

Nutrition Research Reviews

Date of delivery:**Journal and vol/article ref:**

nrr 1500005

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Dietary and pharmacological compounds altering intestinal calcium absorption in humans and animals

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Abstract

The intestine is the only gate for the entry of Ca to the body in humans and mammals. The entrance of Ca occurs via paracellular and intracellular pathways. All steps of the latter pathway are regulated by calcitriol and by other hormones. Dietary and pharmacological compounds also modulate the intestinal Ca absorption process. Among them, dietary Ca and P are known to alter the lipid and protein composition of the brush-border and basolateral membranes and, consequently, Ca transport. Ca intakes are below the requirements recommended by health professionals in most countries, triggering important health problems. Chronic low Ca intake has been related to illness conditions such as osteoporosis, hypertension, renal lithiasis and incidences of human cancer. Carbohydrates, mainly lactose, and prebiotics have been described as positive modulators of intestinal Ca absorption. Apparently, high meat proteins increase intestinal Ca absorption while the effect of dietary lipids remains unclear. Pharmacological compounds such as menadione, DL-butionine-S,R-sulfoximine and ursodeoxycholic acid also modify intestinal Ca absorption as a consequence of altering the redox state of the epithelial cells. The paracellular pathway of intestinal Ca absorption is poorly known and is under present study in some laboratories. Another field that needs to be explored more intensively is the influence of the gene × diet interaction on intestinal Ca absorption. Health professionals should be aware of this knowledge in order to develop nutritional or medical strategies to stimulate the efficiency of intestinal Ca absorption and to prevent diseases.

Key words: Intestinal calcium absorption: Transcellular and paracellular pathways: Hormonal effects: Nutritional factors

Introduction

Ca is the main mineral component of bone and, hence, is essential for achieving optimal peak bone mass in the first decades of life and for maintaining bone mass, later in life⁽¹⁾. It also plays an important role in many physiological processes⁽²⁻⁵⁾. The dysregulation of Ca homeostasis is not only associated with bone disorders, but also with hypertension, insulin resistance, obesity and the metabolic syndrome⁽⁶⁻⁹⁾. Epidemiological and experimental studies have shown an inverse relationship between dietary Ca and risk of breast, colon, prostate and ovarian cancer⁽¹⁰⁻¹³⁾. Therefore, an appropriate Ca homeostasis preserves bone integrity, metabolic balance and avoids epithelial cancers.

Ca metabolism is predominantly regulated by the intestine, kidney, bone and parathyroid glands. Because of their coordinated work, serum Ca concentration is maintained within

a narrow range⁽¹⁴⁾. Intestinal Ca absorption is an essential process that occurs through an active transcellular pathway and a passive non-saturable route, named the paracellular pathway⁽¹⁵⁾. Both routes are regulated by hormones, nutrients and many other factors.

The transcellular pathway is a saturable process, which is prevalent in the proximal small intestine (duodenum and jejunum), vitamin D being the main modulator. This mechanism is energy dependent and implicates Ca movement from the mucosal to serosal side of the intestinal barrier occurring against a concentration gradient. In contrast, the paracellular mechanism occurs throughout the length of the intestine. It is a non-saturable and passive transport and is a linear function of Ca concentration in the lumen⁽¹⁶⁾.

Ca ions are absorbed mainly in the small intestine, which is responsible for about 90 % of overall Ca absorption. The longer residence time in the ileum as compared with the other

Abbreviations: 1,25(OH)₂D₃, 1,25-dihydroxycholecalciferol; Al, aluminium ions; AP, alkaline phosphatase; BMD, bone mineral density; BSO, dl-butionine-S,R-sulfoximine; CB, calbindin; CPP, caseinophosphopeptides; ER, oestrogen receptor; FCA, fractional Ca absorption; FGF-23, fibroblast growth factor-23; GC, glucocorticoid; GSH, glutathione; IGF-1, insulin-like growth factor-1; KO, knockout; MEL, melatonin; MEN, menadione; NaDOC, sodium deoxycholate; NCX1, intestinal Na⁺/Ca²⁺ exchanger; OVX, ovariectomised; PMCA, plasma membrane Ca²⁺ ATPase; PTH, parathyroid hormone; TRP, transient receptor potential; TRPV, transient receptor potential vanilloid; UDCA, ursodeoxycholic acid; VDD, vitamin D deficiency; VDR, vitamin D receptor.

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segments of the small intestine favours Ca absorption in that segment⁽¹⁶⁾. In rat ileum the transit half-time is about 100–120 min, whereas in the duodenum it is about 2–6 min⁽¹⁷⁾. The colon is responsible for less than 10 % of the total Ca absorbed; minor amounts of Ca ions are absorbed from the stomach and large intestine⁽¹⁶⁾. The major contributors to the amount of Ca absorbed are the residence time and the absorption rate in each intestinal segment. The order of Ca absorption rate is: duodenum > jejunum > ileum⁽¹⁶⁾. Ca absorption in the colon is probably very important in pathological conditions such as short bowel syndrome⁽¹⁸⁾.

Intestinal Ca absorption also depends on the physiological needs of Ca. When the requirements increase and/or the intakes are low, there is an improvement in the efficiency of Ca absorption⁽¹⁹⁾. Ageing occurs with a decrease in intestinal Ca absorption⁽²⁰⁾, while growth, pregnancy and lactation promote cation absorption^(21–24). There is little information about the mechanism of lactation-induced intestinal hyperabsorption. Increases have been found in the villous height, villous width and crypt depth with an expansion of the absorptive surface area in the duodenum of 21 d lactating rats. An enhancement in claudin 15 has been also demonstrated in the same animal model⁽²⁵⁾. Recently, Teerapornpantakit *et al.*⁽²⁶⁾ have developed a custom-designed cDNA microarray (CalGene Array) to study the expression of genes related to duodenal nutrient transport, among them those related to bone and Ca metabolism. They have determined the transcriptome responses of duodenal epithelial cells in pregnant and lactating rats; data were subsequently validated by quantitative real-time PCR. They have found that pregnancy and late lactation alter the expression of several transcripts, among them those belonging to Ca transporters.

Transcellular pathway

Epithelial calcium channels

TRPV6 (previously named ECaC2 or CaT1) and TRPV5 (previously named ECaC1 or CaT2) are the two epithelial Ca channels involved in Ca entry to the enterocytes. These channels are homologous members of the transient receptor potential (TRP) superfamily, belonging to the vanilloid subfamily (TRPV), which is different from the canonical (TRPC) and melastatin (TRPM) subfamilies⁽²⁷⁾. TRPV6 and TRPV5 are co-expressed in the human kidney and intestine, but the first one is highly expressed in the intestine and the latter is the major isoform in the kidney. TRPV6 seems to be a major contributor to apical, intestinal Ca absorption, as suggested by a significant reduction in Ca absorption and serum Ca shown in TRPV6 knockout (KO) mice^(28,29). Both channels are also expressed in the pancreas, prostate, and mammary, sweat and salivary glands⁽²⁷⁾. They present a similar structure to other members of the TRP family: six transmembrane domains, a short hydrophobic stretch between segments 5 and 6 involved in the Ca pore and large intracellular N and C terminal tails. The intracellular segments contain phosphorylation sites, post-synaptic density protein motifs and ankyrin repeat domains; all of them are involved in the regulation of channel activity

and trafficking⁽³⁰⁾. It has been demonstrated that the tetrameric structure of TRPV6 and TRPV5 can be combined with each other to form different heterotetrameric channel complexes⁽³¹⁾. Both channels have 75 % homology, share several properties, but have different N and C terminal tails. They are regulated by calcitriol, oestrogen and dietary Ca. Both are inactivated by intracellular Ca, but with a different kinetics. In addition, the affinity of TRPV5 for the inhibitor ruthenium red is 100-fold that of TRPV6⁽³²⁾.

TRPV6 transcripts have been found in duodenum, but not in ileum, human biopsies. The duodenal expression of TRPV6 in men was detected to be vitamin D dependent, whereas in elderly women the TRPV6 and vitamin D receptor (VDR) expressions were low and not vitamin D dependent. This finding could explain, at least in part, the lower intestinal Ca absorption in elderly postmenopausal women⁽³³⁾. In rats, the basal mRNA expression of TRPV6 has been found to be the highest in the duodenum, followed by the colon (46 % of duodenum), and negligible in the jejunum and ileum. The rank order of the basal levels of TRPV6 protein was duodenum > colon (72 % of duodenum) > ileum (25 % of duodenum)⁽³⁴⁾.

Calbindins

These proteins appear to be responsible for carrying Ca from the apical side of the enterocyte to the basal region of the cell. Calbindin (CB) CB_{9k} is present in the intestine of mammals and CB_{28k} in that from avian species⁽³⁵⁾. CB_{9k} has four α -helical regions forming an EF-hand pair consisting of a canonical and a non-canonical/pseudo EF-hand domain, which are joined by a linker region. These EF-hands organised in tandem domains are the physiological relevant structures and two Ca ions bind with positive cooperativity⁽³⁶⁾. CB_{28k} has six EF-hand domains, four of which bind Ca with medium/high affinity⁽³⁷⁾. EF-hand 2 is non-functional and under physiological conditions EF6 most probably is as well. The four medium/high-affinity sites⁽³⁸⁾ are considered Ca specific.

CB also buffer Ca²⁺ ions by keeping intracellular Ca²⁺ concentrations below 10⁻⁷ M, which contribute to the prevention of premature cell death by apoptosis. When there is a down-regulation of CB, an excess of Ca²⁺ is provoked that may trigger apoptosis in the epithelial cells⁽³⁹⁾. Furthermore, it has been reported that CB_{28k} also inhibits apoptosis in osteoblastic cells⁽⁴⁰⁾ and in germ cells from Robertsonian mice^(41,42). In addition, it has been shown in kidney that CB_{28k} regulates the Ca²⁺ concentration in the vicinity of the TRPV5 pore by a direct association with the channel⁽⁴³⁾. This might occur in the intestine and in other tissues with important movements in intracellular Ca²⁺ concentrations.

Genetic studies have provided information that confused the understanding of CB on Ca homeostasis. Mice with ablation of the CB_{28k} gene do not exhibit calcemic abnormalities⁽⁴⁴⁾. In CB_{9k}-null mutant mice as well as mice lacking the epithelial Ca channel TRPV6 it has been demonstrated that the regulation of active intestinal Ca absorption is independent of CB_{9k} and TRPV6. The authors think that in the KO mice there is compensation by another Ca channel or protein and that other novel factors are involved in intestinal Ca absorption⁽⁴⁵⁾. It has

167 been found that an ablation of CB_{9k} alters the expression of
 168 paracellular tight junction genes. The compensatory expression
 169 of paracellular tight junction genes in the duodenum was
 170 associated with CB_{9k}, but not with CB_{28k}⁽⁴⁶⁾. This interaction
 171 between the transcellular and paracellular pathways might
 172 partially explain the variety of gut responses to absorb Ca under
 173 different pathophysiological conditions.

174 *Calcium pump and Na⁺/Ca²⁺ exchanger*

175 Plasma membrane Ca²⁺-ATPase (PMCA) 1 is an ATP-dependent
 176 transporter that pumps Ca out of the cytosol. This protein was
 177 detected in erythrocyte membranes and found to have a high
 178 Ca affinity⁽⁴⁷⁾. It presents four isoforms (PMCA1–4), which are
 179 divided into several subtypes by alternative splicing. PMCA1
 180 is considered as the housekeeping isoform because its mRNA is in
 181 all tissues. The correlation between the regulation of Ca
 182 homeostasis by TRPV6 and PMCA1 and duodenal and renal
 183 function is not well known. Nevertheless, some reports have
 184 described a role for TRPV6 and PMCA1 in the uterus, duode-
 185 num, kidney and brain^(48–52). PMCA has a relative molecular
 186 mass (M_r) of 130 kDa and a K_m for Ca of 0.2 μM in the presence
 187 of calmodulin⁽⁵³⁾. In the intestine, PMCA is located in the
 188 caveolae, which can exist in open and closed forms that
 189 control Ca efflux from the cell⁽⁵⁴⁾. The predominant form in the
 190 intestine is the isoform PMCA_{1b}. We have found that its
 191 expression and activity are higher in enterocytes from the villus
 192 tip than in those from the villus crypt, supporting the idea that
 193 mature enterocytes have the greatest capacity for transcellular
 194 Ca movement⁽⁵⁵⁾.

195 Another novel protein seems to be crucial in the transcellular
 196 Ca pathway. This is the protein 4.1R, which was also first
 197 identified in the erythrocyte membrane skeleton and is
 198 expressed in the epithelia of the intestine. So far, its physio-
 199 logical function remains unknown. Liu *et al.*⁽⁵⁶⁾ have detected
 200 that 4.1R co-localises with PMCA_{1b}. These authors have shown that
 201 4.1R KO mice exhibit deterioration in intestinal Ca absorption.
 202 In 4.1R KO mice, the expression of PMCA_{1b} in enterocytes
 203 was decreased. This finding that the deficiency in the adaptor
 204 protein 4.1R produces an impaired intestinal Ca absorption
 205 suggests that many yet to be defined molecules might also play
 206 important functions in epithelial Ca transport.

207 Apparently, the intestinal Na⁺/Ca²⁺ exchanger (NCX1) is
 208 responsible for about 20 % of Ca exit. However, this protein has
 209 received little attention and many recent reviews ignore it as
 210 another molecule involved in the Ca²⁺ exit from the intestine.
 211 The activity of the intestinal Na⁺/Ca²⁺ exchanger depends on
 212 the gradient created by Na⁺/K⁺-ATPase⁽⁵⁷⁾. There are several
 213 isoforms that result from three different genes⁽⁵⁸⁾, but in the
 214 intestine NCX1 is present mainly in the enterocytes. It has been
 215 detected in rats⁽⁵⁷⁾, mice⁽⁵⁹⁾, chicks⁽⁵⁵⁾, horses⁽⁶⁰⁾ and dogs⁽⁶¹⁾,
 216 but not in rabbits⁽⁶²⁾. NCX1 has a stoichiometry of 3 Na⁺:1 Ca²⁺
 217 and can function in either a forward mode (Ca²⁺ extrusion) or in
 218 a reversed mode (Ca²⁺ entry), depending on the Na⁺ and Ca
 219 gradients and the membrane potential⁽⁶³⁾. We have found that
 220 the expression and activity of NCX1 are quite similar between
 221 mature and immature enterocytes from chick duodenum, but
 222 are slightly higher in the villus tip cells⁽⁵⁵⁾. Recently, it has been

223 found that the gene expression of NCX1, PMCA_{1b} and CB_{9k} was
 224 down-regulated, whereas NCX1 expression was unchanged in
 225 the duodenum of a model of hypoxia in pregnant rats that
 226 shares clinical similarities with humans suffering from pre-
 227 eclampsia or other metabolic diseases. The authors have also
 228 found alterations in Ca transporters from the placenta and
 229 kidney and showed that these changes caused Ca deficiencies
 230 associated with pre-eclampsia⁽⁶⁴⁾. As they pointed out, it is quite
 231 possible that this study may contribute to a better understanding
 232 of the interrelationship between Ca imbalances and metabolic
 233 disturbances during pre-eclampsia pathogenesis.

