Accepted Manuscript

Characterization of a multidrug resistant Citrobacter amalonaticus clinical isolate harboring blaNDM-1 and mcr-1.5 genes

Diego Faccone, Ezequiel Albornoz, Nathalie Tijet, Estefania Biondi, Sonia Gomez, Fernando Pasterán, Miryam Vazquez, Roberto G. Melano, Alejandra Corso



PII: S1567-1348(18)30827-X

DOI: https://doi.org/10.1016/j.meegid.2018.10.020

Reference: MEEGID 3691

To appear in: Infection, Genetics and Evolution

Received date: 25 April 2018
Revised date: 12 October 2018
Accepted date: 26 October 2018

Please cite this article as: Diego Faccone, Ezequiel Albornoz, Nathalie Tijet, Estefania Biondi, Sonia Gomez, Fernando Pasterán, Miryam Vazquez, Roberto G. Melano, Alejandra Corso , Characterization of a multidrug resistant Citrobacter amalonaticus clinical isolate harboring blaNDM-1 and mcr-1.5 genes. Meegid (2018), https://doi.org/10.1016/j.meegid.2018.10.020

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Characterization of a Multidrug Resistant Citrobacter amalonaticus Clinical Isolate

Harboring bla_{NDM-1} and mcr-1.5 Genes

Diego Faccone, a,b Ezequiel Albornoz, Nathalie Tijet, Estefania Biondi, Sonia Gomez, b

Fernando Pasterán, a Miryam Vazquez, Roberto G. Melano, c,e Alejandra Corso H

^aServicio Antimicrobianos, National and Regional Reference Laboratory in Antimicrobial

Resistance, Instituto Nacional de Enfermedades Infecciosas (INEI)-ANLIS "Dr. C. Malbrán"

^bConsejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Buenos Aires, Argentina

^cPublic Health Ontario Laboratory, Toronto, Ontario, Canada

d Hospital de Niños Dr. Ricardo Gutiérrez, Buenos Aires, Argentina

^eDepartment of Laboratory Medicine and Pathobiology, University of Toronto, Toronto,

Ontario, Canada.

Address correspondence to Alejandra Corso, acorso@anlis.gov.ar

Keywords: mcr-1; blaNDM; carbapenem; colistin; Citrobacter amalonaticus

1

ABSTRACT

A multidrug resistant isolate, identified as *Citrobacter amalonaticus* using MALDI-TOF MS and confirmed by genomic analysis, was recovered from a pediatric patient in a hospital from Buenos Aires, Argentina. By whole-genome sequencing a total of 16 resistance genes were detected, including bla_{NDM-1} and mcr-1.5. To the best of our knowledge this is the first description of these two genes together in a clinical isolate of the *Citrobacter* genus.

INTRODUCTION

Citrobacter is a gram-negative coliform bacterial genus comprising 14 species that have been isolated from water, soil, food and the gastrointestinal tract of humans and animals (Clermont et al., 2015). C. freundii and C. koseri are the most common species recovered from human infections while C. amalonaticus is seldom isolated from faecal and urine samples and only occasionally associated with invasive infections, like osteomyelitis, arthritis and peritonitis (Garcia et al., 2016; Maraki et al., 2017; Wong et al., 2012). Citrobacter spp. are usually susceptible to third-generation cephalosporins, carbapenem and colistin (Maraki et al., 2017). However, carbapenem resistant Citrobacter spp. isolates have been reported due to the acquisition of worldwide disseminated carbapenemases, such as New Delhi Metallo-βlactamase (NDM) (Arana et al., 2017; Hammerum et al., 2016; Pasteran et al., 2016). In the last few years, a transferable colistin-resistant mechanism mediated by mcr genes has been recently described (8). mcr-1 gene has been found in several Enterobacteriaceae genus and species but it has mainly been detected in Escherichia coli from animal, food, environmental and human sources (Al-Tawfig et al., 2017; Rapoport et al., 2016; Saavedra et al., 2017). Here, we report the characterization of a clinical multidrug resistant (MDR) C. amalonaticus isolate carrying bla_{NDM-1} and mcr-1.5 genes.

