

pubs.acs.org/IECR Review

Proteins as Promising Biobased Building Blocks for Preparing Functional Hybrid Protein/Synthetic Polymer Nanoparticles

- 3 Luisa Guadalupe Cencha, Mariana Allasia, Ludmila Irene Ronco, Gisela Carina Luque,
- 4 Matías Luis Picchio, Roque J. Minari,* and Luis M. Gugliotta*



Cite This: https://doi.org/10.1021/acs.iecr.0c05958

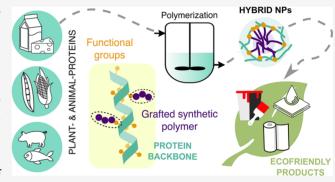


ACCESS

Metrics & More

Article Recommendations

5 ABSTRACT: The pursuit of sustainable and environmentally 6 friendly materials has been powered by environmental concerns 7 and the decline in oil reserves. Among the different routes toward 8 this end, the replacement of oil-based materials by renewable 9 materials stands out. In this way, protein based materials have 10 gained interest. This review article summarizes the progress 11 achieved in the synthesis of hybrid protein/synthetic polymer 12 nanoparticles which have the potential to be used in industrial 13 applications. Although technical achievements and efficacy proofs 14 concerning the increased compatibility of polymer/protein are 15 already available, practical implementation in industry still 16 represents an additional challenge and should be the focus of 17 interest in future research. The available literature supports the



18 potential of hybrid protein/polymer nanoparticles in the production of ecofriendly alternatives for large scale applications as 19 coatings, paints, adhesives and films.

20 INTRODUCTION

21 Polymer materials available today are the result of decades of 22 evolution. In the last decades, the polymer industry has made 23 significant efforts to develop environmentally friendly 24 processes that prevent the emission of volatile organic 25 compounds (VOCs). In this context, waterborne polymers, 26 such as latexes, appear as an ecological alternative to solvent-27 borne resins and occupy a key role in many industries as 28 coatings, paints, adhesives, impact modifiers, inks, packaging, 29 rubber, and so on. 1,2 Furthermore, waterborne latexes in film 30 applications have additional advantages over solvent-borne 31 alternatives, such as faster drying times, less smell, and easier 32 clean up.^{3,4} Additionally, waterborne polymer dispersions are 33 obtained by emulsion polymerization (EP) or miniemulsion 34 polymerization (MEP). This presents several advantages in 35 comparison with processes carried out in bulk or in organic 36 solvents due to the use of water as continuous phase. The 37 environmental friendliness and safety of these polymers are 38 ensured due to the high heat capacity and low viscosity of the 39 emulsion. Thus, the EP process was created and nowadays it is 40 a widespread technique to produce at large scale high-quality 41 materials in a consistent, safe, and environmentally friendly 42 way, with a worldwide production of polymer material of more 43 than 25 million tonnes/year. 5,6 It is forecasted that the global 44 market for waterborne dispersions will increase its value from 45 USD 7.6 billion (2019) to USD 11.8 billion (2027). Despite

the many advantages of waterborne latexes, most of the 46 industrially synthesized dispersed polymers are based on 47 petroleum-based monomers. In recent years, instability of 48 petroleum prices, consumer demands, environmental concerns, 49 and strict regulations about gas emissions have pushed 50 academics and industries toward the development of 51 sustainable and environmentally friendly materials. In this 52 scenario, there is an interest for replacing current petroleum- 53 based polymers by renewable and more sustainable feedstocks 54 aiming to reduce the carbon footprint of this industry and 55 ensure a circular economy.8 In the last decade, many 56 substitution alternatives have been investigated, involving 57 copolymerization of biobased monomers, incorporation of 58 biobased preformed polymers such as alkyds, ^{10–13} and ⁵⁹ biopolymers as carbohydrates, ^{14,15} natural rubber, ¹⁶ proteins, ¹⁷ 60 and lignocellulosic biomass. ^{18–20} Among the alternatives for ⁶¹ substituting petroleum-based polymers, natural proteins 62 represent a promising option due to their superior biodegrad- 63 ability, edibility, low toxicity, convenient absorbability, and in 64

Received: December 3, 2020 Revised: March 12, 2021 Accepted: March 15, 2021



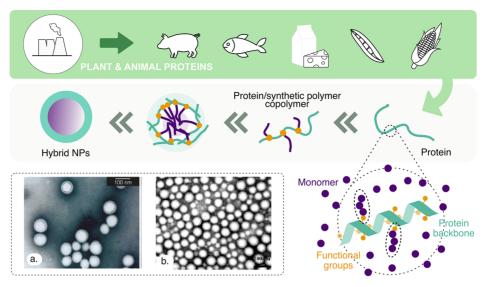


Figure 1. Scheme of the process for producing hybrid protein/polymer NPs. Plant protein and animal protein, usually byproducts or even effluents of others industries, represent a high-potential feedstock to reduce the use of petroleum-based monomers. Waterborne latexes of hybrid NPs are mainly produced by the EP of monomers in the presence of proteins. The inset shows two examples of hybrid NPs produced through EP. (a) Casein/poly(methyl methacrylate). Figure reprinted in part from ref 79. Copyright 2002 American Chemical Society. (b) Caprolactam-modified casein/poly(butyl acrylate). Reproduced with permission from ref 67. Copyright 2013 Elsevier.

Table 1. Characteristics and Comparison of the Main Proteins Included in Hybrid Protein/Synthetic Polymer NPs with Potential to Be Used in Industrial Applications as Functional Materials. The Main Uses Refer to Pure Proteins

		casein		whey protein		soy protein			zein ^b	hydrolyzed collagen c	
origin		animal		animal		vegetal			vegetal	animal	
		acid ⁵³	Rennet ⁵³	WPC ^{a54}	WPI ^{a54}	SF ^{a55}	SPC ^{a55}	SPI ^{a55}			
industrial composition (%)	protein	90	84	25-89	90-95	52-54	62-69	86-87	97	92	
	moisture	12	12			6-8	4-6	4-6		12	
	fat	2	2	9-1	0.5 - 1	0.5 - 1	0.5 - 1	0.5 - 1		5	
	ash	2.5	7.5			5-6	4-6	4-5	0.5	0.8	
	carbohydrates					30-32	19-21	3-4	2.5		
water solubility			yes ^d		yes ^e		yes		nof	yes ^g	
main uses	food	X		X		X		X	X		
	coatings	X		X		X		X			
	adhesives		X	2	X		X				
	cosmetics								X	X	

"WPC: whey protein concentrate; WPI: whey protein isolate; SF: soy flour; SPC: soy protein concentrate; SPI: soy protein isolate. ^bComposition of zein extracted from corn gluten meal and determined on an anhydrous base. ⁵⁶ Composition of commercial bovine gelatin. ⁵⁷ dSoluble for pH > 7.5–9.5 (IEP 4.6). ⁵³ Decreasing solubility with decreasing pH in the range 7 to 4 (IEP 4.5). ⁵⁸ Soluble in aqueous alcohol solutions of ethanol, methanol, and propanol (IEP 6.2). ^{59,60} Native collagen is insoluble in water (IEP of HC 4–6). ⁶¹

65 many cases high availability at good purity and low cost. 21,22 In 66 addition, proteins contain functionalities such as amine, 67 carboxyl, hydroxyl, and thiol, which provide opportunities for 68 introducing structure modifications. 23,24 Functionalities avail-69 able in proteins are also interesting to design hybrid materials 70 with engineered surfaces in order to have specific properties, 71 that is, wettability, adhesion or fouling. 25-27 One limitation of 72 feedstock proteins is the lack of uniformity typically found in 73 natural materials. However, proteins are usually underused, 74 which adds up to the aforementioned advantages to make 75 these biopolymers a promising eco-friendly raw material 76 (Figure 1). Nowadays, several proteins such as casein (CA), 77 collagen derivative proteins, and zein are employed in many 78 film forming applications at the industrial level as adhesives, 79 leather finishing agents, and in paper coating. These materials 80 present as a main drawback a low resistance to microbial attack 81 and to wet rub, and a reduced flexibility and extensibility, 28 and

for this reason protein conjugation with a synthetic polymer 82 appears as the main explored alternative for facing this issue. 83 The synthesis of hybrid latexes containing proteins has gained 84 technological and also biomedical interest because of the 85 possibilities to modify and improve the properties of these 86 natural substances by incorporating synthetic polymers. 29–31 87

Diverse composite materials including proteins and certain 88 organic/inorganic materials such as metals, ^{32–36} clay, ^{37–39} 89 semiconductors, ^{40,41} different types of polymers, ^{42,43} and more 90 have been reported. ^{44,45} Also, the inclusion of proteins in 91 polymeric particles has been extensively investigated and 92 widely addressed in several recent reviews, ^{46–52} focusing on 93 biomedical applications and on the therapeutic role of the 94 employed protein (immunodiagnostic testing, gene treatment, 95 enzymes immobilization, and controlled release, among 96 others). However, these works did not include waterbone 97 high protein-content particles obtained by a scalable process, 98

Table 2. Summary of Protein/Synthetic Polymer Nanoparticles

protein	protein modification	synthetic monomer	synthesis process/initiators	PC (%)	particle diameter (nm)	applications and potential uses	re
casein	neat	MMA	EF-EP/Cu (II)	100-20	96	leather finishing textiles industries biocompatible	95
	neat	MMA	EF-EP/TBHP	25	82	materials nonspecified	79
	neat	MMA	EF-EP/TBHP	25	83	nonspecified	99
	neac	1,11,11	EF-EP/KPS	20	74	nonopeemeu	
			EF-EP/V-50		70		
			EF-EP/AIBN		69		
			EF-EP/BPO				
			EF-EP/TBP				
	neat	MMA	EF-EP/TBHP	3-50	162-113	nonspecified	2
	neat	BA/MMA	EF-EP/TBHP	3-50	194-122	waterborne coatings	1
	HMC	BA/MMA	EF-EP/TBHP	50	165-218	waterborne coatings	8
	CPL-modified	WPU	blend, hydrogen bonds	900-100	77–46	leather finishing film-forming material	6
	CPL-modified	BA	EF-EP/APS EF-EP/KPS EF-EP/APS + NaHSO ₃	70		leather finishing	6
			EF-EP/KPS + NaHSO ₃		80		
	CPL-modified	VAC/BA/AM	EP/APS	30-55	40-50	waterborne printing ink	8
						leather finishing coatings	
	CPL-modified	BA/MMA/ PDMS	EF-EP/APS		50	textiles food packaging	7
hey protein	denatured	PVAc	blending and cross-linking with MDI	30		adhesives	1
оу	neat	MMA	EF-EP/KPS			packaging	1
	neat	St	EP (emulsifier:SDS)/KPS	200-2000	10-30	thermoplastic hybrid materials	1
						poly(styrene) substitute	
	neat/treated with NaOH solution	BA/MMA	MEP (emulsifiers: alkylphenol ethoxylates and SLS)	1.2-6.2		wood adhesives	1
	acylation with NHS	AM/AA	RAFT polymerization	10, 50	66–162	synthetic hybrid latexes	1
	treated with NH ₄ OH solution	diene and vinyl monomers	EF-EP/oil soluble azo initiators	11–43	paper coatings	128	
	neat/carboxylated	Vinyl monomers	EF-EP/azo initiator	4–60	90-600	organic pigments paper coating	1
	functionalization with <i>N</i> -methylolacrylamide	St	EP/APS	535-2674		wood adhesives	1
ollagen	НС	AA/BA	EF-EP/H ₂ O ₂	15-50	500-880	bioadhesives	1
	neat	MMA	EF-EP/TBHP	25	160	nonspecified	7
ein	neat zein/HMC	BA/MMA	EF- MEP/TBHP	5% (25% total protein)	209	waterborne coatings]

99 which represents a current challenge for producing new 100 ecofriendly latexes for novel applications or as alternative for 101 well-established uses. Nevertheless, this article revises the 102 recent advances on hybrid nanoparticles (NPs) composed of 103 proteins and synthetic polymers with the main goal of 104 improving the sustainability of the industrial development of 105 waterborne particles at different large-scale applications such as 106 coatings, adhesives, films, paintings, papermaking, etc. In this 107 context, this review includes the main characteristics of the 108 employed proteins, the proposed strategies of synthesis, and 109 the involved mechanisms for producing different protein/

synthetic polymer hybrid NPs. Moreover, the main features of 110 the considered hybrid NPs are here revised, including their 111 size, protein content, and the degree of compatibilization of 112 the protein/polymer phases. The synthesis routes of the hybrid 113 NPs are analyzed, and a general mechanism for protein-based 114 particles formation is proposed, which involves emulsifier-free 115 emulsion polymerization (EF-EP) as the most preferred 116 process. Moreover, the advances on protein modifications 117 which have proved to increase the compatibility between 118 protein and synthetic polymer, resulting in an improvement in 119 the final properties of the hybrid materials, are emphasized. 120

121 Finally, the main future perspectives and opportunities of 122 proteins-based NPs are outlined.

Proteins as Functional Constituent of Hybrid NPs.
There is a vast assortment of proteins which have been used in the manufacture of several composite materials. Because of their main features, proteins find most of their applications in the field of medicine and bioengineering. In this review, however, the focus relies on polymer/protein hybrid NPs which have the potential to be industrially applied as functional materials. With this aim, the proteins overviewed here are milk proteins (casein and whey protein), soy protein, collagen, and zein. A few more examples including other proteins can be found at the end of this section. A comparison of the origin, industrial composition, water solubility, and main uses of the proteins as raw materials of these mentioned biobuilding locks are summarized in Table 1.

The ultimate goal of protein incorporation into latex 137 138 formulations is the decline in environment pollution and the 139 increase in biodegradability without degrading the good 140 performance of the synthetic polymers. Therefore, a key factor 141 which defines the industrial scalability of the hybrid latexes is 142 the protein content of the NPs. On the other hand, the 143 performance of the hybrid NPs is the result of the balance 144 between the properties of the protein and those of synthetic 145 polymers, which is expected to be optimized when a 146 compatible system is achieved, that is, when protein and 147 polymer remain in intimate contact. Indeed, the microstructure 148 of the hybrid protein/synthetic polymer involves different 149 species, which give place to the formation of multiphase NPs, 150 including the synthetic polymer, the free protein, and the 151 copolymer containing both components chemically bonded 152 (mostly by grafting). With the aim of comparing and 153 discussing the molecular characteristics of these different 154 materials, three parameters are here defined and will be used 155 throughout this review: (i) protein content (PC) based on a 156 synthetic monomer (eq 1), (ii) degree of grafting of a synthetic 157 polymer (GSP, eq 2), and (iii) degree of grafting of protein 158 (GP, eq 3). Note that, GSP and GP are a measure of the 159 degree of compatibility between both materials (protein and 160 synthetic polymer) in the NPs.

$$PC = \frac{\text{wt of loaded protein}}{\text{wt of total loaded synthetic monomer}}$$
(1)

$$GSP = \frac{\text{wt of grafted synthetic polymer}}{\text{wt of total synthetic polymer}}$$
(2)

$$GP = \frac{\text{wt of grafted protein}}{\text{wt of total loaded protein}}$$
(3)

164 Several works including the synthesis or the development of 165 tools with potential for obtaining hybrid protein/polymer NPs 166 are presented next for the aforementioned proteins. A 167 summary of the total content of protein, employed monomers, 168 and main latex properties of the more representative works are 169 presented in Table 2.

Casein. Casein represents ~80% of the proteins in bovine milk, 62 and corresponds to the fraction precipitated when 172 unheated (raw) milk is acidified to pH 4.6 (isoelectric point, 173 IEP). The protein remnant in solution corresponds to whey 174 proteins. Acid casein is precipitated from skim milk by 175 acidification; on the other hand rennet casein is obtained when 176 the milk is treated with rennet. Casein world production,

together with caseinates, is estimated between 430.000 and 177 460.000 tonnes, with New Zealand, The Netherlands, and 178 Germany being its main producers. Casein can be further 179 classified as edible or industrial. The latter is estimated to reach 180 90.2% of the total global casein and caseinate market in 2025, 181 followed by edible grade casein with 9.8%.

Owing to its excellent film-forming capability and good 183 adhesive properties, casein was used as long as thousands of 184 years ago. Nowadays, neat casein and modified casein still find 185 applications in coatings, adhesives, papermaking, leather 186 finishing, and edible films, among others. 64-73 Moreover, due 187 to its biocompatibility and nonimmunogenic characteristics, 188 casein is a striking material to be used in gene-delivery 189 therapies and drug delivery applications. 52,74-76 However, 190 industrial applications of casein are limited mainly due to its 191 poor water-resistance and mechanical strength. A promising 192 approach to tackle these drawbacks is taking advantage of the 193 high quantity of polar groups present in the structure of casein 194 molecules, such as amino, hydroxyl, and carboxyl groups, 195 which give casein the ability to generate physical interactions in 196 blends with other materials, and to be chemically modified by 197 grafting with natural or synthetic polymers.⁷³ Both in- 198 organic ^{73,77,78} and organic materials ^{24,65,79–85} have been 199 proposed to expand casein applicability.

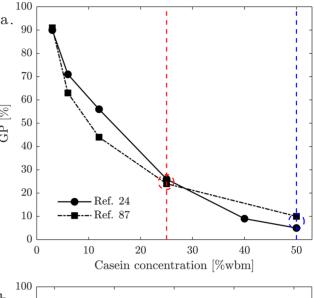
The first published attempts to merge casein with synthetic 201 polymers involved casein incorporation in the EF-EP of 202 conventional monomers initiated with potassium persulfate 203 (KPS) or ammonium persulfate (APS). 86-92 The included 204 monomers were butyl acrylate (BA), vinyl acetate (VAC), 205 butyl methacrylate (BMA), acrylamide (AM), and methyl 206 acrylate (MA), and the polymerizations were carried out in the 207 presence of a wide range of casein amount (PC $\approx 16-220\%$). 208 Despite a grafting mechanism not being reported, it is expected 209 that it involves an unspecific hydrogen abstraction from casein 210 by the sulfate radical species. The grafting extent (measured as 211 the fraction of synthetic polymer containing grafted protein) 212 was studied, and it was found that the GSP increased with the 213 concentrations of initiator, monomer, or casein. One limitation 214 of this method is the use of persulfates initiators that may 215 result in an oxidative degradation of casein, 93,94 producing 216 vellowish products.