234 *Paracellular pathway*

235 The intestinal epithelium is formed by a continuous layer of
 236 individual cells with very narrow spaces between them through
 237 which small molecules and ions diffuse⁽⁶⁵⁾. The epithelium must
 238 regulate this paracellular pathway for the maintenance of
 239 selective permeability. The movement of molecules and ions
 240 through this pathway is regulated by tight junctions. They are
 241 intercellular structures where plasma membranes of adjacent
 242 enterocytes have very close contact. The tight junction proteins
 243 are synthesised in the adjacent cells and they include occludin
 244 (Ocln) and claudins (Cldns). The latter is a protein family
 245 with more than twenty members. Both Ocln and Cldns are
 246 integral proteins having the capability of interacting adhesively
 247 with complementary molecules on adjacent cells and of
 248 co-polymerising laterally⁽⁶⁶⁾. Ca²⁺ movement through the tight
 249 junctions is a passive process that depends on the concentration
 250 and the electric gradient across the epithelium. This transport is
 251 non-saturable and mainly occurs in the jejunum and ileum
 252 under conditions of adequate or high Ca intake⁽⁶⁷⁾. When Ca
 253 intake is high, the sojourn time in the intestine is short and there is
 254 down-regulation of proteins involved in the transcellular pathway,
 255 which switches on the paracellular route⁽⁶⁸⁾ (see Fig. 1).

256 *Hormonal effects*

257 *Calcitriol*

258 Calcitriol or 1,25-dihydroxycholecalciferol (1,25(OH)₂D₃) is the
 259 main stimulus for intestinal Ca absorption. It induces changes in
 260 the structure and function of intestinal epithelial cells, which
 261 enhance Ca transport across the intestine. Calcitriol acts through
 262 genomic and non-genomic pathways, after binding to a VDR.
 263 Most of the studies have been focused on the effect of calcitriol
 264 on the intestinal transcellular Ca pathway. It has been found
 265 that the expression or the activity of all molecules presumably
 266 involved in this route is increased by calcitriol in experimental
 267 animals and even in human subjects^(69–72). Recently, it has been
 268 shown in mice that a single administration of 1,25(OH)₂D₃
 269 produced a 30-fold maximal increase in the ileal TRPV6 mRNA
 270 at 9 h. Multiple dosing of 1,25(OH)₂D₃ increased the ileal
 271 TRPV6 to 200- to 600-fold, being the highest changes observed
 272 at the 3rd and 4th doses. TRPV6 protein levels were increased
 273 1.5-fold throughout the duodenum and ileum after the 3rd and
 274 4th doses, while levels in the colon were increased after

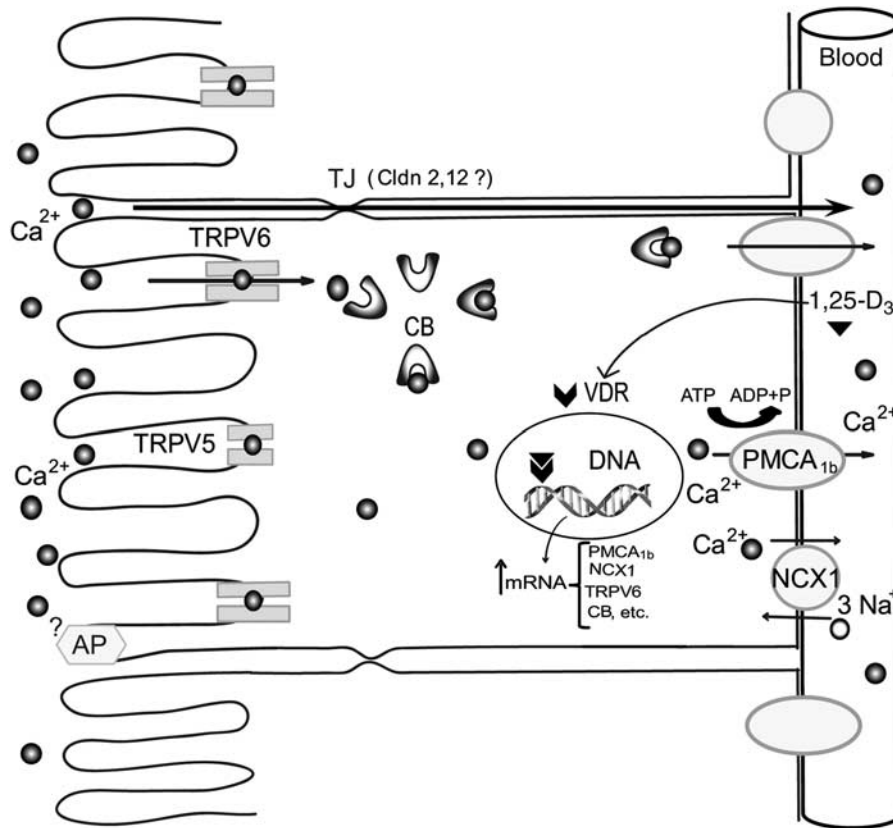


Fig. 1. Schematic model of transepithelial and paracellular calcium transport in the small intestine. The paracellular calcium pathway is carried out through tight junctions (TJ) by an electrochemical gradient (long arrow between cells). The transcellular calcium pathway consists of three steps: (1) apical entry of calcium through epithelial calcium channels TRPV5 and TRPV6 (the second one is the most abundant in intestine); (2) cytosolic diffusion bound to calbindins (CB); and (3) extrusion across the basolateral membranes by plasma membrane Ca^{2+} -ATPase (PMCA_{1b}) and $\text{Na}^+/\text{Ca}^{2+}$ exchanger (NCX1). Calcitriol ($1,25\text{-D}_3$) stimulates the individual steps of transcellular calcium transport. Calcitriol molecules bind to their nuclear receptors (vitamin D receptors; VDR), and the complex $1,25\text{-D}_3$ -VDR interacts with specific DNA sequences inducing transcription and increasing the expression levels of PMCA_{1b} , NCX1, TRPV6 and CB. The real role of the intestinal alkaline phosphatase (AP) enzyme in intestinal calcium absorption has not been elucidated yet. Cldn, claudin.

275 the 4th dose. These changes in protein correlate with a high
 276 TRPV6 mRNA induction in the ileum at the same period⁽³⁴⁾. The
 277 $1,25(\text{OH})_2\text{D}_3$ -enhanced Ca transport in mice was reported to be
 278 inhibited by FGF-23 as well as Ca transport in the colon cancer
 279 Caco-2 cells. Fibroblast growth factor-23 (FGF-23) produced an
 280 abolishment of the enhanced transcellular active Ca fluxes in
 281 both models. Despite the Arrhenius plot indicating that FGF-23
 282 decreased the potential barrier of paracellular Ca movement,
 283 FGF-23 was found to modestly down-regulate the $1,25(\text{OH})_2\text{D}_3$ -
 284 enhanced paracellular Ca transport⁽⁷³⁾. VDR-null mice adapt to
 285 pregnancy by up-regulating duodenal TRPV6 and intestinal Ca
 286 absorption, which enables a rapid normalisation of bone
 287 mineral content. These mice lactate normally and fully restore
 288 bone mineral content after weaning. In other words, VDR
 289 seems not to be required for skeletal adaptation during preg-
 290 nancy, lactation and after weaning⁽⁷⁴⁾. In the elderly, there is an
 291 age-related decrease in Ca absorption and a higher Ca intake is
 292 needed. It seems that increasing Ca intake from dairy products
 293 and Ca-fortified foods is much better than supplements.
 294 **Q3** The combination of vitamin D intake to 800 IU (20 μg) daily
 295 together with a total Ca intake of 1000 mg daily is a simple
 296 and inexpensive strategy that could reduce fractures in aged
 297 individuals by 30 %⁽⁷⁵⁾.

298 Two studies have demonstrated that calcitriol increases
 299 paracellular fluxes across the intestine, mainly in the jejunum
 300 and ileum^(76,77). Fujita *et al.*⁽⁷⁸⁾ have demonstrated that
 301 $1,25(\text{OH})_2\text{D}_3$ significantly increases claudin 2 and claudin
 302 12 mRNA levels in Caco-2 cells. They have also shown that
 303 mRNA and protein levels for these proteins were lower at
 304 12 weeks in the jejunum of VDR KO mice in comparison
 305 with wild-type mice, and small interfering RNA against these
 306 claudins decreased Ca permeability in the Caco-2 cells.

307 An intestinal calcistat has been hypothesised in order to
 308 explain vitamin D deficiency (VDD) with and without the
 309 clinical disease. Not all individuals with varying degree of VDD
 310 present secondary hyperparathyroidism and decreased bone
 311 mineral density (BMD). The intestinal calcistat would control Ca
 312 absorption, independently of parathyroid hormone (PTH)
 313 levels. A protein called Ca receptor (CaR) would dampen the
 314 production of active vitamin D metabolites in intestinal cells and
 315 diminish transcellular Ca transport, but would increase the
 316 paracellular Ca pathway. This local adaptation would adjust the
 317 fractional Ca absorption (FCA) according to the body's needs.
 318 When this local adaptation fails due to decreased Ca intake,
 319 decreased $25\text{-hydroxyvitamin D}_3$ ($25(\text{OH})\text{D}_3$), CaR mutation, the
 320 systemic adaptation comes into play. The systemic adaptations

are an increase in PTH and in active vitamin D metabolites. A rise in PTH is the first indication of VDD with a decrease in BMD depending on the duration of VDD. Therefore, individuals with VDD with normal PTH and BMD should be called subclinical VDD⁽⁷⁹⁾. The beneficial effect of vitamin D supplementation on this group of patients needs to be explored.

Parathyroid hormone

The action of PTH on intestinal Ca absorption occurs by the stimulation of renal CYP27B1 and, hence, increases 1,25(OH)₂D₃-dependent intestinal Ca absorption. Direct effects of PTH on Ca uptake by enterocytes from rat duodenum have been shown. PTH stimulates enterocyte Ca influx, which could be blocked by the Ca channel antagonists verapamil and nitrendipine⁽⁸⁰⁾. PTH/PTHrP receptors have been localised in intestinal epithelial cells along the villus⁽⁸¹⁾. PTH promotes nuclear effects such as a regulation of gene transcription and cell proliferation of enterocytes⁽⁸²⁾. So far, a direct effect of PTH on the global process of intestinal Ca absorption has not been reported yet.

Thyroid hormones

Cross *et al.*⁽⁸³⁾ have demonstrated that thyroid hormone and vitamin D have a cooperative effect on intestinal Ca transport. They have also observed that thyroid hormones increase the genomic action of calcitriol in the intestine. Kumar *et al.*⁽⁸⁴⁾ have reported that hyperthyroid rats show larger Ca uptake by brush-border membrane vesicles and Ca efflux from the basolateral membranes of enterocytes than hypothyroid rats. The authors have also found that the Ca²⁺-ATPase activity is not altered by thyroid hormones, while NCX1 activity is highly increased.

Growth hormone and insulin-like growth factor-1

Growth hormone (GH) can promote intestinal Ca absorption, which would occur indirectly mediated through an activation of renal CYP27B1 and the increase of serum 1,25(OH)₂D₃ concentration⁽⁸⁵⁾. Fleet *et al.*⁽⁸⁶⁾ have shown that GH treatment increases intestinal Ca absorption and duodenal CB_{9k} levels in aged rats without increasing serum 1,25(OH)₂D₃ levels. The effect of GH on Ca absorption is mediated through insulin-like growth factor-1 (IGF-1) and there is evidence that this effect does not depend on vitamin D signalling. Intestinal Ca absorption in adult men has been shown to be positively correlated with IGF-1 and the age-related declines in IGF-1 have a negative impact on Ca absorption, which could not be explained by a decrease in serum 1,25(OH)₂D₃⁽⁸⁷⁾. Since the vitamin D-independent mechanism by which the GH/IGF-1 axis may regulate intestinal Ca absorption is not clear, this issue needs to be investigated.

Oestrogen

Oestrogen corrects the decline in the efficiency of intestinal Ca absorption at the onset of menopause, as suggested by cell

culture studies⁽⁸⁸⁾, but the mechanisms that underlie this effect remain unknown. Colin *et al.*⁽⁸⁹⁾ have shown that oestrogen acts independently of 1,25(OH)₂D₃ in the intestine, while others suggest that oestrogen alters intestinal Ca absorption through the vitamin D endocrine system⁽⁸⁸⁾. Most of the oestrogen studies have been done in ovariectomised (OVX) animals. This ablation decreases endogenous oestrogen, but not totally since the adrenal androgens can be aromatised to oestrogen⁽⁹⁰⁾. Oestrogen receptor (ER) α KO mice showed a decrease in duodenal TRPV6 mRNA expression, without changes in CB_{9k}, PMCA_{1b} and VDR levels. In contrast, ER β KO mice did not alter the genes for intestinal Ca absorption. It seems that the genomic effects of oestrogen in mice are mainly mediated by ER α . This idea should not be extrapolated to humans because it has been shown that in normal colon and cancer colon cells the subtype β is the predominant form of the ER⁽⁹¹⁾. In OVX rats treated with oestradiol, van Abel *et al.*⁽⁹²⁾ have found increased duodenal gene expression of TRPV5, TRPV6, CB_{9k} and PMCA_{1b}. They used CYP27B1 KO mice to analyse the calcitriol dependency of the stimulatory effects of oestradiol on intestinal Ca absorption and demonstrated that the oestradiol treatment increased mRNA levels of duodenal TRPV6.

The effect of two dietary phyto-oestrogens (coumestrol and apigenin) as well as ipriflavone, a synthetic phyto-oestrogen, on Ca absorption has been studied in the human Caco-2 cell line⁽⁹³⁾. A direct effect of these compounds on intestinal Ca absorption was not observed. These controversial results indicate that the mechanism(s) triggered by oestrogen in the intestine requires further investigation⁽⁹⁴⁾.

Glucocorticoids

Osteoporosis is one of the most important side effects after long-term glucocorticoid (GC) treatments. Despite a reduced intestinal Ca absorption being part of the pathogenesis of GC-induced osteoporosis⁽⁹⁵⁾, the mechanisms triggered by GC on the intestine are not clear. A short-term GC treatment in young animals does not affect the expression of genes involved in intestinal Ca absorption⁽⁹⁶⁾, but a sustained dexamethasone suppresses mouse duodenal CB_{9k} expression through the GC receptor pathway⁽⁹⁷⁾. It has been also reported that prednisolone for 10 d diminishes rat intestinal Ca absorption through a decreased expression of the active Ca transporters, which occurs independently of 1,25(OH)₂D₃⁽⁹⁸⁾.

Nutritional factors affecting intestinal calcium absorption

Dietary calcium

Dietary Ca affects the composition of intestinal plasma membranes and alters intestinal Ca transport. All the genes involved in the transcellular pathway are enhanced by a low-Ca diet, which occurs by the activation of the vitamin D endocrine system^(19,55,99,100). The increase in the expression and activity of the intestinal Ca pump and NCX1 caused by a Ca-deficient diet occurs in both mature and immature enterocytes. However, VDR levels are decreased by low-Ca diets, independently of the degree of cell differentiation⁽⁵⁵⁾. We think that high levels of

serum calcitriol provoked by low-Ca diets promote differentiation, which would produce cells more capable of expressing vitamin D-dependent genes required for Ca absorption. The activity of intestinal alkaline phosphatase (AP) is concomitantly increased by dietary Ca restriction, but a real role of this protein in intestinal Ca absorption cannot be discarded⁽⁵⁵⁾. Benn *et al.*⁽¹⁰¹⁾ have found that a low-Ca diet increases intestinal Ca absorption in wild-type, TRPV6 KO and CB_{9k} KO mice. This study indicates that the active intestinal Ca absorption occurs in the absence of TRPV6 and CB_{9k}, which challenges the dogma that both proteins are necessary for vitamin D-induced active intestinal Ca transport. Probably, TRPV6 is not the rate-limiting factor in the transcellular pathway or another factor/s is/are involved in its absence to compensate partially its function.