MATERIALS and METHODS

On February 2016, isolate M21015 was recovered in a pediatric hospital in Buenos Aires from an oncologic four-year old patient, during an active carbapenemase surveillance program for asymptomatic rectal colonization, and confirmed as *C. amalonaticus* by Vitek2, VitekMS (bioMérieux) and MALDI-TOF MS Biotyper 3.0 (Bruker Daltonics). The isolate was sent to the Argentinian National Reference Laboratory for Antimicrobial Resistance for further characterization.

Antimicrobial susceptibility was evaluated by Sensititre ARGNF customized plates (Trek Diagnostic Systems, Thermo Fisher Scientific), and interpreted according to the Clinical and Laboratory Standards Institute or the European Committee on Antimicrobial Susceptibility Testing (colistin and tigecycline) criteria (Clinical and Laboratory Standards Institute (CLSI)., 2017; "The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 7.0, 2017. http://www.eucast.org.," n.d.). The imipenem-EDTA double-disk synergy test was used to detect metallo-ß-lactamase (MBL) activity. The presence of MBL (bla_{NDM} , bla_{IMP} and bla_{VIM}) and mcr-1 genes were detected by in house PCR.

C. amalonaticus M21015 was analysed by whole genome sequencing (WGS) using Nextera XT for library preparation and Illumina compact MiSeq system for sequencing. Plasmids containing *mcr* and bla_{NDM} genes were independently transferred by biparental conjugation to *E. coli* J53, and the transconjugants selected with colistin (2 µg/ml) or meropenem (0.2 µg/ml), respectively, and sodium azide (100 µg/ml)(Gomez et al., 2011). Plasmids were extracted and purified for Illumina sequencing, and analyzed as previously described (Tijet et al., 2016). Antimicrobial resistance genes were predicted using the ResFinder, version 3.0 (Zankari et al., 2012).

RESULTS and DISCUSSION

C. amalonaticus M21015 was resistant (µg/mL) to imipenem (≥16), meropenem (≥32), cefotaxime (≥64), ceftazidime (≥64), cefepime (≥32), amikacin (≥64), gentamicin (≥16), trimethoprim-sulfamethoxazole (≥4) and colistin (≥8), and susceptible to aztreonam (≤8), ciprofloxacin (≤0.06), fosfomycin (≤32), minocycline (≤4) and tigecycline (≤0.5). MBL inhibition was associated with the presence of bla_{NDM} gene. Resistance to colistin was associated with the presence of mcr-1-like gene.

Two plasmids were transferred individually from *C. amalonaticus* M21015 to *E. coli* J53 by conjugation, and subsequently sequenced. One of them, named here pMCR-M21015, harboured the *mcr-1.5* variant(Tijet et al., 2017). pMCR-M21015 (62,891 bp) belonged to the incompatibility group Incl2 sharing >99.9% identity with pMCR-M15049 plasmid (accession number KY471308) of *E. coli* M15049 also isolated in Argentina(Tijet et al., 2017). *mcr-1.5* and *pap2* genes were identified in a 2,749 bp contig. The presence of two copies of IS*Apl1* in the same orientation flanking them was verified by PCR and Sanger sequencing, as well as the 2-bp putative target site duplications, confirming the presence of the composite transposon Tn6330(Liu et al., 2016). No other antimicrobial resistance gene was identified in this plasmid (Suppl. Figure 1). The finding of *mcr-1* in *Citrobacter* spp. is infrequent and to the best of our knowledge has not been described in *C. amalonaticus*.

Sequencing of the second plasmid (named pNDM-M21015) showed that $bla_{\text{NDM-1}}$ is located in a type 1 IncC2 plasmid of 138,998 bp with 99% identity with pKP1-NDM-1 (KF992018). $bla_{\text{NDM-1}}$ was located within a truncated Tn125 structure previously reported paper de Wailan et al. (Wailan et al., 2015). pNDM-M21015 also contains $bla_{\text{CMY-6}}$, rmtC, sul1, and some small MDR genes encoding for the ethidium bromide-methyl viologen resistance protein EmrE and the quaternary ammonium compound-resistance protein SugE, as well as the bleomycin resistance protein (ble_{MBL}) associated to bla_{NDM} (Suppl. Figure 2).

