

Intrauterine Effects of Impaired Lipid Homeostasis in Pregnancy Diseases

R. Higa and A. Jawerbaum*

Laboratory of Reproduction and Metabolism, CEFYBO-CONICET, School of Medicine, University of Buenos Aires, Buenos Aires, Argentina

Abstract: Lipids are crucial structural and bioactive components that sustain embryo, fetal and placental development and growth. Intrauterine development can be disturbed by several diseases that impair maternal lipid homeostasis and lead to abnormal lipid concentrations in the fetal circulation. Deficiency in essential fatty acids can lead to congenital malformations and visual and cognitive problems in the newborn. Either deficient mother-to-fetus lipid transfer or abnormal maternal-fetal lipid metabolism can cause fetal growth restriction. On the other hand, excessive mother-to-fetus fatty acid transfer can induce fetal overgrowth and lipid overaccumulation in different fetal organs and tissues. The placenta plays a fundamental role in the transfer of lipid moieties to the fetal compartment and is affected by maternal diseases associated with impaired lipid homeostasis. Postnatal consequences may be evident in the neonatal period or later in life. Indeed, both defects and excess of different lipid species can lead to the intrauterine programming of metabolic and cardiovascular diseases in the offspring. This review summarizes the lipid impairments induced by different pathologies, including placental insufficiency, malnutrition, obesity and diabetes, and their consequent developmental defects.

Keywords: Pregnancy, lipids, intrauterine growth retardation, obesity, diabetes.

INTRODUCTION

During normal pregnancy, lipids are highly required in the intrauterine compartment and thus efficiently transported through the placenta from maternal circulation. Development implies the rapid generation of millions of new cells that will require lipids both as components and as bioactive molecules, crucial for proper developmental and growth patterns. As most lipid species can be synthesized by the mother and the fetus from other nutrients, caloric restriction can affect the formation of structural and bioactive lipids. Essential fatty acids (EFAs) are the n-3 and n-6 polyunsaturated fatty acids (PUFAs) that cannot be synthesized by either the mother or the fetus, and their deficiency has been largely known to be related to the induction of congenital malformations and visual and neurodevelopmental impairments in the newborn [1]. On the other hand, an excess of circulating free and esterified fatty acids during pregnancy in pathologies such as diabetes and obesity affects feto-placental development and leads to abnormal feto-placental growth through mechanisms that involve an excessive substrate transfer and oxidation, which is related to the induction of oxidative stress and a pro-inflammatory environment [2]. Increased oxidative stress induces loss of bioactive lipids through peroxidation. This includes the PUFAs that are ligands of the nuclear receptors PPARs, master regulators of metabolic and anti-inflammatory processes [3]. Importantly, pregnancy diseases related to both the excess and the deficiency of different lipid species in the intrauterine compartment can lead to the programming of diseases in the offspring [4-6]. This

review summarizes the current knowledge of the damaging effects of impaired lipid homeostasis in intrauterine development and its postnatal consequences.

LIPID IMPAIRMENTS AND THE INDUCTION OF CONGENITAL MALFORMATIONS

Congenital malformations arise during organogenesis and are mostly induced before the seventh week of human gestation [7]. Organogenesis is characterized by continuous and accelerated embryo growth and cell differentiation processes, and both embryo growth and development require lipids as structural components of cell membranes in formation, as oxidative fuels and also as signalling molecules [8]. The required lipids are both derived from the mother and synthesized by the embryo, which is capable of synthesizing both polar and neutral lipids from the early embryonic stages [9].

During the first stage of embryo development, lipids derived from the mother are absorbed by histotrophic mechanisms and the visceral yolk sac is involved in this process [10]. In a later stage, embryonic lipid uptake from maternal decidua or maternal blood requires a process of lipid hydrolysis and re-esterification to translocate across the yolk sac membrane. This involves a repackaging mechanism of both maternal-derived lipids and endogenously synthesized lipids in the yolk sac that lead to the formation of apoB-containing lipoproteins, which are then secreted from yolk sac endoderm cells to viteline circulation or transported by diffusion to the growing embryo [11, 12].

Animal studies have shown that microsomal triglyceride transfer protein is involved in the secretion of apoB-containing lipoproteins, and embryo lethality occurs in null mice for either apoB or microsomal triglyceride transfer protein genes [13, 14]. Differently, in humans, apoB or micro-

*Address correspondence to this author at the Laboratory of Reproduction and Metabolism, CEFYBO-CONICET, School of Medicine, University of Buenos Aires, Buenos Aires, Argentina; Tel: +5411 4508-3680 ext 119; Fax: +5411 4508-3680 ext 106; Email: a.jawerbaum@gmail.com

somal triglyceride transfer protein deficiency does not increase the incidence of early fetal loss, suggesting that in humans, these enzymes are not essential for early nutrient delivery. However, an impaired lipid absorption and peripheral delivery of vitamins A and E have been found in individuals with abetalipoproteinemia [15, 16]. Although no loss-of-function mutations have been reported for scavenger receptor class B member I (SR-BI) in humans, recent studies have shown induction of congenital malformations in SR-BI null mice, suggesting that this receptor is needed for uptake of cholesteryl esters and other lipids from high-density lipoproteins (HDL) during early organogenesis [17].

Smith-Lemli-Opitz Syndrome

Relevant developmental anomalies caused by a deficiency in lipids are the result of impaired function of key enzymes in the cholesterol synthesis pathway. Smith-Lemli-Opitz syndrome (SLO) is a metabolic disorder that leads to a wide spectrum of congenital abnormalities [18]. Individuals with a severe phenotype have multiple major congenital anomalies and may die perinatally, whereas those with a mild phenotype may have learning disorders and minor dysmorphic characteristics [19, 20]. SLO is caused by a deficiency in the activity of 7-dehydrocholesterol reductase. This enzyme catalyzes the reduction of the Δ^7 bond in sterols, reducing 7-dehydrocholesterol to yield cholesterol in the final step of cholesterol biosynthesis. Cholesterol is a major lipid component of cellular membranes and is an important structural component of lipid rafts, which play an essential role in signal transduction. Also, bile acids, steroid hormones, neuroactive steroids and oxysterols are synthesized from cholesterol. SLO impairments are caused at least in part by a deficiency in cholesterol and total sterols, by the toxic effects of the accumulation of 7-dehydrocholesterol or its derivatives, and by a combination of these factors [18].

Some of the malformations induced by SLO are similar to those seen in animals in which sonic hedgehog signalling is impaired [21], suggesting the involvement of this morphogen in SLO-induced malformations. It is important to note that in the pregnant woman, intrauterine identification of SLO syndrome may allow the initiation of maternal treatments with diets enriched in cholesterol that reach the embryo and the fetus and help preventing fetal dysmorphogenesis and cognitive impairments in the offspring later in life [20, 22].

Obesity

As lipid transport is highly facilitated to sustain embryo growth and development, the excess of lipid transfer affects the embryo and may cause congenital anomalies. Indeed, among the adverse fetal outcomes associated with pre-pregnancy obesity in women are neural tube defects (e.g. anencephaly, spina bifida, holoprosencephaly) and congenital heart defects (e.g. atrial septal defects, left and all right ventricular outflow tract obstruction defects, hypoplastic left heart syndrome, tetralogy of Fallot, aortic stenosis and pulmonary stenosis) [23, 24].

Obesity and pregnancy are independently associated with insulin resistance and pro-inflammatory changes, processes that may be enhanced when combined in obese gestations. In

obese pregnant women, there is an earlier shift from an anabolic to a catabolic state and a predominance of lipolysis instead of the increase in adipose tissue lipid storage that normally occurs during early gestation, resulting in increased availability of lipids that reach the intrauterine compartment [25, 26]. Indeed, pre-pregnancy obesity in women leads to an increase in plasma triglyceride and very-low-density lipoprotein (VLDL) and a reduction in HDL [27]. Adipokines, including leptin, resistin and adiponectin, which modulate glucose homeostasis and lipid metabolism, influence insulin action and have an important role in reproductive tissues, are altered in obese women and animal models of obesity [25, 28]. Thus, they may potentially have a role linking obesity with alterations during embryo development.

Excessive oxidation of metabolic substrates is related to the generation of a pro-inflammatory environment and to the induction of congenital malformations, profoundly addressed in maternal diabetes during embryo organogenesis, as detailed later in this section [2, 29]. Similarly, an increase in systemic and local pro-inflammatory responses is found in women with pre-pregnancy obesity. In fact, there is an increase in adipocyte secretory products such as tumor necrosis factor α (TNF- α), interleukin-6 (IL-6) and also c-reactive protein, which play roles in lipid metabolism [27]. Moreover, in an animal model of obesity and pregnancy, nuclear factor κ B (NF κ B) and c-Jun N-terminal kinase signalling pathways are upregulated leading to the expression of pro-inflammatory genes and reducing the expression of antioxidant and mitochondrial genes in embryos from obese rats [30].

Recent studies performed in experimental models of obesity have shown alterations previous to organogenesis in the lipid content of the early embryo and the oocyte, which can be involved in the induction of congenital malformations [31, 32]. In obese ob/ob mice, ovaries show increased lipid content, and lipid droplets are observed in oocytes in which degenerated mitochondria are also evidenced [33]. In addition, several alterations in lipid metabolizing enzymes have been found in blastocysts from obese mice [34]. Importantly, human studies have shown that increased free fatty acid (FFA) concentrations in follicular fluid are correlated with poorer oocyte-cumulus complexes morphology [35].

Diabetes

Maternal diabetes increases two to eight fold the risk for congenital malformations. The malformations induced are central nervous system defects, cardiovascular defects, respiratory and skeletal anomalies, among many others [36, 37]. A common complication of obesity is diabetes, and both pathologies have additive effects on the risk of congenital central nervous system malformations [38].

In maternal diabetes, the maternal metabolic impairments resulting from the deficiency in insulin concentrations and/or signalling lead to an excess of metabolic substrates. The maternal lipid profile has been found altered during diabetic pregnancies both in humans and experimental animal models [39, 40]. In animal models of diabetes and pregnancy, an increase in circulating triglycerides and FFAs has been found during early organogenesis, and triglyceride accumulation is observed in developing embryos [41, 42].

Mechanisms involved in the induction of congenital malformations in diabetes have been mostly evaluated in animal models of diabetes and pregnancy [40]. In the embryo, the excess in metabolic substrates and their consequent oxidation leads to an increase in reactive oxygen species that exert embryo damage by themselves, by leading to the formation of peroxynitrites, and by inducing lipid peroxidation and the loss of the bioactive lipids needed during embryo organogenesis, including n-6 EFAs [2, 43, 44]. Besides, possibly as a compensatory mechanism for the increased transfer from maternal circulation, the *de novo* synthesis of cholesterol, cholesteryl esters, triglycerides, and phospholipids is reduced in embryos from diabetic rats during early organogenesis [42]. Due to the role of lipids as structural components of both plasma and nuclear cell membranes, as morphogens and as signalling molecules during organogenesis, the reduction in their synthesis in the appropriate location and time during organogenesis can impair developmental processes [45].

Arachidonic acid is an n-6 EFA for the embryo required during early organogenesis and is a precursor of prostaglandins (PGs), important bioactive lipids needed for proper morphogenesis [45]. Experimental models of diabetes show reduced arachidonic acid concentrations in embryos from diabetic rats during organogenesis [46]. PGE₂ participates in the process of neural tube closure and its levels are reduced in embryos from diabetic rats during early organogenesis [47, 48]. Indeed, both PGE₂ and arachidonic acid supplementation reduce the embryonic malformations induced by the diabetic condition, both *in vivo* and *in vitro* [46, 49, 50]. It has been reported that PGI₂ can also prevent hyperglycemia-induced embryonic malformations [51]. Interestingly, PGI₂ is an endogenous agonist of the peroxisome proliferator activated receptor δ (PPAR δ), and the PGI₂-PPAR δ signalling pathway is a fundamental pathway for embryo implantation and decidualization [52, 53]. Certain unsaturated lipids, prostaglandins and leukotrienes are endogenous ligands of the nuclear receptor PPARs, transcription factors that regulate metabolic homeostasis, anti-inflammatory pathways and cell differentiation in different tissues [3]. PPARs have critical roles in metabolic diseases such as diabetes and are specifically involved in regulating maternal diabetes-induced reproductive impairments [54]. Both PPAR δ and PGI₂ concentrations have been found reduced in embryos from diabetic rats [55]. PPAR δ activation up-regulates PGI₂ and PGE₂ levels in embryos from diabetic rats and increases the synthesis of phospholipids, highly required to sustain the rapid growth of the neural folds during the process of closure of the neural tube [42, 55].