These oxidative problems were overcame by Li et al. 95 in the 218 synthesis of casein/poly(methyl methacrylate) core—shell NPs, 219 by grafting polymerized methyl methacrylate (MMA) onto 220 casein, using a trace of copper ions as initiator (PC = 20-221 100%). The authors studied the influence of the copper ion/ 222 casein ratio, finding 9.5/1000 as the optimum ratio, below 223 which the conversion of MMA was considerably reduced, and 224 above which particles became very unstable. These results 225 suggested that casein interacts with copper ions to initiate the 226 graft copolymerization of MMA, allowing the formation of 227 core-shell NPs in the absence of surfactant (i.e., in an EF-EP). 228 The latex particles exhibited a mean diameter of 96 nm. 229 Moreover, a typical kinetic behavior for radical polymerization 230 confined in a preformed microenvironment was observed, 231 which is in agreement with a radical polymerization 232 mechanism involving complexes between the polymer and 233 MMA on copper ion.⁹⁶ A redox initiation approach was 234 followed elsewhere to efficiently graft BA⁹⁷ and MMA⁹⁸ onto 235 casein, using potassium diperiodatonickelate (Ni(IV)), and 236 potassium ditelluratocuprate(III) (DTC), respectively, which 237 form redox systems with casein.

Moreover, another novel synthetic method to create casein/ 240 polymer amphiphilic nanoparticles and overcome such 241 oxidative problems of persulfate initiators was developed in 242 the later works of Li and co-workers 79,99 It consists of an EF-243 EP in which an organic hydroperoxide (ROOH) initiator, such 244 as tert-butyl hydroperoxide (TBHP), reacts with the amino 245 groups present in casein, generating radicals in their N 246 atoms. 79 The radicals formed onto casein amine groups initiate 247 the graft copolymerization of MMA, and as these radicals 248 become water insoluble they assemble into polymer NPs 249 precursors. Following this mechanism, stable latexes with solid 250 content up to 31% were produced, with a NPs diameter 251 around 80 nm, having a narrow particle size distribution, and 252 covalent bonding between core (poly(methyl methacrylate) (PMMA)) and shell (casein), that is, compatibilized NPs. The 254 PC and GSP were 25% and 40-50%, respectively, while the 255 grafting percentage of protein was not reported. 79,99 Alter-256 natively, a grafting percentage defined as the ratio between the 257 weights of the grafted PMMA branches to the weight of loaded 258 casein was reported to be 130-190%, 79,99 indicating that the 259 synthetic polymer (PMMA) is the main component of the 260 grafted material. It is worth noting that the authors of this 261 proposal suggested that it is extendible to any water-soluble 262 polymer chains containing amino groups. Moreover, the 263 copolymers obtained are metal ion-free and surfactant-free, 264 which is highly required for extending this method to many 265 biological applications.

The EF-EP process^{79,99} was also applied by Picchio et al.²⁴ 267 to fabricate waterborne acrylic/casein NPs by using the redox 268 initiation method proposed by Li et al. 79 Afterward, Picchio et 269 al. 100 produced hybrid latexes of acrylic (BA/MMA)/casein 270 NPs through EF-EP^{79,99} with the aim of characterizing the 271 synthesis parameters, the NPs microstructure, and the 272 properties of the films obtained from the hybrid latexes. The 273 authors found that PC (ranged from 3 to 50%) strongly affects 274 the grafting percentage of protein, where GP values are 275 reduced according to the increase of casein concentration 276 (Figure 2a). In consequence, a lower casein content allows the 277 formation of hybrid casein-based NPs with higher GP and 278 increased compatibility. 24 A proper content of casein improved 279 the film hardness, resistance to organic solvents, and soil degradability, and the antiblocking property, without degrading the low minimum film formation temperature (MFFT) typical 282 of acrylic binders for coatings. These results demonstrated that casein-based hybrid latexes are proper materials with potential 284 for being used as binders in waterborne coatings.

However, one limiting factor for acrylic/casein hybrid films 286 reaching a commercial scale is the obtained low GP values when high casein concentrations are used in the latex formulation, which is highly desirable for a better exploitation 289 of this renewable resource. With the aim to maximize the GP, 290 casein acrylation with acrylic acid (AA) was studied (see section Synthesis of Hybrid Protein Based NPs), but a low degree 292 of protein modification was reached. 101 A more robust 293 approach to overcome this difficulty was the use of highly 294 methacrylated casein (HMC) in the latexes formula-295 tion, 81,82,102 which can be obtained through an amine—glycidyl 296 ether reaction.⁸¹ With this last procedure, the GP of the hybrid 297 system was controlled by varying the content of methacrylic 298 groups in the casein backbone. HMC containing up to 40 299 methacrylic groups per protein molecule (theoretical value) 300 was synthesized achieving a GP as high as 82%, 82 which highly 301 overpassed the 10% obtained through the traditional grafting



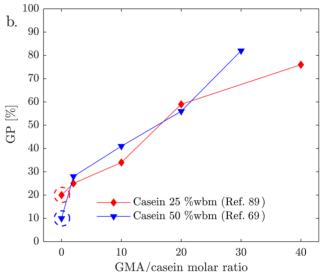


Figure 2. Effect of the casein concentration on the GP when using neat casein in the polymerization of MMA²⁴ and in the copolymerization of MMA/BA¹⁰⁰ (a), compared with the use of HMC (as a function of the methacrylation degree) in the copolymerization of MMA/BA for casein concentrations of 25 and 50% (b). 82,102 The dashed lines in panel a indicate the casein concentrations for which the use of HMC is shown in panel b. The solid content in these latexes was 30%.

method by redox initiation. ¹⁰⁰ A comparison of these values ³⁰² can be found in Figure 2b. Films obtained from high-solids ³⁰³ content hybrid HMC/acrylic latexes exhibited more homoge- ³⁰⁴ neous phase distribution, increased water resistance, and ³⁰⁵ improved block-resistance and open-times comparable to ³⁰⁶ those of organic-solvent based paints (30–40 min). Besides ³⁰⁷ these properties, optimized clear coat formulations with ³⁰⁸ HCM/acrylic hybrid latexes as binders showed also good ³⁰⁹ performance with minimal additive requirements, which ³¹⁰ expands the possibilities of using these latexes as waterborne ³¹¹ eco-friendly binders in industrial applications. ^{81,82,102}

The potential use as adhesives of hybrid HMC/acrylic 313 latexes prepared via the EF-EP route was studied by Aguzin et 314 al. 83 The PC was varied between 5 and 30%, with a solid 315 content of 35%. Moreover, the performance of the final 316

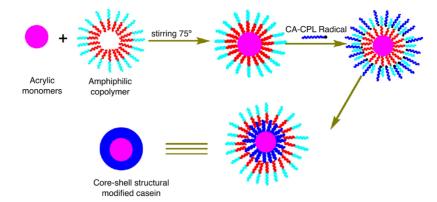


Figure 3. Scheme of the synthesis strategy of CA-CPL/acrylate core—shell NPs via EP. Reprinted with permission from ref 84. Copyright 2015 Elsevier.

317 product produced with neat casein was tested against the use 318 of HMC. Besides maintaining a good adherence of surfaces 319 over time, commercial adhesives are required to be easily 320 removable. For this reason, the removal capacity in alkaline 321 conditions of the hybrid films was tested. It was found that the 322 increased compatibility between the protein and the synthetic 323 polymer, attained by using HMC instead neat casein, resulted 324 in increased removal ability.

A different alternative for casein modification was explored 326 by Ma and co-workers. 65-67,72 These authors studied the 327 ability of caprolactam-modified casein (CA-CPL) to form 328 hybrid protein/polymer NPs in order to achieve an improved 329 applicability as leather finishing agent and film-forming 330 material, without losing its eco-friendly feature. CA-CPL has 331 been used as a leather finishing agent despite having a limited 332 flexibility and water repellency. Ma et al. 66 have blended 333 waterborne polyurethane (WPU) with CA-CPL to produce 334 hybrid NPs mainly formed by the existence of hydrogen bonds 335 between the urethane groups in the WPU and the CA-CPL 336 particles. Particles diameters between 77 and 46 nm were 337 obtained, depending on the WPU/CA-CPL mass ratio used $_{338}$ (PC = 900–100%). The NPs showed improved hydro-339 phobicity and reduced air permeability when compared with 340 CA-CPL particles. Moreover, a superior flexibility and wet-rub 341 resistance was obtained. These characteristics, added up to an 342 acceptable biodegradability, making CA-CPL/WPU NPs a 343 promising eco-friendly film-forming material.

Later, the same authors proposed a second approach to 345 improve CA-CPL performance, preparing latexes with a core-346 shell structure of modified casein/BA via EF-EP.⁶⁷ In this case, 347 the PC was 70%. The effect of different initiators was studied, resulting in the highest grafting extent of monomers for the 349 redox pair KPS-NaHSO3. Particles with a mean diameter of 350 80 nm were obtained, where CPL was the key to forming 351 stable latex particles in the free-emulsifier process. Furthermore, the films exhibited a better hydrophobicity when compared to those produced from pristine casein (contact angle 93.8° versus 23.6°), a good color developing ability, and 355 an improved thermal stability. The authors reported that CA-356 CPL based NPs presented a performance as a leather finishing 357 agent comparable and even better than commercially available products. Finally, the composite showed a preferable 359 biodegradability, making it a promising material for coating 360 applications.

In other work, Ma et al.⁸⁴ synthesized CA-CPL/acrylate NPs by using random amphiphilic copolymers as emulsifier. The

copolymerization of acrylic monomers and CA-CPL was 363 initiated by APS, while a random amphiphilic copolymer, 364 including VAC, AM, and BA blocks, was used as emulsifier. 365 The PC was in the range 30–55%. A scheme of the synthesis 366 route can be seen in Figure 3. The presence of the amphiphilic 367 f3 copolymer helped the formation of micelles swollen with 368 monomer. This hybrid latex has the potential to be an 369 economic and green product as waterborne printing ink 370 binder, leather finishing agent, and coating, among others, due 371 to its biodegradability and excellent performance.

Ma et al. The same also employed the EF-EP with APS as 373 initiator to synthesize hybrid NPs composed by CA-CPL and a 374 synthetic copolymer including BA, MMA, and poly- 375 (dimethylsiloxane) (PDMS). The NPs had a mean diameter 376 of 50 nm with a narrow distribution. During film-formation, a 377 porous structure was developed due to the migration of 378 organic silicone to the coating-air surface (microphase 379 separation). As a result, films with an enhanced water vapor 380 and gas permeation were formed. Moreover, the films showed 381 surface hydrophobicity, biodegradability, and good film- 382 forming properties, being in this way an attractive alternative 383 material with potential to be used in textiles and in the food 384 packaging industry.

Xu et al. 103 synthesized CA-CPL/acrylate/silica latex via 386 double-in situ EP. In this proposal, a condensation reaction 387 occurs between a silane coupling agent (KH750) and a silica 388 precursor (tetraethoxysilane) in the outer region of CA-CPL 389 particles, which were useful to facilitate the emulsification of 390 the acrylate monomers. After the addition of the initiator 391 (APS), a free radical polymerization takes place between silane 392 coupling agent and casein, and between acrylate and casein, 393 resulting in a core—shell structure (mean diameter 80 nm) 394 with an inorganic silica shell and an organic CA-CPL/acrylate 395 core. The hybrid films formed from this latex showed higher 396 hydrophobicity, lower water absorption, superior tensile 397 strength, and reduced flexibility when compared with the 398 protein/acrylate films without silica.

Hollow CA-CPL/acrylate NPs were prepared by Zhang et 400 al. ¹⁰⁴ in a two-step process involving (i) the EP of CA-CPL in 401 the presence of the monomers MMA, BA, and methyl acrylic 402 acid (MAA) initiated by APS; and (ii) the subsequent alkali 403 swelling process in which a migration of the hydrophilic chains 404 in the core of the NPs was produced. The resultant hollow 405 NPs, with a mean diameter of 156 nm, have been tested for 406 opaque coating applications, resulting in tunable transmission 407 between 97% and 57%. Moreover, the coatings exhibited 408

409 excellent antiultraviolet behavior, with almost 0% trans-410 mittance below 200-300 nm light wavelength.

The use of functionalized casein in the hybrid NPs synthesis has shown to be an effective tool to increase the amount of protein in latex formulations, allowing for a better exploitation of this biopolymer resource. A summary of the total amount of casein, casein type, and monomers used in some of the mentioned works is presented in Table 2.

Whey Protein. Whey proteins, mainly obtained as a supproduct of cheese production, 62 represent ~20% of milk proteins. These proteins are a group of globular proteins, which consist primarily of β-lactoglobulin and α-lactalbumin. Other components are immunoglobulins, bovine serum all all Discrete land and all lactoperoxidase. They can be classified as whey protein concentrate (WPC) or whey protein as whey protein concentrate (WPC) or whey protein as from 25 to 90%, and for WPI is higher than 90%. Supproduct to hody requires, being in this way an excellent product to the main applications of this protein family are found in the supproduct in the supproduct in the supproduction of whey protein was supproduction of whey protein was supproduction around 240 million tonnes in 2014, with a value of USD 5.4 billion.

As a byproduct of cheese manufacture, whey protein was 434 considered a polluting effluent from the dairy industry, and 435 discarded in rivers or lands without any treatment. It is an 436 important environmental problem due to its high production 437 volume and organic content, making necessary the expansion 438 of its industrial exploitation, and opening the possibility to 439 introduce whey protein in a second step of the chain values 440 and to produce more eco-friendly materials.

Whey protein is water-soluble—up to 40% by weight—and ti is known to be capable of forming elastic films, which have a good oxygen barrier and moderate moisture permeability. 107 the Beyond the food industry, other current applications include senior environmentally safe adhesives 62,108 (unlike casein, the use of whey protein for adhesives is relatively new), 62,109 biodegradable coatings, 45,110 drug delivery and gene-therapy, 49,50,111,112 among others. 70,116,117 However, due to to the hydrophilicity, whey protein-based materials exhibit poor mechanical and water-vapor barrier properties. 107 The research done in whey protein/synthetic polymer hybrid NPs with an criefly some developed which can be useful for its future timplementation in obtaining of this type of NPs.

One example of these tools is presented in the work by Chan 456 et al., 118 in which polymerizable whey protein was prepared 457 through an amine-based reaction with methacrylic anhydride 458 to introduce (meth)acrylamide moieties onto the protein 459 backbone. Methacrylic anhydride is produced as a byproduct 460 of the amine-anhydride reaction, and its presence did not 461 degrade the mechanical properties of the final product, so 462 purification steps may be avoided. In this work reactive whey 463 protein was obtained, which allows an improved control over 464 chemical cross-linking density in a subsequent free-radical 465 polymerization step. Despite the aim of this research being the 466 synthesis of protein-based thermoset elastomers, the reported 467 method to obtain polymerizable whey protein can be a helpful 468 tool to efficiently develop new hybrid NPs via free radical EP, 469 in much the same way as the use of HMC, where the degree of 470 compatibility with the synthetic polymer is controlled by the 471 presence of methacrylic groups.

Sharma and Luzinov¹¹⁹ produced bioplastics by blending 472 WPI with natural rubber latex (mainly composed of cys-1,4- 473 polyisoprene), and WPI with egg white albumin. They found 474 that the addition of about 10% of any of these biopolymers to 475 the protein improved the composite toughness without 476 compromising the whey protein strength and stiffness. The 477 compatibilization between the protein and rubber/egg white 478 albumin has not been proven in this work. However, the 479 authors suggest that compatibilized latex of natural rubber 480 particles and WPI may have been obtained by the reaction 481 between amino acids groups in both materials.

Gao et al. 120 developed an eco-friendly adhesive of the 483 family of aqueous polymer solution-isocyanate (API) adhe- 484 sives, which are primarily composed of a water based glue and 485 an isocyanate cross-linking agent. In this case, the adhesive 486 formulation included a polyvinyl acetate (PVAc) latex, a 487 solution of thermal denatured whey protein, and a methylene 488 diphenyl diisocyanate (MDI) as cross-linking agent (PC = 489 230%). Despite a latex characterization not being performed, it 490 is assumed that core-shell NPs were formed by blending of 491 the PVAc (core) and the denatured protein (shell). Then, 492 these NPs were cross-linked by the reaction of MDI with the 493 amino and carboxyl groups of the protein. Another explored 494 synthesis route in this work consisted of the addition of 495 poly(vinyl alcohol) (PVA) (3-18 wt %) during the 496 denaturation process of the protein, which resulted in an 497 increased bond durability of the adhesive, due to the 498 interactions between PVA and WPI. It is assumed, as other 499 authors suggested, 121,122 that PVA was physically cross-linked 500 with the protein via hydrogen bonding, thus leading to 501 compatibilized NPs. Moreover, the effect of time and 502 temperature variations of the denaturation process was studied. 503 Generally, it was found that the unfolding of the protein 504 globular structure induced by the denaturation process 505 improved the bonding strength of the adhesive. Also, the 506 addition of nanoscale CaCO₃ powder further increased the 507 bonding strength and durability of the whey protein based API 508 adhesive. Similar strategies varying the plasticizer and cross- 509 linking agent species were presented in a patent of the same 510 authors.

Soy Protein. Soy protein is a relatively inexpensive protein, 512 generated as a byproduct of the soy oil industry. This protein is 513 commercially available in three kinds of products: (i) soy flour 514 (SF), composed by 50% of proteins and carbohydrates; (ii) 515 soy protein concentrate (SPC), composed by 70% of protein 516 and nonwater-soluble carbohydrates; and (iii) soy protein 517 isolate (SPI), having a protein content of ca. 90%. 518 Approximately 90% of soy proteins are storage proteins, 519 mainly globulins, 133 with an abundance of essential amino 520 acids, whereby they have been used mostly in the food 521 industry. In addition, soy protein contains an important 522 proportion of amino acids with polar and reactive groups such 523 as lysine (6.8%), arginine (7.7%), histidine (3.3%), and 524 cysteine (2.4%), 134 which make the protein suitable for 525 chemical modifications.