In addition, other mechanisms of resistance were identified by WGS analysis (Table 1): to aminoglycosides, phenicols, trimethoprim, sulphonamides and β -lactams (including bla_{CdiA} , a

chromosomally encoded class A β -lactamase similar to Sed-1 from *Citrobacter sedlakii*)(Petrella et al., 2001). An efflux pump similar to OqxA/B (~90% nucleotide identity with OqxA/B found on *K. pneumoniae* chromosomes) was also identified, but the MIC for ciprofloxacin suggests that it is not active against fluoroquinolones.

The identification to the species level of *Citrobacter* is controversial (Ribeiro et al., 2015). The conventional biochemical characterization and MALDI-TOF MS methods yield reliable results with approx. 90% and 95% accuracy for each methodology, respectively (Kolínská et al., 2015). The use of 16S rRNA gene sequences displays limited resolution distinguishing only three groups within the genus, where *C. amalonaticus* is included in group II together with *C. farmeri, C. sedlakii* and *C. rodentium* (Clermont et al., 2015). A previously described multilocus sequence analysis of a 2,082 bp concatenated sequence of *fusA*, *pyrG*, *leuS* and *rpoB* genes was used to confirm the isolate identity at the species level (Clermont et al., 2015). A phylogenetic analysis was performed including the sequences of 28 isolates representing 14 well defined *Citrobacter* species (Figure 1). *C. amalonaticus* M21015 grouped together with *C. amalonaticus* FDAARGOS165 and CIP 82.89 in group II (Figure 1), confirming the identification obtained by biochemical and proteomic methods.

C. amalonaticus is an opportunistic pathogen rarely associated with MDR phenotypes (Garcia et al., 2016; Maraki et al., 2017; Wong et al., 2012). To the best of our knowledge this is the first description of both bla_{NDM} and mcr-1-like genes in the Citrobacter genus, co-existing with other 14 determinants of antimicrobial resistance in this clinical isolate of C. amalonaticus. This emergence is coincident with other reports suggesting an increase in the isolation of MDR Citrobacter spp. (Arana et al., 2017; Hammerum et al., 2016; Pasteran et al., 2016). Further studies are required to determine its public health significance and the degree of dissemination since Citrobacter is not frequently included in surveillance programs for MDR organisms. Remarkably, a highly similar Incl2 pMCR plasmid was now described in two genera in Argentina, suggesting horizontal dissemination not only between E. coli isolates(Tijet et al.,

2017) but to other *Enterobacteriaceae*. Furthermore, the presence of *mcr-1.5* in an active composite transposon creates the potential for its dissemination to other plasmids, increasing the risk of a more effective spread of polymyxin resistance.

The following are the supplementary data related to this article.

Supplementary Table 1. Primers used to detect blaNDM, blaIMP, blaVIM and mcr-1 genes.

Suppl. Table 2. Antimicrobial susceptibility of C. amalonaticus M21015 (donor), E. coli J53 (acceptor), and two E. coli transconjugants.

Supplementary figures

KNOWLEDGE

Raw sequence reads of the whole genome of *C. amalonaticus* M21015 were deposited in the NCBI database under accession number PRJNA495344. Additionally plasmid sequences were submitted individually: pMCR-M21015 (submission ID: 2156778); and pNDM-M21015 (submission ID: 2157761).

FUNDING

This work was supported by Préstamo BID-PICT-2016-3154 to D.F. from ANPCYT; the regular federal budget of the National Ministry of Health of Argentina; and Public Health Ontario Laboratory, Toronto, ON, Canada.

TRANSPARENCY DECLARATIONS

Nothing to declare.