Thus, in maternal diabetes, impaired lipid homeostasis is involved in the etiology of congenital malformations in a complex way: damage is due to the excess of oxidizable metabolic substrates in the intrauterine compartment that will increase oxidative stress, to the deficiency in n-6 EFAs that will lead to a deficiency in prostaglandins, and to the impaired PPAR-mediated signalling pathways required during embryogenesis.

In maternal diabetes, the malformation rate varies in different populations, as well as in different rat strains [56, 57]. The genetic background and the environment influence the incidence of diabetic rat embryopathy, partly related to the

embryonic antioxidant defences [58, 59]. Accordingly, congenital malformations are reduced in experimental models of diabetes by moieties with antioxidant effects (e.g. vitamin E and C, folic acid) and with supplementation of diets enriched in mono- and polyunsaturated fatty acids capable of activating PPARs [41, 60-63]. Further work is still needed to address whether congenital malformations are ameliorated by these treatments in diabetic patients. Indeed, as a limitation, data addressing mechanisms of induction of congenital malformations have been mainly obtained through the evaluation of animal models of diabetes and pregnancy [40]. Besides, as congenital malformations can be originated before organogenesis [64], the involvement of oocyte and embryo lipid composition in diabetes-induced congenital malformations has not been clarified and thus deserves further research. Indeed, alterations in arachidonic acid-derived prostaglandins have been found both in oocyte-cumulus complexes and during implantation in experimental models of diabetes and pregnancy and related to alterations in the ovulatory and implantation processes [45].

Summarizing, in the reviewed diseases, congenital malformations can be induced both directly by the deficiency in structural and bioactive lipids, such as cholesterol and EFAs, or indirectly as a result of the generation of a pro-inflammatory environment generated either by the excess of metabolic substrates or by the deficiency in bioactive lipids that regulate both metabolic and anti-inflammatory processes. In contrast to fetal growth effects, the broad spectrum of malformations suggests that the insult occurs very early during embryo development, affecting different tissues and fetal organs.

LIPID IMPAIRMENTS AND FETO-PLACENTAL GROWTH AND DEVELOPMENT

With the establishment of the placenta, nutrition changes from histiotrophic to hemotrophic by the end of the first trimester of pregnancy and further remodelling will occur during the second trimester of pregnancy to allow an efficient exchange of substrates between the maternal and fetal blood according to fetal development [65, 66]. Placental transport of nutrients is exquisitely regulated through specific transporters for the different nutrients, and is especially complex for the transport of lipid moieties [26, 67, 68].

Lipids are provided by maternal circulation and available in increasing concentrations through pregnancy as a result of adaptations in maternal metabolism, which are crucial to provide the lipids to support appropriate fetal growth and development. They involve a first trimester depot of triglycerides in adipose tissue needed for providing sufficient circulating free fatty acids and lipoproteins after mid-pregnancy, when a catabolic and insulin-resistant state driven by pregnancy hormones, will allow providing sufficient energy substrates needed by the fetus and the placenta [26]. EFAs play a fundamental role in fetal development both as components that regulate signalling pathways and as substrates for the generation of other bioactive lipids in cell membranes, cytosolic and nuclear compartments. Their high requirement is fulfilled through an efficient transport of from mother-to-fetus through the placenta, which is assured in physiological conditions [26, 67].

Gestational pathologies either related to nutrient deficiencies or nutrient excesses that affect maternal fat deposition and circulating lipid concentrations have profound consequences in both placental function and fetal development.

Intrauterine Growth Restriction

In experimental models of maternal nutrient restriction, impaired fetal growth is related to the impaired formation of maternal fat depots [69]. In humans, intrauterine growth retardation (IUGR) is related to reduced maternal serum concentrations of lipoproteins (VLDL2, low density lipoprotein (LDL)), although no changes in VLDL1 and HDL and circulating triglycerides and free fatty acids (FFAs) have been reported in various studies [70, 71]. Cholesterol concentrations are reduced when the fetuses have defects in their synthesis, as described in the previous section for the SLO syndrome, but also reduction in maternal-derived cholesterol can affect fetal development [68]. Indeed, IUGR is related to maternal low cholesterol concentrations [72, 73]. Placental insufficiency is an established cause of impaired fetal nutrition and has been related to impairments in fetal lipid metabolism [71, 74]. Also related to placental insufficiency, hypoxia is an important factor involved in the induction of oxidative stress and placental vascular and lipid metabolism impairments [75]. Indeed, increased oxidized LDL, decreased lectin-like oxidized LDL receptor 1 (an oxidized LDL scavenger receptor), increased cholesterol ATP-binding cassette transporter, and increased lipid peroxidation (i.e. TBARS and isoprostanes) are related to hypoxia in preeclampsia and considered relevant in placental damage and dysfunction in this gestational disease [76-78].

Fetal growth and metabolism needs to be a balance of nutrients and oxygen. Indeed, in both human pregnancies and animal models of IUGR, despite the reduction in nutrient transfer, the small for gestational age fetuses have increased circulating triglycerides and even increased liver fat content [79-82]. Besides, studies performed in animal models of IUGR suggest that insulin action is impaired in the fetuses, an alteration that occurs together with reductions in β cell mass and increases in lipid concentrations in the circulation [4].

On the other hand, in fetal growth restriction, reduced concentrations of lipoprotein lipase (LPL) have been described in the human placenta [83], an alteration that may contribute to changes in the transport of lipids to the fetuses and is especially relevant for the transfer of EFAs being carried out in maternal circulation in their esterified form. The relationship of a deficiency in n-6 EFAs to the induction of congenital malformations has been already described in this review. In addition, it is largely known that deficient n-3 EFAs in the last trimester of gestation can affect neurodevelopment, lead to visual defects, and induce cognitive impairments later in life [1, 84]. Indeed, human studies have evidenced in IUGR increased incidence of neurodevelopmental defects, low serum EFAs concentrations in neonates, and reduced capacity to form n-3 PUFAs in infants [85-87]. Besides, in placental insufficiency caused by preeclampsia there are increased concentrations of maternal lipids, an alteration that has been related to the complications of this pathology [88].

Nutrient restriction and placental insufficiency are stressful for the fetus, and pro-inflammatory and pro-oxidant responses have been observed in animal models of these gestational diseases [89]. In placentas from human pregnancies complicated with IUGR, there is increased oxidative and nitrate stress [90]. Pro-inflammatory cytokines like IL-8, IL-12 and TNF α have been found increased in maternal serum, maternal lymphocytes, and cord blood serum in IUGR [91-93]. Therefore, a pro-oxidant and pro-inflammatory intrauterine environment challenges the intrauterine development when fetal nutrients are restricted, similar to that described in those diseases that provide an excess of nutrients, such as obesity and diabetes.

Obesity

In obese patients and obesity experimental models, the mother possesses increased adipose depots, circulating lipids, insulin resistance and a pro-inflammatory state from the pre-pregnancy state [25, 94]. The insulin-resistant state provides an excess of nutrients, including lipids, for transfer to the fetus throughout pregnancy, which is related to fetal overgrowth, and associated with an increase in fat mass rather than lean mass [95]. Besides, in humans and experimental models, not only the mother but also the fetus has been found to be insulin resistant in obese gestations [5, 96]. In part, the presence of insulin resistance in fetuses from obese mothers can be explained by the activation of IRS-1 serine phosphorylation (one of the mechanisms involved in insulin signalling inhibition) and by the increased blood concentrations of pro-inflammatory cytokines [96]. Studies performed in the placenta from obese sheep have shown increased cytokines (TNF α , IL-8, IL-18), NF κ B activation and macrophage infiltration, which are related to an increase in toll-like receptor 4, a receptor which is activated by lipopolysaccharide (LPS) and FFAs [97]. Also in humans, the placenta from obese patients have shown increased pro-inflammatory cytokines, chemokines and macrophage infiltration [98, 99]. On the other hand, in animal models of obesity, changes such as increased placental infarctions, reduced blood flow in the fetal side of the placenta, and reduced fetal and placental junctional zone weights are induced [5]. In humans, impairments of nutrient exchange mechanisms, structural alterations and the pro-inflammatory environment can all be related to the frequent macrosomic phenotype but also to the restricted growth phenotype observed in maternal obesity [94].

Studies performed in obese pregnant women and in experimental models of obesity show that a pro-inflammatory environment in maternal obesity is found not only in the mother and the placenta, but also in the fetuses. Indeed, maternal high fat diets trigger lipotoxicity in the fetal liver of non-human primates [100]. In sheep, maternal obesity is related to a low grade inflammation in the fetus that impairs skeletal muscle development and induces intestine inflammation [101, 102]. In humans, sera obtained from large for gestational age fetuses induce increased matrix metalloproteinase 9 overactivity in cultured human umbilical cord blood endothelial cells (HUVEC) [103]. In overweight patients, amniotic fluid shows increased concentrations of TNF α and C-reactive protein [104]. Both maternal and fetal deficiency of the n-3 PUFA docosahexaenoic acid (DHA) has been

found in both maternal and fetal circulation in non-primates fed high-fat diets, despite normal content of dietary DHA [105]. Moreover, maternal hypercholesterolemia is related to an increase in aortic fatty streaks in the human fetuses from spontaneous abortions/dead preterm newborns [106].

Therefore, maternal obesity leads to metabolic and pro-inflammatory derangements in the intrauterine compartment, similar to that occurring in diabetic gestations.

Diabetes

In both pregestational and gestational diabetes, increased circulating glucose concentrations and increased lipid concentrations resulting from insulin deficiency and/or insulin resistance provide an excess of oxidative substrates related to the generation of a pro-inflammatory state that will affect placental and fetal development [2, 29].

In placentas from both patients and experimental models of diabetes, there are increased markers of oxidative and nitritative stress, increased lipid peroxidation, matrix metalloproteinases overactivity and increased pro-inflammatory cytokines concentrations [29, 40, 90]. Overaccumulation of triglycerides is evident in diabetic placentas, possibly related to increases in placental lipases (LPL, endothelial lipase), lipid moieties transfer proteins (phospholipid transfer protein, fatty acid binding protein) and multiple changes in placental lipid metabolism gene expression that sustain the placental lipid accretion and increased transfer to the fetuses [83, 107-111]. In agreement, FFAs have been directly related to the generation of pro-inflammatory cytokines and lipid droplets in cultured human trophoblasts [112]. Synthesis of different lipid species is reduced in the placenta from diabetic rats, possibly to compensate the increased lipid transport [113]. On the other hand, human studies have shown that concentrations of n-6 and n-3 PUFAs are reduced in maternal circulation in diabetes, although an increased uptake of arachidonic acid in the placenta from diabetic patients has been described [114, 115].