Pure soy protein materials are rigid, brittle, with low thermal 527 stability, poor resistance to moisture absorption and water 528 immersion, and highly susceptible to microbial attack. 529 Therefore, many efforts have been made to improve their 530 mechanical and thermal properties, and moisture resistance. In 531 this context, physical and chemical modifications of soy protein 532 have been widely investigated, including pH adjustment, 533 thermal treatments, incorporation of plasticizers, cross-linking 534

535 reactions, and production of composites by blending with 536 synthetic polymers, which have been extensively re-537 viewed. Also, strategies of chemical modification by 538 grafting with synthetic polymers were reported. 141

Some of these grafting modifications involve the radical 540 polymerization in aqueous dispersion of hydrophobic vinyl 541 monomers, such as MMA, styrene (St), ethyl acrylate, ethyl 542 methacrylate, BMA, hexyl methacrylate, and acrylonitrile 543 (AN), in the presence of the soy protein. 142-149 In general, 544 high PC were used ranging from 50 to 500%, and different 545 initiators such as persulfates, redox systems (ascorbic acid/ 546 KPS), and ammonium cerium nitrate were employed. 547 Although none of these works showed NPs characterization, 548 it can be deduced from the synthetic procedure that hybrid 549 latexes were probably obtained. Instead of the latex application, 550 the grafted copolymer was isolated by solvent extraction and ssi used as either hybrid thermoplastics 144,146,149 or in the ss2 formulation of composites. 145,148 In general, a GSP between 553 15 and 80 was obtained depending on different polymerization 554 parameters such as pH, temperature, polymerization time, and 555 monomer and initiator concentrations, which affected the

In a simple approach to incorporate soy protein to synthetic sss latex, some works explored the physical mixing of both materials. However, nonuniform latexes can lead to heterogeneous films, due to the phase segregation of materials, affecting the application properties. Therefore, a synthetic procedure that promotes the formation of grafted copolymer is desirable to make both phases compatible.

Also, the synthesis of waterborne soy protein-based NPs 565 with an orientation to industrial applications was investigated 566 employing scalable methods such as EP and MEP. Kisku and 567 Swain 124 studied the synthesis of soy protein/PMMA latexes 568 by EF-EP of MMA employing a low content of soy protein 569 (PC ranging from 1 to 5% as emulsifier) and KPS/ammonium 570 ferrous sulfate as radical initiator. The materials obtained after 571 latexes precipitation and drying showed a uniformed 572 distribution of soy protein. These PMMA/soy protein films 573 exhibited reduced oxygen permeability with better flame-574 retardant properties than pure PMMA films. In addition, 575 hybrid materials showed high biodegradation in activated 576 sludge water, with a weight loss of about 65% for the material 577 with the highest PC. Similarly, Liu et al. 125 reported the use of 578 EP for synthesizing St based latexes in the presence of a high 579 content of SPI (PC ranging from 200 to 2000%) with low solid 580 content (5 to 7%) and KPS as initiator. Sodium dodecane 581 sulfonate (SDS) was also incorporated into the emulsion 582 recipe in order to improve the emulsifier capability of this 583 protein. Ionic surfactants, such as SDS, denature the soy 584 protein by disrupting the hydrophobic and electrostatic 585 interaction, resulting in a partially unfolded protein structure. 586 The EP with high content of SPI generated very small latex 587 particles, with diameters between 10 and 30 nm, and coreshell morphology. Figure 4 schematizes the formation of core-589 shell NPs where soy protein is physically linked to the poly(St) 590 core by the interactions with the SDS molecules. This SPI-591 based thermoplastic showed good mechanical properties and 592 importantly improved water resistance.

In all these articles, the formation of grafted copolymer between soy protein and the synthetic polymer was not evaluated. However, it is presumed that some grafting could have happened during these polymerizations, for which persulfates were used as initiators.

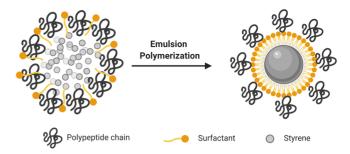


Figure 4. Schematic model of the core—shell structure of soy protein/poly(St) NPs. 125

Zheng et al. 130 have produced an ambient temperature- 598 curable wood adhesive based on a latex of SF/poly(St) 599 synthesized by EP and a polymeric methylenediphenyl 600 diisocyanate as cross-linking agent. SF was previously exposed 601 to a combination of enzymatic and acid treatments to 602 hydrolyze the carbohydrates and to denature the protein, 603 followed by a chemical modification with N-methylolacryla- 604 mide. Functional groups of soy protein, such as carboxylic 605 acids, react with the -OH of N-methylolacrylamide producing 606 a polymerizable protein that promotes the graft copolymeriza- 607 tion with the St during the EP. The concentration of St was 608 varied from 2 to 10% (based on the weight of modified SF) 609 giving a PC between 535 to 2674%. The EP was carried out at 610 75 °C in two consecutive steps: (i) a batch polymerization 611 containing all the modified SF and 1/4 of the mixture of St and 612 initiator (APS), during 20 min; and (ii) a semibatch 613 polymerization dosing the rest of St and initiator during 4 h. 614 In this form, latex with a final solid content between 20 and 615 25% was produced. The formation of SF/poly(St) grafted 616 copolymer was qualitatively confirmed by FTIR but without 617 quantifying the degree of grafting. These hybrid latexes were 618 mixed with a polymeric methylenediphenyl diisocyanate that 619 reacted with hydrophilic groups of the SF and the -OH in the 620 wood, producing a cross-linking adhesive at ambient temper- 621 ature, with improved water resistance and thermal stability. 622

Recently, the synthesis of acrylic/SPI hybrid latexes by 623 emulsion copolymerization of acrylic monomers (MMA, BA, 624 and 2-(methacryloyloxy)ethyl acetoacetate), was reported in 625 order to produce waterborne wood coatings. 153 Latex with 626 variable content of SPI (PC = 1.7-11.4%) and with theoretical 627 solid content between 27 and 29% was produced. Soy protein 628 was previously treated by heating in NaOH solution, followed 629 by the addition of sodium metabisulfite to break the secondary 630 and quaternary structures of the protein. The latex synthesis 631 involved a first semibatch EP of acrylic monomers, using a 632 mixture of sodium dodecyl sulfate and alkylphenolpolyoxy- 633 ethylene ether (OP-10) as surfactants; followed by a second 634 polymerization stage with the addition of the modified soy 635 protein solution, after the monomer feeding was completed. 636 The unreacted monomer, at the moment in which soy protein 637 was added, took part in the formation of grafted soy protein/ 638 acrylic copolymer, giving a higher GSP (ranging from 1.1 to 639 7.5) with the increase of soy protein concentration. The 640 average diameter of hybrid NPs was in the range 35-56 nm, 641 which is lower than the NPs diameter of analogous latex 642 without soy protein (60 nm) possibly indicating the formation 643 of a secondary particle population. The application properties 644 of these hybrid latexes as wood adhesive (wear resistance, 645 adhesion, gloss, and pencil hardness) improved with the 646 protein concentration. The same trend was observed for the 647

648 mechanical properties, except for the hybrid material with the 649 highest PC (11.7%) that was very brittle.

The MEP method was also used to synthesize soy protein/ 651 acrylic NPs by copolymerization of BA and MMA, in order to 652 produced wood adhesives. 126 Soy protein with a previous 653 denaturalization process, by heating in a NaOH solution, or 654 neat soy protein (without denaturalization treatment) was 655 incorporated with a mix of alkylphenol ethoxylates and sodium 656 lauryl sulfate (SLS) as surfactant. This synthetic strategy 657 allowed the production of latex with high solid content, around 658 37%, but with a relatively low concentration of soy protein (PC 659 = 1.2-6.2%). Films obtained from latexes synthesized with 660 neat soy protein showed protein segregation, while an 661 improved phase compatibility and homogeneity resulted 662 when denaturalized soy protein was used. In addition, close 663 mechanical performance to commercial wood adhesives, such 664 as melamine-urea-formaldehyde and phenol-formaldehyde 665 resins, were obtained, with the important environmental 666 advantage of avoiding the formaldehyde emission.

Another synthetic strategy to produce soy protein hybrid 668 NPs was reported by Bhattacharjee and Bong, 127 using 669 reversible addition-fragmentation chain transfer (RAFT) 670 polymerization. In this work, soy protein hydrolyzates-based 671 graft copolymers with poly(AM) and poly(AA)—which form 672 NPs when dispersed in an aqueous medium—were produced. 673 To obtain the grafted copolymers, benzylthiocarbonate 674 moieties were incorporated on the protein surface by acylation 675 of free amino groups with N-ydroxysuccinimide (NHS), 676 creating a protein macrochain transfer agent. Then, RAFT 677 polymerization was carried out in dimethyl sulfoxide (DMSO) 678 with 2,2'-azobis(2-methylpropionitrile) (AIBN) as initiator, 679 and using two concentrations of the protein macrochain 680 transfer agent, 10 and 50%. It was hypothesized that the NP 681 formation by dispersing grafted copolymers in aqueous 682 medium was driven by protein/protein aggregation. In this 683 form, NPs with average diameters between 66 and 162 nm 684 were obtained depending on the composition of grafted 685 copolymers.

Patents also reported the industrial application of soy 687 protein/synthetic polymers NPs. Riley and Coco described 688 the production of soy protein/synthetic polymer latexes with a 689 high solid content (30-43%) for their use as a binder in paper 690 coating formulation. Hybrid latexes were synthesized by EF-691 EP of diene monomers (e.g., 1,3 butadiene, isoprene) and vinyl 692 monomers (e.g., St, alpha methyl styrene) in the presence of 693 soy protein (PC = 11-43%), using oil soluble azo initiators 694 (e.g., 2,2'-azobisisobutyronitrile) and a chain transfer agent 695 (e.g., tert-dodecyl mercaptan). Also, soy protein/acrylic hybrid 696 NPs were fabricated and used as replacement for conventional 697 inorganic pigments, commonly employed in paper coating 698 formulations. 129 The hybrid NPs are formed by the EF-EP 699 process of a monomer—or mixture of monomers—such as St 700 or MMA, and variable concentration of carboxylated SPI (PC 701 = 4-60%). The preferred initiators are azo nitriles. Semibatch 702 polymerization strategies were used, feeding a pre-emulsion of 703 monomers and soy protein. The particle size control was 704 governed by the protein level contained in the pre-emulsion. In 705 this way, final average particle diameters of the obtained hybrid 706 latexes ranged from 90 to 600 nm, the bigger particles being 707 synthesized at lower concentration of protein. The use of these 708 hybrid latexes in combination with an inorganic pigment in 709 paper coating formulations improved gloss and ink holdout, as 710 compared to a coating in which the pigment composition

consisted only of an inorganic material. Moreover, an 711 improved water resistance, water holding, and a potential for 712 reduction in the coating sticking during the paper finishing 713 process were obtained as compared to conventional inorganic 714 polymeric pigments. It is worth noting that the patented 715 method was also successful employing casein instead of soy 716 protein. Moreover, latex containing SPI were produced by the 717 EP of VAC and dioctyl maleate in order to obtain adhesives. 154 718 Depending on the adhesive application, soy protein was added 719 during polymerization, after polymerization, or both. However, 720 the concentration of protein incorporated during the polymer- 721 ization was low, around 1%, and other emulsifiers (anionic 722 emulsifiers based on soybean oil) were used to obtain stable 723 lates.

Collagen. Collagen is the most abundant animal-protein. It 725 provides the principal body-structural and mechanical 726 support. There are 20 genetically distinct members of the 727 collagen family. The majority are fibril forming collagens: type 728 I (tendon, skin, and bone), type II (cartilage), and type III 729 (vasculature and skin). These collagen types are found forming 730 fibrillar structures, which are an essential part of tissue 731 structure. 156-159 Collagen is a highly cross-linked material 732 which is usually insoluble in water and many solvents. The 733 molecule is formed by three chains held together in a triple- 734 helical structure by multiple physical interactions. 160–162 The 735 temperature of thermal denaturation of collagen depends on 736 the pH of the environmental medium, water content, and 737 degree of cross-linking. Hydrolyzed collagen (HC) is a 738 collection of peptides with low molecular weight (between 3 739 and 6 kDa) that can be obtained by enzymatic, acid, or alkaline 740 hydrolysis. HC can be extracted from different sources such as 741 porcine or bovine. HC properties such as molecular weight of 742 the peptide chain, functional activity, and solubility are affected 743 by the type and source of extraction.⁶¹ Moreover, gelatin is 744 obtained by partial acid/alkaline hydrolysis or by thermal or 745 enzymatic degradation of collagen. Gelatin molecules consist 746 of repeating sequences of proline, alanine, and glycine amino 747 acids, which are responsible for the triple helical structure of 748 gelatin. The unique triple helix structure, consisting of three 749 polypeptide chains, is responsible for the high stability of 750 gelatin. 163

According to a new research report by Global Market 752 Insights Inc., the global collagen market is anticipated to 753 exceed USD 6 billion by 2026. The major contributors to 754 the rising demand for gelatin and HC are the food, cosmetic, 755 and beverage industries, due to the gelling capacity 756 (texturizing, gel formation, water binding capacities and 757 thickening) as well as the surface covering (foam formation 758 and stabilization, emulsion, adhesion and cohesion, film 759 forming capacity and protective colloid function), and 760 hydration properties (solubility and swelling) of these 761 materials. 165,166

Different blends^{167–174} and hydrogels^{175–177} involving 763 collagen derivatives with synthetic polymers were reported, 764 but only a few methods have been informed on the production 765 of hybrid NPs. One of the first reports about the synthesis of 766 hybrid NPs was the work by Ramaraj et al., 178 which presented 767 the preparation of collagen based composite by blending a 768 latex of poly(MMA-co-n-BA) with different contents of gelatin. 769 The cross-linking effect of glutaraldehyde on gelatin and 770 diallylphthalate on MMA-co-n-BA polymer was investigated, 771 reporting a higher film tensile strength for those fully cross- 772 linked composites. Also, the gelatin content of the film 773

I

774 composite improved the tensile strength. Deselnicu et al. 174 studied the rheology of blends of vinyl-acrylic copolymer 776 latexes and HC aqueous solutions, which is of interest in the 777 acquisition of biomaterials in industry, agriculture, and 778 cosmetics among others. Moreover, Ye et al. 179 reported the 779 synthesis of core—shell NPs composed by ${\rm AgTiO_2}$ (core) and 780 PMMA with grafting of the HC (shell). The NPs were 781 synthesized by graft polymerization and water-in-oil EP.

The EF-EP proposed by Li et al., 79 previously addressed, 783 was also employed to produce collagen/PMMA hybrid NPs. 784 Following this strategy, Luque et al. 131 recently reported the 785 synthesis of hybrid collagen/acrylic NPs by EF-EP of AA and 786 BA with varied HC content (from 15 to 50%) and H₂O₂ as 787 hydroperoxide initiator. Despite the amount of collagen 788 incorporated being significant, with this method just a small 789 fraction could be grafted, with a GP of approximately 10%. In 790 this case they demonstrated that EF-EP resulted in an adequate 791 technique for synthesizing stable collagen/acrylic latexes with 792 particle size exceeding 500 nm. The obtained latexes presented 793 excellent film formation capability, giving place to materials with different mechanical and adhesive properties, depending on the percentage of collagen, neutralization degree, and 796 moisture content. This characteristic opens the opportunity to obtain switchable biobased materials, where the adhesion is

798 controlled by their moisture content.
799 On the other hand, Zhang et al. 180 synthesized AA/gelatin 800 core—shell NPs via template polymerization, in which occurs 801 the simultaneous polymerization of AA and gelatin. At the 802 reaction conditions (pH = 2) the poly(acrylic acid) is 803 insoluble, allowing the formation of NPs. In this case, the 804 system APS/N,N,N',N'-tetraethylmethylene diamine was used 805 as initiator, while a selective cross-linking of the shell protein 806 was produced with glutaraldehyde in order to lock the NPs 807 structure. Despite the obtained solid content of the latex being 808 very low (3%), the resultant cross-linked PAA/gelatin NPs 809 were pH-responsive and the gelatin shell made the NPs good 810 candidates to be employed in bioapplications.

Gelatin has been employed to produce hybrid NPs for photographic coating applications. The synthesis method involved obtaining polymeric latex by EP having NP diameters between 30 and 500 nm and a further blending with gelatin solutions having low content of Ca ions. Moreover, compatibilized hybrid gelatin/polymer NPs were also achieved, for which covalent bonding between the amine or sulfhydryl groups of gelatin and the carbonyl groups of the polymer was attained through the use of carbamoylonium salts. On the other hand, the EP of hydrophobic monomers in the presence gelatin (protein content up to 30%) was also employed in aqueous photographic coating compositions.

Zein. Zein is the major prolamin of corn and encompasses 824 50% of the total protein of endosperm. Ref. Zein is a 825 heterogeneous mixture of polypeptides classified into four 826 fractions named δ -, γ -, β -, and α -zein (70–85% of total zein) 827 based on their amino acid sequences, molecular weight, 828 solubility, and charge. From 2010, commercial zein was 829 produced as a byproduct from corn gluten meal by two main 830 companies, Flo Chemical Corp. (Ashburnham, MA) and 831 Showa Sangyo (Tokyo, Japan). Then, zein as a byproduct of 832 the ethanol process was introduced to the market by Prairie 833 Gold, Inc. (Bloomington, IL). Nowadays, commercial zein is 834 mostly α -zein with a molecular weight between 19 and 22 kDa, 835 and its composition may vary according to the acquisition 836 method. Ref.