REFERENCES

- Al-Tawfiq, J.A., Laxminarayan, R., Mendelson, M., 2017. How should we respond to the emergence of plasmid-mediated colistin resistance in humans and animals? Int. J. Infect. Dis. 54, 77–84. https://doi.org/10.1016/j.ijid.2016.11.415
- Arana, D.M., Ortega, A., González-Barberá, E., Lara, N., Bautista, V., Gómez-Ruíz, D., Sáez, D., Fernández-Romero, S., Aracil, B., Pérez-Vázquez, M., Campos, J., Oteo, J., Spanish Antibiotic Resistance Surveillance Programme Collaborating Group, 2017.

 Carbapenem-resistant Citrobacter spp. isolated in Spain from 2013 to 2015 produced a variety of carbapenemases including VIM-1, OXA-48, KPC-2, NDM-1 and VIM-2. J. Antimicrob. Chemother. https://doi.org/10.1093/jac/dkx325
- Clermont, D., Motreff, L., Passet, V., Fernandez, J.-C., Bizet, C., Brisse, S., 2015. Multilocus sequence analysis of the genus Citrobacter and description of Citrobacter pasteurii sp. nov. Int. J. Syst. Evol. Microbiol. 65, 1486–1490. https://doi.org/10.1099/ijs.0.000122
- Clinical and Laboratory Standards Institute (CLSI)., 2017. Performance standards for antimicrobial susceptibility testing: Twenty-seventh informational supplement. CLSI document M100-S27:2017.
- Garcia, V., Abat, C., Moal, V., Rolain, J.-M., 2016. Citrobacter amalonaticus human urinary tract infections, Marseille, France. New Microbes New Infect 11, 1–5. https://doi.org/10.1016/j.nmni.2016.01.003
- Gomez, S., Rapoport, M., Togneri, A., Viegas-Caetano, J., Faccone, D., Corso, A., Petroni, A., Pasteran, F., 2011. Emergence of metallo-β-lactamases in Enterobacteriaceae from Argentina. Diagn. Microbiol. Infect. Dis. 69, 94–97. https://doi.org/10.1016/j.diagmicrobio.2010.08.025
- Hammerum, A.M., Hansen, F., Nielsen, H.L., Jakobsen, L., Stegger, M., Andersen, P.S., Jensen, P., Nielsen, T.K., Hansen, L.H., Hasman, H., Fuglsang-Damgaard, D., 2016. Use of WGS data for investigation of a long-term NDM-1-producing Citrobacter freundii outbreak and secondary in vivo spread of blaNDM-1 to Escherichia coli, Klebsiella pneumoniae and Klebsiella oxytoca. J Antimicrob Chemother 71, 3117–3124. https://doi.org/10.1093/jac/dkw289
- Kolínská, R., Spanělová, P., Dřevínek, M., Hrabák, J., Zemličková, H., 2015. Species identification of strains belonging to genus Citrobacter using the biochemical method and MALDI-TOF mass spectrometry. Folia Microbiol. (Praha) 60, 53–59. https://doi.org/10.1007/s12223-014-0340-4
- Liu, Y.-Y., Wang, Y., Walsh, T.R., Yi, L.-X., Zhang, R., Spencer, J., Doi, Y., Tian, G., Dong, B., Huang, X., Yu, L.-F., Gu, D., Ren, H., Chen, X., Lv, L., He, D., Zhou, H., Liang, Z., Liu, J.-H., Shen, J., 2016. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. Lancet Infect Dis 16, 161–168. https://doi.org/10.1016/S1473-3099(15)00424-7
- Maraki, S., Vardakas, K.Z., Mavromanolaki, V.-E., Kyriakidou, M., Spais, G., Kofteridis, D.P., Samonis, G., Falagas, M.E., 2017. In vitro susceptibility and resistance phenotypes in contemporary Citrobacter isolates in a University Hospital in Crete, Greece. Infect Dis (Lond) 49, 532–539. https://doi.org/10.1080/23744235.2017.1297896
- Pasteran, F., Gonzalez, L.J., Albornoz, E., Bahr, G., Vila, A.J., Corso, A., 2016. Triton Hodge Test: Improved Protocol for Modified Hodge Test for Enhanced Detection of NDM and Other Carbapenemase Producers. J. Clin. Microbiol. 54, 640–649. https://doi.org/10.1128/JCM.01298-15
- Petrella, S., Clermont, D., Casin, I., Jarlier, V., Sougakoff, W., 2001. Novel class A beta-lactamase Sed-1 from Citrobacter sedlakii: genetic diversity of beta-lactamases within the Citrobacter genus. Antimicrob. Agents Chemother. 45, 2287–2298. https://doi.org/10.1128/AAC.45.8.2287-2298.2001