The high susceptibility of long chain PUFAs to oxidation affects their role as bioactive lipids [116]. Indeed, deficiency in certain eicosanoids, bioactive lipids derived from arachidonic acid, is observed in placentas and fetuses in maternal diabetes [2]. This could also be due to the reduction in delta5 and delta6 desaturases, insulin-dependent enzymes needed to form arachidonic acid from linoleic acid [117]. Indeed, it has been largely known that the $\text{PGI}_2/\text{TXA}_2$ ratio is reduced, and thus that the vasodilatory/vasoconstrictive balance is directed towards vasoconstriction in the diabetic placenta in humans and experimental models [2]. This, together with defects in the production of bioactive nitric oxide, challenges the placental and umbilical circulation in maternal diabetes [45, 118]. On the other hand, arachidonic acid derivatives that activate the nuclear receptors PPARs are involved in metabolic and anti-inflammatory pathways in the placenta [54]. In the placenta from diabetic patients and from experimental models of diabetes and pregnancy, there are reduced concentrations of $15\text{-deoxy}\Delta^{12,14}\text{PGJ}_2$ (15-dPGJ_2), a prostaglandin that can activate the nuclear receptor $\text{PPAR}\gamma$ and regulate pro-inflammatory pathways such as reduction of excessive nitric oxide and matrix metalloproteinases overproduction [113, 119]. In experimental models of diabetes, $\text{PPAR}\alpha$, the PPAR

isotype mostly involved in lipid catabolism, is reduced in the placenta from diabetic rats, and its activation by leukotriene B_4 (an arachidonic acid metabolite) and fibrates (their pharmacological activators) can regulate both placental lipid content and peroxidation [120]. In addition, PGI_2 , an arachidonic acid derivative with potent vasodilator effects in the placenta and capable of activating the nuclear receptor $\text{PPAR}\delta$, is also reduced in the placenta from diabetic rats, and has the capacity to negatively regulate excessive nitric oxide production and lipid overaccumulation and peroxidation [121].

In maternal diabetes, fetal overgrowth is the result of the increased glucose transferred to the fetus and the consequent increased insulin secretion which lead to anabolic pathways that are sustained by the increase in nutrients provided through the placenta. However, as evidenced in experimental models of diabetes and pregnancy, fetal hyperglycemia can also lead to degranulation of the fetal β cells, resulting in this case in fetal hypoinsulinemia and fetal growth retardation [40].

In the fetuses from diabetic rats, excessive accumulation both of structural lipids such as phospholipids at mid-gestation and of triglycerides and cholesteryl esters in the fetal liver at term provides evidence of the excess of the metabolic substrates [122, 123]. In circulation, increases in triglyceride and cholesterol concentrations are observed in fetuses from diabetic rats [124, 125]. Indeed, fat mass and circulating LDL are increased in infants born to diabetic mothers [126, 127]. Besides, in cord blood from newborns from diabetic patients, FFAs and/or triglycerides have been found increased, although cholesterol concentrations have been found either increased or decreased in newborns from diabetic mothers [39, 128, 129].

In experimental models of diabetes and pregnancy, despite increased fetal fat content, increased lipid synthesis remain increased in fetuses from diabetic rats, an alteration that is regulated by the activation of the nuclear receptor $\text{PPAR}\alpha$ [123]. At term gestation, fetal treatments with the $\text{PPAR}\alpha$ activator LTB_4 have been found to regulate lipid metabolism and pro-inflammatory pathways in the fetal liver and fetal lung from diabetic rats [122, 124]. Besides, fetal activation of $\text{PPAR}\alpha$ has been found to reduce overgrowth of the placenta, the fetus, and fetal organs such as the liver and the lung in experimental models of diabetes and pregnancy [120, 122, 124].

Similar to that occurring in IUGR and in maternal obesity, deficiency of PUFAs occurs in diabetic gestations. This deficiency can be observed in cord blood from type 1, type 2 and gestational diabetic patients [115, 130, 131]. Indeed, EFAs deficiency is related to the induction of congenital malformations, as described earlier in this review, and is possibly related to the induction of neurodevelopmental defects, which are enhanced in the offspring of diabetic mothers [84, 132].

Therefore, fetuses and placentas are affected by pathologies that change maternal lipid concentrations, not only directly due to the altered transfer of lipids to the fetus, but also indirectly as a result of the damage caused by the pro-inflammatory environment and the loss of bioactive lipids that regulate metabolic and anti-inflammatory processes during development.

INTRAUTERINE LIPID IMPAIRMENTS AND POST-NATAL CONSEQUENCES

Postnatal consequences arise both at the perinatal period and in the long term in pregnancy diseases associated with intrauterine lipid impairments. IUGR as well as diabetes and obesity are causes of increased infant morbidity and mortality in the infancy and childhood, and also have long-term impact on adult life [95, 133-135]. Growing evidence relates both small and large for gestational age newborns with the programming of metabolic and cardiovascular diseases [4, 5]. Due to its role in nutrient transport and as a determinant of fetal weight, the placenta has a relevant role in the intrauterine programming of these diseases [74, 136, 137]. The concept of intrauterine programming is supported by epidemiological evidence that shows that the *in utero* environment influences the onset of diseases later in life. The mechanisms involved in programming the susceptibility of diseases in the offspring are a current focus of research and remain to be fully elucidated.

Programming of Type 2 Diabetes

Human studies have established an association between intrauterine growth retardation and increased risk of glucose intolerance and type 2 diabetes in later life [138, 139]. In experimental models of IUGR, impaired insulin action and increased catch-up growth in the offspring occur in parallel with enhanced fat deposition in the early postnatal life and are related to the later development of glucose intolerance [140].

Pre-pregnancy obesity and excessive gestational weight gain are implicated in the offspring's increased risk of multiple metabolic abnormalities, including an increase in plasma levels of insulin, FFAs and triglycerides, insulin resistance, an increase in hepatic gluconeogenic pathways, accelerated neonatal catch-up growth, glucose intolerance and obesity, as observed in both humans and animal models [100, 141-143]. Besides, in different species (rodents, lambs and non-human primates) experimental models of obesity have shown hepatic steatosis, fatty pancreas disease and nonalcoholic fatty liver disease in the offspring [100, 144, 145].

Experimental studies in diabetes and pregnancy have clearly established the increase in mother-to-fetus glucose transfer and the hypertrophy and hyperplasia of insulin-producing β cells in fetuses from diabetic mothers, as well as the resulting impaired glucose tolerance when the offspring become adult [146]. This abnormal glucose homeostasis in the offspring of diabetic mothers has also been reported in epidemiological studies [147-149]. Impaired adaptive responses of β cell mass are implicated in the increased risk for diabetes in later life not only in maternal diabetes, but also in maternal overnutrition, intrauterine growth restriction and uteroplacental insufficiency [150, 151].

Other factors related to the intrauterine programming of adulthood diseases in maternal obesity, diabetes and intrauterine growth restriction and to the impaired lipid metabolism is the adipokine leptin [5, 151]. The intrauterine and perinatal changes in leptin concentrations and/or leptin resistance in these pathologies program an altered brain appetite regulation and energy balance in the offspring. In IUGR, low levels of fetal leptin have been shown to be followed by in-

creases in leptin in the infants, children and adults, affecting satiety, adiposity and weight gain [152]. In the offspring from animals fed high-fat diets, hyperphagia and increased energy intake are observed, and these changes are associated with increases in serum lipid concentrations and increases in the expression of orexigenic peptides, which result from the proliferation and differentiation of neurons that express orexigenic peptides during development [153-155]. Experimental and human studies have shown altered leptin concentrations and altered programming of orexigenic and anorexigenic hypothalamic neurons function in the offspring from diabetic mothers [156, 157].

It is important that in both humans and animal models, programming of type 2 diabetes often shows gender-specific effects on the type and degree of metabolic alterations [158-160]. In experimental IUGR models, this is partly related to gender-dependent impairments in hepatic fatty acid metabolism and triglyceride concentrations that specifically affect the male offspring [161, 162].

Programming of Cardiovascular Diseases

Pregnancy diseases that lead to lipid metabolism impairments are clearly related to the programming of cardiovascular diseases. Indeed, human and experimental studies have shown that fetal nutrient restriction or excess can induce cardiac and vascular impairments in the offspring [163, 164]. In rats fed a saturated fatty acid diet, the offspring shows an abnormal fatty acid composition of the aorta [165], whereas in obese mice, the offspring show hypertension and resistance artery endothelial dysfunction [166]. Besides, in humans, maternal hypercholesterolemia induces early markers of cardiovascular disease such as endothelial dysfunction, vascular reactivity and atherosclerosis in the offspring [167]. The risk of developing cardiovascular disease is increased for both the mother and the offspring in diabetic patients [168]. Interestingly, in rats, not only excessive lipid concentrations but also n-3 PUFAs deficiency during fetal development are found related to an increased blood pressure in the offspring [169].

PPARs

Nuclear receptors PPARs, master regulators of metabolic homeostasis and inflammatory processes that are activated by PUFAs and their derived eicosanoids, have also been related to the intrauterine programming of metabolic diseases. Indeed, experimental models of both intrauterine growth restriction and maternal obesity induced by high-fat diet lead to an enhanced activity of PPAR γ in adipose tissue of offspring due to two different mechanisms: upregulation of PPAR coactivators in IUGR and downregulation of PPAR corepressors in obese pregnancies, leading in both cases to a subsequent increase in fat mass [170]. Low intramuscular expression of PPAR γ and high expression of fatty acid transporters, which correlate with intramuscular triglycerides, are found in lambs born to obese mothers [171]. On the other hand, adult offspring from high-fat fed rats show increased hepatic PPAR α expression, an alteration associated with an increased expression of key genes that regulate fatty acid oxidation, with the expression of insulin growth factor-2 (IGF-2) and with changes in the expression of several microRNAs that could be involved in the impaired regulation of the gene expression observed [172].

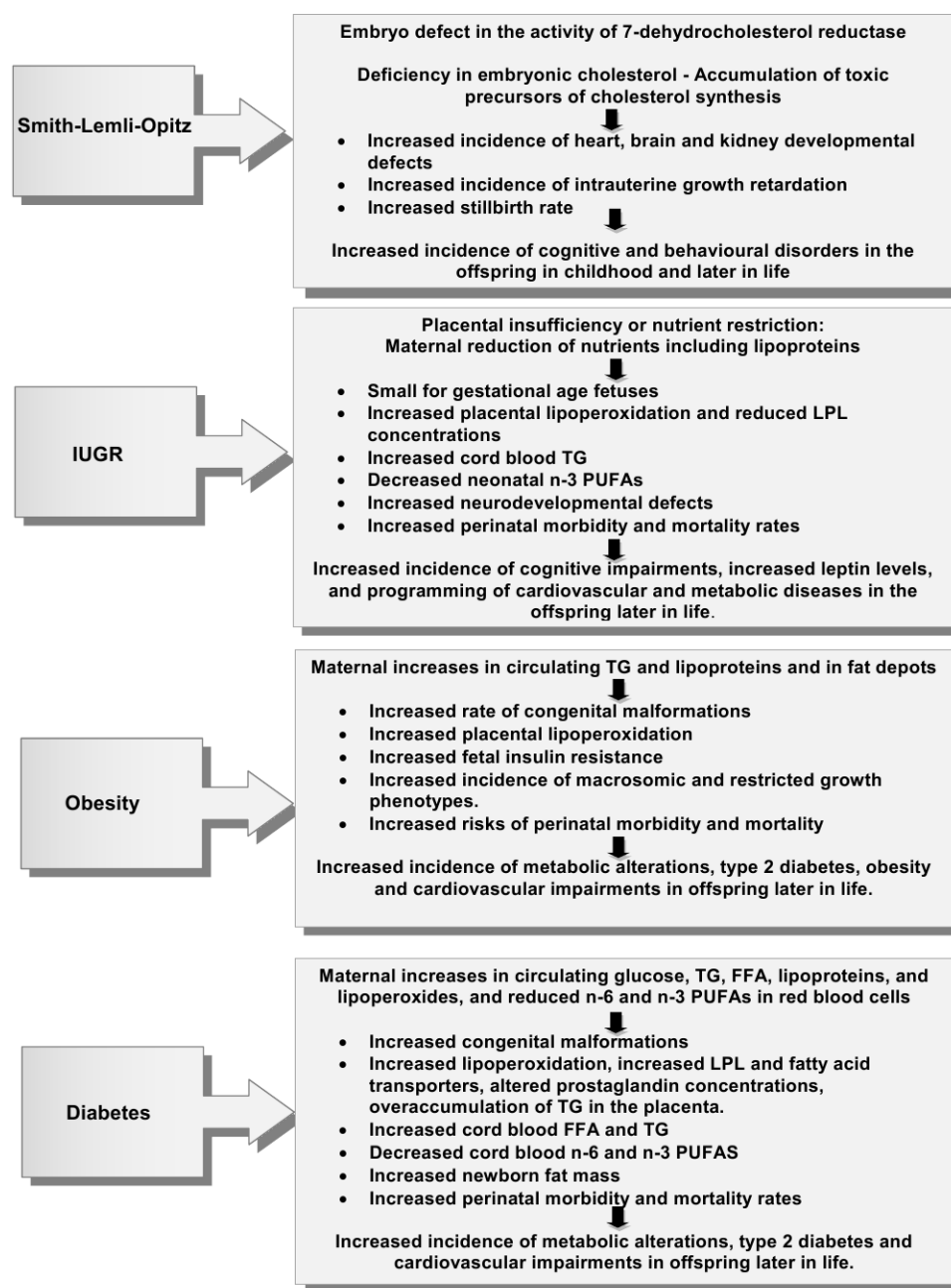


Fig. (1). Main clinical features related to lipid impairments in Smith-Lemli-Opitz and gestational diseases.