Compared to other plant-based proteins, zein has significant 837 and unique properties due to its amino acid sequence, which is 838 mainly composed by nonpolar and uncharged amino acids as 839 glutamine (21–26%), leucine (20%), proline (10%), and 840 alanine (10%). Possibly due to this structure, zein is 841 relatively heat- and pH-stable, and water insoluble, but soluble 842 in aqueous ethanol. Although this protein has both hydro-843 phobic and hydrophilic side domains in its structure, it is 844 frequently considered to be a hydrophobic protein due to its 845 insolubility in water. 185,187

Since zein is not an indispensable protein for human 847 consumption due to its poor water solubility and low 848 nutritional value (it lacks in the essential amino acids as lysine 849 and tryptophan)60 and it is obtained as a byproduct of well- 850 established processes, this biopolymer has attracted many 851 industries to explore its potential as a feedstock material. In 852 addition, the attention given to zein-based polymers is 853 primarily attributed to their ability to form films, biocompat- 854 ibility, biodegradability, and ability to encapsulate both 855 hydrophobic and hydrophilic actives. Moreover, zein was 856 approved as a generally recognized as safe (GRAS) material by 857 the United States Food and Drug Administration. 186,188,189 As 858 a result, reports on zein-based biomaterials for cosmetic, 859 biodegradable plastics, coatings, tissue engineering, biomedical 860 applications, and control drug delivery applications have 861 appeared, with zein being in different formats such as fibers, 862 micelles, nanoparticles, nanospheres, or films, where zein is 863 applied pure, in physical mixtures with other materials or as 864 part of a hybrid material. 52,184,186,190–199 However, much less 865 has been done with the incorporation of zein in the synthesis 866 of hybrid NPs with synthetic monomers. One exception is the 867 work by Allasia et al., 132 where a dispersion of hybrid acrylic/ 868 zein/casein NPs with average diameter of 209 nm and a solid 869 content of 35% were obtained through emulsifier-free MEP. 870 The proposed synthesis strategy involved the use of the 871 stabilization power of casein to form stable miniemulsion 872 droplets, which contain both acrylic monomers (BA and 873 MMA) and zein. Two kinds of initiators were employed in 874 order to have thermal initiation with KPS and the redox 875 TBHP/NH₂-casein radical formation.⁷⁹ The hybrid material 876 had a total PC of 25%, in which only 5% of that protein was 877 zein. Despite the low concentration of hydrophobic protein, 878 the hybrid material showed improved water resistance (2.4 879 times less of water absorption) and a more hydrophobic 880 surface (contact angle 83°) compared with zein-free hybrid 881 material (Figure 5), due to the high compatibility achieved 882 f5 between the different phases (GP = 75%). In addition, these 883 hybrid materials were partially biodegradable in compost, 884 losing 15% of their initial weight after 14 days of being buried. 885

On the other hand, zein based particles could be formed by 886 stabilizing it with different biomaterials using the antisolvent 887 method. Biomaterials such as tea polysaccharide (TPS), 200 888 pectin, 201 carboxymethylated short-chain amylose (CSA), 889 alginate, 203 sodium caseinate (SC), 204–208 carboxymethyl 890 chitosan (CMCS), 190 chitosan, 209,210 starch, 211 and tannic 891 acid 212 act as electrostatic and steric stabilizer for zein-based 892 particles, offering a good challenge for obtaining fully biobased 893 NPs with excellent final properties based on the synergy of the 894 combined biopolymers. A summary of these hybrid NPs, 895 including the biomaterials used, composition, size, particle 896 yield (ratio of NPs produced to initial biomaterial amount), 897 and its applications are presented in Table 3. As can be seen, it 898 t3 is possible to synthesize various NPs based on zein, having 899

Figure 5. Water absorption of hybrid casein based film with and without zein and static contact angles of water onto its surfaces. The contact angle of film obtained from casein/zein/acrylic hybrid NPs is 83°, close to that of pristine acrylic film.

900 different diameters and different zein/biomaterial ratios with 901 high particle yields (70–90%). Figure 6 shows image examples 902 of the morphology of the hybrid NPs of zein-SC, zein-CMCS, 903 and zein/tannic acid. In addition, some research has focused 904 on the development of a procedure that can be scalable and 905 economically attractive for the industry to obtain stable NPs 906 using some variants of the antisolvent precipitation method, as 907 flash nano precipitation (FNP), 213 new four-stream FNP 908 configuration, 189 dual channel microfluidization method, 214 909 and the combination of FNP with spray-drying as fabrication 910 scalable processes. 215 Although most of the applications of the 911 aforementioned NPs are aimed at control drug delivery 912 systems (Table 3), these zein based NPs could be employed

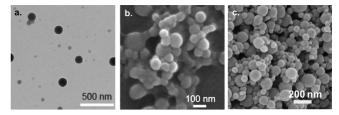


Figure 6. Examples of the morphology of some zein/biomaterial hybrid NPs. (a) TEM image of zein/SC NPs. Reprinted in part with permission from ref 205. Copyright 2017 Dovepress. (b) SEM images of VD3 encapsulated zein-CMCS NPs complex with calcium. Reprinted from ref 190. Copyright 2012 American Chemical Society. (c) Field emission scanning electron microscopy image of zein/tannic acid (1:0.2). Reprinted from ref 212. Copyright 2015 American Chemical Society.

as precursors for the formation of new protein/synthetic 913 polymer hybrid NPs, paving the way for the use of this 914 biomaterial in other industrial applications.

Other Proteins. A few more examples of hybrid protein/ 916 polymer NPs—or available tools for their development— 917 employing other less common proteins are given in this 918 section. For each case a brief description of the protein, the 919 method of synthesis of the hybrid NPs, and the NPs 920 application is presented. An interesting method for the 921 development of protein/polymer NPs is given in the work 922 by Xue et al., 216 in which hybrid NPs including BSA were 923 synthesized and used to stabilize water-in-water (w/w) 924 emulsions. Albumins are a family of water-soluble proteins 925 which can be found in egg, milk, and blood. This protein 926 contains many ionizable groups, which makes albumin one of 927 the most soluble plasma proteins.²¹⁷ It has found many 928 applications in medicine, food industry, and in research studies 929 as a protein model. Emulsions are interesting systems for the 930 synthesis of functional materials, and in particular w/w 931 emulsions are attractive for an eco-friendly process, because 932 the use of organic solvents is avoided. However, the 933

Table 3. Summary of Zein Based NPs Synthesized Using Some Variants of the Antisolvent Precipitation Method

	•	·		-	
system	zein/X ^a ratio	particle yield (%)	size (nm)	applications	ref
zein/TPS	1:0.07 -1:0.7	20-70	130-495	anticancer drug delivery system	200
zein/pectin	1:0.32	>92	250	controlled release of dietary supplements	201
zein/CSA	1:2 - 1:13	nonspecified	140-200	oral drug delivery	202
zein/alginate	1:0.25	95	160	nanodelivery systems for bioactive molecules	203
Zein/SC	1:1	nonspecified	110-500	encapsulation and/or controlled release of therapeutic drugs, bioactive components, and food pigments	205 [†]
	1:1-5:1	nonspecified	100-130		206
	1:0.1-1:2	nonspecified	120-150		204
	1:0.75 -1:1.25	74-82	177-240		207
zein/CMCS	1:0.5-1:2	nonspecified	86-200	encapsulation and controlled release of fat-soluble vitamin	190 [‡]
zein/chitosan	1:0.05 -1:0.2	nonspecified	211-862	controlled release of bioactive agent	209
	1:0.02 -1:0.1	nonspecified	60-1000	pickering emulsion stabilizer	210
zein/SC with PDC- CA	1:0.25 -1:0.8	nonspecified	141-421	oral drug delivery	208
zein/starch	1:2-1:10	nonspecified	115-564	pickering emulsion stabilizer	211
zein/tannic acid	1:0.1-1:0.5	nonspecified	96-250	pickering emulsion stabilizer	212

[&]quot;X represents the biomaterial used to produce the zein-based particles. Corresponding NPs morphologies are shown in †Figure 6a, ‡6b, and |6c, respectively.

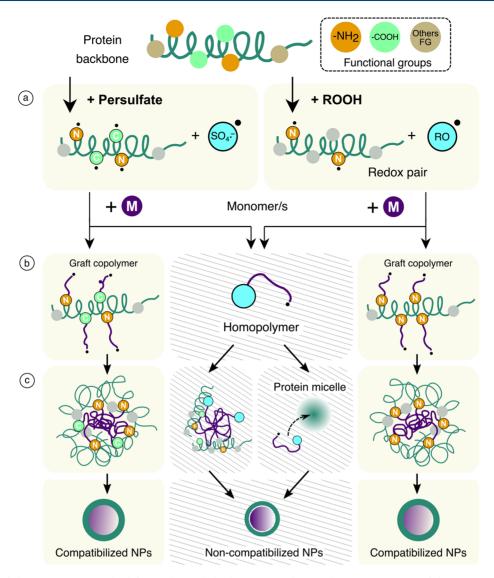


Figure 7. Scheme of the main routes involved for producing hybrid protein-synthetic polymer NPs; EF-EP of the protein in the presence of a monomer or mix of monomers, using persulfates or hydro peroxides as polymerization initiators. The main stages of the process are initiation (a), propagation in water phase (b), and NPs nucleation (c).

934 stabilization of w/w emulsions is not straightforward due to 935 the low tension of water—water interfaces and the associated 936 large interfacial thickness. Xue et al., 216 have demonstrated that 937 methoxy polyethylene glycol (mPEG)/BSA NPs have the 938 ability to stabilize w/w emulsions. In their work, mPEG/939 protein conjugate was achieved by reacting BSA protein with 940 mPEG-acetaldehyde via Schiff base chemistry at a pH near to 941 the protein IEP. Moreover, they showed that the NPs diameter 942 (200–500 nm) can be easily tuned with the reaction time.

A method to fabricate core—shell particles, where the shell is made of an albumin and the core is a water-insoluble material, was reported in a patent by Jaromir and Atkins. The method consists in dispersing a water-immiscible material in an aqueous solution of a coagulable protein, such as egg albumin step involves the coagulation of the protein onto the surface of the particles as a result of an increase in the temperature of the particles. The rise in temperature is achieved by the previous incorporation of carbon black infrared absorbers in the water-simmiscible material, and a further exposure to infrared radiation for a certain time. The size of the core—shell

particles is set by the size of the water-immiscible particle 955 (liquid or solid) in the oil-in-water emulsion. This method was 956 proposed a priori as a useful tool to encapsulate materials, but 957 depending on the employed materials it could be suitable for 958 other ends.

Another interesting combination of an animal-derived 960 protein with other materials (such as synthetic polymers and 961 inorganic compounds) is given by silk protein-based composite 962 materials. Silk proteins are mainly obtained from the silkworm 963 specie *Bombyx mori*, and from spiders such as the Araneus 964 diadematus or Nephila clavipes. Silk protein materials have 965 been used for applications such as papers, strings, textiles, and 966 tissues due to the excellent mechanical properties and good 967 biocompatibility of silk. Because these materials are 968 acquired naturally, their application at the industrial scale has 969 not been achieved. However, spider silk-like proteins can be 970 produced using recombinant DNA technology. Hybrid 971 particles composed of *Bombyx mori* fibroin and poly- 972 (acrylonitrile-co-methyl acrylate) have been obtained in the 973 work by Sun et al. 221 These particles were simply formed by 974

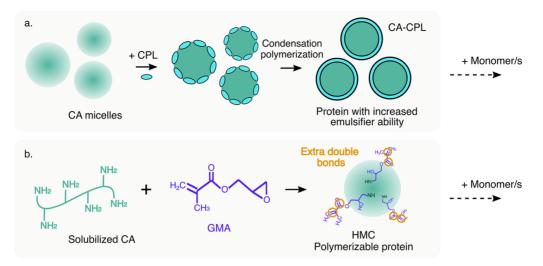


Figure 8. Scheme of CA modification strategies previously to EP process. Route employed for obtaining casein with an increased emulsifier capability through its reaction with CPL (a), 67,72,84 and HMC by the glycidyl ether reaction of CA (b). 82,132

975 blending both components, establishing hydrogen bonds 976 between the protein and the polymer particles.

Synthesis of Hybrid Protein Based NPs. The main 978 applications of the hybrid latexes were film forming material, 979 coatings, adhesives, leather finishing agents, pigments, and 980 textile. A summary of the synthesis method, PC, mean diameter, and applications of the NPs are summarized in Table 2. The EP process has been the main route to obtain 983 hybrid protein/synthetic polymer latexes. Most of the proposed strategies involve a free-emulsifier mechanism, in which the amphiphilic character of proteins or the grafted 986 molecule formed with the synthetic polymer self-assemble to 987 form the precursors of core-shell compatibilized particles in 988 an EP. Also, an important improvement in the performance of 989 hybrid latex is reached when compatibilization between the 990 protein and synthetic polymer is enhanced by protein 991 modification previous to the EP. In this direction, this session 992 summarizes the main features of EF-EP, involving hybrid NP 993 formation (Figure 7), and the more efficient and scalable 994 protein modification proposals (Figure 8).

The EP process includes a monomer-in-water emulsion 996 leading, with the addition of a radical initiator, to a dispersion 997 of colloidal polymer particles (latex). Several mechanisms have 998 been proposed to explain the formation of particles during EP. 999 Among them, the homogeneous nucleation mechanism was 1000 able to explain the EP of water-soluble monomers. 222,223 It 1001 involves the formation of oligoradicals by the interaction of a 1002 radical initiator and the monomer in the aqueous phase, which become insoluble and form particle precursors by precipitation. On the other hand, in the micellar (or heterogeneous) 1005 nucleation the radicals diffuse into monomer swollen micelles where the polymerization occur. 222,223 For a given polymerization system, the determination of the nucleation mechanism 1008 is not straightforward since it is a function of several 1009 characteristics, including the monomers, initiator, emulsifier, 1010 temperature, and kinetics parameters among others. Moreover, 1011 the EP process is highly complex because it involves multiple 1012 steps on different time and length scales. A detailed description 1013 of the method and proposed nucleation mechanisms can be 1014 found in the literature. 1

1015 In this scenario, most of the approaches for obtaining 1016 protein based NPs by EF-EP included in this review involve

the particle formation mechanisms summarized in Figure 7. In 1017 all the cases, EF-EP proceeds with the protein soluble in the 1018 water phase (or its water-soluble fraction), containing different 1019 functional and reactive groups, which depend on its origin 1020 and/or its previous chemical modification. As schematized in 1021 Figure 7 different pathways of particle formation can be found, 1022 which are governed by the employed protein, that is, its 1023 hydrophilic/hydrophobic balance (capability to form micelles), 1024 concentration, and its chemical modification, as well as the 1025 type of initiation system. The use of persulfate initiators, such 1026 as APS and KPS, produces sulfate radicals' ions which could 1027 easily abstract labile hydrogen from protein functional groups 1028 and form radical sites onto the protein backbone. Also, the 1029 interaction between persulfate and carboxylate in the aqueous 1030 phase forming radical carboxyl species is well accepted.²²⁵ 1031 These radical species on protein backbone could initiate the 1032 polymerization of the available monomer in the water phase, 1033 leading to the grafting between the protein and the synthetic 1034 polymer (Figure 7b, left). The amphiphilic protein/polymer 1035 chains then self-assemble to form compatibilized core-shell 1036 NPs (Figure 7c, left). The presence of sulfate radical anions 1037 may also lead to the aqueous phase polymerization of the 1038 water-soluble monomers, until being absorbed into the protein 1039 micelles swollen with the monomers or its precipitation 1040 followed by the stabilization with ungrafted protein (Figure 1041 7b,c, middle). These two alternatives promote the formation of 1042 noncompatibilized NPs. The nucleation of noncompatibilized 1043 NPs was not proven for the case of persulfates initiators, since 1044 the amount of grafted and ungrafted polymer and protein was 1045 not experimentally assessed.2

On the other hand, when hydroperoxides or peroxides 1047 (ROOH) are employed as initiators, a redox pair between the 1048 protein and the ROOH specie is created (Figure 7, right). Li et 1049 al. Proposed this method in which amino radicals in the 1050 protein backbone and alkyl radicals are formed. At the 1051 temperature at which the reaction was explored, the ROOH 1052 initiator does not thermally decompose, and therefore it reacts 1053 mainly with the amino groups of the protein generating a 1054 nitrogen centered radical and an alkoxy radical (RO·). Then, 1055 the amino radicals promoted the graft copolymerization of 1056 vinyl monomers in the water phase by forming the grafted 1057 copolymer protein/synthetic polymer which acts as a macro- 1058

1059 surfactant in the self-assembling of the microdomains (Figure 1060 7b,c, right). Monomer polymerization takes place inside the 1061 hydrophobic microdomains to form amphiphilic core—shell 1062 NPs. On the other hand, the homopolymerization of the 1063 monomer initiated by the alkoxy radicals is also present during 1064 the process. This is related to a second method of nucleation in 1065 which the formation of noncompatibilized particles occurs by 1066 hydrophobic radicals' coagulation or absorption into the 1067 protein micelles, as identified by Picchio et al. 24 for the 1068 synthesis of casein/acrylate latexes (Figure 7c, middle).

When Cu(III) and Ni(IV) are used as initiators ^{97,98} a similar ¹⁰⁷⁰ situation to the propagation and nucleation stages of the ¹⁰⁷¹ hydroperoxides route is developed. The metals induce the ¹⁰⁷² formation of radicals in the protein backbone, creating sites of ¹⁰⁷³ grafting and polymerization of the synthetic monomers. Then, ¹⁰⁷⁴ the formation of compatibilized NPs is promoted in a ¹⁰⁷⁵ subsequent stage of particle nucleation.

As it could be observed in Figure 7, it was found that the 1077 incorporation of water-soluble initiators (persulfates and 1078 alkylhydroperoxides) in an EF-EP and in the presence of a 1079 protein could promote one or another of these two 1080 competitive mechanisms, which can be favored by tuning the 1081 protein concentration, as it was identified by Picchio et al. 24 In 1082 the cases where the protein concentration is low, grafting 1083 extent is high and the biomaterial availability is limited for 1084 micelles formation or stabilization of noncompatibilized NPs. 1085 Consequently, the predominant way of progress of the 1086 polymerization is the compatibilized particles, resulting in 1087 highly compatibilized hybrid latexes. In contrast, when the protein concentration is high, formation of noncompatibilized 1089 NPs is promoted by the available ungrafted protein, which acts 1090 as a macro-emulsifier. Although it has been not appropriately 1091 addressed, these nucleation pathways are expected to also 1092 occur when supernormal valence transitional metals are used as 1093 initiators. Thus, the development of protein chemical 1094 modification strategies to improve compatibility of protein-1095 based hybrid NPs with high biomaterial content is highly 1096 desirable for reducing the products carbon footprint and a 1097 better exploitation of renewable resources.

In this direction, two principal strategies were identified. The 1099 first one consisted in increasing the emulsifying capability of 1100 the protein in order to create stable micelles which serve as a 1101 substrate for a subsequent monomer polymerization inside 1102 them, as schematized in Figure 8a. This route was followed by 1103 Ma and co-workers 67,72,84 in the EF-EP of casein in the 1104 presence of VAC, acrylate monomers, and PDMS. This protein 1105 modification approach consisted in the condensation polymer-1106 ization between CPL and the carboxyl and amine groups of 1107 casein, through the ring opening of the CPL molecules. The 1108 incorporated pendant groups in the casein backbone acted as 1109 blockers of the polar groups of the protein increasing its 1110 emulsifier capability, resulting in more stable casein micelles. In 1111 this way, CA-CPL micelles acted as self-emulsifier and as 1112 substrate for the polymerization of the hydrophobic mono-1113 mers.