- Rapoport, M., Faccone, D., Pasteran, F., Ceriana, P., Albornoz, E., Petroni, A., MCR Group, Corso, A., 2016. First Description of mcr-1-Mediated Colistin Resistance in Human Infections Caused by Escherichia coli in Latin America. Antimicrob. Agents Chemother. 60, 4412–4413. https://doi.org/10.1128/AAC.00573-16
- Ribeiro, T.G., Novais, Â., Branquinho, R., Machado, E., Peixe, L., 2015. Phylogeny and Comparative Genomics Unveil Independent Diversification Trajectories of qnrB and Genetic Platforms within Particular Citrobacter Species. Antimicrob. Agents Chemother. 59, 5951–5958. https://doi.org/10.1128/AAC.00027-15
- Saavedra, S.Y., Diaz, L., Wiesner, M., Correa, A., Arévalo, S.A., Reyes, J., Hidalgo, A.M., de la Cadena, E., Perenguez, M., Montaño, L.A., Ardila, J., Ríos, R., Ovalle, M.V., Díaz, P., Porras, P., Villegas, M.V., Arias, C.A., Beltrán, M., Duarte, C., 2017. Genomic and Molecular Characterization of Clinical Isolates of Enterobacteriaceae Harboring mcr-1 in Colombia, 2002 to 2016. Antimicrob. Agents Chemother. 61. https://doi.org/10.1128/AAC.00841-17
- Tamura, K., Stecher, G., Peterson, D., Filipski, A., Kumar, S., 2013. MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. Mol Biol Evol 30, 2725–2729. https://doi.org/10.1093/molbev/mst197
- The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 7.0, 2017. http://www.eucast.org., n.d.
- Tijet, N., Faccone, D., Rapoport, M., Seah, C., Pasterán, F., Ceriana, P., Albornoz, E., Corso, A., Petroni, A., Melano, R.G., 2017. Molecular characteristics of mcr-1-carrying plasmids and new mcr-1 variant recovered from polyclonal clinical Escherichia coli from Argentina and Canada. PLoS ONE 12, e0180347. https://doi.org/10.1371/journal.pone.0180347
- Tijet, N., Muller, M.P., Matukas, L.M., Khan, A., Patel, S.N., Melano, R.G., 2016. Lateral dissemination and inter-patient transmission of blaKPC-3: role of a conjugative plasmid in spreading carbapenem resistance. J. Antimicrob. Chemother. 71, 344–347. https://doi.org/10.1093/jac/dkv356
- Wailan, A.M., Sartor, A.L., Zowawi, H.M., Perry, J.D., Paterson, D.L., Sidjabat, H.E., 2015.
 Genetic Contexts of blaNDM-1 in Patients Carrying Multiple NDM-Producing Strains.
 Antimicrob. Agents Chemother. 59, 7405–7410. https://doi.org/10.1128/AAC.01319-15
- Wong, M.Y., Lau, S.K.P., Tang, S.C.W., Curreem, S.O.T., Woo, P.C.Y., Yuen, K.Y., 2012. First report of peritoneal dialysis-related peritonitis caused by Citrobacter amalonaticus. Perit Dial Int 32, 224–225. https://doi.org/10.3747/pdi.2011.00136
- Zankari, E., Hasman, H., Cosentino, S., Vestergaard, M., Rasmussen, S., Lund, O., Aarestrup, F.M., Larsen, M.V., 2012. Identification of acquired antimicrobial resistance genes. J. Antimicrob. Chemother. 67, 2640–2644. https://doi.org/10.1093/jac/dks261