Epigenetics

There are exciting advances in the involvement of epigenetic regulation in intrauterine programming of metabolic diseases. Several studies in experimental models and some human studies have shown that an excess or restriction of nutrients can epigenetically modify expression of certain genes by altering the methylation status of gene promoter regions or by the post-translational modification of histone tail residues through methylation, acetylation, phosphorylation, sumoylation and/or ubiquitination, resulting in changes in chromatin condensation and gene expression [25, 151, 173]. Undernutrition in rats during pregnancy induces hypomethylation of the PPAR α promoter and increases the expression of this key regulator of lipid metabolism in the

liver and heart of the offspring [174, 175]. Besides, intrauterine growth retardation in rats is related to an epigenetic modification in the PDX-1 gene that codifies for pancreatic and duodenal homeobox 1 (PDX-1) transcription factor, critical for β cell development and plasticity and function after birth [176]. Interestingly, intervention studies in IUGR rat neonates provide reversal of epigenetic changes in the PDX-1 gene with postnatal treatments with exendin-4, treatments also associated with a reduction in lipid peroxidation in the liver [177].

On the other hand, in primates, maternal high-fat diets lead to an increase in histone H3 acetylation and a decrease in histone deacetylase activity in the liver, in correlation with an altered lipid metabolism in the fetus [173]. In a recent human

study performed in umbilical cord tissue, methylation of the promoter of retinoid X receptor (RXR), an obligate heterodimer of PPAR nuclear receptors, has been found associated with children later adiposity [178]. Interestingly, in rats, high fat fed fathers have been shown to program β cell dysfunction by altering the expression of multiple genes in the β cell of female offspring, at least in part through epigenetic modifications [179]. Besides, recently, in an experimental model of diabetes in pregnancy, glucose intolerance in both F1 and F2 has been associated with an abnormal expression and hypermethylation of IGF-2/H19, a paternal imprinted growth factor and its regulator, which are involved in β cell development and function [180, 181]. Besides, epigenetic changes in histone acetylation in embryos from diabetic rats during embryo organogenesis suggest a possible epigenetic role in diabetic embryopathy, and these changes are different from those found in embryos from rats fed a high-fat diet [182].

Overall, lipid homeostasis is highly relevant in fetal and perinatal stages to sustain growth and development, to regulate metabolic and signalling pathways and to lead to a proper gene transcription that will be relevant in the infant and adult offspring. Therefore, impaired lipid homeostasis during pregnancy diseases is a relevant component in the mechanisms of programming of intrauterine diseases that warrants further evaluation.

CONCLUSIONS

The clinical relevance of impaired lipid homeostasis in pregnancy diseases is summarized in (Fig. 1). It is interesting to note that common pathways related to impaired maternal lipid homeostasis have been associated with different gestational pathologies and with two extreme phenotypes, the IUGR and the overweight offspring. Although the mechanisms that relate the fetal growth-related disturbances to lipid derangements are far to be completely understood, it is clear that there are roles for the mother, the placenta and the fetus, which differentially lead to the altered lipid concentrations and metabolism here reviewed. Indeed, relevant roles of the abnormal formation of adipose depots, the generation of a pro-inflammatory state, the insulin-resistant state, the deficiency in EFAs and alterations in the placental transfer and utilization of lipids by the fetus have been here described and related to the alterations in fetal growth and the programming of intrauterine diseases. Also, congenital malformations are related to impaired lipid homeostasis, and the availability of EFAs and cholesterol play a fundamental role. Although experimental studies have shown that supplementation with antioxidants and PUFAs has beneficial effects in some of these pregnancy pathologies, there are still no clear beneficial results in clinical studies [6, 29, 54, 84, 132, 183]. Thus, to what extent essential fatty acids deficiency may lead to abnormalities of embryonic, fetal or post-natal development still remains unresolved. Overall, further research is needed for a better understanding of the mechanisms involved in abnormal lipid moieties-induced damage during gestation and for further translation of this knowledge to clinical assays.

CONFLICT OF INTEREST

The author(s) confirm that this article content has no conflicts of interest.

ACKNOWLEDGEMENTS

The author's work was funded by grants from Agencia de Promoción Científica y Tecnológica de Argentina (PICT 2010-00034) and from Consejo Nacional de Investigaciones Científicas y Técnicas (PIP 11220100100002).

ABBREVIATIONS

15dPGJ2	=	15deoxy $\Delta^{12,14}$ PGJ ₂
DHA	=	Docosahexaenoic Acid
EFAs	=	Essential Fatty Acids
FFAs	=	Free Fatty Acids
HDL	=	High-Density Lipoprotein
HUVEC	=	Human Umbilical Cord Blood Endothelial Cells
IGF-2	=	Insulin Growth Factor-2
IL	=	Interleukin
IUGR	=	Intrauterine Growth Retardation
LDL	=	Low Density Lipoprotein
LPL	=	Lipoprotein Lipase
LPS	=	Lipopolysaccharide
NF κ B	=	Nuclear Factor Kappa B
PDX-1	=	Pancreatic and Duodenal Homeobox-1
PGs	=	Prostaglandins
PPAR	=	Peroxisome Proliferator Activated Receptor
PUFAs	=	Polyunsaturated Fatty Acids
RXR	=	Retinoid X Receptor
SLO	=	Smith-Lemli-Opitz Syndrome
TNF α	=	Tumor Necrosis Factor α
VLDL	=	Very-Low-Density Lipoprotein

REFERENCES

- [1] Innis, S.M. Dietary (n-3) fatty acids and brain development. *J Nutr*, **2007**, 137, 855-9.
- [2] Jawerbaum, A.; Gonzalez, E. Diabetic pregnancies: the challenge of developing in a pro-inflammatory environment. *Curr Med Chem*, **2006**, 13, 2127-38.
- [3] Desvergne, B.; Michalik, L.; Wahli, W. Be fit or be sick: peroxisome proliferator-activated receptors are down the road. *Mol Endocrinol*, **2004**, 18, 1321-32.
- [4] Gatford, K.L.; Simmons, R.A.; De Blasio, M.J.; Robinson, J.S.; Owens, J.A. Review: Placental programming of postnatal diabetes and impaired insulin action after IUGR. *Placenta*, **2010**, 31 Suppl, S60-5.
- [5] Li, M.; Sloboda, D.M.; Vickers, M.H. Maternal obesity and developmental programming of metabolic disorders in offspring: evidence from animal models. *Exp Diabetes Res*, **2011**, 2011, 592408.
- [6] Yessoufou, A.; Moutairou, K. Maternal diabetes in pregnancy: early and long-term outcomes on the offspring and the concept of "metabolic memory". *Exp Diabetes Res*, **2011**, 2011, 218598.
- [7] Mills, J.L.; Baker, L.; Goldman, A.S. Malformations in infants of diabetic mothers occur before the seventh gestational week. Implications for treatment. *Diabetes*, **1979**, 28, 292-3.
- [8] Vrablik, T.L.; Watts, J.L. Emerging roles for specific fatty acids in developmental processes. *Genes Dev*, **2012**, 26, 631-7.
- [9] Pratt, H.P. Phospholipid synthesis in the preimplantation mouse embryo. *J Reprod Fertil*, **1980**, 58, 237-48.

