. australe and G. applanatum complexes (1/76). The third one is a small subclade of 2 reddish including G. laccate species zonatum G. ecuadoriense (0.96/66). Studied Uruguayan

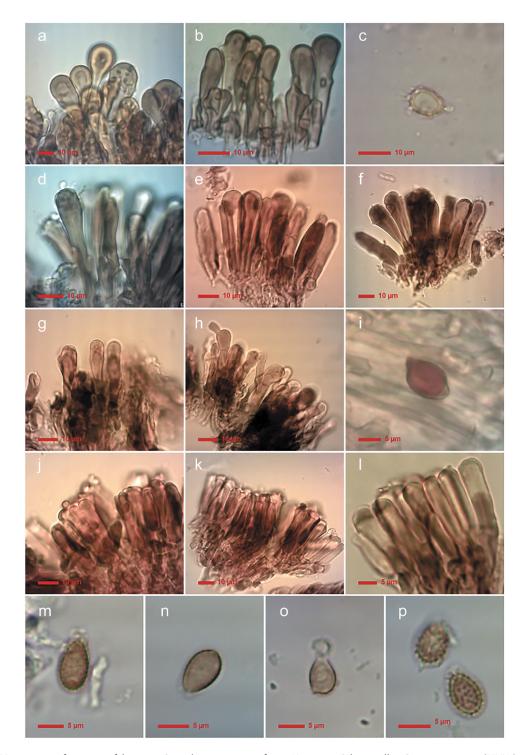


Figure 2. Microscopic features of laccate *Ganoderma* species from Uruguay. Pilear cells: *G. martinicense* (MVHC 5635): (a,b), *G. platense* (MVHC 5687): (d–f), *G. mexicanum* (MVHC 5652): (g–h), *G. dorsale* (MVHC 5655): (j–l). Chlamydospores from fresh *G. martinicense* (MVHC 5635) culture in MEA: (c) and dextrinoid characteristic chlamydospores from the context of *G. mexicanum* (MVHC 5652): (i). Basidiospores from *G. martinicense* (MVHC 5635): (m), *G. platense* (MVHC 5687): (n), *G. mexicanum* (MVHC 5652): (o) and *G. dorsale* (MVHC 5655): (p).

Ganoderma specimens are distributed in six clades representing species whose taxonomic resolutions are discussed below.

Ganoderma martinicense is characterized by developing short stipitate basidiomata of large dimensions (up to 30 cm upper view), with the pilear surface concentrically zonate, with melanoid incrustations in the context, pores 5–6/mm, basidiospores measuring 10– 12×5 –7 µm and non-amyloid pear-shaped to shortly

cylindrical cutis cells. The species is distributed from SW North America through the Caribbean and the Neotropical Atlantic Rain Forests, [11,56], reaching Uruguay and NE Argentina. Ten of the studied specimens fit very well with that morphological description. Moreover, they are grouped with *G. martinicense* specimens, including the type in a strongly supported clade (1/88). Other names were used to label sequences of specimens that are conspecific with sequences of this