We have also observed that a low-Ca diet increases the reactivity and availability of sulfhydryl groups from intestinal brush-border membrane proteins of chicks⁽¹⁰²⁾. It is quite possible that the sulfhydryl status of the brush-border membrane proteins is involved in the vitamin D-dependent intestinal Ca absorption. With regard to the lipid composition, we have detected minor changes in the fatty acid content of the basolateral membrane, but the lipid fluidity of these membranes is highly increased by dietary Ca restriction⁽¹⁹⁾.

Recently, it has been reported that luminal Ca controls the intestinal Ca absorption through the modification of intestinal AP activity⁽¹⁰³⁾. Authors have found that luminal Ca concentration increases the activity of AP and simultaneously decreases percentage Ca absorption, acting as a minute-to-minute regulatory mechanism of Ca entry.

Some studies have found an inverse association between Ca intake and adiposity^(104–106). However, not all studies have observed this relationship⁽¹⁰⁷⁾. The mechanisms underlying these responses are not quite clear.

Most patients with idiopathic hypercalciuria show an increased intestinal Ca absorption. This has been demonstrated in studies using radiolabelled Ca and comparing the intestinal Ca absorption in patients with idiopathic hypercalciuria with that in normal subjects⁽¹⁰⁸⁾. One possible mechanism for the increased Ca absorption might be the highest levels of serum calcitriol found in these patients as compared with normal individuals⁽⁶⁵⁾. Apparently, this response is independent of Ca intake and reflects an enhancement in active Ca transport by the intestine⁽¹⁰⁹⁾.

With regard to the risk of kidney calculi formation in postmenopausal women, it is not clear if it is associated with Ca intake and vitamin D supplements. Haghighi *et al.*⁽¹¹⁰⁾ have shown that the administration of 1000 mg/d of dietary Ca and vitamin D had a weak association with the formation of kidney calculi (only 1.9 % of fifty-three patients). Jackson *et al.*⁽¹¹¹⁾ have reported that in a study with 36 282 postmenopausal women treated with 1000 mg of Ca and 400 IU (10 µg) of vitamin D/d, a 17 % higher rate of kidney calculi was shown in the treated group after 7 years of follow-up. In contrast, another study contradicted this result, suggesting no association between Ca and vitamin D consumption and kidney calculi formation⁽¹¹²⁾. A low-Ca diet enhances the absorption of oxalate in the gut from normal individuals and kidney stone formers leading to an increase in urinary oxalate excretion, which has an important role in Ca stone formation^(113,114).

There is evidence that dietary Ca restriction is a risk factor for cancer incidence, particularly for colorectal cancer⁽¹¹⁵⁾. By contrast, a high intake of dairy products and Ca intake protects against the distal colon and rectal tumours as compared with the proximal colon and reduces the risk of colon cancer⁽¹¹⁶⁾. It is also quite possible that a low Ca intake contributes to the development of renal, gastric, pancreatic, ovarian, endometrial and lung cancer as well as multiple myeloma, but there is no conclusive evidence⁽¹¹⁷⁾.

Several studies have shown that a poor Ca intake is associated with an increased risk of breast cancer^(118–120). Vergne *et al.*⁽¹²¹⁾ have recently demonstrated that Ca intake is associated with a high DNA repair capacity level, which in turn decreases the development of breast cancer.

Observational studies have identified an inverse relationship between maternal Ca intake and the incidence of pre-eclampsia^(122,123). When Ca supplements are administered during pregnancy, the incidence of pre-eclampsia and its consequences are reduced, including severe maternal morbidity and death⁽¹²⁴⁾.

Several meta-analyses of randomised controlled trials have also indicated that Ca supplementation can lead to a small reduction in systolic and diastolic blood pressure; therefore, Ca supplements might be beneficial in patients with hypertension^(125,126). Varenna *et al.*⁽¹²⁷⁾ have confirmed an association between hypertension and osteoporosis. There is a higher prevalence of hypertension in women with osteoporosis and a higher prevalence of osteoporosis in women with hypertension. The authors suggest that a low dairy Ca intake could be associated with an increased risk of both diseases and be a possible pathogenic link between the two conditions.

It is well known that a reduced intestinal Ca absorption is a risk factor for osteoporosis. There is evidence that the use of Ca supplements decreases bone turnover by about 20 %, and this is associated with a reduction in bone loss in postmenopausal women⁽¹²⁸⁾. Low Ca intake was significantly associated with low BMD and increased risk of osteoporosis. However, the association between Ca and BMD was not consistently linear, and a sufficient vitamin D level might compensate for the negative influence of low Ca intake on bone⁽¹²⁹⁾. Zhou *et al.*⁽¹³⁰⁾ have observed that a higher cumulative Ca and vitamin D intake in adult women was associated with better bone health, as indicated by BMD at multiple sites.

Other ions

Aluminium. Several authors have reported that aluminium ions (Al), at pharmacological doses, are able to reduce intestinal Ca absorption via the vitamin D-dependent transcellular pathway in the small intestine of humans and animals^(131–135). Orihuela *et al.*⁽¹³⁵⁾ have suggested that Al might interfere with Ca uptake by enterocytes through an effect on cell membranes. In addition, Al would decrease the intestinal glutathione (GSH) level affecting CB function and/or synthesis, which would lead to a reduced transcellular Ca absorption. Apparently, the inhibitory effect of Al varies according to thyroid hormone status⁽¹³⁶⁾. In late pregnancy and mainly during the middle lactation of rats, Al has also been shown to reduce transcellular Ca absorption in the duodenum

538 by interfering with the mechanisms of Ca transport partially
539 mediated by a high serum level of oestrogen and prolactin⁽¹³⁷⁾.

540 **Phosphate.** Severe dietary P deficiency is rare in humans and
541 occurs only in conditions of severe starvation. Nevertheless,
542 P deficiency can occur in alcoholics, patients with malabsorp-
543 tion syndromes, and those taking excessive amounts of
544 Ca supplements⁽¹³⁸⁾. Dietary P deficiency affects intestinal
545 Ca absorption. Under this deficiency, serum 1,25(OH)₂D₃
546 content increases^(139,140) as well as the levels of CB and CB
547 mRNA synthesis⁽¹⁴¹⁾. However, in some fashion, a low-P diet
548 can stimulate CB formation and intestinal Ca absorption in the
549 absence of an increased production of 1,25(OH)₂D₃⁽¹⁴²⁾. The
550 reactivity and availability of sulphhydryl groups of chick intestinal
551 brush-border membranes have been also shown to increase by
552 a low-P diet⁽¹⁰²⁾, but it remains unknown whether this response
553 is related to increased intestinal Ca absorption. Meyer *et al.*⁽¹⁴³⁾
554 have demonstrated that chickens fed a low-P diet displayed an
555 increase in intestinal VDR mRNA. The authors have reported
556 that a complex regulation of VDR expression occurs in that
557 low-P restriction enhances VDR mRNA levels, possibly via
558 increased serum 1,25(OH)₂D₃.

559 **Magnesium.** For several decades, it has been known that there
560 is interaction between Ca and Mg²⁺ in the intestine, which
561 varies throughout the intestine. Mg²⁺ has been found to inhibit
562 Ca absorption primarily in the duodenum, whereas Ca inhibits
563 Mg²⁺ transport in the ileum but not in the duodenum⁽¹⁴⁴⁾. In
564 studies of short-term uptake in rat duodenal mucosa, O'Donnell
565 & Smith⁽¹⁴⁵⁾ have demonstrated that Mg²⁺ inhibited the time-
566 dependent uptake of Ca, but Ca did not alter Mg²⁺ uptake.
567 Increasing the Mg²⁺ concentration to 1.25 mmol/l decreased the
568 mucosal-to-serosal flux of Ca by 50 % and abolished net Ca
569 absorption, mainly due to a depression in the paracellular
570 pathway⁽¹⁴⁶⁾. In patients with idiopathic hypercalciuria and
571 renal Ca stone disease, oral supplementation of Mg²⁺ has been
572 shown to be favourable because it decreases Ca absorption and
573 increases Mg²⁺ absorption, which may reduce risk factors
574 for renal Ca stone formation⁽¹⁴⁷⁾. By contrast, Mg²⁺ deficiency
575 significantly increased enterocyte content of Ca⁽¹⁴⁸⁾. The
576 mechanism is difficult to understand because Mg²⁺ deprivation
577 has been associated with a low production of 1,25(OH)₂D₃, the
578 main stimulator of intestinal Ca absorption⁽¹⁴⁹⁾. However, some
579 studies contradict the previous data. Fine *et al.*⁽¹⁵⁰⁾ have found
580 that increasing dietary Mg²⁺ had no effect on Ca absorption.
581 Kosakai *et al.*⁽¹⁵¹⁾ have demonstrated in sheep that the apparent
582 Ca absorption tended to increase when the dietary Mg²⁺ content
583 was increased, which was accompanied without an alteration
584 in the plasma Ca concentration and increased urinary Ca
585 excretion. Bae *et al.*⁽¹⁵²⁾ have shown that the consumption
586 of seaweed Ca extract or inorganic calcium carbonate with
587 Mg²⁺ oxide in OVX rats produced a similar intestinal Ca
588 absorption, but only the seaweed Ca extract caused an increase
589 in femoral BMD and strength in OVX rats. They concluded
590 that seaweed Ca extract is a good Ca and Mg²⁺ source for
591 improving bone health as compared with synthetic Ca and
592 Mg²⁺ supplementation.

Lipids

593 At present, the effects of dietary lipids on intestinal Ca
594 absorption are not clear. Hessov *et al.*⁽¹⁵³⁾ have demonstrated
595 that low-fat-diets increase intestinal Ca absorption in nine
596 patients with fat malabsorption. Steatorrhoea has been
597 associated with an inhibition of intestinal Ca absorption⁽¹⁵⁴⁾.
598 Jewell *et al.*⁽¹⁵⁵⁾ have found that both the paracellular
599 Ca transport and transcellular Ca transport across monolayers of
600 Caco-2 cells were significantly increased after exposure to
601 conjugated linoleic acid. Murphy *et al.*⁽¹⁵⁶⁾ have found that zona
602 occludens-1, Ocldn, and Cldn-4 were all up-regulated while
603 Cldn-1 was down-regulated by *trans*-10, *cis*-12-conjugated
604 linoleic acid, which explains the increase in the paracellular
605 route. However, they did not find any effect on genes involved
606 in the transcellular pathway, which should be further explored.
607 Fatty acids directly contribute to increasing intestinal Ca absorption
608 via a cation exchange mechanism between cellular H⁺ for
609 luminal Ca favoured by exchanger activity 2H⁺/Ca²⁺⁽¹⁵⁷⁾. Fatty
610 acids would increase the proliferation of colonic epithelial cells,
611 producing a trophic effect on the mucosa; they contribute to
612 increasing the absorptive surface^(158,159).

613 Recently, in high-fat diet-fed mice, Xiao *et al.*⁽¹⁶⁰⁾ have found
614 a marked decrease in the intestinal Ca absorption, which is
615 accompanied by redox imbalance and increased duodenal
616 oxidative damage, an effect that was avoided by lipoic acid
617 or Ca supplementation. In addition, they have also shown
618 that a high-fat diet down-regulates the gene expression of
619 molecules involved in the transcellular pathway of intestinal
620 Ca absorption, independently of calcitriol regulation. The
621 authors think that the main cause for high-fat-diet-induced
622 inhibitory intestinal Ca absorption is not Ca soap but duodenal
623 oxidative stress.
624

Carbohydrates

625 Lactose is a disaccharide found in milk and dairy products that
626 enhances Ca homeostasis, which should be beneficial for bone
627 health⁽¹⁶¹⁾. It is well established that lactose stimulates the
628 intestinal Ca absorption that seems to occur by passive transport
629 in the small intestine⁽¹⁶²⁾. Apparently, lactose promotes intestinal
630 Ca absorption, independently of the vitamin D endocrine system.
631 Natsuko *et al.*⁽¹⁶³⁾ have found that lactose alters intestinal AP
632 in rats not only in a direct way, but also indirectly through a
633 regulation of intestinal AP expression, mainly in the jejunum.
634 Epilactose (a rare disaccharide in cows' milk) has been
635 demonstrated to increase Ca transport in everted small-intestinal
636 sacs by promoting the generation of SCFA and other organic
637 acids⁽¹⁶⁴⁾. Later on, the same group of investigators has found
638 that the epilactose-mediated promotion of intestinal Ca absorption
639 involves the paracellular route in the rat small intestine through
640 the induction of myosin regulatory light chain phosphorylation
641 via myosin light chain kinase and Rho-associated kinase⁽¹⁶⁵⁾.
642 They have also shown that epilactose improves intestinal
643 Ca absorption in gastrectomised rats. They think that the resulting
644 SCFA production by intestinal microbes is responsible for this
645 effect, as well as the increase in the caecal mucosa area and the
646 soluble Ca concentration⁽¹⁶⁶⁾.
647

648 Some sugar alcohols such as erythritol, xylitol, sorbitol, maltitol,
 649 palatinol or lactitol have been found to enhance Ca transport from
 650 rat small and large intestine epithelium *in vitro*. Differences in
 651 Ca transport were shown in different segments of the intestine,
 652 but not between the sugar alcohols tested⁽¹⁶⁷⁾. Recently, Xiao
 653 *et al.*⁽¹⁶⁸⁾ have found that mannitol improves the absorption and
 654 retention of Ca and Mg²⁺ in growing rats, an effect that occurs
 655 through the fermentation of mannitol in the caecum.

656 Prebiotics

657 The fortification of milk with milk Ca or Ca salts is among
 658 the strategies suggested to increase Ca intake or absorption or
 659 both, but the availability of Ca salts in milk has not been well
 660 characterised⁽¹⁶⁹⁾. In addition, several food ingredients such as
 661 fructo-oligosaccharides and caseinophosphopeptides (CPP) have
 662 been proposed as enhancers of the absorption of Ca from milk or
 663 other foods. Fructo-oligosaccharides belong to the group of
 664 non-digestible oligosaccharides (NDOs), which includes inulin,
 665 oligofructose and galacto-oligosaccharides. They can be digested
 666 by the colonic microflora, which produces SCFA that decreases
 667 intestinal pH and increases Ca solubility leading to an enhance-
 668 ment of paracellular and transcellular Ca transport^(170,171). It has
 669 also been suggested that NDOs increase active Ca transport by
 670 the activation of CB_{9k}⁽¹⁷²⁾. Fukushima *et al.*⁽¹⁷³⁾ have demon-
 671 strated in rats that fructo-oligosaccharide consumption increases
 672 the gene expression of TRPV6 and CB_{9k} through the SCFA
 673 formed in the fermentation. The fibres formed mainly by inulin
 674 have positive chronic effects on Ca metabolism related to changes
 675 in the intestine, resulting in improvement of bone health⁽¹⁷⁴⁾.
 676 In addition, galacto-oligosaccharides have also potential for
 677 improving mineral balance and bone properties⁽¹⁷⁵⁾. With regard
 678 to CPP, originating from casein digestion, it has been demon-
 679 strated that their addition to Ca-fortified milk increases intestinal
 680 Ca absorption in growing rats⁽¹⁷⁶⁾, but the mechanisms involved
 681 in this response were not elucidated. Erba *et al.*⁽¹⁷⁷⁾ have studied
 682 the influence of different four CPP/Ca ratios and three mineral
 683 concentrations on the amount of passive Ca absorbed across the
 684 everted distal small intestine of rats. The positive effect was
 685 dependent on the relative amount of both species in the intestinal
 686 lumen, the ratio 15 being the most efficient at increasing mineral
 687 transport. Nevertheless, in a randomised cross-over trial under-
 688 taken in fifteen adults, no effect of CPP was found on intestinal
 689 Ca absorption⁽¹⁷⁸⁾. Cosentino *et al.*⁽¹⁷⁹⁾ have demonstrated that
 690 both intestinal human HT-29 and Caco-2 cells have the ability to
 691 take up extracellular Ca under CPP stimulation. Recently,
 692 Colombini *et al.*⁽¹⁸⁰⁾ did not find effects of CPP on paracellular Ca
 693 absorption and on TRPV6 mRNA expression in intestinal human
 694 HT-29 and Caco-2 cell lines.