- [10] Zohn, I.E.; Sarkar, A.A. The visceral yolk sac endoderm provides for absorption of nutrients to the embryo during neurulation. *Birth Defects Res A Clin Mol Teratol*, **2010**, 88, 593-600.
- [11] Terasawa, Y.; Cases, S.J.; Wong, J.S.; Jamil, H.; Jothi, S.; Traber, M.G.; Packer, L.; Gordon, D.A.; Hamilton, R.L.; Farese, R.V., Jr. Apolipoprotein B-related gene expression and ultrastructural characteristics of lipoprotein secretion in mouse yolk sac during embryonic development. *J Lipid Res*, **1999**, 40, 1967-77.
- [12] Herz, J.; Farese, R.V., Jr. The LDL receptor gene family, apolipoprotein B and cholesterol in embryonic development. *J Nutr*, **1999**, 129, 473S-475S.
- [13] Farese, R.V., Jr.; Ruland, S.L.; Flynn, L.M.; Stokowski, R.P.; Young, S.G. Knockout of the mouse apolipoprotein B gene results in embryonic lethality in homozygotes and protection against diet-induced hypercholesterolemia in heterozygotes. *Proc Natl Acad Sci U S A*, **1995**, 92, 1774-8.
- [14] Shelton, J.M.; Lee, M.H.; Richardson, J.A.; Patel, S.B. Microsomal triglyceride transfer protein expression during mouse development. *J Lipid Res*, **2000**, 41, 532-7.
- [15] Linton, M.F.; Farese, R.V., Jr.; Young, S.G. Familial hypobetalipoproteinemia. *J Lipid Res*, **1993**, 34, 521-41.
- [16] Raabe, M.; Kim, E.; Veniant, M.; Nielsen, L.B.; Young, S.G. Using genetically engineered mice to understand apolipoprotein-B deficiency syndromes in humans. *Proc Assoc Am Physicians*, **1998**, 110, 521-30.
- [17] Santander, N.G.; Contreras-Duarte, S.; Awad, M.F.; Lizama, C.; Passalacqua, I.; Rigotti, A.; Busso, D. Developmental abnormalities in mouse embryos lacking the HDL receptor SR-BI. *Hum Mol Genet*, **2012**.
- [18] Porter, F.D. Smith-Lemli-Opitz syndrome: pathogenesis, diagnosis and management. *Eur J Hum Genet*, **2008**, 16, 535-41.
- [19] Quelin, C.; Loget, P.; Verloes, A.; Bazin, A.; Bessieres, B.; Laquerriere, A.; Patrier, S.; Grigorescu, R.; Encha-Razavi, F.; Delahaye, S.; Jouannic, J.M.; Carbonne, B.; D'Herve, D.; Aubry, M.C.; Mace, G.; Harvey, T.; Ville, Y.; Viot, G.; Joye, N.; Odent, S.; Attie-Bitach, T.; Wolf, C.; Chevy, F.; Benlian, P.; Gonzales, M. Phenotypic spectrum of fetal Smith-Lemli-Opitz syndrome. *Eur J Med Genet*, **2012**, 55, 81-90.
- [20] Diaz-Stransky, A.; Tierney, E. Cognitive and behavioral aspects of Smith-Lemli-Opitz syndrome. *Am J Med Genet C Semin Med Genet*, **2012**, 160C, 295-300.
- [21] Porter, J.A.; Young, K.E.; Beachy, P.A. Cholesterol modification of hedgehog signaling proteins in animal development. *Science*, **1996**, 274, 255-9.
- [22] Woollett, L.A. Maternal cholesterol in fetal development: transport of cholesterol from the maternal to the fetal circulation. *Am J Clin Nutr*, **2005**, 82, 1155-61.
- [23] Mills, J.L.; Troendle, J.; Conley, M.R.; Carter, T.; Druschel, C.M. Maternal obesity and congenital heart defects: a population-based study. *Am J Clin Nutr*, **2010**, 91, 1543-9.
- [24] Stothard, K.J.; Tennant, P.W.; Bell, R.; Rankin, J. Maternal overweight and obesity and the risk of congenital anomalies: a systematic review and meta-analysis. *Jama*, **2009**, 301, 636-50.
- [25] Heerwagen, M.J.; Miller, M.R.; Barbour, L.A.; Friedman, J.E. Maternal obesity and fetal metabolic programming: a fertile epigenetic soil. *Am J Physiol Regul Integr Comp Physiol*, **2010**, 299, R711-22.
- [26] Herrera, E.; Amusquivar, E.; Lopez-Soldado, I.; Ortega, H. Maternal lipid metabolism and placental lipid transfer. *Horm Res*, **2006**, 65 Suppl 3, 59-64.
- [27] Ramsay, J.E.; Ferrell, W.R.; Crawford, L.; Wallace, A.M.; Greer, I.A.; Sattar, N. Maternal obesity is associated with dysregulation of metabolic, vascular, and inflammatory pathways. *J Clin Endocrinol Metab*, **2002**, 87, 4231-7.
- [28] Mitchell, M.; Armstrong, D.T.; Robker, R.L.; Norman, R.J. Adipokines: implications for female fertility and obesity. *Reproduction*, **2005**, 130, 583-97.
- [29] Lappas, M.; Hiden, U.; Desoye, G.; Froehlich, J.; Hauguel-de Mouzon, S.; Jawerbaum, A. The role of oxidative stress in the pathophysiology of gestational diabetes mellitus. *Antioxid Redox Signal*, **2011**, 15, 3061-100.
- [30] Shankar, K.; Zhong, Y.; Kang, P.; Lau, F.; Blackburn, M.L.; Chen, J.R.; Borengasser, S.J.; Ronis, M.J.; Badger, T.M. Maternal obesity promotes a proinflammatory signature in rat uterus and blastocyst. *Endocrinology*, **2011**, 152, 4158-70.
- [31] Jungheim, E.S.; Schoeller, E.L.; Marquard, K.L.; Loudon, E.D.; Schaffer, J.E.; Moley, K.H. Diet-induced obesity model: abnormal oocytes and persistent growth abnormalities in the offspring. *Endocrinology*, **2010**, 151, 4039-46.
- [32] Van Hoes, V.; Sturmey, R.G.; Bermejo-Alvarez, P.; Rizos, D.; Gutierrez-Adan, A.; Leese, H.J.; Bols, P.E.; Leroy, J.L. Elevated non-esterified fatty acid concentrations during bovine oocyte maturation compromise early embryo physiology. *PLoS One*, **2011**, 6, e23183.
- [33] Serke, H.; Nowicki, M.; Kosacka, J.; Schroder, T.; Kloting, N.; Bluher, M.; Kallendrusch, S.; Spanel-Borowski, K. Leptin-deficient (ob/ob) mouse ovaries show fatty degeneration, enhanced apoptosis and decreased expression of steroidogenic acute regulatory enzyme. *Int J Obes (Lond)*, **2012**, 36, 1047-53.
- [34] Konigsdorf, C.A.; Navarrete Santos, A.; Schmidt, J.S.; Fischer, S.; Fischer, B. Expression profile of fatty acid metabolism genes in preimplantation blastocysts of obese and non-obese mice. *Obes Facts*, **2012**, 5, 575-86.
- [35] Jungheim, E.S.; Macones, G.A.; Odem, R.R.; Patterson, B.W.; Lanzendorf, S.E.; Ratts, V.S.; Moley, K.H. Associations between free fatty acids, cumulus oocyte complex morphology and ovarian function during *in vitro* fertilization. *Fertil Steril*, **2011**, 95, 1970-4.
- [36] Martinez-Frias, M.L. Epidemiological analysis of outcomes of pregnancy in diabetic mothers: identification of the most characteristic and most frequent congenital anomalies. *Am J Med Genet*, **1994**, 51, 108-13.
- [37] Correa, A.; Gilboa, S.M.; Besser, L.M.; Botto, L.D.; Moore, C.A.; Hobbs, C.A.; Cleves, M.A.; Riehle-Colarusso, T.J.; Waller, D.K.; Reece, E.A. Diabetes mellitus and birth defects. *Am J Obstet Gynecol*, **2008**, 199, 237 e1-9.
- [38] Anderson, J.L.; Waller, D.K.; Canfield, M.A.; Shaw, G.M.; Watkins, M.L.; Werler, M.M. Maternal obesity, gestational diabetes, and central nervous system birth defects. *Epidemiology*, **2005**, 16, 87-92.
- [39] Herrera, E.; Ortega-Senovilla, H. Disturbances in lipid metabolism in diabetic pregnancy - Are these the cause of the problem? *Best Pract Res Clin Endocrinol Metab*, **2010**, 24, 515-25.
- [40] Jawerbaum, A.; White, V. Animal models in diabetes and pregnancy. *Endocr Rev*, **2010**, 31, 680-701.
- [41] Higa, R.; Kurtz, M.; Mazzucco, M.B.; Musikant, D.; White, V.; Jawerbaum, A. Folic acid and safflower oil supplementation interacts and protects embryos from maternal diabetes-induced damage. *Mol Hum Reprod*, **2012**, 18:253-64.
- [42] Sinner, D.; Caviglia, J.M.; Jawerbaum, A.; Igal, R.A.; Gonzalez, E. Lipid metabolism in the embryos of diabetic rats during early organogenesis: modulatory effect of prostaglandin E2. *Reprod Fertil Dev*, **2003**, 15, 75-80.
- [43] Zabihi, S.; Loeken, M.R. Understanding diabetic teratogenesis: where are we now and where are we going? *Birth Defects Res A Clin Mol Teratol*, **2010**, 88, 779-90.
- [44] Eriksson, U.J. Congenital anomalies in diabetic pregnancy. *Semin Fetal Neonatal Med*, **2009**, 14, 85-93.
- [45] Jawerbaum, A.; Gonzalez, E. The role of alterations in arachidonic acid metabolism and nitric oxide homeostasis in rat models of diabetes during early pregnancy. *Curr Pharm Des*, **2005**, 11, 1327-33.
- [46] Reece, E.A.; Wu, Y.K.; Wiznitzer, A.; Homko, C.; Yao, J.; Borenstein, M.; Sloskey, G. Dietary polyunsaturated fatty acid prevents malformations in offspring of diabetic rats. *Am J Obstet Gynecol*, **1996**, 175, 818-23.
- [47] Piddington, R.; Joyce, J.; Dhanasekaran, P.; Baker, L. Diabetes mellitus affects prostaglandin E2 levels in mouse embryos during neurulation. *Diabetologia*, **1996**, 39, 915-20.
- [48] Jawerbaum, A.; Sinner, D.; White, V.; Pustovrh, C.; Capobianco, E.; Gimeno, M.A.; Gonzalez, E.T. Modulation of PGE2 generation in the diabetic embryo: effect of nitric oxide and superoxide dismutase. *Prostaglandins Leukot Essent Fatty Acids*, **2001**, 64, 127-33.
- [49] Wentzel, P.; Eriksson, U.J. A diabetes-like environment increases malformation rate and diminishes prostaglandin E(2) in rat embryos: reversal by administration of vitamin E and folic acid. *Birth Defects Res A Clin Mol Teratol*, **2005**, 73, 506-11.
- [50] Goto, M.P.; Goldman, A.S.; Uhing, M.R. PGE2 prevents anomalies induced by hyperglycemia or diabetic serum in mouse embryos. *Diabetes*, **1992**, 41, 1644-50.
- [51] Baker, L.; Piddington, R.; Goldman, A.; Egler, J.; Moehring, J. Myo-inositol and prostaglandins reverse the glucose inhibition of