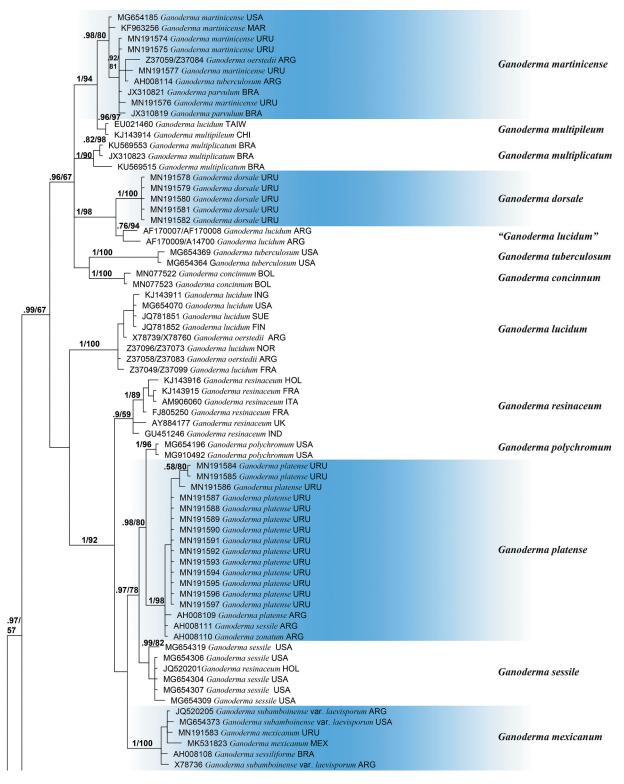


Figure 3. Consensus Bayesian Inference tree based on ITS sequence data used for positioning Ganoderma species found in Uruguay (highlighted in light blue). Values of Bayesian and Maximum likelihood are presented as PPB/BS. Cristataspora coffeata (FLOR 50933) and Foraminispora rugosa (FLOR 52191 and HUEFS DHCR560) were used as outgroup.

lineage (Figure 3), i.e. G. tuberculosum [7], G. oerstedii (Fr.) Murrill [13] and G. parvulum Murrill [57]. Ganoderma tuberculosum and G. parvulum form two different, distant and unrelated lineages [6,11]. Ganoderma oerstedii has been described as presenting a context lacking resinous lines, longer basidiospores with semi-rough columnar ornamentations: 12-15 \times 8–10 μm [12] and 9–14 \times 6–9 μm [19]. The pileipellis is formed by irregular, lobed and branched cells with up to seven short, wide protuberances [25]. Sequences of specimens from the type locality (San Juan de Puerto Rico) need to be included to determine

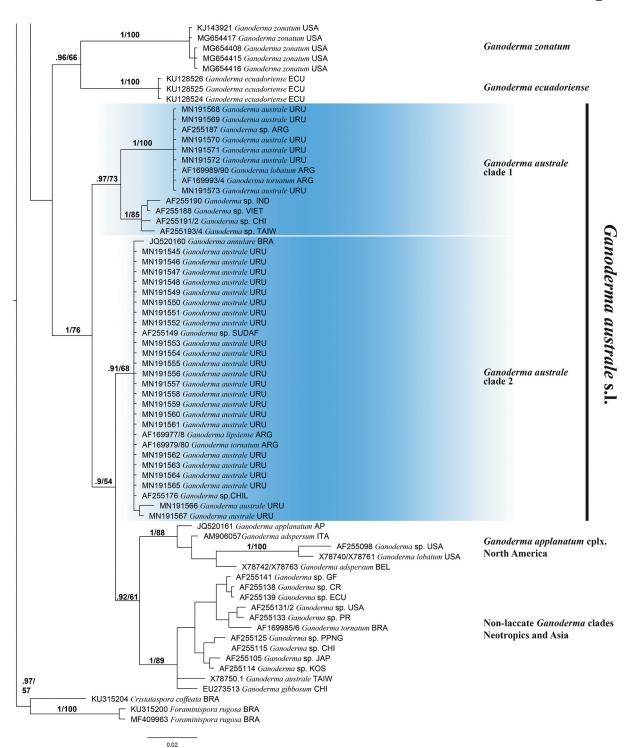


Figure 3. Continued.

the taxonomic status and the phylogenetic relationships of *G oerstedii*. The closest relative of *Ganoderma martinicense* is *G. multipileum* (1/94), an Asian species with small pores: 6–8/mm [9].