695 A recent study has shown that the daily intake of soluble
 696 maize fibre, a well-tolerated prebiotic fibre, increases short-term
 697 Ca absorption in adolescents consuming less than the
 698 recommended amounts of Ca⁽¹⁸¹⁾.

699 Probiotics

700 Probiotics are viable microbes that alter the microflora in a
 701 compartment of the host exerting beneficial health effects

in this host⁽¹⁸²⁾. Most probiotic products contain lactic acid-
 producing bacteria, which mainly belong to the genera
Lactobacillus and *Bifidobacterium*. In growing rats, it has
 been demonstrated that probiotic yoghurt containing strains of
Lactobacillus casei, *L. reuteri* and *L. gasseri* increase intestinal
 Ca absorption and bone mineral content⁽¹⁸³⁾. Besides, it has
 been observed that in Caco-2 cells, the probiotic *L. salivarius*
 causes an increase in Ca uptake⁽¹⁸⁴⁾.

Synbiotics

Synbiotics are defined as products containing prebiotics and
 probiotics, in which the prebiotic compound favours the
 probiotic compound⁽¹⁸²⁾. It has been shown in OVX rats that
 intestinal Ca absorption tended to be higher in the synbiotic
 group and was significantly higher in the prebiotic group in
 comparison with the control group⁽¹⁸⁵⁾.

Proteins

Proteins are essential to bone, but the Ca-wasting effect of a
 high protein intake constitutes a point of debate. It has been
 known for many years that increasing dietary protein enhances
 urinary Ca either in human subjects or in rats⁽¹⁸⁶⁾. The idea was
 that the additional Ca excretion was of skeletal origin as a result
 of buffering in bones the metabolic acid load imposed by higher
 protein intake⁽¹⁸⁷⁾. The theory was that a high-protein diet,
 mainly meat, creates a higher acid load due to the high content
 of amino acids containing sulfur. This acid load cannot be
 neutralised by the kidneys and the body pulls Ca from the
 skeleton to balance pH at the expense of bone, causing an
 enhancement of urinary Ca⁽¹⁸⁸⁾. However, clinical studies have
 demonstrated that a short-term high-protein diet (2.1 g/kg)
 significantly increases the intestinal Ca absorption as compared
 with a medium-protein diet (1 g/kg) and the increment in
 urinary Ca is quantitatively explained by an increase in intestinal
 Ca absorption efficiency⁽¹⁸⁹⁾. The transcellular route, the para-
 cellular pathway or a combination of both mechanisms of
 intestinal Ca absorption might be involved in response to high
 dietary protein. By using duodenal brush-border membrane
 vesicles, Gaffney-Stomberg *et al.*⁽¹⁹⁰⁾ have demonstrated that the
 transcellular component of Ca absorption was accelerated in rats
 fed a high-protein diet, which was due to an enhancement in
 maximum velocity, without affecting the Michaelis-Menten
 constant. However, they did not study whether the gene or
 protein expression of the molecules involved in the transcellular
 pathway was modified. They did not find increased bone
 resorption or changes in serum PTH and calcitriol levels.

Since more research is necessary to resolve the protein debate,
 it has been suggested not to reduce the protein intake below the
 dietary reference intake because it could be detrimental to bone
 health, especially in old individuals⁽¹⁹¹⁾, and protein intakes and
 balance of different protein sources with a variety of different
 foods constitutes appropriate dietary advice⁽¹⁹²⁾.

Black tea

Black tea (*Camellia sinensis*) is a medicinal plant with a rich
 flavonoid content and a plethora of health-promoting effects^(193,194).

755 Das *et al.*⁽¹⁹⁵⁾ have studied the ability of black tea extract as a
 756 suitable alternative adjunct for Ca supplementation in treating
 757 an OVX rat model of early osteoporosis. The results suggest that
 758 black tea could stimulate intestinal Ca absorption, which is
 759 associated with an increased activity of AP and Ca²⁺-ATPase.
 760 Black tea's effectiveness in maintaining bone health was detected
 761 to be similar to 17 β -oestradiol. Therefore, this study suggests that
 762 a simultaneous use of black tea is promising as a prospective
 763 candidate for adjunctive therapies for Ca supplementation in the
 764 early stage of menopausal bone changes.

765 Coffee

766 Coffee drinking is a popular habit worldwide. It is consumed in
 767 considerable amounts every day. Caffeine, a methylxanthine
 768 present in coffee, has been considered to be responsible for an
 769 increased risk of osteoporosis in coffee drinkers^(196,197). At
 770 present, data are inconsistent^(198–200), hence, the effect of
 771 caffeine on intestinal Ca absorption is not well established. It
 772 has been demonstrated in rats that intestinal Ca absorption is
 773 stimulated by the increase in 1,25(OH)₂D₃ production after
 774 chronic administration of caffeine⁽²⁰¹⁾. In addition, urinary and
 775 faecal excretion is also increased. In postmenopausal osteo-
 776 porotic women, a coffee intake in excess of 1000 ml could
 777 induce an extra Ca loss of 1.6 mmol Ca/d, while 1–2 cups of
 778 coffee/d would have little impact on Ca balance⁽²⁰²⁾. Metabolic
 779 balance studies show a weak negative effect of caffeine on the
 780 efficiency of intestinal Ca absorption. However, the effect of
 781 caffeine is small enough to be fully offset by 1–2 tablespoons
 782 (15–30 ml) of milk⁽¹³⁸⁾. A recent study in OVX rats has shown
 783 that low to moderate caffeine intake may exert some beneficial
 784 effects on the skeleton, increasing bone mineralisation, and
 785 improving the strength and structure of cancellous bone and the
 786 mechanical properties of compact bone; however, it did not
 787 cause any significant effect in rats with normal oestrogen
 788 levels⁽²⁰³⁾. The continuous debate has weakened interest in the
 789 study of coffee as a risk factor for osteoporosis, which has been
 790 reinforced by the non-inclusion of coffee in the list of risk
 791 factors in the predictive scale for fracture informed by the
 792 WHO⁽²⁰⁴⁾. The understanding of physiological effects of coffee
 793 consumption is difficult because of the vast array of components
 794 included in the brewed product and the varied effects of each
 795 compound. Recently, an unfavourable effect of trigonelline, an
 796 alkaloid present in coffee, has been demonstrated on bone
 797 mechanical properties in oestrogen-deficient rats, but not in
 798 control rats⁽²⁰⁵⁾. It is quite possible that the effects on intestinal Ca
 799 absorption and the Ca economy by caffeine or other bioactive
 800 compounds present in coffee depend on the amount and
 801 frequency of coffee intake. This is another issue that merits to
 802 be more investigated due to the considerable number of coffee
 803 drinkers and the rising life expectancy that will increase bone
 804 disorders in the next years.

805 Pharmacological compounds altering intestinal calcium 806 absorption

807 Almost two decades ago, we reported that the intactness of
 808 the steady-state levels of intestinal glutathione (GSH) seemed

to be critical for Ca absorption. By using DL-buthionine-(S,R)-
 sulfoximine (BSO), a specific inhibitor of γ -glutamylcysteine
 synthetase, we have shown that the Ca transfer from lumen to
 blood in vitamin D-supplemented chicks was inhibited, an
 effect that did not occur in vitamin D-deficient chicks. At that
 time we concluded that the effects of BSO on intestinal Ca
 absorption were dependent on the vitamin D status of the
 animal. One explanation that we gave was that GSH depletion
 might increase the reactive oxygen species and other sub-
 stances that could deteriorate intestinal Ca absorption⁽¹⁹⁾. Since
 intestinal AP was one of the best candidates to suffer oxidative
 stress⁽²⁰⁶⁾, we later studied the effect of BSO on the activity of
 this enzyme in chicks fed a commercial diet. In fact, the AP
 activity declined after BSO treatment, which was dose and time
 dependent. The effect occurred either *in vivo* or *in vitro* but
 was not direct; it was first necessary to deplete GSH in order to
 produce free hydroxyl radicals and an increment in the protein
 carbonyl content. The reversibility of the BSO effect was proved
 by the addition of GSH monoester to the duodenal loop⁽²⁰⁷⁾.
 Menadione (MEN) is another pharmacological compound
 that alters chick intestinal Ca absorption. It is a quinone that is
 clinically relevant because of its anti-tumour properties⁽²⁰⁸⁾ and
 its use in the treatment of osteoporosis⁽²⁰⁹⁾. MEN metabolism
 involves redox cycling, resulting in the release of various
 reactive oxygen species including free hydroxyl radicals⁽²¹⁰⁾.
 We have demonstrated 30 min after a single large dose of MEN
 that the intestinal Ca absorption was inhibited, which lasted for
 9 h. The inhibition affected the transcellular pathway as judged
 by the inhibition of Ca pump activity, the main protein involved
 in Ca extrusion from the enterocyte to the lamina propria.
 Intestinal AP activity was also inhibited, but not that from other
 brush-border membrane enzymes. GSH depletion, enhance-
 ment in the protein carbonyl content as well as the appearance
 of free hydroxyl radicals were indications that MEN caused
 oxidative stress provoking deleterious consequences on
 intestinal Ca absorption. The oral administration of GSH
 monoester prevented the inhibition of intestinal Ca absorption
 and the GSH deprivation produced by MEN⁽²¹¹⁾. As mitochondria
 are the major source of reactive oxygen species⁽²¹²⁾, we have
 investigated the role of these organelles in the inhibition
 of the intestinal Ca absorption caused by MEN. The quinone
 produced mitochondrial dysfunction as shown by inhibition of
 enzymes from Krebs' cycle, DNA fragmentation, release of
 cytochrome c, alteration of membrane potential and enhance-
 ment of Mn²⁺-superoxide dismutase activity. The mitochondrial
 dysfunction would be a consequence of mitochondrial GSH
 depletion, which would alter the membrane permeability
 triggering the release of apoptotic molecules leading to DNA
 fragmentation. The oxidant effects would alter the transcellular
 Ca pathway affecting the global process of intestinal
 Ca absorption⁽²¹³⁾. Since the flavonol quercetin has antioxidant
 properties⁽²¹⁴⁾, we have studied the ability of quercetin to protect
 the chick intestine against the inhibition of intestinal Ca absorption
 caused by MEN. Effectively, quercetin abrogated the inhibitory
 effect of MEN on chick intestinal Ca absorption through the
 restoration of intestinal redox state, blockage of alterations
 in the mitochondrial membrane permeability, and abolition
 of the FasL/Fas/caspase-3 signalling pathway activation⁽²¹⁵⁾.

867 In addition, the hormone melatonin (MEL) was also able to
868 restore chick intestinal Ca absorption inhibited by MEN. MEL is
869 known as a direct scavenger of free radicals with the ability
870 to remove singlet oxygen, the superoxide anion radical and
871 hydroperoxide. It has also an indirect antioxidant action
872 through a modulation of antioxidant enzyme activities. MEL by
873 itself did not alter intestinal Ca absorption and other variables
874 influencing that process. The MEL protective mechanism seems
875 to be switched on under oxidative stress conditions produced
876 by MEN, leading cells to the normal redox status. MEL
877 administration after MEN injection returned rapidly the intestinal
878 GSH and protein carbonyl contents to control values as well
879 as the SOD and CAT activities. Concomitantly, intestinal
880 Ca absorption went up to normal values, suggesting that the
881 restoration of redox status of the gut by MEL allowed the
882 recovering of the intestinal capability to absorb the cation
883 properly. MEL not only normalised the redox status of
884 the enterocytes but also rescued the epithelial cells from
885 MEN-induced apoptosis⁽²¹⁶⁾. Therefore, MEL could be a
886 potential drug of choice for the treatment of impaired intestinal
887 Ca absorption caused by oxidative stress and exacerbated
888 apoptosis, which occurs in certain pathophysiological conditions
889 (ageing, coeliac disease, intestinal bowel disease, cancer and
890 others) or after intake of drugs causing oxidation.

891 We have also shown that a single high concentration of
892 sodium deoxycholate (NaDOC) inhibits intestinal Ca absorption
893 through a down-regulation of proteins involved in the trans-
894 cellular pathway, as a consequence of triggering oxidative stress
895 and mitochondria-mediated apoptosis⁽²¹⁷⁾. This inhibitory effect
896 on intestinal Ca absorption produced by NaDOC has been
897 shown to be abolished by the concomitant use of the anti-
898 oxidant quercetin, which clearly indicates that the response of
899 NaDOC was mediated by the oxidative stress. Deoxycholic acid
900 or its salt, NaDOC, is the major secondary bile acid in humans
901 and is toxic in high concentrations causing liver damage during
902 cholestasis and acting as a promoter of colon cancer in
903 experimental animals⁽²¹⁸⁾. It is well known that its concentration
904 varies according to the diet; a high-fat diet is associated with an
905 increased secretion of NaDOC⁽²¹⁹⁾. This bile salt perturbs the
906 membrane structures by alteration of membrane microdomains
907 and decreases the transepithelial electrical resistance in the
908 Caco-2 cell line through reactive oxygen species generation and
909 other signalling mechanisms^(220,221). Therefore, the tight junctions
910 constitute another target of NaDOC in the intestine, suggesting
911 that the paracellular pathway of intestinal Ca absorption might be
912 also affected by this bile salt. Based on the knowledge that a
913 minor bile acid, ursodeoxycholic acid (UDCA), has beneficial
914 effects of protection against cytotoxicity due to more toxic bile
915 acids, we have tried to ascertain the potentiality of UDCA to
916 prevent the inhibition of intestinal Ca absorption caused by
917 NaDOC. In addition, we have studied the effects of UDCA alone
918 on intestinal Ca absorption either in chicks or rats. The data
919 have shown that UDCA not only prevented the inhibition of
920 intestinal Ca absorption caused by NaDOC either in chicks or
921 rats, but also UDCA alone enhanced that process. This was an
922 unpredictable finding, which together with the previous data
923 indicate that NaDOC is a bad whereas UDCA is a good bile acid
924 for intestinal Ca absorption. The interesting point is that the

combination of both bile acids neutralises the response of each
925 other, probably because UDCA protects the intestine against the
926 GSH depletion and protein carbonyl increment produced by
927 NaDOC. Both NaDOC and UDCA altered protein and gene
928 expression of molecules involved in the transcellular pathway
929 of intestinal Ca absorption, but in the opposite way. NaDOC
930 decreased the protein expression of PMCA1b, NCX1 and CB,
931 whereas UDCA increased the protein expression of all of them.
932 The expression of these molecules was identical to those from
933 the control group when the combined treatment was used. The
934 gene expression of *pmca1b*, *ncx1* and *cb* was increased by
935 UDCA and UDCA + NaDOC. In contrast, NaDOC decreased the
936 gene expression of *pmca1b* and *cb* without modifying that of
937 *ncx1*. UDCA also increased the protein and gene expression of
938 VDR, which suggests that VDR is involved in the enhancement
939 of intestinal Ca absorption produced by UDCA⁽²²²⁾. The rela-
940 tionship between UDCA and VDR is not surprising because it
941 has been demonstrated that VDR also binds bile acids^(223,224)
942 and the increase in the cathelicidin expression in biliary
943 epithelial cells from human liver caused by UDCA is mediated
944 by VDR activation, an effect that is blunted by a small interfering
945 RNA strategy⁽²²⁵⁾.
946

947 There are conflicting data with regard to the effects of proton
948 pump inhibitors and osteoporotic fracture risk. Presumably,
949 they increase the risk through hypochlorhydria and decreased
950 FCA. Hansen *et al.*⁽²²⁶⁾ have evaluated the effect of proton
951 pump inhibitor therapy on FCA using the dual stable isotope
952 method. Participants underwent three 24 h FCA studies; two
953 of them were accomplished 1 month apart to establish the
954 baseline of FCA, the third one was after taking omeprazole
955 (40 mg/d for 30 d). The data revealed that age, gastric pH,
956 serum omeprazole levels, adherence to omeprazole and
957 25-hydroxyvitamin D levels were not related to changes in FCA
958 between visits 2 and 3. The level of serum 1,25(OH)₂D₃ was the
959 only variable associated with the change in FCA between
960 visits 2 and 3. More studies are necessary to elucidate the
961 mechanisms by which proton pump inhibitors increase osteo-
962 porotic fracture risk.