- neural tube fusion in cultured mouse embryos. *Diabetologia*, **1990**, 33, 593-6.
- [52] Lim, H.; Dey, S.K. PPAR delta functions as a prostacyclin receptor in blastocyst implantation. *Trends Endocrinol Metab*, **2000**, 11, 137-42.
- [53] Wang, H.; Xie, H.; Sun, X.; Tranguch, S.; Zhang, H.; Jia, X.; Wang, D.; Das, S.K.; Desvergne, B.; Wahli, W.; DuBois, R.N.; Dey, S.K. Stage-specific integration of maternal and embryonic peroxisome proliferator-activated receptor delta signaling is critical to pregnancy success. *J Biol Chem*, **2007**, 282, 37770-82.
- [54] Jawerbaum, A.; Capobianco, E. Review: Effects of PPAR activation in the placenta and the fetus: Implications in maternal diabetes. *Placenta*, **2011**, 35:S212-7.
- [55] Higa, R.; Gonzalez, E.; Pustovrh, M.C.; White, V.; Capobianco, E.; Martinez, N.; Jawerbaum, A. PPARdelta and its activator PGI2 are reduced in diabetic embryopathy: involvement of PPARdelta activation in lipid metabolic and signalling pathways in rat embryo early organogenesis. *Mol Hum Reprod*, **2007**, 13, 103-10.
- [56] Reece, E.A. Diabetes-induced birth defects: what do we know? What can we do? *Curr Diab Rep*, **2012**, 12, 24-32.
- [57] Eriksson, U.J.; Cederberg, J.; Wentzel, P. Congenital malformations in offspring of diabetic mothers—animal and human studies. *Rev Endocr Metab Disord*, **2003**, 4, 79-93.
- [58] Ejdesjo, A.; Wentzel, P.; Eriksson, U.J. Influence of maternal metabolism and parental genetics on fetal maldevelopment in diabetic rat pregnancy. *Am J Physiol Endocrinol Metab*, **2012**, 302, E1198-209.
- [59] Ejdesjo, A.; Wentzel, P.; Eriksson, U.J. Genetic and environmental influence on diabetic rat embryopathy. *Am J Physiol Endocrinol Metab*, **2011**, 300, E454-67.
- [60] Higa, R.; White, V.; Martinez, N.; Kurtz, M.; Capobianco, E.; Jawerbaum, A. Safflower and olive oil dietary treatments rescue aberrant embryonic arachidonic acid and nitric oxide metabolism and prevent diabetic embryopathy in rats. *Mol Hum Reprod*, **2010**, 16, 286-95.
- [61] Cederberg, J.; Siman, C.M.; Eriksson, U.J. Combined treatment with vitamin E and vitamin C decreases oxidative stress and improves fetal outcome in experimental diabetic pregnancy. *Pediatr Res*, **2001**, 49, 755-62.
- [62] Wentzel, P.; Gareskog, M.; Eriksson, U.J. Folic acid supplementation diminishes diabetes- and glucose-induced dysmorphogenesis in rat embryos *in vivo* and *in vitro*. *Diabetes*, **2005**, 54, 546-53.
- [63] Reece, E.A.; Wu, Y.K.; Zhao, Z.; Dhanasekaran, D. Dietary vitamin and lipid therapy rescues aberrant signaling and apoptosis and prevents hyperglycemia-induced diabetic embryopathy in rats. *Am J Obstet Gynecol*, **2006**, 194, 580-5.
- [64] Wyman, A.; Pinto, A.B.; Sheridan, R.; Moley, K.H. One-cell zygote transfer from diabetic to nondiabetic mouse results in congenital malformations and growth retardation in offspring. *Endocrinology*, **2008**, 149, 466-9.
- [65] Brosens, I.; Pijnenborg, R.; Vercruysse, L.; Romero, R. The "Great Obstetrical Syndromes" are associated with disorders of deep placentation. *Am J Obstet Gynecol*, **2011**, 204, 193-201.
- [66] Burton, G.J.; Watson, A.L.; Hempstock, J.; Skepper, J.N.; Jauniaux, E. Uterine glands provide histiotrophic nutrition for the human fetus during the first trimester of pregnancy. *J Clin Endocrinol Metab*, **2002**, 87, 2954-9.
- [67] Gil-Sanchez, A.; Demmelmair, H.; Parrilla, J.J.; Koletzko, B.; Larque, E. Mechanisms involved in the selective transfer of long chain polyunsaturated Fatty acids to the fetus. *Front Genet*, **2011**, 2, 57.
- [68] Woollett, L.A. Review: Transport of maternal cholesterol to the fetal circulation. *Placenta*, **2011**, 32 Suppl 2, S218-21.
- [69] Luther, J.; Aitken, R.; Milne, J.; Matsuzaki, M.; Reynolds, L.; Redmer, D.; Wallace, J. Maternal and fetal growth, body composition, endocrinology, and metabolic status in undernourished adolescent sheep. *Biol Reprod*, **2007**, 77, 343-50.
- [70] Cetin, I.; Alvino, G.; Cardellicchio, M. Long chain fatty acids and dietary fats in fetal nutrition. *J Physiol*, **2009**, 587, 3441-51.
- [71] Desoye, G.; Gauster, M.; Wadsack, C. Placental transport in pregnancy pathologies. *Am J Clin Nutr*, **2011**, 94:1896S-1902S.
- [72] Sattar, N.; Greer, I.A.; Galloway, P.J.; Packard, C.J.; Shepherd, J.; Kelly, T.; Mathers, A. Lipid and lipoprotein concentrations in pregnancies complicated by intrauterine growth restriction. *J Clin Endocrinol Metab*, **1999**, 84, 128-30.
- [73] Wadsack, C.; Tabano, S.; Maier, A.; Hiden, U.; Alvino, G.; Cozzi, V.; Huttering, M.; Schneider, W.J.; Lang, U.; Cetin, I.; Desoye, G. Intrauterine growth restriction is associated with alterations in placental lipoprotein receptors and maternal lipoprotein composition. *Am J Physiol Endocrinol Metab*, **2007**, 292, E476-84.
- [74] Myatt, L. Placental adaptive responses and fetal programming. *J Physiol*, **2006**, 572, 25-30.
- [75] Heazell, A.E.; Brown, M.; Worton, S.A.; Dunn, W.B. Review: The effects of oxygen on normal and pre-eclamptic placental tissue—insights from metabolomics. *Placenta*, **2011**, 32 Suppl 2, S119-24.
- [76] Chigusa, Y.; Tatsumi, K.; Kondoh, E.; Fujita, K.; Nishimura, F.; Mogami, H.; Konishi, I. Decreased lectin-like oxidized LDL receptor 1 (LOX-1) and low Nr1h2 activation in placenta are involved in preeclampsia. *J Clin Endocrinol Metab*, **2012**, 97, E1862-70.
- [77] Plosch, T.; Gellhaus, A.; van Straten, E.M.; Wolf, N.; Huijckman, N.C.; Schmidt, M.; Dunk, C.E.; Kuipers, F.; Winterhager, E. The liver X receptor (LXR) and its target gene ABCA1 are regulated upon low oxygen in human trophoblast cells: a reason for alterations in preeclampsia? *Placenta*, **2010**, 31, 910-8.
- [78] Borrego-Diaz, E.; Rosales, J.C.; Proverbio, T.; Teppa-Garran, A.; Andaluz, R.; Abad, C.; Marin, R.; Proverbio, F. Effect of placental hypoxia on the plasma membrane Ca-ATPase (PMCA) activity and the level of lipid peroxidation of syncytiotrophoblast and red blood cell ghosts. *Placenta*, **2008**, 29, 44-50.
- [79] Rueda-Clausen, C.F.; Dolinsky, V.W.; Morton, J.S.; Proctor, S.D.; Dyck, J.R.; Davidge, S.T. Hypoxia-induced intrauterine growth restriction increases the susceptibility of rats to high-fat diet-induced metabolic syndrome. *Diabetes*, **2011**, 60, 507-16.
- [80] Rodie, V.A.; Caslake, M.J.; Stewart, F.; Sattar, N.; Ramsay, J.E.; Greer, I.A.; Freeman, D.J. Fetal cord plasma lipoprotein status in uncomplicated human pregnancies and in pregnancies complicated by pre-eclampsia and intrauterine growth restriction. *Atherosclerosis*, **2004**, 176, 181-7.
- [81] Merzouk, H.; Meghelli-Bouchenak, M.; el-Korso, N.; Belleville, J.; Prost, J. Low birth weight at term impairs cord serum lipoprotein compositions and concentrations. *Eur J Pediatr*, **1998**, 157, 321-6.
- [82] Yamada, M.; Wolfe, D.; Han, G.; French, S.W.; Ross, M.G.; Desai, M. Early onset of fatty liver in growth-restricted rat fetuses and newborns. *Congenit Anom (Kyoto)*, **2011**, 51, 167-73.
- [83] Magnusson, A.L.; Waterman, I.J.; Wennergren, M.; Jansson, T.; Powell, T.L. Triglyceride hydrolase activities and expression of fatty acid binding proteins in the human placenta in pregnancies complicated by intrauterine growth restriction and diabetes. *J Clin Endocrinol Metab*, **2004**, 89, 4607-14.
- [84] Campoy, C.; Escolano-Margarit, M.V.; Anjos, T.; Szajewska, H.; Uauy, R. Omega 3 fatty acids on child growth, visual acuity and neurodevelopment. *Br J Nutr*, **2012**, 107 Suppl 2, S85-106.
- [85] Baschat, A.A. Neurodevelopment following fetal growth restriction and its relationship with antepartum parameters of placental dysfunction. *Ultrasound Obstet Gynecol*, **2011**, 37, 501-14.
- [86] Llanos, A.; Lin, Y.; Mena, P.; Salem, N., Jr.; Uauy, R. Infants with intrauterine growth restriction have impaired formation of docosahexaenoic acid in early neonatal life: a stable isotope study. *Pediatr Res*, **2005**, 58, 735-40.
- [87] Crawford, M.A.; Doyle, W.; Leaf, A.; Leighfield, M.; Ghebremeskel, K.; Phylactos, A. Nutrition and neurodevelopmental disorders. *Nutr Health*, **1993**, 9, 81-97.
- [88] Rodie, V.A.; Freeman, D.J.; Sattar, N.; Greer, I.A. Pre-eclampsia and cardiovascular disease: metabolic syndrome of pregnancy? *Atherosclerosis*, **2004**, 175, 189-202.
- [89] Leduc, L.; Delvin, E.; Ouellet, A.; Garofalo, C.; Grenier, E.; Morin, L.; Dube, J.; Bouity-Voubou, M.; Moutquin, J.M.; Fouron, J.C.; Klam, S.; Levy, E. Oxidized low-density lipoproteins in cord blood from neonates with intra-uterine growth restriction. *Eur J Obstet Gynecol Reprod Biol*, **2011**, 156, 46-9.
- [90] Myatt, L. Review: Reactive oxygen and nitrogen species and functional adaptation of the placenta. *Placenta*, **2010**, 31 Suppl, S66-9.
- [91] Laskowska, M.; Laskowska, K.; Leszczynska-Gorzela, B.; Oleszczuk, J. Comparative analysis of the maternal and umbilical interleukin-8 levels in normal pregnancies and in pregnancies complicated by preeclampsia with intrauterine normal growth and intrauterine growth retardation. *J Matern Fetal Neonatal Med*, **2007**, 20, 527-32.
- [92] Holcberg, G.; Huleihel, M.; Sapir, O.; Katz, M.; Tsadkin, M.; Furman, B.; Mazor, M.; Myatt, L. Increased production of tumor ne-