Five sequences of Uruguayan specimens are grouped together in a distinct, well-supported clade (1/100), unrelated to any available sequence. Specimens of this clade are characterized by a stylized basidiomata, laterally to centrally slender stipite, shiny reddish orange to violet colored radial and concentric zonate pilear surface, and melanoid bands in the context. Microscopically, the cutis is

composed of clavate cells (Figure 2) and basidiospores with thick, often anastomosed endosporic ornamentations. The macro-morphological features suggest some Neotropical taxa including *Ganoderma elegantum* Ryvarden [12], *G. concinnum* [23] and *G. dorsale* [58]. *Ganoderma elegantum* is characterized by pores 6–7/mm, branched cutis cells (with up to six branches) and basidiospores with thin endosporic ornamentations [12]. *Ganoderma concinnum*, recorded in Colombia and Bolivia, is characterized by the formation of basidiomata with slender stipes of up to 20 cm long and it forms a different

phylogenetic lineage [12,53]. Ganoderma dorsale (= G. lucidum var. dorsale in accordance with Torres-Torres et al. [25]) was described from Río Grande Do Sul in southern Brazil [59]. The macro and micro morphological characteristics of the specimens of this lineage agree with the description of the type specimen of G. dorsale [22,25] and fit well with the studied specimens: basidiomata shape and size, presence of melanoid lines in the context, basidiospores' shape and size, chemical reactions and shapes of pileipellis cells. Thus, considering morphology and distribution, Ganoderma dorsale is the most suitable name for the new clade of Uruguayan specimens. The closest relative is a phylogenetic clade named "G. lucidum" from NW Argentinean Yungas [7]. More sequences and morphological reassessments are necessary to evaluate the taxonomic status of the "G. lucidum" clade sensu Gottlieb et al. [7].

Ganoderma multiplicatum, also occurring in the Neotropics, is morphologically differentiated by its cutis cells with numerous protuberances [25,57,60,61]. The phylogenetic relationship between G. multiplicatum, G. martinicense, G. multipileum, G. dorsale, "G. lucidum", G. tuberculosum and G. concinnum remains unsolved.

Ganoderma platense is characterized by developing sessile basidiomata, usually imbricated, with a semiorbicular zonate pilear surface context, usually with inconspicuous and discontinuous melanoid deposits. thin dissepiments and large pores (2-4/mm), pileipellis formed by cylindrical to slightly claviform elements, with apical constrictions and basidiospores measuring 9-13 × 5-8 um, with thin, endosporic ornamentations [7,22,62]. The morphological characteristics of the studied specimens constituting this clade (Figures 1 and 2) agree with the description of G. platense and this name is hereinafter applied to the clade. Originally described from Buenos Aires (Argentina), it is currently known for growing on Platanus acerifolia trees of urban ecosystems of its type locality and on stumps of gallery forests of the Parana and Uruguay rivers [7,22,62]. Within the G. platense clade, two sequences correspond to specimens labelled as G. sessile and G. zonatum [7]; however, both species were phylogenetically circumscribed to distinct North American clades and are characterized by growing on hardwoods wood and monocots, respectively [11]. Ganoderma platense is grouped in the so called "resinaceum clade", together with G. resinaceum s.l. in Cabarroi-Hernández et al. [6], G. polychromum and G. sessile sensu Loyd et al. [11]. Ganoderma resinaceum encompasses North American, European and Asian populations, including more than one phylogenetic species. Nevertheless G. resinaceum sensu auctores from Eurasian descriptions present cutis cells with laterally diverticulate branches [22,25,63,64]. It contrasts with G. platense, which presents cylindrical to slightly clavate cells with apical constrictions. Ganoderma polychromum (Copel.) Murrill is distributed in North America, growing

on hardwoods and characterized by a context without melanoid deposits, smaller pores (4-5 mm) and larger basidiospores measuring $10.8-13.2 \times 6-7.5 \mu m$ [11].

Ganoderma mexicanum is known for its occurrence in Brazil, Colombia, Costa Rica, Cuba, French Guiana, Mexico, Nicaragua, South-eastern USA (Florida) [6], and now reported in Uruguay.

Morphologically, the studied specimen presents the diagnostic characters of light-colored context with variably abundant dextrinoid, smooth chlamydospores [6].