963 Wahl *et al.*⁽²²⁷⁾ have demonstrated reduced FCA in patients
964 under anticonvulsant treatment. The possible mechanism
965 underlying this process is complex. It has been demonstrated
966 that phenytoin and carbamazepine inhibit active Ca transport
967 from the apical to the basolateral side of Caco-2 cells under
968 physiological Ca conditions and vitamin D improves the anti-
969 epileptic drug-induced decrease in Ca permeability⁽²²⁸⁾.

970 Restraint stress significantly down-regulates the mRNA
971 expressions of TRPV6 and Ca_v1.3, CB-D_{9k}, and PMCA_{1b}, but not
972 the expression of TRPV5 or NCX1. In contrast, the mRNA
973 expressions of paracellular genes, ZO-1, occludin and claudin-3,
974 are not modified by restraint stress. Since several antidepressant
975 or anxiolytic drugs alleviate stress-induced depressive and
976 anxiety symptoms, Charoenphandhu *et al.*⁽²²⁹⁾ have hypothe-
977 sised that these drugs might also enhance Ca transporter gene
978 expression in stressed rats. In fact, a 4-week daily administration
979 of 10 mg/kg fluoxetine, 10 mg/kg reboxetine or 10 mg/kg
980 venlafaxine differentially increased the duodenal Ca transporter
981 genes in stressed rats, whereas 2 mg/kg diazepam had no
982 such effect. These findings might be applied to help ameliorate

983 the stress-induced bone loss and osteoporosis by restoring
984 intestinal Ca absorption.

985 Octyphenol, a degradative product used to produce rubber,
986 pesticides and paints, and bisphenol A (BPA), an organic
987 compound used for manufacturing polycarbonate plastic and
988 epoxy resins, are known as endocrine disruptors. The effect of
989 both on serum Ca levels and expressions of Ca transport genes
990 in the duodenum and kidney was studied in pregnant mice.
991 Either octyphenol or BPA decreased serum Ca levels. Both
992 drugs decreased the levels of TRPV5 and CB_{9k} in the kidney and
993 the levels of TRPV6 in the duodenum. Gene expression and
994 protein expression of CB_{9k} were decreased in the duodenum by
995 BPA but increased by octyphenol at high doses. These results
996 indicate that decreased serum Ca levels caused by these
997 disruptors might be a consequence of the alteration in the
998 expression of genes related to Ca transport⁽²³⁰⁾.

999 *Gene × diet interactions influence intestinal calcium* 1000 *absorption*

1001 Dietary Ca restriction increases the efficiency of intestinal
1002 Ca absorption, but the impact of genetics on this adaptive
1003 response is not clear. In humans, the efficiency of intestinal Ca
1004 absorption varies from 7 to 75 %⁽²³¹⁾. The large variation is
1005 probably owing to the influence of multiple physiological
1006 factors (for example, growth, pregnancy, lactation, ageing)
1007 and environmental variables (for example, dietary Ca intake,
1008 vitamin D). Little information is available for the impact of
1009 genetics on the efficiency of intestinal Ca absorption and the
1010 adaptive up-regulation of Ca absorption to a low dietary Ca
1011 intake. Two laboratories have studied the efficiency of intestinal
1012 Ca absorption in different mice and have found that it is
1013 higher in C3H/HeJ mice in comparison with C57BL/6J mice,
1014 which suggests that genetic background might influence this
1015 trait⁽²³²⁾. In addition, racial differences in the ability of adoles-
1016 cent girls to increase Ca absorption efficiency during a low
1017 Ca intake also indicate that this adaptive response has a genetic
1018 component^(233,234). In agreement with this concept, adolescent
1019 black girls have been shown to exhibit higher intestinal
1020 Ca absorption as compared with white girls⁽²³⁵⁾, and this may
1021 contribute to the higher bone deposition found in black girls⁽²³⁶⁾.

1022 Replogle *et al.*⁽²³⁷⁾ have examined eleven inbred lines of
1023 mice fed on defined diets containing either high or low Ca
1024 concentration from weaning to 12 weeks of age. The authors
1025 have shown that genetic variation and gene × diet interactions
1026 affect not only the active intestinal Ca absorption, but also its
1027 relationship to bone. These interactions are partially explained
1028 by variations in the traditional cellular mediators (i.e. TRPV6,
1029 CB_{9k}, PMCA_{1b}, mRNA) and in the main hormonal regulator,
1030 1,25(OH)₂D₃, of intestinal Ca absorption. This field is relatively
1031 new, hence many efforts are required to bring more light for the
1032 understanding of this knowledge.

1033 **Conclusions**

1034 Ca is an ion involved in multiple physiological functions.
1035 Absorption through the intestinal epithelium is a complex

process regulated by an intricate network of hormones and
1036 nutritional factors. At present, the Western diet model adopted
1037 is poor in Ca content and at the same time interferes with the
1038 proper absorption of the cation. Some diseases such as osteo-
1039 porosis, hypertension and cancer are associated with dietary Ca
1040 restriction. Current recommendations are to obtain Ca from the
1041 diet in preference to supplements since dietary Ca intake has
1042 not been associated with the adverse effects of supplements,
1043 probably because Ca is provided in smaller boluses absorbed
1044 more slowly⁽²³⁸⁾. Milk and dairy products are the best sources of
1045 Ca. There have been advances in food industry attempts to
1046 compensate for the Ca shortage through the introduction of
1047 prebiotics and probiotics in the basic diet. Certain dietary habits
1048 such as an increased protein intake remain a point of debate. It
1049 is important to take into account that alterations in the redox
1050 state of the intestinal epithelium produced by some medications
1051 such as MEN, BSO and UDCA also modify the intestinal
1052 Ca absorption. Therefore, intestinal Ca absorption must be
1053 carefully attended, so it is necessary to consider not only the
1054 intake, but also possible interactions with other ions, the genetic
1055 background, the effect of diet and the use of certain medications.
1056 Health professionals should be aware of this knowledge in order
1057 to develop nutritional or medical strategies to stimulate the
1058 efficiency of intestinal Ca absorption and to prevent diseases.
1059

Acknowledgements

There are no conflicts of interest to declare.

References

1. Ma J, Johns RA & Stafford RS (2007) Americans are not
meeting current calcium recommendations. *Am J Clin Nutr*
85, 1361–1366.
2. Eisner V, Csordás G & Hajnóczky G (2013) Interactions
between sarco-endoplasmic reticulum and mitochondria in
cardiac and skeletal muscle – pivotal roles in Ca²⁺ and
reactive oxygen species signaling. *J Cell Sci* **15**, 2965–2978.
3. Szadujkis-Szadurska K, Szadujkis-Szadurski R, Szadujkis-
Szadurski L, *et al.* (2010) The role of calcium in modulating
the reactivity of the smooth muscle cells during ischemia/
reperfusion. Part 1. *Postepy Hig Med Dosw* **15**, 188–194.
4. Koklic T, Majumder R & Lentz BR (2014) Ca²⁺ switches the
effect of PS-containing membranes on factor Xa from
activating to inhibiting: implications for initiation of blood
coagulation. *Biochem J* **462**, 591–601.
5. Chaigne-Delalande B & Lenardo MJ (2014) Divalent cation
signaling in immune cells. *Trends Immunol* **35**, 332–344.
6. Zemel MB (2001) Calcium modulation of hypertension and
obesity: mechanisms and implications. *J Am Coll Nutr* **20**,
428S–435S.
7. Appel LJ, Brands MW, Daniels SR, *et al.* (2006) Dietary
approaches to prevent and treat hypertension: a scientific
statement from the American Heart Association. *Hypertension*
47, 296–308.
8. Laraichi S, Parra P, Zamanillo R, *et al.* (2013) Dietary supple-
mentation of calcium may counteract obesity in mice mediated
by changes in plasma fatty acids. *Lipids* **48**, 817–826.
9. Bartlett PJ, Gaspers LD, Pierobon N, *et al.* (2014) Calcium-
dependent regulation of glucose homeostasis in the liver.
Cell Calcium **55**, 306–316.

- 1093 10. Lin J, Manson JE, Lee IM, *et al.* (2007) Intakes of calcium and
1094 vitamin D and breast cancer risk in women. *Arch Intern Med*
1095 **167**, 1050–1059.
- 1096 11. Ju J, Kwak Y, Hao X, *et al.* (2012) Inhibitory effects of
1097 calcium against intestinal cancer in human colon cancer cells
1098 and *Apc*^{Min/+} mice. *Nutr Res Pract* **6**, 396–404.
- 1099 12. Williams CD, Whitley BM, Hoyo C, *et al.* (2012) Dietary
1100 calcium and risk for prostate cancer: a case–control study
1101 among US veterans. *Prev Chronic Dis* **9**, 110125.
- 1102 13. Merritt MA, Cramer DW, Vitonis AF, *et al.* (2013) Dairy foods
1103 and nutrients in relation to risk of ovarian cancer and major
1104 histological subtypes. *Int J Cancer* **132**, 1114–1124.
- 1105 14. Fleet JC & Schoch RD (2010) Molecular mechanisms for
1106 regulation of intestinal calcium absorption by vitamin D and
1107 other factors. *Crit Rev Clin Lab Sci* **47**, 181–195.
- 1108 15. Bronner F (1998) Calcium absorption – a paradigm for
1109 mineral absorption. *J Nutr* **128**, 917–920.
- 1110 16. Wasserman RH (2004) Vitamin D and the dual processes of
1111 intestinal calcium absorption. *J Nutr* **134**, 3137–3139.
- 1112 17. Marcus CS & Lengemann FW (1962) Absorption of Ca⁴⁵ and
1113 Sr⁸⁵ from solid and liquid food at various levels of the
1114 alimentary tract of the rat. *J Nutr* **77**, 155–160.
- 1115 18. Wali RK, Baum CL, Sitrin MD, *et al.* (1990) 1,25(OH)₂ vitamin
1116 D₃ stimulates membrane phosphoinositide turnover,
1117 activates protein kinase C, and increases cytosolic calcium in
1118 rat colonic epithelium. *J Clin Invest* **85**, 1296–1303.
- 1119 19. Tolosa de Talamoni N (1996) Calcium and phosphorous
1120 deficiencies alter the lipid composition and fluidity of
1121 intestinal basolateral membranes. *Comp Biochem Physiol A*
1122 *Physiol* **115**, 309–315.
- 1123 20. Nordin BE, Need AG, Morris HA, *et al.* (2004) Effect of age
1124 on calcium absorption in postmenopausal women. *Am J*
1125 *Clin Nutr* **80**, 998–1002.
- 1126 21. O'Brien KO, Nathanson MS, Mancini J, *et al.* (2003) Calcium
1127 absorption is significantly higher in adolescents during
1128 pregnancy than in the early postpartum period. *Am J Clin*
1129 *Nutr* **78**, 1188–1193.
- 1130 22. Prentice A (2000) Maternal calcium metabolism and bone
1131 mineral status. *Am J Clin Nutr* **71**, 1312S–1316S.
- 1132 23. Zhu Y, Goff JP, Reinhardt TA, *et al.* (1998) Pregnancy and
1133 lactation increase vitamin D-dependent intestinal membrane
1134 calcium adenosine triphosphatase and calcium binding
1135 protein messenger ribonucleic acid expression. *Endocrinology*
1136 **139**, 3520–3524.
- 1137 24. Liesegang A, Riner K & Boos A (2007) Effects of gestation
1138 and lactation on vitamin D receptor amounts in goats
1139 and sheep. *Domest Anim Endocrinol* **33**, 190–202.
- 1140 25. Wongdee K, Teerapornpantakit J, Siangpro C, *et al.* (2013)
1141 Duodenal villous hypertrophy and upregulation of claudin-15
1142 protein expression in lactating rats. *J Mol Histol* **44**, 103–109.
- 1143 26. Teerapornpantakit J, Klanchui A, Karoonuthaisiri N, *et al.*
1144 (2014) Expression of transcripts related to intestinal ion
1145 and nutrient absorption in pregnant and lactating rats as
1146 determined by custom-designed cDNA microarray. *Mol Cell*
1147 *Biochem* **391**, 103–116.
- 1148 27. van Abel M, Hoenderop JG & Bindels RJ (2005) The
1149 epithelial calcium channels TRPV5 and TRPV6: regulation
1150 and implications for disease. *Nuunyn Schmiedebergs Arch*
1151 *Pharmacol* **371**, 295–306.
- 1152 28. Bianco SD, Peng JB & Takanaga H (2007) Marked
1153 disturbance of calcium homeostasis in mice with targeted
1154 disruption of the *Trpv6* calcium channel gene. *J Bone Miner*
1155 *Res* **22**, 274–285.
- 1156 29. Cui M, Li Q, Johnson R, *et al.* (2012) Villin promoter-mediated
1157 transgenic expression of transient receptor potential cation
1158 channel, subfamily V, member 6 (TRPV6) increases intestinal
calcium absorption in wild-type and vitamin D receptor
knockout mice. *J Bone Miner Res* **27**, 2097–2107.
30. Den Dekker E, Hoenderop JG, Nilius B, *et al.* (2003)
The epithelial calcium channels, TRPV5 & TRPV6: from
identification towards regulation. *Cell Calcium* **33**, 497–507.
31. Hoenderop JG, Voets T, Hoefs S, *et al.* (2003) Homo- and
heterotetrameric architecture of the epithelial Ca²⁺ channels
TRPV5 and TRPV6. *EMBO J* **17**, 776–785.
32. Hoenderop JG, Vennekens R & Müller D (2001) Function and
expression of the epithelial Ca²⁺ channel family: comparison
of mammalian ECaC1 and 2. *J Physiol* **15**, 747–761.
33. Walters JR, Balesaria S, Chavele KM, *et al.* (2006) Calcium
channel TRPV6 expression in human duodenum: different
relationships to the vitamin D system and aging in men
and women. *J Bone Miner Res* **21**, 1770–1777.
34. Chow EC, Quach HP, Vieth R, *et al.* (2013) Temporal changes
in tissue 1 α ,25-dihydroxyvitamin D₃, vitamin D receptor target
genes, and calcium and PTH levels after 1,25(OH)₂D₃ treatment
in mice. *Am J Physiol Endocrinol Metab* **304**, E977–E989.
35. Tolosa de Talamoni N, Pérez A & Alisio A (1998) Effect of
cholecalciferol on intestinal epithelial cells. *Trends Comp*
Biochem Physiol **5**, 179–185.
36. Schwaller B (2010) Cytosolic Ca²⁺ buffers. *Cold Spring Harb*
Perspect Biol **2**, a004051.
37. Cheung WT, Richards DE & Rogers JH (1993) Calcium
binding by chick calretinin and rat calbindin D28k synthesised
in bacteria. *Eur J Biochem* **15**, 401–410.
38. Nägerl UV, Novo D, Mody I, *et al.* (2000) Binding kinetics of
calbindin-D_{28k} determined by flash photolysis of caged Ca²⁺.
Biophys J **79**, 3009–3018.
39. Choi KJ, Cho DS, Kim JY, *et al.* (2011) Ca-induced Ca release
from internal stores in INS-1 rat insulinoma cells. *Korean*
J Physiol Pharmacol **15**, 53–59.
40. Bellido T, Huening M, Raval-Pandya M, *et al.* (2000)
Calbindin-D_{28k} is expressed in osteoblastic cells and
suppresses their apoptosis by inhibiting caspase-3 activity.
J Biol Chem **275**, 26328–26332.
41. Merico V, de Barboza GD, Vasco C, *et al.* (2008)
A mitochondrial mechanism is involved in apoptosis of
Robertsonian mouse male germ cells. *Reproduction* **135**,
797–804.
42. Rodriguez V, Diaz de Barboza G, Ponce R, *et al.* (2010)
Spermatocyte apoptosis, which involves both intrinsic
and extrinsic pathways, explains the sterility of *Graomys*
griseoflavus × *Graomys centralis* male hybrids. *Reprod*
Fertil Dev **22**, 478–488.
43. Lambers TT, Mahieu F, Oancea E, *et al.* (2006) Calbindin-
D28K dynamically controls TRPV5-mediated Ca²⁺ transport.
EMBO J **12**, 2978–2988.
44. Airaksinen MS, Eilers J & Garaschuk O (1997) Ataxia and
altered dendritic calcium signaling in mice carrying a
targeted null mutation of the calbindin D28k gene. *Proc Natl*
Acad Sci U S A **18**, 1488–1493.
45. Christakos S, Dhawan P, Ajibade D, *et al.* (2010) Mechanisms
involved in vitamin D mediated intestinal calcium absorption
and in non-classical actions of vitamin D. *J Steroid Biochem*
Mol Biol **121**, 183–187.
46. Hwang I, Yang H, Kang HS, *et al.* (2013) Alteration of tight
junction gene expression by calcium- and vitamin D-deficient
diet in the duodenum of calbindin-null mice. *Int J Mol Sci* **14**,
22997–23010.
47. Schatzmann HJ (1966) ATP-dependent Ca⁺⁺-extrusion from
human red cells. *Experientia* **15**, 364–365.
48. Barley NF, Howard A, O'Callaghan D, *et al.* (2001) Epithelial
calcium transporter expression in human duodenum. *Am J*
Physiol Gastrointest Liver Physiol **280**, G285–G290.