Another approach to increase the grafting efficiency between protein and synthetic polymer consisted in adding vinyl functionalities to the protein, which then serve as propagating sites in a radical polymerization. With this aim Picchio et al. 101 synthesized functionalized casein through the reaction of protein with AA, in the presence of N-(3-dimethylaminoprop-1120 yl)-N'-ethylcarbodiimine hydrochloride (EDC). In this way, an 1121 amide bond is formed between the casein and the AA, 226

providing new acrylic functionality able to propagate via radical 1122 polymerization. This modified casein improved the compatibi- 1123 lization degree of hybrid NPs obtained by EP of acrylic 1124 monomers. However, significantly enhanced results were 1125 obtained in the latter work by Picchio and co-workers^{82,102} where HMC was used in the EF-EP in the presence of MMA 1127 and BA. HMC was prepared through a glycidyl ether reaction 1128 to incorporate pendant methacrylic groups onto the protein 1129 backbone (Figure 8b). These pendant groups and the primary 1130 amino groups serve as propagation sites during the polymer- 1131 ization, favoring the formation of compatibilized hybrid NPs 1132 following the mechanisms summarized in Figure 7. Also, 1133 Zheng et al. 130 proposed the vinyl functionality incorporation 1134 to soy protein using N-methylolacrylamide as functional agent. 1135 A possible esterification reaction between carboxylic acid of the 1136 protein and hydroxyl groups of N-methylolacrylamide was 1137 proposed.

Although EF-EP was the main polymerization strategy 1139 employed for obtaining protein based hybrid NPs, also a few 1140 proposals employed MEP. 227 Fapeng et al. 126 used MEP to 1141 obtain soy protein/acrylic NPs, which were then used to 1142 fabricate wood adhesive films. Neat and modified soy proteins 1143 were used, where the latter case increased the compatibility 1144 with the polymer and the films homogeneity. The emulsifiers 1145 employed to stabilize the miniemulsion were alkylphenol 1146 ethoxylates and SLS. N-Hexadecane was employed as 1147 cosurfactant, and the polymerization reaction was initiated by 1148 using APS. This approach also involved the modification of soy 1149 protein by the denaturation of the protein in aqueous solution 1150 of sodium hydroxide at 70 °C. The modified protein showed 1151 an increased surface energy due to the rise of unsaturated 1152 bonds, which provided more reaction sites able to be 1153 combined with the acrylate monomers in the posterior MEP 1154

Another example of the use of MEP was given by Allasia et 1156 al., 132 where a emulsifier-free latex of acrylic/zein/casein NPs 1157 having a high solids content (35%) was obtained. The use of 1158 neat and methacrylated casein was tested as stabilizer, instead 1159 of emulsifier, resulting in hybrid latexes with improved 1160 compatibility and water resistance.

SUMMARY AND PERSPECTIVES

Waterborne polymeric NPs are nowadays a widespread 1163 product for obtaining high-quality materials at large scale. 1164 However, despite their many advantages, polymeric NPs are 1165 still mainly produced from petroleum-based monomers. The 1166 continuous demand for this resource, the reserves of which are 1167 of exhaustible nature is nonsustainable, and for this reason the 1168 exploration of renewable feedstocks to prepare a new 1169 generation of waterborne NPs with low carbon footprint and 1170 environmental impact is imperative for both academics and 1171 industries. In this context, this review collects the works done 1172 to obtain hybrid protein/polymer NPs, which have been 1173 employed or have the potential to be used in industrial 1174 applications. The incorporation of proteins in the waterborne 1175 NPs formulation puts latex technology at a higher level, 1176 offering the possibility to improve its structural, functional, and 1177 biodegradability features. In that sense, hybrid protein-based 1178 latexes have been used to produce and study final products, 1179 including coatings, 82,84,100,128,129,132 adhesives, 120,126,131 and 1180 films, 66,72,124 which have shown improved properties in 1181 comparison with the synthetic polymer latexes (without 1182

1275

1276

1282

1288

1289

1291

1292

1303

1304

1305

1183 protein). Moreover, properties comparable to existent 1184 commercial products were obtained in some cases. 67,102,126 The main synthesis route for obtaining these protein/ 1186 synthetic polymer latexes involves the emulsion polymerization 1187 in the absence of emulsifier, ^{24,67,79,99,124} which paves the way 1188 for its industrial implementation without significant techno-1189 logical upgrades. The efforts made have allowed the achieve-1190 ment of producing high solids hybrid latexes with elevated 1191 protein content 82,83,99,132,153 and compatibility, 82,132 highly 1192 desirable for an efficient exploitation of this sustainable 1193 resource. These features were in most cases the result of a 1194 prepolymerization stage of protein modification, which pursues 1195 introducing extra active sites onto the protein backbone to 1196 promote its grafting with the synthetic polymer^{82,132} or to 1197 modify its hydrophilic-hydrophobic balance. 67,72,84 In this 1198 scenario, highly compatibilized hybrid materials showed 1199 superior properties than the simple constituents' addition, 1200 demonstrating a synergetic contribution of both components 1201 due to the intimate contact between phases. Some of these 1202 synergetic improvements observed in the applications of 1203 protein-based hybrid NPs were partial biodegradabil-1204 ity, 66,67,72,84,124,132 thermal stability, 67 film formation with 1205 low requirement of formulation agents (better NPs coales-1206 cence), 24,131 solvent resistance, 24 water resist-1207 ance, 72,82,102,103,125 and mechanical strength 103,125,126,153

1208 among others. The most explored incorporated protein was casein, and 1210 hence the more advanced reported developments were with 1211 this protein. 66,67,102,104 This is because casein is a commonly 1212 employed reagent in the formulation of many industrial 1213 products, due to its excellent film-forming features, and 1214 therefore it is highly available in large volume and with 1215 acceptable purity. There are many protein alternatives which, 1216 though less explored, offer the opportunity to incorporate 1217 additional functionalities into waterborne hybrid latexes and 1218 put many biomaterials obtained as byproducts of other 1219 industries into a second step of the chain values. In this 1220 regard, it is remarkable the few efforts for incorporating whey 1221 protein, which is obtained from a polluting effluent, to 1222 formulate biobased products such as bioplastics and coat-1223 ings. 119,120

One drawback reported for many protein-based hybrid 1225 products was their low resistance to waterborne solvents, 1226 because most of these biomaterials are hydrophilic. Many 1227 attempts focused on solving this limitation involved the 1228 chemical modification of the protein to improve its 1229 compatibility with the synthetic phase. One unexplored 1230 alternative was the formulation of cross-linkable hybrid latexes, which is a commonly used strategy in the coating and adhesive 1232 industry for obtaining high-performance products. 228,229 It is 1233 expected that the incorporation of functionalities into protein-1234 based hybrid NPs that promote cross-linking during film-1235 forming, that is, a biobased self-cross-linkable latex, could open 1236 the opportunity to increase their biocomponent content and 1237 reach improved features for producing high-performance 1238 alternative materials.

One important point arising from the composition of 1240 protein-based hybrid latexes is that the explored systems 1241 considered the incorporation of only one protein. An exception 1242 is the work by Allasia et al., 132 in which the synergetic balance 1243 between a hydrophilic and a hydrophobic protein with acrylic-1244 based NPs was proposed. The incorporation of a low fraction 1245 of zein counteracted the hydrophilic character of casein, by

reaching a film surface hydrophobicity similar to that of pure 1246 acrylics. Under this context, a material design involving an 1247 accurate combination of proteins, or otherwise a protein with 1248 other biomaterials, may provide the opportunity to produce 1249 novel waterborne hybrid latexes with high biocontent and an 1250 end-use performance governed by the synergetic joining of 1251 their functional components. Following this basis, many 1252 authors have investigated the formation of zein-based particles 1253 stabilized with different biopolymers. 189,190,194,200-213,230,231 Although these zein-based NPs offer a good challenge for 1255 obtaining fully biobased materials, this strategy could be 1256 exploited as precursors for producing a new generation of 1257 proteins/synthetic polymer hybrid NPs in many industrial 1258 applications.

Finally, although most of the proposed strategies pursue 1260 reducing the employment of oil-based materials by incorporat- 1261 ing an industrially available protein, few articles focused on 1262 investigating the involved mechanism of hybrid particles 1263 nucleation, 24,79 the effect of protein interaction on the 1264 polymerization process and its kinetics, 95 and the influence 1265 of protein content on the end-use materials and their final 1266 properties. 24,65,95,125,127,129,131 It is expected that advances on 1267 the fundamental knowledge about the interplay of protein with 1268 synthetic polymer will contribute to rationalize the design of 1269 both the waterborne hybrid NPs and the synthetic pathway. In 1270 this regard, future research should aim to expand the 1271 assortment of proteins explored up to now, encompassing 1272 those which are of great interest in biological catalysis and 1273 other technological applications.

AUTHOR INFORMATION

Corresponding Authors

Roque J. Minari - Polymer Reaction Engineering Group, INTEC, Santa Fe S3000, Argentina; Facultad de Ingeniería 1278 Ouímica, Universidad Nacional del Litoral, Santa Fe, Santa 1279 Fe S3000, Argentina; o orcid.org/0000-0003-3645-5317; 1280 Email: rjminari@santafe-conicet.gov.ar

Luis M. Gugliotta - Polymer Reaction Engineering Group, INTEC, Santa Fe S3000, Argentina; Facultad de Ingeniería 1283 Química, Universidad Nacional del Litoral, Santa Fe, Santa 1284 Fe S3000, Argentina; Email: lgug@intec.unl.edu.ar

Authors 1286 1287

Luisa Guadalupe Cencha – Polymer Reaction Engineering Group, INTEC, Santa Fe S3000, Argentina; Facultad de Ingeniería y Ciencias Hídricas, Universidad Nacional del Litoral, Santa Fe, Santa Fe S3000, Argentina; oorcid.org/ 1290 0000-0003-2858-6827

Mariana Allasia – Polymer Reaction Engineering Group, INTEC, Santa Fe S3000, Argentina

Ludmila Irene Ronco – Polymer Reaction Engineering Group, 1294 INTEC, Santa Fe S3000, Argentina; Facultad de Ingeniería 1295 Química, Universidad Nacional del Litoral, Santa Fe, Santa 1296 Fe S3000, Argentina; o orcid.org/0000-0001-6954-3159 1297

Gisela Carina Luque - Polymer Reaction Engineering Group, 1298 INTEC, Santa Fe S3000, Argentina; Facultad de Ingeniería 1299 Química, Universidad Nacional del Litoral, Santa Fe, Santa 1300 Fe S3000, Argentina

Matías Luis Picchio - Departamento de Química Orgánica, 1302 Facultad de Ciencias Químicas, Universidad Nacional de Córdoba, IPQA—CONICET, Córdoba, Córdoba X5000, Argentina; orcid.org/0000-0003-3454-5992

Complete contact information is available at:

1307 https://pubs.acs.org/10.1021/acs.iecr.0c05958

1308 Notes

1309 The authors declare no competing financial interest.

1310 **ACKNOWLEDGMENTS**

1311 The author acknowledge the financial support received from 1312 CONICET, UNL (CAI+D 50420150100100LI), and ANP-1313 CyT (PICT-2016-3876) (all of Argentina).

1314 REFERENCES

- 1315 (1) Chern, C. S. Emulsion Polymerization Mechanisms and Kinetics. 1316 *Prog. Polym. Sci.* **2006**, *31*, 443–486.
- 1317 (2) Ramli, R. A.; Laftah, W. A.; Hashim, S. Core—shell polymers: a 1318 review. *RSC Adv.* **2013**, *3*, 15543.
- 1319 (3) Overbeek, A.; Bückmann, F.; Martin, E.; Steenwinkel, P.; 1320 Annable, T. New Generation Decorative Paint Technology. *Prog. Org.* 1321 *Coat.* **2003**, *48*, 125–139.
- 1322 (4) Goldschmidt, A.; Streitberger, H.-J. In *BASF Handbook on Basics* 1323 of Coating Technology; Andrew, W., Ed.; American Coatings 1324 Literature; Vincentz Network, 2003.
- 1325 (5) Chou, C.-S.; Weier, J. E. In *Polymer Dispersions and Their* 1326 *Industrial Applications*; Urban, D., Takamura, K., Eds.; John Wiley & 1327 Sons, Ltd, 2002; Chapter 14, pp 355–382.
- 1328 (6) Rodrigo, A. B. Development of Waterborne Polymeric Dispersions 1329 Based on Biobased Monomers for Their Application as PSAs and 1330 Coatings. Ph.D. Thesis, University of the Basque Country UPV/EHU, 1331 2020.
- 1332 (7) Polymer Dispersion Market Global Industry Analysis, Size, Share, 1333 Growth, Trends, and Forecast 2019 2027; Report Linker, 2020.
- 1334 (8) Harris, S.; Staffas, L.; Rydberg, T.; Eriksson, E. Renewable 1335 Materials in the Circular Economy; IVL Swedish Environmental 1336 Research Institute Ltd, 2018.
- 1337 (9) Molina-Gutiérrez, S.; Ladmiral, V.; Bongiovanni, R.; Caillol, S.; 1338 Lacroix-Desmazes, P. Radical Polymerization of Biobased Monomers 1339 in Aqueous Dispersed Media. *Green Chem.* **2019**, *21*, 36–53.
- 1340 (10) Goikoetxea, M.; Minari, R. J.; Beristain, I.; Paulis, M.; 1341 Barandiaran, M. J.; Asua, J. M. Polymerization Kinetics and 1342 Microstructure of Waterborne Acrylic/Alkyd Nanocomposites 1343 Synthesized by Miniemulsion. *J. Polym. Sci., Part A: Polym. Chem.* 1344 **2009**, 47, 4871.
- 1345 (11) Lim, K.; Ching, Y.; Gan, S. Effect of Palm Oil Bio-Based 1346 Plasticizer on the Morphological, Thermal and Mechanical Properties 1347 of Poly(Vinyl Chloride). *Polymers* **2015**, *7*, 2031–2043.
- 1348 (12) Fei, G.; Sun, L.; Wang, H.; Gohar, F.; Ma, Y.; Kang, Y.-M. 1349 Rational design of phosphorylated poly(vinyl alcohol) grafted 1350 polyaniline for waterborne bio-based alkyd nanocomposites with 1351 high performance. *Prog. Org. Coat.* 2020, 140, 105484.
- 1352 (13) Pathan, S.; Ahmad, S. Synergistic Effects of Linseed Oil Based 1353 Waterborne Alkyd and 3-Isocynatopropyl Triethoxysilane: Highly 1354 Transparent, Mechanically Robust, Thermally Stable, Hydrophobic, 1355 Anticorrosive Coatings. ACS Sustainable Chem. Eng. 2016, 4, 3062— 1356 3075.
- 1357 (14) Smeets, N. M.; Imbrogno, S.; Bloembergen, S. Carbohydrate 1358 functionalized hybrid latex particles. *Carbohydr. Polym.* **2017**, *173*, 1359 233–252.
- 1360 (15) Shamshina, J. L.; Berton, P.; Rogers, R. D. Advances in 1361 Functional Chitin Materials: A Review. *ACS Sustainable Chem. Eng.* 1362 **2019**, *7*, 6444–6457.
- 1363 (16) Mastalygina, E.; Varyan, I.; Kolesnikova, N.; Gonzalez, M. I. C.; 1364 Popov, A. Effect of Natural Rubber in Polyethylene Composites on 1365 Morphology, Mechanical Properties and Biodegradability. *Polymers* 1366 **2020**, *12*, 437.
- 1367 (17) Capezza, A. J.; Newson, W. R.; Olsson, R. T.; Hedenqvist, M. 1368 S.; Johansson, E. Advances in the Use of Protein-Based Materials: 1369 Toward Sustainable Naturally Sourced Absorbent Materials. ACS 1370 Sustainable Chem. Eng. 2019, 7, 4532–4547.

- (18) Rajinipriya, M.; Nagalakshmaiah, M.; Robert, M.; Elkoun, S. 1371 Importance of Agricultural and Industrial Waste in the Field of 1372 Nanocellulose and Recent Industrial Developments of Wood Based 1373 Nanocellulose: A Review. ACS Sustainable Chem. Eng. 2018, 6, 2807—1375 2828.
- (19) Thakur, V. K.; Thakur, M. K.; Raghavan, P.; Kessler, M. R. 1376 Progress in Green Polymer Composites from Lignin for Multifunc- 1377 tional Applications: A Review. ACS Sustainable Chem. Eng. 2014, 2, 1378 1072–1092.
- (20) Isikgor, F. H.; Becer, C. R. Lignocellulosic biomass: a 1380 sustainable platform for the production of bio-based chemicals and 1381 polymers. *Polym. Chem.* **2015**, *6*, 4497–4559.
- (21) Guilbert, S.; Cuq, B. 11. Material formed from proteins; De 1383 Gruyter: Berlin, Boston, 2020; pp 299–338.
- (22) Zhang, L.; Zeng, M. Monomers, Polymers and Composites from 1385 Renewable Resources; Elsevier, 2008; pp 479–493.
- (23) Shi, K.; Huang, Y.; Yu, H.; Lee, T.-C.; Huang, Q. Reducing the 1387 Brittleness of Zein Films through Chemical Modification. *J. Agric.* 1388 Food Chem. **2011**, *59*, 56–61.
- (24) Picchio, M. L.; Minari, R. J.; Gonzalez, V. D. G.; Passeggi, M. 1390 C.; Vega, J. R.; Barandiaran, M. J.; Gugliotta, L. M. Waterborne 1391 Acrylic-Casein Nanoparticles. Nucleation and Grafting. *Macromol.* 1392 *Symp.* 2014, 344, 76–85.
- (25) Lai, M.; Cai, K.; Zhao, L.; Chen, X.; Hou, Y.; Yang, Z. Surface 1394 Functionalization of TiO2Nanotubes with Bone Morphogenetic 1395 Protein 2 and Its Synergistic Effect on the Differentiation of 1396 Mesenchymal Stem Cells. *Biomacromolecules* 2011, 12, 1097–1105. 1397
- (26) Qi, H.; Zheng, W.; Zhang, C.; Zhou, X.; Zhang, L. Novel 1398 Mussel-Inspired Universal Surface Functionalization Strategy: Pro- 1399 tein-Based Coating with Residue-Specific Post-Translational Mod- 1400 ification in Vivo. ACS Appl. Mater. Interfaces 2019, 11, 12846–12853. 1401
- (27) Liu, R.; Zhao, J.; Han, Q.; Hu, X.; Wang, D.; Zhang, X.; Yang, 1402 P. One-Step Assembly of a Biomimetic Biopolymer Coating for 1403 Particle Surface Engineering. *Adv. Mater.* **2018**, *30*, 1802851.
- (28) Silvernail, L. H.; Bain, W. M. Synthetic and Protein Adhesives for 1405 Paper Coating; TAPPI monograph series, no. 22; Technical 1406 Association of the Pulp and Paper Industry. Coating Committee., 1407 1961.
- (29) Kickelbick, G. Hybrid Materials: Synthesis, Characterization, and 1409 Applications; John Wiley & Sons, 2021.
- (30) Johnson, R. P.; John, J. V.; Kim, I. Recent developments in 1411 polymer-block-polypeptide and protein-polymer bioconjugate 1412 hybrid materials. *Eur. Polym. J.* **2013**, *49*, 2925–2948.
- (31) Gupta, P.; Nayak, K. K. Characteristics of protein-based 1414 biopolymer and its application. *Polym. Eng. Sci.* **2015**, *SS*, 485–498. 1415
- (32) Saif, B.; Zhang, W.; Zhang, X.; Gu, Q.; Yang, P. Sn-Triggered 1416 Two-Dimensional Fast Protein Assembly with Emergent Functions. 1417 ACS Nano 2019, 13, 7736–7749.
- (33) Qin, R.; Liu, Y.; Tao, F.; Li, C.; Cao, W.; Yang, P. Protein- 1419 Bound Freestanding 2D Metal Film for Stealth Information 1420 Transmission. *Adv. Mater.* **2018**, 1803377.
- (34) Wang, L.; Deng, L.; Liu, Y.-N. Protein-Metal-Ion Networks: A 1422 Unique Approach toward Metal Sulfide Nanoparticles Embedded In 1423 Situ in Nanocomposites. *Chem. Eur. J.* **2019**, *25*, 904–912.
- (35) Palomo, J. M. Nanobiohybrids: a new concept for metal 1425 nanoparticles synthesis. *Chem. Commun.* **2019**, *55*, 9583–9589. 1426
- (36) Chawla, S. S.; Gorakshakar, A. C.; Ghosh, K. K.; Madkaikar, M. 1427 R.; Devarajan, P. V. Fabrication of gelatin functionalized silver 1428 nanoparticles for blood group profiling. *Nanotechnology* **2020**, *31*, 1429 295102.
- (37) Sothornvit, R.; Rhim, J.-W.; Hong, S.-I. Effect of nano-clay type 1431 on the physical and antimicrobial properties of whey protein isolate/ 1432 clay composite films. *J. Food Eng.* **2009**, *91*, 468–473.
- (38) Swain, S. K.; Priyadarshini, P. P.; Patra, S. K. Soy Protein/Clay 1434 Bionanocomposites as Ideal Packaging Materials. *Polym.-Plast.* 1435 Technol. Eng. 2012, 51, 1282–1287.
- (39) Kiryukhin, M. V.; Lau, H. H.; Lim, S. H.; Salgado, G.; Fan, C.; 1437 Ng, Y. Z.; Leavesley, D. I.; Upton, Z. Arrays of Biocompatible and 1438