- crosis factor- α TNF- α by IUGR human placentae. *Eur J Obstet Gynecol Reprod Biol*, **2001**, 94, 69-72.
- [93] Raghupathy, R.; Al-Azemi, M.; Azizieh, F. Intrauterine growth restriction: cytokine profiles of trophoblast antigen-stimulated maternal lymphocytes. *Clin Dev Immunol*, **2012**, 2012, 734865.
- [94] Higgins, L.; Greenwood, S.L.; Wareing, M.; Sibley, C.P.; Mills, T.A. Obesity and the placenta: A consideration of nutrient exchange mechanisms in relation to aberrant fetal growth. *Placenta*, **2011**, 32, 1-7.
- [95] Catalano, P.M. Obesity, insulin resistance, and pregnancy outcome. *Reproduction*, **2010**, 140, 365-71.
- [96] Catalano, P.M.; Presley, L.; Minium, J.; Hauguel-de Mouzon, S. Fetuses of obese mothers develop insulin resistance in utero. *Diabetes Care*, **2009**, 32, 1076-80.
- [97] Zhu, M.J.; Du, M.; Nathanielsz, P.W.; Ford, S.P. Maternal obesity up-regulates inflammatory signaling pathways and enhances cytokine expression in the mid-gestation sheep placenta. *Placenta*, **2010**, 31, 387-91.
- [98] Roberts, K.A.; Riley, S.C.; Reynolds, R.M.; Barr, S.; Evans, M.; Statham, A.; Hor, K.; Jabbour, H.N.; Norman, J.E.; Denison, F.C. Placental structure and inflammation in pregnancies associated with obesity. *Placenta*, **2011**, 32, 247-54.
- [99] Challier, J.C.; Basu, S.; Bintein, T.; Minium, J.; Hotmire, K.; Catalano, P.M.; Hauguel-de Mouzon, S. Obesity in pregnancy stimulates macrophage accumulation and inflammation in the placenta. *Placenta*, **2008**, 29, 274-81.
- [100] McCurdy, C.E.; Bishop, J.M.; Williams, S.M.; Grayson, B.E.; Smith, M.S.; Friedman, J.E.; Grove, K.L. Maternal high-fat diet triggers lipotoxicity in the fetal livers of nonhuman primates. *J Clin Invest*, **2009**, 119, 323-35.
- [101] Tong, J.F.; Yan, X.; Zhu, M.J.; Ford, S.P.; Nathanielsz, P.W.; Du, M. Maternal obesity downregulates myogenesis and beta-catenin signaling in fetal skeletal muscle. *Am J Physiol Endocrinol Metab*, **2009**, 296, E917-24.
- [102] Yan, X.; Huang, Y.; Wang, H.; Du, M.; Hess, B.W.; Ford, S.P.; Nathanielsz, P.W.; Zhu, M.J. Maternal obesity induces sustained inflammation in both fetal and offspring large intestine of sheep. *Inflamm Bowel Dis*, **2011**, 17, 1513-22.
- [103] Johannsson, E.; Henriksen, T.; Iversen, P.O. Increase in matrix metalloproteinases from endothelial cells exposed to umbilical cord plasma from high birth weight newborns. *Am J Physiol Regul Integr Comp Physiol*, **2007**, 292, R1563-8.
- [104] Bugatto, F.; Fernandez-Deudero, A.; Bailen, A.; Fernandez-Macias, R.; Hervias-Vivancos, B.; Bartha, J.L. Second-trimester amniotic fluid proinflammatory cytokine levels in normal and overweight women. *Obstet Gynecol*, **2010**, 115, 127-33.
- [105] Grant, W.F.; Gillingham, M.B.; Batra, A.K.; Fewkes, N.M.; Comstock, S.M.; Takahashi, D.; Braun, T.P.; Grove, K.L.; Friedman, J.E.; Marks, D.L. Maternal high fat diet is associated with decreased plasma n-3 fatty acids and fetal hepatic apoptosis in nonhuman primates. *PLoS One*, **2011**, 6, e17261.
- [106] Napoli, C.; D'Armiento, F.P.; Mancini, F.P.; Postiglione, A.; Witztum, J.L.; Palumbo, G.; Palinski, W. Fatty streak formation occurs in human fetal aortas and is greatly enhanced by maternal hypercholesterolemia. Intimal accumulation of low density lipoprotein and its oxidation precede monocyte recruitment into early atherosclerotic lesions. *J Clin Invest*, **1997**, 100, 2680-90.
- [107] Radaelli, T.; Lepercq, J.; Varastehpour, A.; Basu, S.; Catalano, P.M.; Hauguel-De Mouzon, S. Differential regulation of genes for fetoplacental lipid pathways in pregnancy with gestational and type 1 diabetes mellitus. *Am J Obstet Gynecol*, **2009**, 201, 209 e1-209 e10.
- [108] Lindegaard, M.L.; Damm, P.; Mathiesen, E.R.; Nielsen, L.B. Placental triglyceride accumulation in maternal type 1 diabetes is associated with increased lipase gene expression. *J Lipid Res*, **2006**, 47, 2581-8.
- [109] Shafir, E.; Barash, V. Placental function in maternal-fetal fat transport in diabetes. *Biol Neonate*, **1987**, 51, 102-12.
- [110] Scholler, M.; Wadsack, C.; Lang, I.; Etschmaier, K.; Schweinzer, C.; Marsche, G.; Dieber-Rotheneder, M.; Desoye, G.; Panzenboeck, U. Phospholipid transfer protein in the placental endothelium is affected by gestational diabetes mellitus. *J Clin Endocrinol Metab*, **2012**, 97, 437-45.
- [111] Scifres, C.M.; Chen, B.; Nelson, D.M.; Sadovsky, Y. Fatty acid binding protein 4 regulates intracellular lipid accumulation in human trophoblasts. *J Clin Endocrinol Metab*, **2011**, 96, E1083-91.
- [112] Pathmaperuma, A.N.; Mana, P.; Cheung, S.N.; Kugathas, K.; Josiah, A.; Koina, M.E.; Broomfield, A.; Delghingaro-Augusto, V.; Ellwood, D.A.; Dahlstrom, J.E.; Nolan, C.J. Fatty acids alter glycerolipid metabolism and induce lipid droplet formation, syncytialisation and cytokine production in human trophoblasts with minimal glucose effect or interaction. *Placenta*, **2010**, 31, 230-9.
- [113] Capobianco, E.; Jawerbaum, A.; Romanini, M.C.; White, V.; Pustovrh, C.; Higa, R.; Martinez, N.; Mugnaini, M.T.; Sonez, C.; Gonzalez, E. 15-Deoxy-Delta12,14-prostaglandin J2 and peroxisome proliferator-activated receptor gamma (PPARgamma) levels in term placental tissues from control and diabetic rats: modulatory effects of a PPARgamma agonist on nitridergic and lipid placental metabolism. *Reprod Fertil Dev*, **2005**, 17, 423-33.
- [114] Kuhn, D.C.; Crawford, M.A.; Stuart, M.J.; Botti, J.J.; Demers, L.M. Alterations in transfer and lipid distribution of arachidonic acid in placentas of diabetic pregnancies. *Diabetes*, **1990**, 39, 914-8.
- [115] Min, Y.; Lowy, C.; Ghebreskel, K.; Thomas, B.; Offley-Shore, B.; Crawford, M. Unfavorable effect of type 1 and type 2 diabetes on maternal and fetal essential fatty acid status: a potential marker of fetal insulin resistance. *Am J Clin Nutr*, **2005**, 82, 1162-8.
- [116] Negre-Salvayre, A.; Auge, N.; Ayala, V.; Basaga, H.; Boada, J.; Brenke, R.; Chapple, S.; Cohen, G.; Feher, J.; Grune, T.; Lengyel, G.; Mann, G.E.; Pamplona, R.; Poli, G.; Portero-Otin, M.; Riahi, Y.; Salvayre, R.; Sasson, S.; Serrano, J.; Shamni, O.; Siems, W.; Siow, R.C.; Wiswedel, I.; Zarkovic, K.; Zarkovic, N. Pathological aspects of lipid peroxidation. *Free Radic Res*, **2010**, 44, 1125-71.
- [117] Brenner, R.R. Hormonal modulation of delta6 and delta5 desaturases: case of diabetes. *Prostaglandins Leukot Essent Fatty Acids*, **2003**, 68, 151-62.
- [118] San Martin, R.; Sobrevia, L. Gestational diabetes and the adenosine/L-arginine/nitric oxide (ALANO) pathway in human umbilical vein endothelium. *Placenta*, **2006**, 27, 1-10.
- [119] Pustovrh, M.C.; Capobianco, E.; Martinez, N.; Higa, R.; White, V.; Jawerbaum, A. MMP/ TIMP balance is modulated *in vitro* by 15dPGJ(2) in fetuses and placentas from diabetic rats. *Eur J Clin Invest*, **2009**, 39, 1082-90.
- [120] Martinez, N.; Kurtz, M.; Capobianco, E.; Higa, R.; White, V.; Jawerbaum, A. PPARalpha agonists regulate lipid metabolism and nitric oxide production and prevent placental overgrowth in term placentas from diabetic rats. *J Mol Endocrinol*, **2011**, 47, 1-12.
- [121] Kurtz, M.; Capobianco, E.; Martinez, N.; Fernandez, J.; Higa, R.; White, V.; Jawerbaum, A. Carbaprostacyclin, a PPARdelta agonist, ameliorates excess lipid accumulation in diabetic rat placentas. *Life Sci*, **2010**, 86, 781-90.
- [122] Martinez, N.; Kurtz, M.; Higa, R.; Capobianco, E.; Jawerbaum, A. Activation of the nuclear receptor PPARalpha regulates lipid metabolism in foetal liver from diabetic rats: implications in diabetes-induced foetal overgrowth. *Diabetes Metab Res Rev*, **2011**, 27, 35-46.
- [123] Martinez, N.; Capobianco, E.; White, V.; Pustovrh, M.C.; Higa, R.; Jawerbaum, A. Peroxisome proliferator-activated receptor alpha activation regulates lipid metabolism in the feto-placental unit from diabetic rats. *Reproduction*, **2008**, 136, 95-103.
- [124] Kurtz, M.; Martinez, N.; Capobianco, E.; Higa, R.; Fornes, D.; White, V.; Jawerbaum, A. Increased nitric oxide production and gender-dependent changes in PPARalpha expression and signaling in the fetal lung from diabetic rats. *Mol Cell Endocrinol*, **2012**.
- [125] Damasceno, D.C.; Kiss, A.C.; Sinzato, Y.K.; de Campos, K.E.; Rudge, M.V.; Calderon, I.M.; Volpato, G.T. Maternal-fetal outcome, lipid profile and oxidative stress of diabetic rats neonatally exposed to streptozotocin. *Exp Clin Endocrinol Diabetes*, **2011**, 119, 408-13.
- [126] Lindegaard, M.L.; Svarrer, E.M.; Damm, P.; Mathiesen, E.R.; Nielsen, L.B. Increased LDL cholesterol and CRP in infants of mothers with type 1 diabetes. *Diabetes Metab Res Rev*, **2008**, 24, 465-71.
- [127] Lawlor, D.A.; Fraser, A.; Lindsay, R.S.; Ness, A.; Dabelea, D.; Catalano, P.; Davey Smith, G.; Sattar, N.; Nelson, S.M. Association of existing diabetes, gestational diabetes and glycosuria in pregnancy with macrosomia and offspring body mass index, waist and fat mass in later childhood: findings from a prospective pregnancy cohort. *Diabetologia*, **2010**, 53, 89-97.
- [128] Merzouk, H.; Madani, S.; Korso, N.; Bouchenak, M.; Prost, J.; Belleville, J. Maternal and fetal serum lipid and lipoprotein concen-