- 1225 49. Peng JB, Chen XZ, Berger UV, *et al.* (1999) Molecular
1226 cloning and characterization of a channel-like transporter
1227 mediating intestinal calcium absorption. *J Biol Chem* **274**,
1228 22739–22746.
- 1229 50. Kim HJ, Lee GS, Ji YK, *et al.* (2006) Differential expression of
1230 uterine calcium transporter 1 and plasma membrane Ca²⁺
1231 ATPase 1b during rat estrous cycle. *Am J Physiol Endocrinol*
1232 *Metab* **291**, E234–E241.
- 1233 51. Stauffer TP, Guerini D, Celio MR, *et al.* (1997) Immuno-
1234 localization of the plasma membrane Ca²⁺ pump isoforms in
1235 the rat brain. *Brain Res* **748**, 21–29.
- 1236 52. Tribe RM, Moriarty P & Poston L (2000) Calcium homeostatic
1237 pathways change with gestation in human myometrium. *Biol*
1238 *Reprod* **63**, 748–755.
- 1239 53. Ghijsen WE, De Jong MD & Van Os CH (1982)
1240 ATP-dependent calcium transport and its correlation with
1241 Ca²⁺-ATPase activity in basolateral plasma membranes of rat
1242 duodenum. *Biochim Biophys Acta* **28**, 327–336.
- 1243 54. Anderson RG (1993) Caveolae: where incoming and outgoing
1244 messengers meet. *Proc Natl Acad Sci U S A* **90**, 10909–10913.
- 1245 55. Centeno VA, Díaz de Barboza GE, Marchionatti AM, *et al.*
1246 (2004) Dietary calcium deficiency increases Ca²⁺ uptake and
1247 Ca²⁺ extrusion mechanisms in chick enterocytes. *Comp*
1248 *Biochem Physiol A Mol Integr Physiol* **139**, 133–141.
- 1249 56. Liu C, Weng H, Chen L, *et al.* (2013) Impaired intestinal
1250 calcium absorption in protein 4.1R-deficient mice due to
1251 altered expression of plasma membrane calcium ATPase_{1b}
1252 (PMCA_{1b}). *J Biol Chem* **19**, 11407–11415.
- 1253 57. Ghijsen WE, De Jong MD & Van Os CH (1983) Kinetic
1254 properties of Na⁺/Ca²⁺ exchange in basolateral plasma
1255 membranes of rat small intestine. *Biochim Biophys Acta*
1256 **730**, 85–94.
- 1257 58. Philipson KD, Nicoll DA, Matsuoka S, *et al.* (1996) Molecular
1258 regulation of the Na⁺-Ca²⁺ exchanger. *Ann N Y Acad Sci*
1259 **779**, 20–28.
- 1260 59. Dong H, Sellers ZM, Smith A, *et al.* (2005) Na⁺/Ca²⁺
1261 exchange regulates Ca²⁺-dependent duodenal mucosal
1262 ion transport and HCO₃⁻ secretion in mice. *Am J Physiol*
1263 *Gastrointest Liver Physiol* **288**, G457–G465.
- 1264 60. Hwang I, Jung EM, Yang H, *et al.* (2011) Tissue-specific
1265 expression of the calcium transporter genes TRPV5, TRPV6,
1266 NCX1, and PMCA1b in the duodenum, kidney and heart of
1267 *Equus caballus*. *J Vet Med Sci* **73**, 1437–1444.
- 1268 61. Kim JA, Yang H, Hwang I, *et al.* (2011) Expression patterns
1269 and potential action of the calcium transport genes *Trpv5*,
1270 *Trpv6*, *Ncx1* and *Pmca1b* in the canine duodenum, kidney
1271 and uterus. *In Vivo* **25**, 773–780.
- 1272 62. Hoenderop JG, Hartog A, Stuiver M, *et al.* (2000) Localization
1273 of the epithelial Ca²⁺ channel in rabbit kidney and intestine.
1274 *J Am Soc Nephrol* **11**, 1171–1178.
- 1275 63. Blaustein MP & Lederer WJ (1999) Sodium/calcium exchange:
1276 its physiological implications. *Physiol Rev* **79**, 763–854.
- 1277 64. Yang H, Lei C, Cheng C, *et al.* (2012) The antiapoptotic effect
1278 of galectin-3 in human endometrial cells under the regulation
1279 of estrogen and progesterone. *Biol Reprod* **87**, 39.
- 1280 65. Hoenderop JG, Nilius B & Bindels RJ (2005) Calcium
1281 absorption across epithelia. *Physiol Rev* **85**, 373–422.
- 1282 66. González-Mariscal L, Betanzos A, Nava P, *et al.* (2003) Tight
1283 junction proteins. *Prog Biophys Mol Biol* **81**, 1–44.
- 1284 67. Bronner F & Pansu D (1999) Nutritional aspects of calcium
1285 absorption. *J Nutr* **129**, 9–12.
- 1286 68. Bronner F (2003) Mechanisms of intestinal calcium absorption.
1287 *J Cell Biochem* **88**, 387–393.
- 1288 69. Bouillon R, Lieben L, Mathieu C, *et al.* (2013) Vitamin D
1289 action: lessons from VDR and Cyp27b1 null mice. *Pediatr*
1290 *Endocrinol Rev* **10**, 354–366.
70. Wasserman RH, Smith CA, Brindak ME, *et al.* (1992) Vitamin D
1291 and mineral deficiencies increase the plasma membrane
1292 calcium pump of chicken intestine. *Gastroenterology* **102**,
1293 886–894.
- 1294 71. Centeno V, Picotto G & Pérez A (2011) Intestinal Na⁺/Ca²⁺
1295 exchanger protein and gene expression are regulated by
1296 1,25(OH)₂D₃ in vitamin D-deficient chicks. *Arch Biochem*
1297 *Biophys* **15**, 191–196.
- 1298 72. Balesaria S, Sangha S & Walters JR (2009) Human duodenum
1299 responses to vitamin D metabolites of TRPV6 and other genes
1300 involved in calcium absorption. *Am J Physiol Gastrointest*
1301 *Liver Physiol* **297**, G1193–G1197.
- 1302 73. Khuittuan P, Wongdee K, Jantarajit W, *et al.* (2013) Fibroblast
1303 growth factor-23 negates 1,25(OH)₂D₃-induced intestinal
1304 calcium transport by reducing the transcellular and para-
1305 cellular calcium fluxes. *Arch Biochem Biophys* **536**, 46–52.
- 1306 74. Fudge NJ & Kovacs CS (2010) Pregnancy up-regulates
1307 intestinal calcium absorption and skeletal mineralization
1308 independently of the vitamin D receptor. *Endocrinology*
1309 **151**, 886–895.
- 1310 75. Gallagher JC (2013) Vitamin D and aging. *Endocrinol Metab*
1311 *Clin North Am* **42**, 319–332.
- 1312 76. Sheikh MS, Schiller LR, Fordtran JS, *et al.* (1990) *In vivo*
1313 intestinal absorption of calcium in humans. *Miner Electrolyte*
1314 *Metab* **16**, 130–146.
- 1315 77. Karbach U (1992) Paracellular calcium transport across the
1316 small intestine. *J Nutr* **122**, 672–677.
- 1317 78. Fujita H, Sugimoto K, Inatomi S, *et al.* (2008) Tight junction
1318 proteins claudin-2 and -12 are critical for vitamin D-dependent
1319 Ca²⁺ absorption between enterocytes. *Mol Biol Cell* **19**,
1320 1912–1921.
- 1321 79. Garg MK, Kalra S & Mahalle N (2013) Defining vitamin D
1322 deficiency using surrogate markers. *Indian J Endocrinol*
1323 *Metab* **17**, 784–786.
- 1324 80. Picotto G, Massheimer V & Boland R (1997) Parathyroid
1325 hormone stimulates calcium influx and the cAMP messenger
1326 system in rat enterocytes. *Am J Physiol* **273**, C1349–C1353.
- 1327 81. Gentili C, Morelli S & de Boland AR (2003) Characterization
1328 of PTH/PTHrP receptor in rat duodenum: effects of ageing.
1329 *J Cell Biochem* **88**, 1157–1167.
- 1330 82. Nemere I & Larsson D (2002) Does PTH have a direct effect
1331 on intestine? *J Cell Biochem* **86**, 29–34.
- 1332 83. Cross HS, Debiec H & Peterlik M (1990) Thyroid hormone
1333 enhances the genomic action of calcitriol in the small
1334 intestine. *Prog Clin Biol Res* **332**, 163–180.
- 1335 84. Kumar V & Prasad R (2003) Thyroid hormones stimulate
1336 calcium transport systems in rat intestine. *Biochim Biophys*
1337 *Acta* **1639**, 185–194.
- 1338 85. Zoidis E, Gosteli-Peter M, Ghirlanda-Keller C, *et al.* (2002)
1339 IGF-I and GH stimulate Phex mRNA expression in lungs
1340 and bones and 1,25-dihydroxyvitamin D₃ production in
1341 hypophysectomized rats. *Eur J Endocrinol* **146**, 97–105.
- 1342 86. Fleet JC, Bruns ME, Hock JM, *et al.* (1994) Growth hormone and
1343 parathyroid hormone stimulate intestinal calcium absorption in
1344 aged female rats. *Endocrinology* **134**, 1755–1760.
- 1345 87. Fatayerji D, Mawer EB & Eastell R (2000) The role of insulin-
1346 like growth factor I in age-related changes in calcium
1347 homeostasis in men. *J Clin Endocrinol Metab* **85**, 4657–4662.
- 1348 88. Cotter AA & Cashman KD (2006) Effect of 17β-oestradiol on
1349 transepithelial calcium transport in human intestinal-like Caco-2
1350 cells and its interactions with 1,25-dihydroxycholecalciferol and
1351 9-*cis* retinoic acid. *Eur J Nutr* **45**, 234–241.
- 1352 89. Colin EM, Van Den Bemd GJ, Van Aken M, *et al.* (1999)
1353 Evidence for involvement of 17β-estradiol in intestinal calcium
1354 absorption independent of 1,25-dihydroxyvitamin D₃ level in
1355 the rat. *J Bone Miner Res* **14**, 57–64.
- 1356