The sequences of non-laccate Ganoderma specimens collected in Uruguay were distributed in two clades: one of them with Southern Hemisphere specimens and the other one with specimens from South America and Asia [5]. Although the morphological analysis did not allow us to discriminate the specimens of each clade, the pilear surface generally appears tubercular and non-zonate in G. australe M1 and concentrically zonate G. australe M2. In addition, there seems to be ecological differences regarding host preferences: G. australe M1 specimens develop basidiomata on living hosts, whereas G. australe M2 on dead hosts. It was established that species of the G. australe/ applanatum complexes could have a recent origin (not earlier than 30 Ma), with a distribution pattern explained by a large-scale, episodic colonization model and subsequent distance isolation [5]. The recent origin, in addition to the remaining interfertility between specimens from both clades [5], may be the plausible explanation of the crypticity and lack of clear morphological differences for the specimens of the two clades of G. australe. Many names were proposed for sequence of specimens grouped in each clade: G. tornatum (Pers.) Bres., G. lobatum (Schwein.) G.F. Atk., G. annulare (Lloyd) Boedijn and G. lipsiense [7]. Ganoderma annulare and G. tornatum are largely considered synonyms of G. australe, and Ganoderma lipsiense was long considered a synonym of G. applanatum [21,25]. Ganoderma applanatum type locality is in the Northern Hemisphere and previous phylogenetic analyses suggested that it could be represented by a Northern Hemisphere clade [5.8]. Morphologically, G. applanatum lacks melanoid lines in the context [25], whereas all Uruguayan specimens present melanoid elements in the context. From the morphological, ecological and distribution data, M1 and M2 specimens should remain as G. australe s.l.

In this context, the previous records of Ganoderma in Uruguay (Table 1) should be questioned, and due to the absence of herbarium specimens, only speculations can be made in relation to published descriptions. Records of G. lorenzianum [65.66] have characteristics that resemble G. mexicanum due their morphological similarities regarding light colored context (picture in reference [67]), stipe and ovoid, smooth basidiospores (9–10 \times 6–7 μm). G. lorenzianum is an



earlier name than G. mexicanum but the description and picture offered in bibliography [67] and the low number of specimens of G. mexicanum analyzed in Uruguay is insufficient to assess the epitypification of these species. Focused studies on specimens corresponding to this species are urgently needed. In Ganoderma orbiforme (Fr.) Ryvarden (=G. fornicatum (Fr.) Pat.) was recorded by Felippone [66], but it currently forms a different phylogenetic clade [57] and no specimens or sequences corresponding to this species were recorded in the country. Ganoderma nitens was recorded by Patouillard [65] characterized by warty basidiospores, $10 \times 7 \mu m$, the type is currently lost [68], so this record could be attributed to G. dorsale, a single reddish laccate Ganoderma species of Uruguay with warty spores.

Conclusions

An integrated comprehensive approach was carried out using morphological, molecular and ecological evidence to assess the diversity of Ganoderma in Uruguay. A total of five species (forming six supported clades) were found through the analysis of 163 basidiomata and 53 sequences. Among those species, G. martinicense, G. mexicanum, G. platense and G. dorsale were confirmed and identified for the first time in Uruguay. Particularly, G. platense and G. dorsale were first recovered in the phylogenetic analyses. On the other hand, non-laccate specimens were distributed in two clades and so far considered as G. australe s.l.

Acknowledgments

The authors kindly acknowledge Anaclara Cabrera Varela for her technical support in improving figures editions. Curators of CORD, MVM and MVHC herbaria are acknowledged for the loan of collections for this study. Daniel Newman, (ORCID: 0000-0002-5400-3691) is kindly acknowledged for discussions, comments and proofreading the English version of the manuscript. The assistance of Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) and Universidad Nacional de Córdoba, both of which supported the facilities used in this project, is also acknowledged. Authorities that granted permits to collect in Uruguay are kindly acknowledged.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported by Agencia Nacional de Investigación e Innovación [ANII, POS_NAC_2016_1_130911], Pedeciba, Universidad de la República, Uruguay, IDEA WILD, FONCYT [PICT 0830 to G. Robledo], and Fundación Fungicosmos.

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