- 1439 Mechanically Robust Microchambers Made of Protein-Polyphenol-1440 Clay Multilayer Films. ACS Biomater. Sci. Eng. 2020, 6, 5653-5661.
- 1441 (40) Kaseem, M.; Ko, Y. G. A novel hybrid composite composed of 1442 albumin, WO3, and LDHs film for smart corrosion protection of Mg 1443 alloy. *Composites, Part B* **2021**, 204, 108490.
- 1444 (41) Ravi, S. K.; Tan, S. C. Solar Energy Harvesting with 1445 Photosynthetic Pigment-Protein Complexes; Springer: Singapore, 2020; 1446 pp 65–77.
- 1447 (42) Corrado, I.; Abdalrazeq, M.; Pezzella, C.; Girolamo, R. D.; 1448 Porta, R.; Sannia, G.; Giosafatto, C. V. L. Design and characterization 1449 of poly (3-hydroxybutyrate-co-hydroxyhexanoate) nanoparticles and 1450 their grafting in whey protein-based nanocomposites. *Food Hydro-* 1451 colloids 2021, 110, 106167.
- 1452 (43) Peng, J.; Calabrese, V.; Ainis, W. N.; Scager, R.; Velikov, K. P.; 1453 Venema, P.; van der Linden, E. Mixed gels from whey protein isolate 1454 and cellulose microfibrils. *Int. J. Biol. Macromol.* **2019**, *124*, 1094–1455 1105.
- 1456 (44) Saveleva, M. S.; Eftekhari, K.; Abalymov, A.; Douglas, T. E. L.; 1457 Volodkin, D.; Parakhonskiy, B. V.; Skirtach, A. G. Hierarchy of 1458 Hybrid Materials—The Place of Inorganics-in-Organics in it, Their 1459 Composition and Applications. *Front. Chem.* **2019**, *7*, 179.
- 1460 (45) Motelica, L.; Ficai, D.; Ficai, A.; Oprea, O. C.; Kaya, D. A.; 1461 Andronescu, E. Biodegradable Antimicrobial Food Packaging: Trends 1462 and Perspectives. *Foods* **2020**, *9*, 1438.
- 1463 (46) Palivan, C. G.; Fischer-Onaca, O.; Delcea, M.; Itel, F.; Meier, 1464 W. Protein—polymer nanoreactors for medical applications. *Chem.* 1465 Soc. Rev. **2012**, 41, 2800—2823.
- 1466 (47) Wu, Y.; Ng, D. Y. W.; Kuan, S. L.; Weil, T. Protein—polymer 1467 therapeutics: a macromolecular perspective. *Biomater. Sci.* **2015**, 3, 1468 214—230.
- 1469 (48) Boyer, C.; Huang, X.; Whittaker, M. R.; Bulmus, V.; Davis, T. 1470 P. An overview of protein—polymer particles. *Soft Matter* **2011**, *7*, 1471 1599—1614.
- 1472 (49) Ha, H.-K.; Rankin, S.; Lee, M.-R.; Lee, W.-J. Development and 1473 Characterization of Whey Protein-Based Nano-Delivery Systems: A 1474 Review. *Molecules* **2019**, *24*, 3254.
- 1475 (50) Saallah, S.; Lenggoro, I. W. Nanoparticles Carrying Biological 1476 Molecules: Recent Advances and Applications. *KONA Powder Part. J.* 1477 **2018**, 35, 89–111.
- 1478 (51) DeFrates, K.; Markiewicz, T.; Gallo, P.; Rack, A.; Weyhmiller, 1479 A.; Jarmusik, B.; Hu, X. Protein Polymer-Based Nanoparticles: 1480 Fabrication and Medical Applications. *Int. J. Mol. Sci.* **2018**, *19*, 1717.
- 1481 (52) Martínez-López, A. L.; Pangua, C.; Reboredo, C.; Campión, R.; 1482 Morales-Gracia, J.; Irache, J. M. Protein-based nanoparticles for drug delivery purposes. *Int. J. Pharm.* **2020**, *581*, 119289.
- 1484 (53) Sarode, A.; Sawale, P.; Khedkar, C.; Kalyankar, S.; Pawshe, R. 1485 *Encyclopedia of Food and Health*; Elsevier, 2016; pp 676–682.
- 1486 (54) Dhillon, G. Protein Byproducts: Transformation from Environ-1487 mental Burden Into Value-Added Products; Elsevier Science, 2016.
- 1488 (55) Hettiarachchy, N.; Kalapathy, U. Soybeans; Springer, 1997; pp 1489 379-411.
- 1490 (56) Cook, R. B.; Mallee, F. M.; Shulman, M. L. Purification of zein 1491 from corn gluten meal. WO Patent WO1993012667A1, 1993.
- 1492 (57) Widyasari, R.; Rawdkuen, S. Extraction and characterization of 1493 gelatin from chicken feet by acid and ultrasound assisted extraction. 1494 *FABJ.* **2014**, *2*, 85–97.
- 1495 (58) Pelegrine, D.; Gasparetto, C. Whey proteins solubility as 1496 function of temperature and pH. *Food Sci. Technol.* **2005**, 38, 77–80. 1497 (59) Shukla, R.; Cheryan, M. Zein: the industrial protein from corn.
- 1497 (S9) Shukla, R.; Cheryan, M. Zein: the industrial protein from 1498 *Ind. Crops Prod.* **2001**, *13*, 171–192.
- 1499 (60) Anderson, T. J.; Lamsal, B. P. REVIEW: Zein Extraction from 1500 Corn, Corn Products, and Coproducts and Modifications for Various 1501 Applications: A Review. *Cereal Chem.* **2011**, *88*, 159–173.
- 1502 (61) León-López, A.; Morales-Peñaloza, A.; Martínez-Juárez, V. M.; 1503 Vargas-Torres, A.; Zeugolis, D. I.; Aguirre-Álvarez, G. Hydrolyzed
- 1503 Vargas-Torres, A.; Zeugolis, D. I.; Aguirre-Alvarez, G. Hydrolyze 1504 Collagen—Sources and Applications. *Molecules* **2019**, 24, 4031.
- 1505 (62) Guo, M.; Wang, G. Milk Protein Polymer and Its Application in 1506 Environmentally Safe Adhesives. *Polymers* **2016**, *8*, 324.

- (63) Global Casein and Caseinate Market to Account for US\$ 3564.4 1507 Mn by 2025. Process Industry Match (accessed on 2020/11/23). 1508
- (64) Wusigale; Liang, L.; Luo, Y. Casein and pectin: Structures, 1509 interactions, and applications. *Trends Food Sci. Technol.* **2020**, 97, 1510 391–403.
- (65) Ma, J. Z.; Xu, Q. N.; Gao, D. G. Study on Synthesis and 1512 Performances of Casein Resin Grafting Modified by Caprolactam/ 1513 Acrylic Esters/Vinyl Acetate/Organic Silicone. *Adv. Mater. Res.* **2010**, 1514 123–125, 1267–1270.
- (66) Ma, J.; Xu, Q.; Gao, D.; Zhou, J.; Zhang, J. Blend composites of 1516 caprolactam-modified casein and waterborne polyurethane for film- 1517 forming binder: Miscibility, morphology and properties. *Polym.* 1518 *Degrad. Stab.* **2012**, *97*, 1545–1552.
- (67) Ma, J.; Xu, Q.; Zhou, J.; Gao, D.; Zhang, J.; Chen, L. Nano- 1520 scale core—shell structural casein based coating latex: Synthesis, 1521 characterization and its biodegradability. *Prog. Org. Coat.* **2013**, *76*, 1522 1346–1355.
- (68) Barreto, P.; Pires, A.; Soldi, V. Thermal degradation of edible 1524 films based on milk proteins and gelatin in inert atmosphere. *Polym.* 1525 *Degrad. Stab.* **2003**, *79*, 147–152.
- (69) Sohail, S. S.; Wang, B.; Biswas, M. A. S.; Oh, J.-H. Physical, 1527 Morphological, and Barrier Properties of Edible Casein Films with 1528 Wax Applications. *J. Food Sci.* **2006**, *71*, C255–C259.
- (70) Audic, J.-L.; Chaufer, B.; Daufin, G. Non-food applications of 1530 milk components and dairy co-products: A review. *Lait* **2003**, 83, 1531 417–438.
- (71) Ghosh, A.; Ali, M. A.; Dias, G. J. Effect of Cross-Linking on 1533 Microstructure and Physical Performance of Casein Protein. 1534 Biomacromolecules 2009, 10, 1681–1688.
- (72) Ma, J.; Zhang, F.; Qiao, Y.; Xu, Q.; Zhou, J.; Zhang, J. Vi- 1536 PDMS incorporated with protein-based coatings designed for 1537 permeability-enhanced applications. *J. Appl. Polym. Sci.* **2018**, *135*, 1538 46501.
- (73) Fan, Q.; Ma, J.; Xu, Q.; Zhang, J.; Simion, D.; Carmen, G.; 1540 Guo, C. Animal-derived natural products review: Focus on novel 1541 modifications and applications. *Colloids Surf., B* **2015**, 128, 181–190. 1542
- (74) Narayanan, S.; Pavithran, M.; Viswanath, A.; Narayanan, D.; 1543 Mohan, C. C.; Manzoor, K.; Menon, D. Sequentially releasing dual- 1544 drug-loaded PLGA—casein core/shell nanomedicine: Design, syn- 1545 thesis, biocompatibility and pharmacokinetics. *Acta Biomater.* **2014**, 1546 10, 2112—2124.
- (75) Costa, J. P.; Carvalho, S.; Jesus, S.; Soares, E.; Marques, A. P.; 1548 Borges, O. Optimization of Chitosan- α -casein Nanoparticles for 1549 Improved Gene Delivery: Characterization, Stability, and Trans- 1550 fection Efficiency. *AAPS PharmSciTech* **2019**, *20*, 132.
- (76) Picchio, M. L.; Cuggino, J. C.; Nagel, G.; Wedepohl, S.; Minari, 1552 R. J.; Igarzabal, C. I. A.; Gugliotta, L. M.; Calderón, M. Crosslinked 1553 casein-based micelles as a dually responsive drug delivery system. 1554 *Polym. Chem.* **2018**, *9*, 3499–3510.
- (77) Sahba, R.; Seyed, M.; Sajjadi, A. A.; Farhadyar, N.; Sadeghi, B. 1556 Preparation and characterization of friendly colloidal Hydroxyapatite 1557 based on natural Milk's casein. *Int. J. Nanodimens.* **2018**, *9*, 238–245. 1558
- (78) Wang, Y.; Ma, J.; Xu, Q.; Zhang, J. Fabrication of antibacterial 1559 casein-based ZnO nanocomposite for flexible coatings. *Mater. Des.* 1560 **2017**, 113, 240–245.
- (79) Li, P.; Zhu, J.; Sunintaboon, P.; Harris, F. W. New Route to 1562 Amphiphilic Core-Shell Polymer Nanospheres: Graft Copolymeriza- 1563 tion of Methyl Methacrylate from Water-Soluble Polymer Chains 1564 Containing Amino Groups. *Langmuir* **2002**, *18*, 8641–8646.
- (80) Blackley, D. Emulsion Polymerization: Theory and practice; 1566 Springer, 1975.
- (81) Picchio, M. L.; Bohórquez, S. J.; van den Berg, P. G. C. A.; 1568 Barandiaran, M. J.; Gugliotta, L. M.; Minari, R. J. Waterborne Casein- 1569 Based Latexes with High Solids Content and Their High-Throughput 1570 Coating Optimization. *Ind. Eng. Chem. Res.* **2016**, *55*, 10271–10277. 1571 (82) Picchio, M. L.; Minari, R. J.; Gugliotta, L. M. Enhancing the 1572 coating properties of acrylic/casein latexes with high protein content. 1573

J. Coat. Technol. Res. 2017, 14, 543-553.

- 1575 (83) Aguzin, A.; Jerkovich, J. I.; Trucone, J.; Ronco, L. I.; Minari, R. 1576 J.; Gugliotta, L. M. Acrylic-Casein Latexes with Potential Application 1577 as Adhesives. *Lat. Am. Appl. Res.* **2020**, *50*, 115–120.
- 1578 (84) Ma, J.; Gan, C.; Xu, Q.; Zhou, J.; Zhang, J. Amphiphilic 1579 copolymer stabilized core—shell structural casein-based emulsion. 1580 *Colloids Surf., A* **2015**, *471*, 65–72.
- 1581 (85) Ma, J.; Xu, Q.; Zhou, J.; Zhang, J.; Zhang, L.; Tang, H.; Chen, 1582 L. Synthesis and biological response of casein-based silica nano-1583 composite film for drug delivery system. *Colloids Surf., B* **2013**, *111*, 1584 257–263.
- 1585 (86) Mohan, D.; Radhakrishnan, G.; Nagabhushanam, T. Synthesis 1586 of casein—g—poly(butyl acrylate). *J. Appl. Polym. Sci.* **1980**, 25, 1799— 1587 1806.
- 1588 (87) Mohan, D.; Radhakrishnan, G.; Rajadurai, S. Synthesis of 1589 Casein-g-Poly(methyl Acrylate). *J. Macromol. Sci., Chem.* **1983**, 20, 1590 201–212.
- 1591 (88) Mohan, D.; Radhakrishnan, G.; Rajadurai, S. Synthesis of 1592 Casein-g-Poly(n-butyl Methacrylate). *J. Macromol. Sci., Chem.* **1985**, 1593 22, 75–83.
- 1594 (89) Mohan, D.; Radhakrishnan, G.; Rajadurai, S. Synthesis of 1595 Casein-g-Poly(Ethyl Acrylate). *Leather Sci.* **1986**, 33, 242.
- 1596 (90) Mohan, D.; Radhakrishnan, G.; Rajadurai, S.; Rao, K. V.; 1597 Cameron, G. G. Graft copolymerization of acrylamide onto casein: A 1598 kinetic study. *J. Polym. Sci., Part A: Polym. Chem.* **1989**, 27, 2123–1599 2133.
- 1600 (91) Mohan, D.; Radhakrishnan, G.; Rajadurai, S. Synthesis of 1601 casein-g-poly(methyl acrylate). II. *J. Appl. Polym. Sci.* **1990**, 39, 1507—1602 1518.
- 1603 (92) Somanathan, N.; Sanjeevi, R.; Reddy, C. R.; Radhakrishnan, N. 1604 Graft co-polymerization of casein with acrylonitrile and n-butyl 1605 methacrylate. *Eur. Polym. J.* **1987**, 23, 489–492.
- 1606 (93) Dong, Q.; Hsieh, Y.-L. Acrylonitrile graft copolymerization of 1607 casein proteins for enhanced solubility and thermal properties. *J. Appl.* 1608 *Polym. Sci.* **2000**, *77*, 2543–2551.
- 1609 (94) Somanathan, N.; Jeevan, R. G.; Sanjeevi, R. Syhthesis of Casein 1610 Graft Poly(acrylonitrile). *Polym. J.* **1993**, 25, 939–946.
- 1611 (95) Li, P.; Liu, J. H.; Wang, Q.; Wu, C. Copper-mediated graft 1612 copolymerization of methyl methacrylate onto casein. *Macromol.* 1613 *Symp.* **2000**, *151*, 605–610.
- 1614 (96) Imoto, M.; Ouchi, T. Radical Polymerization of Vinyl 1615 Monomers by Hydrophilic Macromolecules. I. Uncatalyzed Polymer-1616 ization in the Absence of Copper(ll) Ions. *J. Macromol. Sci., Polym.* 1617 *Rev.* 1982, 22, 261–302.
- 1618 (97) Liu, Y.; Zhang, Y.; Liu, Z.; Deng, K. Graft copolymerization of 1619 butyl acrylate onto casein initiated by potassium diperiodatonickelate-1620 (IV) in alkaline medium. *Eur. Polym. J.* **2002**, 38, 1619–1625.
- 1621 (98) Liu, Y.; Li, J.; Yang, L.; Shi, Z. Graft Copolymerization of 1622 Methyl Methacrylate onto Casein Initiated by Potassium 1623 Ditelluratocuprate(III). J. Macromol. Sci., Part A: Pure Appl.Chem. 1624 2004, 41, 305–316.
- 1625 (99) Zhu, J.; Li, P. Synthesis and characterization of poly(methyl 1626 methacrylate)/casein nanoparticles with a well-defined core-shell 1627 structure. *J. Polym. Sci., Part A: Polym. Chem.* **2003**, *41*, 3346–3353. 1628 (100) Picchio, M. L.; Passeggi, M. C.; Barandiaran, M. J.; Gugliotta,
- 1629 L. M.; Minari, R. J. Waterborne acrylic—casein latexes as eco-friendly 1630 binders for coatings. *Prog. Org. Coat.* **2015**, 88, 8–16.
- 1631 (101) Picchio, M. L.; Minari, R. J.; Gonzalez, V. D. G.; Barandiaran, 1632 M. J.; Gugliotta, L. M. New strategy to improve acrylic/casein 1633 compatibilization in waterborne hybrid nanoparticles. *J. Appl. Polym.* 1634 Sci. 2015, 132, 42421 DOI: 10.1002/app.42421.
- 1635 (102) Picchio, M. L.; Passeggi, M. C.; Barandiaran, M. J.; Gugliotta, 1636 L. M.; Minari, R. J. Acrylic/casein latexes with controlled degree of 1637 grafting and improved coating performance. *Prog. Org. Coat.* **2016**, 1638 *101*, 587–596.
- 1639 (103) Xu, Q.; Ma, J.; Zhou, J.; Wang, Y.; Zhang, J. Bio-based core—1640 shell casein-based silica nano-composite latex by double-in situ 1641 polymerization: Synthesis, characterization and mechanism. *Chem.* 1642 *Eng. J.* **2013**, 228, 281–289.