- 1357 90. Bouillon R, Carmeliet G & Van Cromphaut S (2005) Intestinal
1358 calcium absorption: lessons from knockout mice and men.
1359 In *Vitamin D*, 2nd ed., pp. 429–452 [D Feldman, FH Glorieux
1360 and JW Pike, editors]. San Diego, CA: Academic Press.
- 1361 91. Campbell-Thompson M, Lynch IJ, Bhardwaj B, *et al.* (2001)
1362 Expression of estrogen receptor (ER) subtypes and ER β
1363 isoforms in colon cancer. *Cancer Res* **61**, 632–640.
- 1364 92. van Abel M, Hoenderop JG, van der Kemp AW, *et al.* (2003)
1365 Regulation of the epithelial Ca²⁺ channels in small intestine
1366 as studied by quantitative mRNA detection. *Am J Physiol*
1367 *Gastrointest Liver Physiol* **285**, G78–G85.
- 1368 93. Cotter AA & Cashman KD (2005) The effect of two dietary
1369 and a synthetic phytoestrogen on transepithelial calcium
1370 transport in human intestinal-like Caco-2 cells. *Eur J Nutr* **44**,
1371 72–78.
- 1372 94. Park CY & Weaver CM (2012) Vitamin D interactions with
1373 soy isoflavones on bone after menopause: a review. *Nutrients*
1374 **4**, 1610–1621.
- 1375 95. Reid IR (1997) Glucocorticoid osteoporosis – mechanisms
1376 and management. *Eur J Endocrinol* **137**, 209–217.
- 1377 96. Van Cromphaut SJ, Stockmans I, Torrekens S, *et al.* (2007)
1378 Duodenal calcium absorption in dexamethasone-treated
1379 mice: functional and molecular aspects. *Arch Biochem*
1380 *Biophys* **460**, 300–305.
- 1381 97. Lee GS, Choi KC, Jeung EB, *et al.* (2006) Glucocorticoids
1382 differentially regulate expression of duodenal and renal
1383 calbindin-D_{9k} through glucocorticoid receptor-mediated
1384 pathway in mouse model. *Am J Physiol Endocrinol Metab*
1385 **290**, E299–E307.
- 1386 98. Huybers S, Naber TH, Bindels RJ, *et al.* (2007) Prednisolone-
1387 induced Ca²⁺ malabsorption is caused by diminished
1388 expression of the epithelial Ca²⁺ channel TRPV6. *Am J*
1389 *Physiol Gastrointest Liver Physiol* **292**, G92–G97.
- 1390 99. Christakos S, Dhawan P & Liu Y (2003) New insights into the
1391 mechanisms of vitamin D action. *J Cell Biochem* **88**, 695–705.
- 1392 100. Brown AJ, Krits I & Armbrrecht HJ (2005) Effect of age,
1393 vitamin D, and calcium on the regulation of rat intestinal
1394 epithelial calcium channels. *Arch Biochem Biophys* **437**,
1395 51–58.
- 1396 101. Benn BS, Ajibade D & Porta A (2008) Active intestinal
1397 calcium transport in the absence of transient receptor
1398 potential vanilloid type 6 and calbindin-D_{9k}. *Endocrinology*
1399 **149**, 3196–3205.
- 1400 102. Tolosa de Talamoni N, Mykkanen H & Wasserman RH (1990)
1401 Enhancement of sulfhydryl group availability in the intestinal
1402 brush border membrane by deficiencies of dietary calcium
1403 and phosphorus in chicks. *J Nutr* **120**, 1198–1204.
- 1404 103. Brun LR, Brance ML & Rigalli A (2012) Luminal calcium
1405 concentration controls intestinal calcium absorption by
1406 modification of intestinal alkaline phosphatase activity. *Br J*
1407 *Nutr* **108**, 229–233.
- 1408 104. Zemel MB, Shi H, Greer B, *et al.* (2000) Regulation of
1409 adiposity by dietary calcium. *FASEB J* **14**, 1132–1138.
- 1410 105. Zemel MB (2005) The role of dairy foods in weight
1411 management. *J Am Coll Nutr* **24**, 537S–546S.
- 1412 106. Huang JY & Qi SJ (2015) Childhood obesity and food intake.
1413 *World J Pediatr* **11**, 101–107.
- 1414 107. Barr S (2003) Increased dairy product or calcium intake: is
1415 body weight or composition affected in humans? *J Nutr* **133**,
1416 245S–248S.
- 1417 108. Coe FL, Favus MJ & Asplin JR (2004) Nephrolithiasis. In *The*
1418 *Kidney*, 7th ed., pp. 1819–1866 [BM Brenner and FC Rector,
1419 editors]. Philadelphia, PA: Elsevier.
- 1420 109. Lemann J (2002) Idiopathic hypercalciuria. In *Disorders of*
1421 *Bone and Mineral Metabolism*, pp. 673–697 [FL Coe and M
1422 Favus, editors]. Philadelphia, PA: Lippincott.
110. Haghghi A, Samimagham H & Gohardehi G (2013) Calcium
1423 and vitamin D supplementation and risk of kidney stone
1424 formation in postmenopausal women. *Iran J Kidney Dis* **7**,
1425 210–213.
111. Jackson RD, LaCroix AZ, Gass M, *et al.* (2006) Calcium plus
1427 vitamin D supplementation and the risk of fractures. *N Engl J*
1428 *Med* **354**, 669–683.
112. Diaz-Lopez B & Cannata-Andia JB (2006) Supplementation
1430 of vitamin D and calcium: advantages and risks. *Nephrol Dial*
1431 *Transplant* **21**, 2375–2377.
113. Curhan GC, Willett WC, Rimm EB, *et al.* (1993) A prospective
1433 study of dietary calcium and other nutrients and the risk of
1434 symptomatic kidney stones. *N Engl J Med* **328**, 833–838.
114. Borghi L, Schianchi T, Meschi T, *et al.* (2002) Comparison of
1436 two diets for the prevention of recurrent stones in idiopathic
1437 hypercalciuria. *N Engl J Med* **346**, 77–84.
115. Park Y, Leitzmann MF, Subar A, *et al.* (2009) Dairy food,
1439 calcium, and risk of cancer in the NIH-AARP diet and
1440 health study. *Arch Intern Med* **169**, 391–401.
116. Tàrraga López PJ, Albero JS & Rodríguez-Montes JA (2014)
1442 Primary and secondary prevention of colorectal cancer. *Clin*
1443 *Med Insights Gastroenterol* **7**, 33–46.
117. Peterlik M, Grant WB & Cross HS (2009) Calcium, vitamin D
1445 and cancer. *Anticancer Res* **29**, 3687–3698.
118. McCullough ML, Rodriguez C, Diver WR, *et al.* (2005) Dairy,
1447 calcium, and vitamin D intake and postmenopausal breast
1448 cancer risk in the Cancer Prevention Study II Nutrition Cohort.
1449 *Cancer Epidemiol Biomarkers Prev* **14**, 2898–2904.
119. Hong Z, Tian C & Zhang X (2012) Dietary calcium intake,
1451 vitamin D levels, and breast cancer risk: a dose–response
1452 analysis of observational studies. *Breast Cancer Res Treat*
1453 **136**, 309–312.
120. Chen P, Hu P, Xie D, *et al.* (2010) Meta-analysis of vitamin D,
1455 calcium and the prevention of breast cancer. *Breast Cancer*
1456 *Res Treat* **121**, 469–477.
121. Vergne Y, Matta J, Morales L, *et al.* (2007) Intakes of calcium
1458 and vitamin D and breast cancer risk in women. *Arch Intern*
1459 *Med* **167**, 1050–1059.
122. Belizan JM & Villar J (1980) The relationship between
1461 calcium intake and edema-, proteinuria-, and hypertension-
1462 getosis: an hypothesis. *Am J Clin Nutr* **33**, 2202–2210.
123. Belizan JM, Villar J & Repke J (1988) The relationship between
1464 calcium intake and pregnancy-induced hypertension: up-to-date
1465 evidence. *Am J Obstet Gynecol* **158**, 898–902.
124. Camargo EB, Moraes LF, Souza CM, *et al.* (2013) Survey of
1467 calcium supplementation to prevent preeclampsia: the gap
1468 between evidence and practice in Brazil. *BMC Pregnancy*
1469 *Childbirth* **13**, 206.
125. Griffith LE, Guyatt GH & Cook RJ (1999) The influence of
1471 dietary and nondietary calcium supplementation on blood
1472 pressure: an updated metaanalysis of randomized controlled
1473 trials. *Am J Hypertens* **12**, 84–92.
126. van Mierlo LA, Arends LR, Streppel MT, *et al.* (2006) Blood
1475 pressure response to calcium supplementation: a meta-
1476 analysis of randomized controlled trials. *J Hum Hypertens*
1477 **20**, 571–580.
127. Varenna M, Manara M, Galli L, *et al.* (2013) The association
1479 between osteoporosis and hypertension: the role of a low
1480 dairy intake. *Calcif Tissue Int* **93**, 86–92.
128. Reid IR, Mason B, Horne A, *et al.* (2006) Randomized
1482 controlled trial of calcium in healthy older women. *Am J Med*
1483 **119**, 777–785.
129. Kim KM, Choi SH, Lim S, *et al.* (2014) Interactions between
1485 dietary calcium intake and bone mineral density or
1486 bone geometry in a low calcium intake population. *J Clin*
1487 *Endocrinol Metab* **99**, 2409–2417.

- 1489 130. Zhou W, Langsetmo L, Berger C, *et al.* (2013) Longitudinal
1490 changes in calcium and vitamin D intakes and relationship to
1491 bone mineral density in a prospective population-based
1492 study: the Canadian Multicentre Osteoporosis Study
1493 (CaMos). *J Musculoskelet Neuronal Interact* **13**, 470–479.
- 1494 131. Adler AJ & Berlyne GM (1985) Duodenal aluminum
1495 absorption in the rat: effect of vitamin D. *Am J Physiol* **249**,
1496 G209–G213.
- 1497 132. Dunn MA, Johnson NE, Liew MY, *et al.* (1993) Dietary
1498 aluminum chloride reduces the amount of intestinal calbindin
1499 D-28K in chicks fed low calcium or low phosphorus diets.
1500 *J Nutr* **123**, 1786–1793.
- 1501 133. Cox KA & Dunn MA (2001) Aluminum toxicity alters the
1502 regulation of calbindin-D28k protein and mRNA expression
1503 in chick intestine. *J Nutr* **131**, 2007–2013.
- 1504 134. Orihuela D, Meichtry V, Pregi N, *et al.* (2005) Short-term oral
1505 exposure to aluminium decreases glutathione intestinal
1506 levels and changes enzyme activities involved in its meta-
1507 bolism. *J Inorg Biochem* **99**, 1871–1878.
- 1508 135. Orihuela D, Meichtry V & Pizarro M (2005) Aluminium-
1509 induced impairment of transcellular calcium absorption in
1510 the small intestine: calcium uptake and glutathione influence.
1511 *J Inorg Biochem* **99**, 1879–1886.
- 1512 136. Orihuela D (2009) Inhibitory effect of aluminium on calcium
1513 absorption in small intestine of rats with different thyroid
1514 hormone status. *J Inorg Biochem* **103**, 1542–1547.
- 1515 137. Orihuela D (2007) Effect of aluminium on duodenal calcium
1516 transport in pregnant and lactating rats treated with bromo-
1517 criptine. *J Inorg Biochem* **101**, 1270–1274.
- 1518 138. Heaney RP & Nordin BE (2002) Calcium effects on
1519 phosphorus absorption: implications for the prevention and
1520 co-therapy of osteoporosis. *J Am Coll Nutr* **21**, 239–244.
- 1521 139. Ribovich ML & DeLuca HF (1978) Effect of dietary calcium
1522 and phosphorus on intestinal calcium absorption and vitamin
1523 D metabolism. *Arch Biochem Biophys* **188**, 145–156.
- 1524 140. Gray RW & Napoli JL (1983) Dietary phosphate deprivation
1525 increases 1,25-dihydroxyvitamin D₃ synthesis in rat kidney
1526 *in vitro*. *J Biol Chem* **258**, 1152–1155.
- 1527 141. Meyer RA Jr, Tenenhouse HS, Meyer MH, *et al.* (1989) The
1528 renal phosphate transport defect in normal mice parabiosed
1529 to X-linked hypophosphatemic mice persists after para-
1530 thyroidectomy. *J Bone Miner Res* **4**, 523–532.
- 1531 142. Bar A & Wasserman RH (1973) Control of calcium absorption
1532 and intestinal calcium-binding protein synthesis. *Biochem*
1533 *Biophys Res Commun* **54**, 191–196.
- 1534 143. Meyer J, Fullmer CS, Wasserman RH, *et al.* (1992) Dietary
1535 restriction of calcium, phosphorus, and vitamin D elicits
1536 differential regulation of the mRNAs for avian intestinal
1537 calbindin-D28k and the 1,25-dihydroxyvitamin D₃ receptor.
1538 *J Bone Miner Res* **7**, 441–448.
- 1539 144. Hendrix ZJ, Alcock NW & Archibald RM (1963) competition
1540 between calcium, strontium, and magnesium for absorption
1541 in the isolated rat intestine. *Clin Chem* **12**, 734–744.
- 1542 145. O'Donnell JM & Smith MW (1973) Uptake of calcium and
1543 magnesium by rat duodenal mucosa analysed by means of
1544 competing metals. *J Physiol* **229**, 733–749.
- 1545 146. Hardwick LL, Jones MR, Brautbar N, *et al.* (1991) Magnesium
1546 absorption: mechanisms and the influence of vitamin D,
1547 calcium and phosphate. *J Nutr* **121**, 13–23.
- 1548 147. de Swart PM, Sokole EB & Wilmink JM (1998) The
1549 interrelationship of calcium and magnesium absorption in
1550 idiopathic hypercalciuria and renal calcium stone disease.
1551 *J Urol* **159**, 669–672.
- 1552 148. Planells E, Sánchez-Morito N, Montellano MA, *et al.* (2000)
1553 Effect of magnesium deficiency on enterocyte Ca, Fe, Cu, Zn,
1554 Mn and Se content. *J Physiol Biochem* **56**, 217–222.
149. Dimai H-P, Porta S, Wirnsberger G, *et al.* (1998) Daily oral
1555 magnesium supplementation suppresses bone turnover in
1556 young adult males. *J Clin Endocrinol Metab* **83**, 2742–2748.
1557
150. Fine KD, Santa Ana CA, Porter JL, *et al.* (1991) Intestinal
1558 absorption of magnesium from food and supplements. *J Clin*
1559 *Invest* **88**, 396–402.
1560
151. Kozakai T, Uozumi N, Katoh K, *et al.* (2002) Dietary magnesium
1561 increases calcium absorption of ovine small intestine *in vivo*
1562 and *in vitro*. *Reprod Nutr Dev* **42**, 25–33.
1563
152. Bae YJ, Bu SY, Kim JY, *et al.* (2011) Magnesium supple-
1564 mentation through seaweed calcium extract rather than
1565 synthetic magnesium oxide improves femur bone mineral
1566 density and strength in ovariectomized rats. *Biol Trace Elem*
1567 *Res* **144**, 992–1002.
1568
153. Hessov I, Andersson H & Isaksson B (1983) Effects of a low-
1569 fat diet on mineral absorption in small-bowel disease. *Scand*
1570 *J Gastroenterol* **18**, 551–554.
1571
154. Haderslev KV, Jeppesen PB, Mortensen PB, *et al.* (2000)
1572 Absorption of calcium and magnesium in patients with
1573 intestinal resections treated with medium chain fatty acids.
1574 *Gut* **46**, 819–823.
1575
155. Jewell C, Cusack S & Cashman KD (2005) The effect of
1576 conjugated linoleic acid on transepithelial calcium transport
1577 and mediators of paracellular permeability in human
1578 intestinal-like Caco-2 cells. *Prostaglandins Leukot Essent*
1579 *Fatty Acids* **72**, 163–171.
1580
156. Murphy EF, Jewell C, Hooiveld GJ, *et al.* (2006) Conjugated
1581 linoleic acid enhances transepithelial calcium transport in
1582 human intestinal-like Caco-2 cells: an insight into molecular
1583 changes. *Prostaglandins Leukot Essent Fatty Acids* **74**,
1584 295–301.
1585
157. Coxam V (2007) Current data with inulin-type fructans
1586 and calcium, targeting bone health in adults. *J Nutr* **137**,
1587 2527S–2533S.
1588
158. Raschka L & Daniel H (2005) Diet composition and age
1589 determine the effects of inulin-type fructans on intestinal
1590 calcium absorption in rat. *Eur J Nutr* **44**, 360–364.
1591
159. Lobo AR, Cocato ML & Jorgetti V (2009) Changes in bone
1592 mass, biomechanical properties, and microarchitecture of
1593 calcium- and iron-deficient rats fed diets supplemented with
1594 inulin-type fructans. *Nutr Res* **29**, 873–881.
1595
160. Xiao Y, Cui J, Shi YH, *et al.* (2010) Effects of duodenal redox
1596 status on calcium absorption and related genes expression in
1597 high-fat diet-fed mice. *Nutrition* **26**, 1188–1194.
1598
161. Buchowski MS & Miller DD (1991) Lactose, calcium source
1599 and age affect calcium bioavailability in rats. *J Nutr* **121**,
1600 1746–1754.
1601
162. Dupuis Y, Tardivel S, Porembka Z, *et al.* (1991) Effect of
1602 some alkaline phosphatase inhibitors on intestinal calcium
1603 transfer. *Int J Biochem* **23**, 175–180.
1604
163. Sogabe N, Mizoi L, Asahi K, *et al.* (2004) Enhancement by
1605 lactose of intestinal alkaline phosphatase expression in rats.
1606 *Bone* **35**, 249–255.
1607
164. Nishimukai M, Watanabe J, Taguchi H, *et al.* (2008) Effects of
1608 epilactose on calcium absorption and serum lipid metabo-
1609 lism in rats. *J Agric Food Chem* **56**, 10340–10345.
1610
165. Suzuki T, Nishimukai M, Shinoki A, *et al.* (2010) Ingestion of
1611 epilactose, a non-digestible disaccharide, improves post-
1612 gastrectomy osteopenia and anemia in rats through the
1613 promotion of intestinal calcium and iron absorption. *J Agric*
1614 *Food Chem* **58**, 10787–10792.
1615
166. Suzuki T, Nishimukai M, Takechi M, *et al.* (2010) The
1616 nondigestible disaccharide epilactose increases paracellular
1617 Ca absorption via rho-associated kinase- and myosin light
1618 chain kinase-dependent mechanisms in rat small intestines.
1619 *J Agric Food Chem* **58**, 1927–1932.
1620