- trations and compositions in type 1 diabetic pregnancy: relationship with maternal glycemic control. *J Lab Clin Med*, **2000**, 136, 441-8.
- [129] Gordon, Y.; Patterson, T.M.; Parker, C.R., Jr. Relation of glucose control in diabetic pregnancy to fetal cholesterol homeostasis. *J Matern Fetal Neonatal Med*, **2009**, 22, 8-12.
- [130] Thomas, B.A.; Ghebremeskel, K.; Lowy, C.; Offley-Shore, B.; Crawford, M.A. Plasma fatty acids of neonates born to mothers with and without gestational diabetes. *Prostaglandins Leukot Essent Fatty Acids*, **2005**, 72, 335-41.
- [131] Ortega-Senovilla, H.; Alvino, G.; Taricco, E.; Cetin, I.; Herrera, E. Gestational Diabetes Mellitus Impairs the Proportion of Fatty Acids in Umbilical Artery but not in Umbilical Vein Plasma. *Diabetes Care*, **2008**.
- [132] Larque, E.; Gil-Sanchez, A.; Prieto-Sanchez, M.T.; Koletzko, B. Omega 3 fatty acids, gestation and pregnancy outcomes. *Br J Nutr*, **2012**, 107 Suppl 2, S77-84.
- [133] Reece, E.A.; Leguizamón, G.; Wiznitzer, A. Gestational diabetes: the need for a common ground. *Lancet*, **2009**, 373, 1789-97.
- [134] Simeoni, U.; Barker, D.J. Offspring of diabetic pregnancy: long-term outcomes. *Semin Fetal Neonatal Med*, **2009**, 14, 119-24.
- [135] Gardosi, J. Intrauterine growth restriction: new standards for assessing adverse outcome. *Best Pract Res Clin Obstet Gynaecol*, **2009**, 23, 741-9.
- [136] Jansson, T.; Powell, T.L. Role of the placenta in fetal programming: underlying mechanisms and potential interventional approaches. *Clin Sci (Lond)*, **2007**, 113, 1-13.
- [137] Sibley, C.P.; Brownbill, P.; Dilworth, M.; Glazier, J.D. Review: Adaptation in placental nutrient supply to meet fetal growth demand: implications for programming. *Placenta*, **2010**, 31 Suppl, S70-4.
- [138] Harder, T.; Rodekamp, E.; Schellong, K.; Dudenhausen, J.W.; Plagemann, A. Birth weight and subsequent risk of type 2 diabetes: a meta-analysis. *Am J Epidemiol*, **2007**, 165, 849-57.
- [139] Kaijser, M.; Bonamy, A.K.; Akre, O.; Cnattingius, S.; Granath, F.; Norman, M.; Ekblom, A. Perinatal risk factors for diabetes in later life. *Diabetes*, **2009**, 58, 523-6.
- [140] De Blasio, M.J.; Gatford, K.L.; McMillen, I.C.; Robinson, J.S.; Owens, J.A. Placental restriction of fetal growth increases insulin action, growth, and adiposity in the young lamb. *Endocrinology*, **2007**, 148, 1350-8.
- [141] Wrotniak, B.H.; Shultz, J.; Butts, S.; Stettler, N. Gestational weight gain and risk of overweight in the offspring at age 7 y in a multicenter, multiethnic cohort study. *Am J Clin Nutr*, **2008**, 87, 1818-24.
- [142] Srinivasan, M.; Katewa, S.D.; Palaniyappan, A.; Pandya, J.D.; Patel, M.S. Maternal high-fat diet consumption results in fetal malprogramming predisposing to the onset of metabolic syndrome-like phenotype in adulthood. *Am J Physiol Endocrinol Metab*, **2006**, 291, E792-9.
- [143] Nivoit, P.; Morens, C.; Van Assche, F.A.; Jansen, E.; Poston, L.; Remacle, C.; Reusens, B. Established diet-induced obesity in female rats leads to offspring hyperphagia, adiposity and insulin resistance. *Diabetologia*, **2009**, 52, 1133-42.
- [144] Bouanane, S.; Merzouk, H.; Benkalfat, N.B.; Soulimane, N.; Merzouk, S.A.; Gresti, J.; Tessier, C.; Narce, M. Hepatic and very low-density lipoprotein fatty acids in obese offspring of overfed dams. *Metabolism*, **2010**, 59, 1701-9.
- [145] Oben, J.A.; Mouralidarane, A.; Samuelsson, A.M.; Matthews, P.J.; Morgan, M.L.; McKee, C.; Soeda, J.; Fernandez-Twinn, D.S.; Martin-Gronert, M.S.; Ozanne, S.E.; Sigala, B.; Novelli, M.; Poston, L.; Taylor, P.D. Maternal obesity during pregnancy and lactation programs the development of offspring non-alcoholic fatty liver disease in mice. *J Hepatol*, **2010**, 52, 913-20.
- [146] Holemans, K.; Aerts, L.; Van Assche, F.A. Lifetime consequences of abnormal fetal pancreatic development. *J Physiol*, **2003**, 547, 11-20.
- [147] Gautier, J.F.; Wilson, C.; Weyer, C.; Mott, D.; Knowler, W.C.; Cavaghan, M.; Polonsky, K.S.; Bogardus, C.; Pratley, R.E. Low acute insulin secretory responses in adult offspring of people with early onset type 2 diabetes. *Diabetes*, **2001**, 50, 1828-33.
- [148] Singh, R.; Pearson, E.; Avery, P.J.; McCarthy, M.I.; Levy, J.C.; Hitman, G.A.; Sampson, M.; Walker, M.; Hattersley, A.T. Reduced beta cell function in offspring of mothers with young-onset type 2 diabetes. *Diabetologia*, **2006**, 49, 1876-80.
- [149] Dabelea, D.; Pettitt, D.J. Intrauterine diabetic environment confers risks for type 2 diabetes mellitus and obesity in the offspring, in addition to genetic susceptibility. *J Pediatr Endocrinol Metab*, **2001**, 14, 1085-91.
- [150] Portha, B.; Chavey, A.; Movassat, J. Early-life origins of type 2 diabetes: fetal programming of the beta-cell mass. *Exp Diabetes Res*, **2011**, 2011, 105076.
- [151] Warner, M.J.; Ozanne, S.E. Mechanisms involved in the developmental programming of adulthood disease. *Biochem J*, **2010**, 427, 333-47.
- [152] Briana, D.D.; Malamitsi-Puchner, A. Intrauterine growth restriction and adult disease: the role of adipocytokines. *Eur J Endocrinol*, **2009**, 160, 337-47.
- [153] Chang, G.Q.; Gaysinskaya, V.; Karatayev, O.; Leibowitz, S.F. Maternal high-fat diet and fetal programming: increased proliferation of hypothalamic peptide-producing neurons that increase risk for overeating and obesity. *J Neurosci*, **2008**, 28, 12107-19.
- [154] Rajia, S.; Chen, H.; Morris, M.J. Maternal overnutrition impacts offspring adiposity and brain appetite markers-modulation by postweaning diet. *J Neuroendocrinol*, **2010**, 22, 905-14.
- [155] Bouret, S.G. Early life origins of obesity: role of hypothalamic programming. *J Pediatr Gastroenterol Nutr*, **2009**, 48 Suppl 1, S31-8.
- [156] Franke, K.; Harder, T.; Aerts, L.; Melchior, K.; Fahrenkrog, S.; Rodekamp, E.; Ziska, T.; Van Assche, F.A.; Dudenhausen, J.W.; Plagemann, A. 'Programming' of orexigenic and anorexigenic hypothalamic neurons in offspring of treated and untreated diabetic mother rats. *Brain Res*, **2005**, 1031, 276-83.
- [157] Hauguel-de Mouzon, S.; Lepercq, J.; Catalano, P. The known and unknown of leptin in pregnancy. *Am J Obstet Gynecol*, **2006**, 194, 1537-45.
- [158] Kautzky-Willer, A.; Handisurya, A. Metabolic diseases and associated complications: sex and gender matter! *Eur J Clin Invest*, **2009**, 39, 631-48.
- [159] Elahi, M.M.; Cagampang, F.R.; Mukhtar, D.; Anthony, F.W.; Ohri, S.K.; Hanson, M.A. Long-term maternal high-fat feeding from weaning through pregnancy and lactation predisposes offspring to hypertension, raised plasma lipids and fatty liver in mice. *Br J Nutr*, **2009**, 102, 514-9.
- [160] Flanagan, D.E.; Moore, V.M.; Godsland, I.F.; Cockington, R.A.; Robinson, J.S.; Phillips, D.I. Fetal growth and the physiological control of glucose tolerance in adults: a minimal model analysis. *Am J Physiol Endocrinol Metab*, **2000**, 278, E700-6.
- [161] Zambrano, E.; Bautista, C.J.; Deas, M.; Martinez-Samayo, P.M.; Gonzalez-Zamorano, M.; Ledesma, H.; Morales, J.; Larrea, F.; Nathanielsz, P.W. A low maternal protein diet during pregnancy and lactation has sex- and window of exposure-specific effects on offspring growth and food intake, glucose metabolism and serum leptin in the rat. *J Physiol*, **2006**, 571, 221-30.
- [162] van Straten, E.M.; Bloks, V.W.; van Dijk, T.H.; Baller, J.F.; Huijkman, N.C.; Kuipers, I.; Verkade, H.J.; Plosch, T. Sex-dependent programming of glucose and fatty acid metabolism in mouse offspring by maternal protein restriction. *Gend Med*, **2012**, 9, 166-179 e13.
- [163] Crispi, F.; Bijlens, B.; Figueras, F.; Bartrons, J.; Eixarch, E.; Le Noble, F.; Ahmed, A.; Gratacos, E. Fetal growth restriction results in remodeled and less efficient hearts in children. *Circulation*, **2010**, 121, 2427-36.
- [164] Drake, A.J.; Reynolds, R.M. Impact of maternal obesity on offspring obesity and cardiometabolic disease risk. *Reproduction*, **2010**, 140, 387-98.
- [165] Ghosh, P.; Bitsanis, D.; Ghebremeskel, K.; Crawford, M.A.; Poston, L. Abnormal aortic fatty acid composition and small artery function in offspring of rats fed a high fat diet in pregnancy. *J Physiol*, **2001**, 533, 815-22.
- [166] Samuelsson, A.M.; Matthews, P.A.; Argenton, M.; Christie, M.R.; McConnell, J.M.; Jansen, E.H.; Piersma, A.H.; Ozanne, S.E.; Twinn, D.F.; Remacle, C.; Rowlerson, A.; Poston, L.; Taylor, P.D. Diet-induced obesity in female mice leads to offspring hyperphagia, adiposity, hypertension, and insulin resistance: a novel murine model of developmental programming. *Hypertension*, **2008**, 51, 383-92.
- [167] Napoli, C.; Palinski, W. Maternal hypercholesterolemia during pregnancy influences the later development of atherosclerosis: clinical and pathogenic implications. *Eur Heart J*, **2001**, 22, 4-9.
- [168] Garcia-Vargas, L.; Addison, S.S.; Nistala, R.; Kurukulasuriya, D.; Sowers, J.R. Gestational Diabetes and the Offspring: Implications

- in the Development of the Cardiorenal Metabolic Syndrome in Offspring. *Cardiorenal Med*, **2012**, 2, 134-142.
- [169] Weisinger, H.S.; Armitage, J.A.; Sinclair, A.J.; Vingrys, A.J.; Burns, P.L.; Weisinger, R.S. Perinatal omega-3 fatty acid deficiency affects blood pressure later in life. *Nat Med*, **2001**, 7, 258-9.
- [170] Desai, M.; Ross, M.G. Fetal programming of adipose tissue: effects of intrauterine growth restriction and maternal obesity/high-fat diet. *Semin Reprod Med*, **2011**, 29, 237-45.
- [171] Yan, X.; Huang, Y.; Zhao, J.X.; Long, N.M.; Uthlaut, A.B.; Zhu, M.J.; Ford, S.P.; Nathanielsz, P.W.; Du, M. Maternal obesity-impaired insulin signaling in sheep and induced lipid accumulation and fibrosis in skeletal muscle of offspring. *Biol Reprod*, **2011**, 85, 172-8.
- [172] Zhang, J.; Zhang, F.; Didelot, X.; Bruce, K.D.; Cagampang, F.R.; Vatish, M.; Hanson, M.; Lehnert, H.; Ceriello, A.; Byrne, C.D. Maternal high fat diet during pregnancy and lactation alters hepatic expression of insulin like growth factor-2 and key microRNAs in the adult offspring. *BMC Genomics*, **2009**, 10, 478.
- [173] Aagaard-Tillery, K.M.; Grove, K.; Bishop, J.; Ke, X.; Fu, Q.; McKnight, R.; Lane, R.H. Developmental origins of disease and determinants of chromatin structure: maternal diet modifies the primate fetal epigenome. *J Mol Endocrinol*, **2008**, 41, 91-102.
- [174] Lillycrop, K.A.; Phillips, E.S.; Jackson, A.A.; Hanson, M.A.; Burdge, G.C. Dietary protein restriction of pregnant rats induces and folic acid supplementation prevents epigenetic modification of hepatic gene expression in the offspring. *J Nutr*, **2005**, 135, 1382-6.
- [175] Slater-Jefferies, J.L.; Lillycrop, K.A.; Townsend, P.A.; Torrens, C.; Hoile, S.P.; Hanson, M.A.; Burdge, G.C. Feeding a protein-restricted diet during pregnancy induces altered epigenetic regulation of peroxisomal proliferator-activated receptor-alpha in the heart of the offspring. *J Dev Orig Health Dis*, **2011**, 2, 250-255.
- [176] Park, J.H.; Stoffers, D.A.; Nicholls, R.D.; Simmons, R.A. Development of type 2 diabetes following intrauterine growth retardation in rats is associated with progressive epigenetic silencing of Pdx1. *J Clin Invest*, **2008**, 118, 2316-24.
- [177] Stoffers, D.A.; Desai, B.M.; DeLeon, D.D.; Simmons, R.A. Neonatal exendin-4 prevents the development of diabetes in the intrauterine growth retarded rat. *Diabetes*, **2003**, 52, 734-40.
- [178] Godfrey, K.M.; Sheppard, A.; Gluckman, P.D.; Lillycrop, K.A.; Burdge, G.C.; McLean, C.; Rodford, J.; Slater-Jefferies, J.L.; Garratt, E.; Crozier, S.R.; Emerald, B.S.; Gale, C.R.; Inskip, H.M.; Cooper, C.; Hanson, M.A. Epigenetic gene promoter methylation at birth is associated with child's later adiposity. *Diabetes*, **2011**, 60, 1528-34.
- [179] Ng, S.F.; Lin, R.C.; Laybutt, D.R.; Barres, R.; Owens, J.A.; Morris, M.J. Chronic high-fat diet in fathers programs beta-cell dysfunction in female rat offspring. *Nature*, **2010**, 467, 963-6.
- [180] Ding, G.L.; Wang, F.F.; Shu, J.; Tian, S.; Jiang, Y.; Zhang, D.; Wang, N.; Luo, Q.; Zhang, Y.; Jin, F.; Leung, P.C.; Sheng, J.Z.; Huang, H.F. Transgenerational glucose intolerance with Igf2/H19 epigenetic alterations in mouse islet induced by intrauterine hyperglycemia. *Diabetes*, **2012**, 61, 1133-42.
- [181] Calderari, S.; Gangnerau, M.N.; Thibault, M.; Meile, M.J.; Kassis, N.; Alvarez, C.; Portha, B.; Serradas, P. Defective IGF2 and IGF1R protein production in embryonic pancreas precedes beta cell mass anomaly in the Goto-Kakizaki rat model of type 2 diabetes. *Diabetologia*, **2007**, 50, 1463-71.
- [182] Salbaum, J.M.; Kappen, C. Responses of the embryonic epigenome to maternal diabetes. *Birth Defects Res A Clin Mol Teratol*, **2012**, 94:774-81.
- [183] Poston, L.; Igosheva, N.; Mistry, H.D.; Seed, P.T.; Shennan, A.H.; Rana, S.; Karumanchi, S.A.; Chappell, L.C. Role of oxidative stress and antioxidant supplementation in pregnancy disorders. *Am J Clin Nutr*, **2011**, 94, 1980S-1985S.