- (104) Zhang, F.; Ma, J.; Xu, Q.; Zhou, J.; Simion, D.; Carmen, G.; 1643 Wang, J.; Li, Y. Hollow Casein-Based Polymeric Nanospheres for 1644 Opaque Coatings. ACS Appl. Mater. Interfaces 2016, 8, 11739–11748. 1645 (105) Ramos, O.; Pereira, R.; Rodrigues, R.; Teixeira, J.; Vicente, A.; 1646 Malcata, F. Encyclopedia of Food and Health; Elsevier, 2016; pp 498–1647 505.
- (106) Global Whey Protein Market Growth, Trends And Forecast 1649 (2015–2020). https://www.prnewswire.com/news-releases/global-1650 whey-protein-market---growth-trends-and-forecast-2015-2020-1651 300155575.html (accessed 2021/01/12).
- (107) Gautam, R. B.; Kumar, S. Development of Protein Based 1653 Films with Nanoparticle as Strengthening Material for Biodegradable 1654 Packaging - A Review. *Int. J. Agric. Innov. Res.* **2017**, *5*, 790–805. 1655
- (108) Guo, M.; Wang, G. Whey protein polymerisation and its 1656 applications in environmentally safe adhesives. *Int. J. Dairy Technol.* 1657 **2016**, 69, 481–488.
- (109) Tschabold, G. L.; Mueller, D. L. Adhesive from whey and a 1659 method of making it. US Patent US2624679A, 1953.
- (110) Le Tien, C.; Letendre, M.; Ispas-Szabo, P.; Mateescu, M. A.; 1661 Delmas-Patterson, G.; Yu, H.-L.; Lacroix, M. Development of 1662 Biodegradable Films from Whey Proteins by Cross-Linking and 1663 Entrapment in Cellulose. *J. Agric. Food Chem.* **2000**, *48*, 5566–5575. 1664
- (111) Khan, A.; Wang, C.; Sun, X.; Killpartrick, A.; Guo, M. 1665 Physicochemical and Microstructural Properties of Polymerized Whey 1666 Protein Encapsulated 3,30-Diindolylmethane Nanoparticles. *Molecules* 1667 **2019**, 24, 702.
- (112) Verma, D.; Gulati, N.; Kaul, S.; Mukherjee, S.; Nagaich, U. 1669 Protein Based Nanostructures for Drug Delivery. J. Pharm. (Cairo) 1670 **2018**, 2018, 1–18.
- (113) Shendurse, A. M.; Gopikrishna, G.; Patel, A. C.; Pandya, A. J. 1672 Milk protein based edible films and coatings—preparation, properties 1673 and food applications. *J. Nutr. Health Food Eng.* **2018**, *8*, 219—226. 1674
- (114) Oymaci, P.; Altinkaya, S. A. Improvement of barrier and 1675 mechanical properties of whey protein isolate based food packaging 1676 films by incorporation of zein nanoparticles as a novel bionano- 1677 composite. *Food Hydrocolloids* **2016**, *54*, 1–9.
- (115) Ramos, Ö. L.; Reinas, I.; Silva, S. I.; Fernandes, J. C.; 1679 Cerqueira, M. A.; Pereira, R. N.; Vicente, A. A.; Poças, M. F.; Pintado, 1680 M. E.; Malcata, F. X. Effect of whey protein purity and glycerol 1681 content upon physical properties of edible films manufactured 1682 therefrom. *Food Hydrocolloids* **2013**, *30*, 110–122.
- (116) Han, J. H.; Krochta, J. M. Wetting Properties and Water 1684 Vapor Permeability of Whey-Protein-Coated Paper. *Trans. ASAE* 1685 **1999**, 42, 1375–1382.
- (117) Hidalgo, J.; Jost, R. Kosmetische Oder Therapeutische 1687 Zusammensetzung Fuer Topische Anwendung (Cosmetic or therapeutic 1688 composition for topical use). DE Patent DE3001300C2, 1988.
- (118) Chan, W. Y.; Bochenski, T.; Schmidt, J. E.; Olsen, B. D. 1690 Peptide Domains as Reinforcement in Protein-Based Elastomers. ACS 1691 Sustainable Chem. Eng. 2017, 5, 8568–8578.
- (119) Sharma, S.; Luzinov, I. Whey based binary bioplastics. *J. Food* 1693 *Eng.* **2013**, *119*, 404–410.
- (120) Gao, Z.; Yu, G.; Bao, Y.; Guo, M. Whey-protein based 1695 environmentally friendly wood adhesives. *Pigm. Resin Technol.* **2011**, 1696 40, 42–48.
- (121) Srinivasa, P.; Ramesh, M.; Kumar, K.; Tharanathan, R. 1698 Properties and sorption studies of chitosan–polyvinyl alcohol blend 1699 films. *Carbohydr. Polym.* **2003**, *53*, 431–438.
- (122) Lacroix, M.; Le, T.; Ouattara, B.; Yu, H.; Letendre, M.; 1701 Sabato, S.; Mateescu, M.; Patterson, G. Use of -irradiation to produce 1702 films from whey, casein and soya proteins: structure and functionals 1703 characteristics. *Radiat. Phys. Chem.* **2002**, *63*, 827–832.
- (123) Guo, M.; Vayda, M. E.; Gao, Z. Whey-protein based 1705 environmentally friendly wood adhesives and methods of producing and 1706 using the same. US Patent US20120183794A1, 2012.
- (124) Kisku, S. K.; Swain, S. K. Poly(methyl methacrylate)/soy 1708 protein green composites as gas barrier materials. *Chin. J. Polym. Sci.* 1709 **2012**, 30, 397–404.

- 1711 (125) Liu, D.; Tian, H.; Zeng, J.; Chang, P. R. Core-Shell 1712 Nanoblends from Soy Protein/Polystyrene by Emulsion Polymer-1713 ization. *Macromol. Mater. Eng.* **2008**, 293, 714–721.
- 1714 (126) Fapeng, W.; Jifu, W.; Chunpeng, W.; Fuxiang, C.; Xiaohuan,
- 1715 L.; Jiuyin, P. Fabrication of soybean protein-acrylate composite mini-1716 emulsion toward wood adhesive. *Eur. J. Wood Wood Prod.* **2018**, *76*, 1717 305–313
- 1718 (127) Bhattacharjee, S.; Bong, D. Protein-Polymer Grafts via a Soy 1719 Protein Derived Macro-RAFT Chain Transfer Agent. *J. Polym.* 1720 Environ. **2011**, 19, 203–208.
- 1721 (128) Riley, R. R.; Coco, C. E. Grafted soy protein latex. US Patent 1722 US4607089A. 1986.
- 1723 (129) Coco, C. E.; Scacciaferro, L. M. Polymeric Pigments Used in 1724 Paper Coating Compositions and a Process for Their Preparation. US 1725 Patent US4963604A, 1990.
- 1726 (130) Zheng, P.; Zeng, Q.; Lin, Q.; Fan, M.; Zhou, J.; Rao, J.; Chen, 1727 N. Investigation of an ambient temperature-curable soy-based 1738 adhesive for wood composites. Int. J. Adhes. Adhes. 2019, 95, 102429.
- 1728 adhesive for wood composites. *Int. J. Adhes. Adhes.* 2019, 95, 102429. 1729 (131) Luque, G. C.; Stürtz, R.; Passeggi, M. C.; Gugliotta, L. M.;
- 1730 Gonzalez, V. D.; Minari, R. J. New hybrid acrylic/collagen 1731 nanocomposites and their potential use as bio-adhesives. *Int. J.*

1732 Adhes. Adhes. 2020, 100, 102624.

- 1733 (132) Allasia, M.; Passeggi, M. C. G.; Gugliotta, L. M.; Minari, R. J. 1734 Waterborne Hybrid Acrylic/Protein Nanocomposites with Enhanced 1735 Hydrophobicity by Incorporating a Water Repelling Protein. *Ind. Eng.* 1736 *Chem. Res.* **2019**, *58*, 21070–21079.
- 1737 (133) Kinsella, J. E. Functional properties of soy proteins. *J. Am. Oil* 1738 *Chem. Soc.* **1979**, *56*, 242–258.
- 1739 (134) Mateos-Aparicio, I.; Cuenca, A. R.; Villanueva-Suárez, M. J.; 1740 Zapata-Revilla, M. A. Soybean, a promising health source. *Nutr. Hosp.* 1741 **2008**, 23, 305–312.
- 1742 (135) Song, F.; Tang, D.-L.; Wang, X.-L.; Wang, Y.-Z. Biodegradable 1743 Soy Protein Isolate-Based Materials: A Review. *Biomacromolecules* 1744 **2011**, *12*, 3369–3380.
- 1745 (136) Deepmala, K. M.; Singh, V. K.; Chauhan, S.; Jain, N. Soy 1746 Protein Based Green Composite: A Review. Res. Rev. J. Mater. Sci. 1747 **2017**, 5, 66–77.
- 1748 (137) Tian, H.; Guo, G.; Fu, X.; Yao, Y.; Yuan, L.; Xiang, A. 1749 Fabrication, properties and applications of soy-protein-based materi-1750 als: A review. *Int. J. Biol. Macromol.* **2018**, *120*, 475–490.
- 1751 (138) Wang, Z.; Zhao, S.; Pang, H.; Zhang, W.; Zhang, S.; Li, J. 1752 Developing Eco-friendly High-Strength Soy Adhesives with Improved 1753 Ductility through Multiphase Core—Shell Hyperbranched Polysilox-

1754 ane. ACS Sustainable Chem. Eng. 2019, 7, 7784-7794.

- 1755 (139) Rahman, M. M.; Netravali, A. N.; Tiimob, B. J.; Rangari, V. K. 1756 Bioderived "Green" Composite from Soy Protein and Eggshell 1757 Nanopowder. ACS Sustainable Chem. Eng. 2014, 2, 2329–2337.
- 1758 (140) Tian, H.; Guo, G.; Xiang, A.; Zhong, W.-H. Intermolecular 1759 interactions and microstructure of glycerol-plasticized soy protein 1760 materials at molecular and nanometer levels. *Polym. Test.* **2018**, *67*, 1761 197–204.
- 1762 (141) Thakur, M. K.; Thakur, V. K.; Gupta, R. K.; Pappu, A. 1763 Synthesis and Applications of Biodegradable Soy Based Graft 1764 Copolymers: A Review. ACS Sustainable Chem. Eng. 2016, 4, 1–17.
- 1765 (142) Xi, D.; Yang, C.; Liu, X.; Chen, M.; Sun, C.; Xu, Y. Graft 1766 polymerization of styrene on soy protein isolate. *J. Appl. Polym. Sci.* 1767 **2005**, 98, 1457–1461.
- 1768 (143) Kaith, B. S.; Jindal, R.; Bhatia, J. K. Morphological and thermal 1769 evaluation of soy protein concentrate on graft copolymerization with 1770 ethylmethacrylate. *J. Appl. Polym. Sci.* **2011**, *120*, 2183–2190.
- 1771 (144) Thakur, V. K.; Thunga, M.; Madbouly, S. A.; Kessler, M. R. 1772 PMMA-g-SOY as a sustainable novel dielectric material. *RSC Adv.* 1773 **2014**, *4*, 18240.
- 1774 (145) Thakur, V. K.; Kessler, M. R. Synthesis and Characterization 1775 of AN-g-SOY for Sustainable Polymer Composites. *ACS Sustainable* 1776 *Chem. Eng.* **2014**, *2*, 2454–2460.
- 1777 (146) Shi, Z.; Reddy, N.; Shen, L.; Hou, X.; Yang, Y. Grafting 1778 soyprotein isolates with various methacrylates for thermoplastic 1779 applications. *Ind. Crops Prod.* **2014**, *60*, 168–176.

- (147) Retna, A. M.; Sophia, A.; Theivasanthi, T.; Gopinath, S. C. 1780 Performance of Biodegradable Soy-based Polymer and Nano- 1781 composite with Reduced Moisture Absorptivity. *Micro Nanosyst.* 1782 **2018**, 10, 40–46.
- (148) Thakur, V. K.; Kessler, M. R. Free radical induced graft 1784 copolymerization of ethyl acrylate onto Soy for multifunctional 1785 materials. *Mater. Today Commun.* **2014**, *1*, 34–41.
- (149) González, A.; Igarzabal, C. I. A. Study of Graft 1787 Copolymerization of Soy Protein-Methyl Methacrylate: Preparation 1788 and Characterization of Grafted Films. *J. Polym. Environ.* **2017**, 25, 1789 214–220.
- (150) Jong, L. Reinforcement Effect of Soy Protein/Carbohydrate 1791 Ratio in Styrene—Butadiene Polymer. *J. Elastomers Plast.* **2011**, 43, 1792 99–117.
- (151) Qi, G.; Sun, X. S. Soy Protein Adhesive Blends with Synthetic 1794 Latex on Wood Veneer. J. Am. Oil Chem. Soc. 2011, 88, 271–281. 1795
- (152) Wang, F.; Wang, J.; Chu, F.; Wang, C.; Jin, C.; Wang, S.; 1796 Pang, J. Combinations of soy protein and polyacrylate emulsions as 1797 wood adhesives. *Int. J. Adhes. Adhes.* **2018**, *82*, 160–165.
- (153) Feng, B.; Wang, D.; Li, Y.; Qian, J.; Yu, C.; Wang, M.; Luo, 1799 D.; Wei, S. Mechanical Properties of a Soy Protein Isolate—Grafted—1800 Acrylate (SGA) Copolymer Used for Wood Coatings. *Polymers* **2020**, 1801 12. 1137.
- (154) Browning, J. D.; Vuayendran, B. R. Protein stabilized latex 1803 polymer emulsions, method of making, and adhesives containing such 1804 emulsions. WO Patent WO2008112644A1, 2008.
- (155) Paul, R. G.; Bailey, A. J. Chemical Stabilisation of Collagen as 1806 a Biomimetic. Sci. World J. 2003, 3, 138–155.
- (156) Nishi, Y.; Doi, m.; Uchiyama, S.; Nishiuchi, Y.; Nakazawa, T.; 1808 Ohkubo, T.; Kobayashi, Y. Stabilization mechanism of triple helical 1809 structure of collagen molecules. *Lett. Pept. Sci.* **2003**, *10*, 533–537. 1810
- (157) Orgel, J. P. R. O.; Antonio, J. D. S.; Antipova, O. Molecular 1811 and structural mapping of collagen fibril interactions. *Connect. Tissue* 1812 *Res.* **2011**, *52*, 2–17.
- (158) Usha, R.; Ramasami, T. Structure and conformation of 1814 intramolecularly cross-linked collagen. *Colloids Surf., B* **2005**, *41*, 21–1815 24.
- (159) Nakamura, Y. Structure of type I collagen dimers. *Int. J. Biol.* 1817 *Macromol.* **1987**, *9*, 281–290.
- (160) Goh, K.; Hiller, J.; Haston, J.; Holmes, D.; Kadler, K.; 1819 Murdoch, A.; Meakin, J.; Wess, T. Analysis of collagen fibril diameter 1820 distribution in connective tissues using small-angle X-ray scattering. 1821 Biochim. Biophys. Acta, Gen. Subj. 2005, 1722, 183–188.
- (161) Fraser, R.; MacRae, T.; Suzuki, E. Chain conformation in the 1823 collagen molecule. *J. Mol. Biol.* 1979, 129, 463–481.
- (162) Bella, J. A new method for describing the helical conformation 1825 of collagen: Dependence of the triple helical twist on amino acid 1826 sequence. *J. Struct. Biol.* **2010**, *170*, 377–391.
- (163) Flory, P. J.; Weaver, E. S. Helix [UNK] Coil Transitions in 1828 Dilute Aqueous Collagen Solutions 1. J. Am. Chem. Soc. 1960, 82, 1829 4518–4525.
- (164) Collagen Market revenue to hit \$6 billion by 2026, Says Global 1831 Market Insights, Inc. https://www.globenewswire.com/news-release/ 1832 2020/05/21/2036847/0/en/Collagen-Market-revenue-to-hit-6- 1833 billion-by-2026-Says-Global-Market-Insights-Inc.html, (accessed 1834 2021/02/20). 1835
- (165) Schmidt, M. M.; Dornelles, R. C. P.; Mello, R. O.; Kubota, E. 1836 H.; Mazutti, M. A.; Kempka, A. P.; Demiate, I. M. Collagen extraction 1837 process. *Int. Food Res. J.* **2016**, 23, 913–922.
- (166) Rashid, T. U.; Sharmeen, S.; Biswas, S.; Ahmed, T.; Mallik, A. 1839 K.; Shahruzzaman, M.; Sakib, M. N.; Haque, P.; Rahman, M. M. 1840 *Polymers and Polymeric Composites: A Reference Series*; Springer 1841 International Publishing, 2018; pp 1–41.
- (167) Sionkowska, A. Current research on the blends of natural and 1843 synthetic polymers as new biomaterials: Review. *Prog. Polym. Sci.* 1844 **2011**, 36, 1254–1276.
- (168) Barbani, N.; Bertoni, F.; Ciardelli, G.; Cristallini, C.; Silvestri, 1846 D.; Coluccio, M.; Giusti, P. Bioartificial materials based on blends of 1847