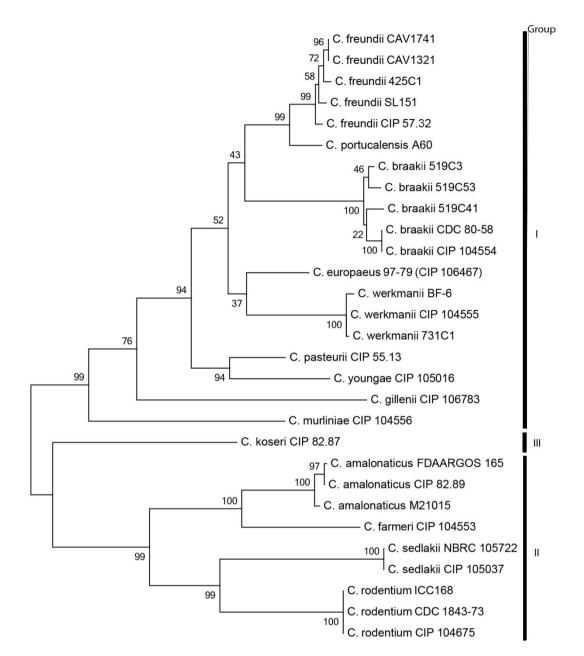


Figure 1. Phylogenetic analysis by maximum likelihood method of Citrobacter species.

FOOTNOTE. The evolutionary history was inferred by using the Maximum Likelihood method based on the Jukes-Cantor model. Branch lengths measured in the number of substitutions per site. The analysis included 29 nucleotide sequences representing 14 *Citrobacter* species. Groups I, II and III were described in the text. Phylogenetic analyses were conducted in MEGA6 (Tamura et al., 2013).

0.01

Table 1. Summary of resistance genes detected by plasmid and whole-genome sequencing

Antibiotic affected	Resistance gene	e Identity (%)	GenBank Accession no.
Polymyxins	mcr-1.5	100	KP347127
ß-Lactams	bla _{NDM-1}	100	FN396876
	bla _{CMY-6}	100	AJ011293
	bla _{TEM-1B}	100	JF910132
	$bla_{ extsf{SED-1}}$	82.64	AF321608
Aminoglycosides	rmtC	100	AB194779
	aadA1	100	JQ414041
	aadA2	100	JQ364967
	aacA4	100	KM278199
	aph(3')-Ia	100	V00359
Phenicols	floR	98.11	AF118107
	cmlA1	99.92	M64556
Sulphonamide	sul1	100	CP002151
	sul2	100	GQ421466
	sul3	100	AJ459418
Trimethoprim	dfrA12	100	AB571791

Highlights.

- 1. Multidrug Resistant Citrobacter amalonaticus Clinical Isolate
- 2. C. amalonaticus harbouring bla_{NDM-1} and mcr-1.5 genes
- 3. Specie identification was performed using MALDI-TOF MS and confirmed by sequencing



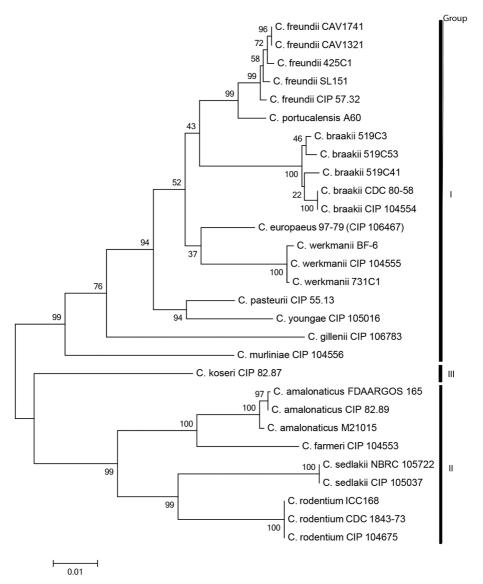


Figure 1