- 1848 dextran and poly(vinyl alcohol-co-acrylic acid). Eur. Polym. J. 2005, 1849 41, 3004–3010.
- 1850 (169) Sionkowska, A.; Kaczmarek, H.; Wiśniewski, M.; Kowalonek, 1851 J.; Skopinska, J. Surface characteristics of UV-irradiated collagen/PVP 1852 blended films. *Surf. Sci.* **2004**, *566*–*568*, 608–612.
- 1853 (170) Alexy, P.; Bakoš, D.; Hanzelová, S.; Kukolíková, L.; Kupec, J.; 1854 Charvátová, K.; Chiellini, E.; Cinelli, P. Poly(vinyl alcohol)—collagen 1855 hydrolysate thermoplastic blends: I. Experimental design optimization
- 1856 and biodegradation behaviour. Polym. Test. 2003, 22, 801-809.
- 1857 (171) Sionkowska, A.; Skopinska-Wisniewska, J.; Wisniewski, M. 1858 Collagen—synthetic polymer interactions in solution and in thin films. 1859 *J. Mol. Liq.* **2009**, *145*, 135–138.
- 1860 (172) Rogovina, S. Z.; Vikhoreva, G. A. Polysaccharide-based 1861 polymer blends: Methods of their production. *Glycoconjugate J.* **2006**, 1862 23, 611–618.
- 1863 (173) Sam, S. T.; Nuradibah, M. A.; Ismail, H.; Noriman, N. Z.; 1864 Ragunathan, S. Recent Advances in Polyolefins/Natural Polymer 1865 Blends Used for Packaging Application. *Polym.-Plast. Technol. Eng.* 1866 **2014**, 53, 631–644.
- 1867 (174) Deselnicu, V.; Deselnicu, D. C.; Vasilescu, A. M.; Crudu, M.; 1868 Albu, L. Polymer collagen Biocomposites. *Mater. Plast.* **2015**, *52*, 1869 159–164.
- 1870 (175) Wang, H.; Boerman, O. C.; Sariibrahimoglu, K.; Li, Y.; Jansen, 1871 J. A.; Leeuwenburgh, S. C. Comparison of micro- vs. nanostructured 1872 colloidal gelatin gels for sustained delivery of osteogenic proteins: 1873 Bone morphogenetic protein-2 and alkaline phosphatase. *Biomaterials* 1874 **2012**, 33, 8695–8703.
- 1875 (176) Giusti, P.; Lazzeri, L.; Barbani, N.; Narducci, P.; Bonaretti, A.; 1876 Palla, M.; Lelli, L. Hydrogels of poly(vinyl alcohol) and collagen as 1877 new bioartificial materials. *J. Mater. Sci.: Mater. Med.* **1993**, *4*, 538–1878 542.
- 1879 (177) Hoch, E.; Schuh, C.; Hirth, T.; Tovar, G. E. M.; Borchers, K. 1880 Stiff gelatin hydrogels can be photo-chemically synthesized from low 1881 viscous gelatin solutions using molecularly functionalized gelatin with 1882 a high degree of methacrylation. *J. Mater. Sci.: Mater. Med.* **2012**, 23, 1883 2607–2617.
- 1884 (178) Ramaraj, B.; Rajalingam, P.; Radhakrishnan, G. Crosslinked 1885 latex blends based on gelatin: Synthesis, morphology, thermal, and 1886 mechanical properties. *J. Appl. Polym. Sci.* **1991**, 43, 23–28.
- 1887 (179) Ye, X.; Zhou, Y.; Chen, J.; Sun, Y. Synthesis and infrared 1888 emissivity study of collagen-g-PMMA/Ag@TiO2 composite. *Mater.* 1889 *Chem. Phys.* **200**7, *106*, 447–451.
- 1890 (180) Zhang, Y.; Wang, Z.; Wang, Y.; Zhao, J.; Wu, C. Facile 1891 preparation of pH-responsive gelatin-based core—shell polymeric 1892 nanoparticles at high concentrations via template polymerization. 1893 *Polymer* **2007**, *48*, 5639–5645.
- 1894 (181) O'Connor, K. M.; Szajewski, R. P.; Bagchi, P. Control of 1895 pressure-fog with gelatin-grafted and case-hardened gelatin-grafted 1896 soft polymer latex particles. US Patent US5066572A, 1991.
- 1897 (182) Honan, J. S.; Walters, J. B.; Whitesides, T. H. Gelatin and 1898 polymer latex dispersion coating compositions. US Patent 1899 US5731134A, 1998.
- 1900 (183) Meyer, R.; Fowler, J. W. F. Vinylidene chloride-ethylenically 1901 unsaturated monomer-ethylenically unsaturated acid-gelatin emulsion 1902 polymerized coating composition. US Patent US3403116A, 1968.
- 1903 (184) Luo, Y.; Wang, T. Protein Byproducts; Elsevier, 2016; pp 147–1904 160.
- 1905 (185) Nonthanum, P.; Lee, Y.; Padua, G. W. Effect of -Zein on the 1906 Rheological Behavior of Concentrated Zein Solutions. *J. Agric. Food* 1907 *Chem.* **2012**, *60*, 1742–1747.
- 1908 (186) Demir, M.; Ramos-Rivera, L.; Silva, R.; Nazhat, S. N.; 1909 Boccaccini, A. R. Zein-based composites in biomedical applications. *J.* 1910 *Biomed. Mater. Res., Part A* **2017**, *105*, 1656–1665.
- 1911 (187) Bouman, J.; Belton, P.; Venema, P.; van der Linden, E.; de 1912 Vries, R.; Qi, S. Controlled Release from Zein Matrices: Interplay of 1913 Drug Hydrophobicity and pH. *Pharm. Res.* **2016**, *33*, *673*–685.
- 1914 (188) Dong, J.; Sun, Q.; Wang, J.-Y. Basic study of corn protein, 1915 zein, as a biomaterial in tissue engineering, surface morphology and 1916 biocompatibility. *Biomaterials* **2004**, *25*, 4691–4697.

- (189) Weissmueller, N. T.; Lu, H. D.; Hurley, A.; Prud'homme, R. 1917 K. Nanocarriers from GRAS Zein Proteins to Encapsulate Hydrophobic Actives. *Biomacromolecules* **2016**, *17*, 3828–3837.
- (190) Luo, Y.; Teng, Z.; Wang, Q. Development of Zein 1920 Nanoparticles Coated with Carboxymethyl Chitosan for Encapsula- 1921 tion and Controlled Release of Vitamin D3. *J. Agric. Food Chem.* **2012**, 1922 60, 836–843.
- (191) Davidov-Pardo, G.; Joye, I. J.; McClements, D. J. Advances in 1924 Protein Chemistry and Structural Biology; Elsevier, 2015; pp 293–325. 1925
- (192) Paliwal, R.; Palakurthi, S. Zein in controlled drug delivery and 1926 tissue engineering. *J. Controlled Release* **2014**, *189*, 108–122.
- (193) Chen, H.; Wang, J.; Cheng, Y.; Wang, C.; Liu, H.; Bian, H.; 1928 Pan, Y.; Sun, J.; Han, W. Application of Protein-Based Films and 1929 Coatings for Food Packaging: A Review. *Polymers* **2019**, *11*, 2039.
- (194) Kasaai, M. R. Zein and zein -based nano-materials for food 1931 and nutrition applications: A review. *Trends Food Sci. Technol.* **2018**, 1932 79, 184–197.
- (195) Tian, H.; Fu, X.; Zheng, M.; Wang, Y.; Li, Y.; Xiang, A.; 1934 Zhong, W.-H. Natural polypeptides treat pollution complex: 1935 Moisture-resistant multi-functional protein nanofabrics for sustainable 1936 air filtration. *Nano Res.* **2018**, *11*, 4265–4277.
- (196) Lee, H.; Xu, G.; Kharaghani, D.; Nishino, M.; Song, K. H.; 1938 Lee, J. S.; Kim, I. S. Electrospun tri-layered zein/PVP-GO/zein 1939 nanofiber mats for providing biphasic drug release profiles. *Int. J.* 1940 *Pharm.* **2017**, *531*, 101–107.
- (197) Khatri, M.; Khatri, Z.; El-Ghazali, S.; Hussain, N.; Qureshi, U. 1942 A.; Kobayashi, S.; Ahmed, F.; Kim, I. S. Zein nanofibers via deep 1943 eutectic solvent electrospinning: tunable morphology with super 1944 hydrophilic properties. *Sci. Rep.* **2020**, *10*, 15307.
- (198) Deng, L.; Zhang, X.; Li, Y.; Que, F.; Kang, X.; Liu, Y.; Feng, 1946 F.; Zhang, H. Characterization of gelatin/zein nanofibers by hybrid 1947 electrospinning. *Food Hydrocolloids* **2018**, 75, 72–80.
- (199) Ansari, A. Q.; Ansari, S. J.; Khan, M. Q.; Khan, M. F.; Qureshi, 1949 U. A.; Khatri, Z.; Ahmed, F.; Kim, I. S. Electrospun Zein nanofibers as 1950 drug carriers for controlled delivery of Levodopa in Parkinson 1951 syndrome. *Mater. Res. Express* **2019**, *6*, 075405.
- (200) Li, S.; Wang, X.; Li, W.; Yuan, G.; Pan, Y.; Chen, H. 1953 Preparation and characterization of a novel conformed bipolymer 1954 paclitaxel-nanoparticle using tea polysaccharides and zein. *Carbohydr.* 1955 *Polym.* **2016**, 146, 52–57.
- (201) Hu, K.; Huang, X.; Gao, Y.; Huang, X.; Xiao, H.; 1957 McClements, D. J. Core-shell biopolymer nanoparticle delivery 1958 systems: Synthesis and characterization of curcumin fortified zein—1959 pectin nanoparticles. *Food Chem.* 2015, 182, 275–281.
- (202) Ji, N.; Hong, Y.; Gu, Z.; Cheng, L.; Li, Z.; Li, C. Preparation 1961 and Characterization of Insulin-Loaded Zein/Carboxymethylated 1962 Short-Chain Amylose Complex Nanoparticles. *J. Agric. Food Chem.* 1963 **2018**, *66*, 9335–9343.
- (203) Hu, K.; McClements, D. J. Fabrication of biopolymer 1965 nanoparticles by antisolvent precipitation and electrostatic deposition: 1966 Zein-alginate core/shell nanoparticles. *Food Hydrocolloids* **2015**, 44, 1967 101–108.
- (204) Patel, A. R.; Bouwens, E. C. M.; Velikov, K. P. Sodium 1969 Caseinate Stabilized Zein Colloidal Particles. *J. Agric. Food Chem.* 1970 **2010**, *58*, 12497–12503.
- (205) Li, F.; Chen, Y.; Liu, S.; Qi, J.; Wang, W.; Wang, C.; Zhong, 1972 R.; Chen, Z.; Li, X.; Guan, Y.; Kong, W.; Zhang, Y. Size-controlled 1973 fabrication of zein nano/microparticles by modified anti-solvent 1974 precipitation with/without sodium caseinate. *Int. J. Nanomed.* **2017**, 1975 12, 8197–8209.
- (206) Li, H.; Xu, Y.; Sun, X.; Wang, S.; Wang, J.; Zhu, J.; Wang, D.; 1977 Zhao, L. Stability, bioactivity, and bioaccessibility of fucoxanthin in 1978 zein-caseinate composite nanoparticles fabricated at neutral pH by 1979 antisolvent precipitation. *Food Hydrocolloids* **2018**, *84*, 379–388.
- (207) Li, K.-K.; Yin, S.-W.; Yang, X.-Q.; Tang, C.-H.; Wei, Z.-H. 1981 Fabrication and Characterization of Novel Antimicrobial Films 1982 Derived from Thymol-Loaded Zein—Sodium Caseinate (SC) Nano- 1983 particles. J. Agric. Food Chem. 2012, 60, 11592—11600.

- 1985 (208) Bao, X.; Qian, K.; Yao, P. Oral delivery of exenatide-loaded 1986 hybrid zein nanoparticles for stable blood glucose control and -cell 1987 repair of type 2 diabetes mice. *J. Nanobiotechnol.* **2020**, *18*, 67.
- 1988 (209) Luo, Y.; Zhang, B.; Whent, M.; Yu, L. L.; Wang, Q. 1989 Preparation and characterization of zein/chitosan complex for 1990 encapsulation of -tocopherol, and its in vitro controlled release 1991 study. *Colloids Surf., B* **2011**, *85*, 145–152.
- 1992 (210) Wang, L.-J.; Yin, S.-W.; Wu, L.-Y.; Qi, J.-R.; Guo, J.; Yang, X.-1993 Q. Fabrication and characterization of Pickering emulsions and oil 1994 gels stabilized by highly charged zein/chitosan complex particles 1995 (ZCCPs). Food Chem. 2016, 213, 462–469.
- 1996 (211) Li, S.; Huang, L.; Zhang, B.; Chen, C.; Fu, X.; Huang, Q. 1997 Fabrication and characterization of starch/zein nanocomposites with 1998 pH-responsive emulsion behavior. *Food Hydrocolloids* **2021**, *112*, 1999 106341.
- 2000 (212) Zou, Y.; Guo, J.; Yin, S.-W.; Wang, J.-M.; Yang, X.-Q. 2001 Pickering Emulsion Gels Prepared by Hydrogen-Bonded Zein/Tannic 2002 Acid Complex Colloidal Particles. *J. Agric. Food Chem.* **2015**, *63*, 2003 7405–7414.
- 2004 (213) Li, K.-K.; Zhang, X.; Huang, Q.; Yin, S.-W.; Yang, X.-Q.; Wen, 2005 Q.-B.; Tang, C.-H.; Lai, F.-R. Continuous preparation of zein colloidal 2006 particles by Flash NanoPrecipitation (FNP). *J. Food Eng.* **2014**, *127*, 2007 103–110.
- 2008 (214) Ebert, S.; Koo, C. K.; Weiss, J.; McClements, D. J. Continuous 2009 production of core-shell protein nanoparticles by antisolvent 2010 precipitation using dual-channel microfluidization: Caseinate-coated 2011 zein nanoparticles. *Food Res. Int.* **2017**, *92*, 48–55.
- 2012 (215) Feng, J.; Zhang, Y.; McManus, S. A.; Qian, R.; Ristroph, K. D.; 2013 Ramachandruni, H.; Gong, K.; White, C. E.; Rawal, A.; Prud'homme, 2014 R. K. Amorphous nanoparticles by self-assembly: processing for 2015 controlled release of hydrophobic molecules. *Soft Matter* **2019**, *15*, 2016 2400–2410.
- 2017 (216) Xue, L.-H.; Xie, C.-Y.; Meng, S.-X.; Bai, R.-X.; Yang, X.; Wang, 2018 Y.; Wang, S.; Binks, B. P.; Guo, T.; Meng, T. Polymer—Protein 2019 Conjugate Particles with Biocatalytic Activity for Stabilization of 2020 Water-in-Water Emulsions. ACS Macro Lett. 2017, 6, 679—683.
- 2021 (217) Zhang, M. Biological and Medical Physics, Biomedical 2022 Engineering; Springer Berlin Heidelberg, 2004; pp 83–143.
- 2023 (218) Jaromir, K.; Atkins, G. M. Encapsulation. US Patent 2024 US3406119A, 1968.
- 2025 (219) Hardy, J. G.; Scheibel, T. R. Composite materials based on silk 2026 proteins. *Prog. Polym. Sci.* **2010**, *35*, 1093–1115.
- 2027 (220) Vendrely, C.; Scheibel, T. Biotechnological Production of 2028 Spider-Silk Proteins Enables New Applications. *Macromol. Biosci.* 2029 **2007**, 7, 401–409.
- 2030 (221) Sun, Y.; Shao, Z.; Hu, P.; Yu, T. Hydrogen bonds in silk 2031 fibroin-poly (acrylonitrile-co-methyl acrylate) blends: FT-IR study. *J.* 2032 *Polym. Sci., Part B: Polym. Phys.* **1997**, *35*, 1405–1414.
- 2033 (222) Meira, G.; Gugliotta, L. M. Polimeros. Introducción a Su 2034 Caracterización y a La Ingeniería de La Polimerización (Polymers. 2035 Introduction to Its Characterization and Polymerization Engineering); 2036 Cátedra Ediciones UNL: Santa Fe, Argentina, 2019.
- 2037 (223) Principles and Applications of Emulsion Polymerization; John 2038 Wiley & Sons, Inc., pp 53–94.
- 2039 (224) van Herk, A. Chemistry and Technology of Emulsion 2040 Polymerisation; Wiley, 2013.
- 2041 (225) Kolthoff, I. M.; Miller, I. K. The Chemistry of Persulfate. II. 2042 The Reaction of Persulfate with Mercaptans Solubilized in Solutions 2043 of Saturated Fatty Acid Soaps1. *J. Am. Chem. Soc.* **1951**, 73, 5118–2044 5122
- 2045 (226) Khorana, H. G. The Chemistry of Carbodiimides. *Chem. Rev.* 2046 **1953**, 53, 145–166.
- 2047 (227) Asua, J. M. Miniemulsion polymerization. *Prog. Polym. Sci.* 2048 **2002**, 27, 1283–1346.
- 2049 (228) Córdoba, C. A.; Ronco, L. I.; Barrios, C. E.; Gugliotta, L. M.; 2050 Minari, R. J. High Solid Acrylic–Melamine Latexes with Tunable 2051 Crosslinking Capability. *Macromol. React. Eng.* **2019**, *13*, 1800063.

- (229) Kessel, N.; Illsley, D. R.; Keddie, J. L. The diacetone 2052 acrylamide crosslinking reaction and its influence on the film 2053 formation of an acrylic latex. *J. Coat. Technol. Res.* **2008**, *5*, 285–297. 2054 (230) Chen, H.; Zhong, Q. Processes improving the dispersibility of 2055 spray-dried zein nanoparticles using sodium caseinate. *Food Hydro-2056 colloids* **2014**, *35*, 358–366.
- (231) Chang, C.; Wang, T.; Hu, Q.; Zhou, M.; Xue, J.; Luo, Y. 2058 Pectin coating improves physicochemical properties of caseinate/zein 2059 nanoparticles as oral delivery vehicles for curcumin. *Food Hydro-2060 colloids* **2017**, *70*, 143–151.