- 1621 167. Mineo H, Hara H & Tomita F (2002) Sugar alcohols enhance
1622 calcium transport from rat small and large intestine epithelium
1623 *in vitro*. *Dig Dis Sci* **47**, 1326–1333.
- 1624 168. Xiao J, Li X, Min X, *et al.* (2013) Mannitol improves absorption
1625 and retention of calcium and magnesium in growing rats.
1626 *Nutrition* **29**, 325–331.
- 1627 169. López-Huertas E, Teucher B, Boza JJ, *et al.* (2006) Absorption
1628 of calcium from milks enriched with fructo-oligosaccharides,
1629 caseinophosphopeptides, tricalcium phosphate, and milk
1630 solids. *Am J Clin Nutr* **83**, 310–316.
- 1631 170. Scholz-Ahrens KE, Schaafsma G, van den Heuvel EG, *et al.*
1632 (2001) Effects of prebiotics on mineral metabolism. *Am J Clin*
1633 *Nutr* **73**, 459S–464S.
- 1634 171. Suzuki T & Hara H (2004) Various non-digestible saccharides
1635 increase intracellular calcium ion concentration in rat small-
1636 intestinal enterocytes. *Br J Nutr* **92**, 751–755.
- 1637 172. Takasaki M, Inaba H, Ohta A, *et al.* (2000) Dietary short-
1638 chain fructooligosaccharides increase calbindin-D9k levels
1639 only in the large intestine in rats independent of dietary
1640 calcium deficiency or serum 1,25 dihydroxy vitamin D levels.
1641 *Int J Vitam Nutr Res* **70**, 206–213.
- 1642 173. Fukushima A, Aizaki Y & Sakuma K (2009) Short-chain fatty
1643 acids induce intestinal transient receptor potential vanilloid
1644 type 6 expression in rats and Caco-2 cells. *J Nutr* **139**, 20–25.
- 1645 174. Legette LL, Lee W, Martin BR, *et al.* (2012) Prebiotics enhance
1646 magnesium absorption and inulin-based fibers exert chronic
1647 effects on calcium utilization in a postmenopausal
1648 rodent model. *J Food Sci* **77**, H88–H94.
- 1649 175. Weaver CM, Martin BR, Nakatsu CH, *et al.* (2011)
1650 Galactooligosaccharides improve mineral absorption and
1651 bone properties in growing rats through gut fermentation.
1652 *J Agric Food Chem* **59**, 6501–6510.
- 1653 176. Tsuchita H, Suzuki T & Kuwata T (2001) The effect of casein
1654 phosphopeptides on calcium absorption from calcium-
1655 fortified milk in growing rats. *Br J Nutr* **85**, 5–10.
- 1656 177. Erba D, Ciappellano S & Testolin G (2002) Effect of the ratio
1657 of casein phosphopeptides to calcium (w/w) on passive
1658 calcium transport in the distal small intestine of rats. *Nutrition*
1659 **18**, 743–746.
- 1660 178. Teucher B, Majsak-Newman G, Dainty JR, *et al.* (2006) Calcium
1661 absorption is not increased by caseinophosphopeptides. *Am J*
1662 *Clin Nutr* **84**, 162–166.
- 1663 179. Cosentino S, Gravaghi C, Donetti E, *et al.* (2010)
1664 Caseinophosphopeptide-induced calcium uptake in human
1665 intestinal cell lines HT-29 and Caco2 is correlated to cellular
1666 differentiation. *J Nutr Biochem* **21**, 247–254.
- 1667 180. Colombini A, Perego S, Ardoino I, *et al.* (2013) Evaluation of
1668 a possible direct effect by casein phosphopeptides on
1669 paracellular and vitamin D controlled transcellular calcium
1670 transport mechanisms in intestinal human HT-29 and Caco2
1671 cell lines. *Food Funct* **4**, 1195–1203.
- 1672 181. Whisner CM, Martin BR & Nakatsu CH (2014) Soluble maize
1673 fibre affects short-term calcium absorption in adolescent
1674 boys and girls: a randomised controlled trial using dual stable
1675 isotopic tracers. *Br J Nutr* **112**, 446–456.
- 1676 182. Schrezenmeir J & de Vrese M (2001) Probiotics, prebiotics,
1677 and synbiotics – approaching a definition. *Am J Clin Nutr* **73**,
1678 361S–364S.
- 1679 183. Ghanem KZ, Badawy IH & Abdel-Samam AM (2004)
1680 Influence of yogourt and probiotic yogourt on the absorption
1681 of calcium, magnesium, iron and bone mineralization in rats.
1682 *Milchwissenschaft* **59**, 472–475.
- 1683 184. Gilman J & Cashman KD (2006) The effect of probiotic
1684 bacteria on transepithelial calcium transport and calcium
1685 uptake in human intestinal-like Caco-2 cells. *Curr Issues*
1686 *Intest Microbiol* **7**, 1–5.
185. Scholz-Ahrens K, Ade P, Marten B, *et al.* (2007) Prebiotics,
1687 probiotics, and synbiotics affect mineral absorption, bone
1688 mineral content, and bone structure. *J Nutr* **137**, 838S–846S.
1689
186. Kerstetter JE, O'Brien KO & Insogna KL (2003) Low protein
1690 intake: the impact on calcium and bone homeostasis
1691 in humans. *J Nutr* **133**, 855S–861S.
1692
187. Johnson NE, Alcantara EN & Linkswiler H (1970) Effect
1693 of level of protein intake on urinary and fecal calcium
1694 and calcium retention of young adult males. *J Nutr* **100**,
1695 1425–1430.
1696
188. Darling AL, Millward DJ, Torgerson DJ, *et al.* (2009) Dietary
1697 protein and bone health: a systematic review and meta-
1698 analysis. *Am J Clin Nutr* **90**, 1674–1692.
1699
189. Kerstetter JE, O'Brien KO, Caseria DM, *et al.* (2005) The
1700 impact of dietary protein on calcium absorption and kinetic
1701 measures of bone turnover in women. *J Clin Endocrinol*
1702 *Metab* **90**, 26–31.
1703
190. Gaffney-Stomberg E, Sun BH, Cucchi CE, *et al.* (2010) The
1704 effect of dietary protein on intestinal calcium absorption
1705 in rats. *Endocrinology* **151**, 1071–1078.
1706
191. Skipper A (2010) Nutrition Care Manual. Chicago, IL.
1707 American Dietetic Association, American Dietetic Association
1708 Nutrition Care Manual. <http://www.nutritioncaremanual.org>
1709 (accessed September 2015).
1710
192. Marcason W (2010) What is the effect of a high-protein diet
1711 on bone health? *J Am Diet Assoc* **110**, 812.
1712
193. Das AS, Das D, Mukherjee M, *et al.* (2005) Phytoestrogenic
1713 effects of black tea extract (*Camellia sinensis*) in an
1714 oophorectomized rat (*Rattus norvegicus*) model of osteo-
1715 porosis. *Life Sci* **77**, 3049–3057.
1716
194. Sharma V & Rao LJ (2009) A thought on the biological
1717 activities of black tea. *Crit Rev Food Sci Nutr* **49**, 379–404.
1718
195. Das AS, Banerjee M, Das D, *et al.* (2013) Black tea may be a
1719 prospective adjunct for calcium supplementation to prevent
1720 early menopausal bone loss in a rat model of osteoporosis.
1721 *J Osteoporos* **2013**, 760586.
1722
196. Hernandez-Avila M, Colditz GA, Stampfer MJ, *et al.* (1991)
1723 Caffeine, moderate alcohol intake, and risk of fractures of the
1724 hip and forearm in middle-aged women. *Am J Clin Nutr* **54**,
1725 157–163.
1726
197. Welch AA, Bingham SA, Reeve J, *et al.* (2007) More acidic
1727 dietary acid–base load is associated with reduced calcaneal
1728 broadband ultrasound attenuation in women but not in men:
1729 results from the EPIC-Norfolk cohort study. *Am J Clin Nutr*
1730 **85**, 1134–1141.
1731
198. Cooper C, Atkinson EJ, Wahner HW, *et al.* (1992) Is caffeine
1732 consumption a risk factor for osteoporosis? *J Bone Miner Res*
1733 **7**, 465–471.
1734
199. Huopio J, Kröger H, Honkanen R, *et al.* (2000) Risk factors for
1735 perimenopausal fractures: a prospective study. *Osteoporos Int*
1736 **11**, 219–227.
1737
200. Hallström H, Byberg L, Glynn A, *et al.* (2013) Long-term
1738 coffee consumption in relation to fracture risk and bone
1739 mineral density in women. *Am J Epidemiol* **178**, 898–909.
1740
201. Yeh JK & Aloia JF (1986) Differential effect of caffeine
1741 administration on calcium and vitamin D metabolism in
1742 young and adult rats. *J Bone Miner Res* **1**, 251–258.
1743
202. Hasling C, Søndergaard K, Charles P, *et al.* (1992) Calcium
1744 metabolism in postmenopausal osteoporotic women is
1745 determined by dietary calcium and coffee intake. *J Nutr* **122**,
1746 1119–1126.
1747
203. Folwarczna J, Pytlik M, Zych M, *et al.* (2013) Favorable effect
1748 of moderate dose caffeine on the skeletal system in
1749 ovariectomized rats. *Mol Nutr Food Res* **57**, 1772–1784.
1750
204. Cano-Marquina A, Tarín JJ & Cano A (2013) The impact of
1751 coffee on health. *Maturitas* **75**, 7–21.
1752

- 1753 205. Folwarczna J, Zych M, Nowińska B, *et al.* (2014) Unfavorable
1754 effect of trigonelline, an alkaloid present in coffee and
1755 fenugreek, on bone mechanical properties in estrogen-
1756 deficient rats. *Mol Nutr Food Res* **58**, 1457–1464.
- 1757 206. Lowe M, Strauss AW, Alpers R, *et al.* (1990) Molecular cloning
1758 and expression of a cDNA encoding the membrane-associated
1759 rat intestinal alkaline phosphatase. *Biochim Biophys Acta*
1760 **1037**, 170–177.
- 1761 207. Marchionatti A, Alisio A, Díaz de Barboza G, *et al.* (2001)
1762 DL-Buthionine-S,R-sulfoximine affects intestinal alkaline
1763 phosphatase activity. *Comp Biochem Physiol C Toxicol*
1764 *Pharmacol* **129**, 85–91.
- 1765 208. Chiou TJ & Tzeng WF (2000) The roles of glutathione and
1766 antioxidant enzymes in menadione-induced oxidative stress.
1767 *Toxicology* **154**, 75–84.
- 1768 209. Shiraki M, Shiraki Y, Aoki C, *et al.* (2000) Vitamin K₂
1769 (menatetrenone) effectively prevents fractures and sustains
1770 lumbar bone mineral density in osteoporosis. *J Bone Miner*
1771 *Res* **15**, 515–521.
- 1772 210. Sata N, Klonowski-Stumpe H, Han B, *et al.* (1997) Men-
1773 adione induces both necrosis and apoptosis in rat pancreatic
1774 acinar AR4-2J cells. *Free Radic Biol Med* **23**, 844–850.
- 1775 211. Marchionatti AM, Díaz de Barboza GE, Centeno VA, *et al.*
1776 (2003) Effects of a single dose of menadione on the intestinal
1777 calcium absorption and associated variables. *J Nutr Biochem*
1778 **14**, 466–472.
- 1779 212. Higuchi Y (2004) Glutathione depletion-induced chromosomal
1780 DNA fragmentation associated with apoptosis and necrosis.
1781 *J Cell Mol Med* **8**, 455–464.
- 1782 213. Marchionatti AM, Perez AV, Diaz de Barboza GE, *et al.*
1783 (2008) Mitochondrial dysfunction is responsible for the
1784 intestinal calcium absorption inhibition induced by menadione.
1785 *Biochim Biophys Acta* **1780**, 101–107.
- 1786 214. Suzuki T & Hara H (2009) Quercetin enhances intestinal
1787 barrier function through the assembly of zonula [corrected]
1788 occludens-2, occludin, and claudin-1 and the expression of
1789 claudin-4 in Caco-2 cells. *J Nutr* **139**, 965–974.
- 1790 215. Marchionatti AM, Pacciaroni A & Tolosa de Talamoni NG
1791 (2013) Effects of quercetin and menadione on intestinal
1792 calcium absorption and the underlying mechanisms. *Comp*
1793 *Biochem Physiol A Mol Integr Physiol* **164**, 215–220.
- 1794 216. Carpentieri A, Marchionatti A, Areco V, *et al.* (2014) Anti-
1795 oxidant and antiapoptotic properties of melatonin restore
1796 intestinal calcium absorption altered by menadione. *Mol Cell*
1797 *Biochem* **387**, 197–205.
- 1798 217. Rivoira MA, Marchionatti AM, Centeno VA, *et al.* (2012)
1799 Sodium deoxycholate inhibits chick duodenal calcium
1800 absorption through oxidative stress and apoptosis. *Comp*
1801 *Biochem Physiol A Mol Integr Physiol* **162**, 397–405.
- 1802 218. Lamireau T, Zoltowska M, Levy E, *et al.* (2003) Effects of bile
1803 acids on biliary epithelial cells: proliferation, cytotoxicity, and
1804 cytokine secretion. *Life Sci* **72**, 1401–1411.
- 1805 219. Kawano A, Ishikawa H, Kamano T, *et al.* (2010) Significance
1806 of fecal deoxycholic acid concentration for colorectal tumor
1807 enlargement. *Asian Pac J Cancer Prev* **11**, 1541–1546.
- 1808 220. Jean-Louis S, Akare S, Ali MA, *et al.* (2006) Deoxycholic acid
1809 induces intracellular signaling through membrane perturbations.
1810 *J Biol Chem* **281**, 14948–14960.
221. Araki Y, Katoh T, Ogawa A, *et al.* (2005) Bile acid modulates
1811 transepithelial permeability via the generation of reactive
1812 oxygen species in the Caco-2 cell line. *Free Radic Biol Med*
1813 **39**, 769–780.
222. Rodríguez V, Rivoira M, Marchionatti A, *et al.* (2013)
1814 Ursodeoxycholic and deoxycholic acids: a good and a bad
1815 bile acid for intestinal calcium absorption. *Arch Biochem*
1816 *Biophys* **540**, 19–25.
223. Makishima M, Lu TT, Xie W, *et al.* (2002) Vitamin D receptor
1817 as an intestinal bile acid sensor. *Science* **296**, 1313–1316.
224. Krasowski MD, Ni A, Hagey LR, *et al.* (2011) Evolution of
1818 promiscuous nuclear hormone receptors: LXR, FXR, VDR,
1819 PXR, and CAR. *Mol Cell Endocrinol* **334**, 39–48.
225. D'Aldebert E, Biyeyeme Bi Mve MJ, Mergey M, *et al.* (2009)
1820 Bile salts control the antimicrobial peptide cathelicidin
1821 through nuclear receptors in the human biliary epithelium.
1822 *Gastroenterology* **136**, 1435–1443.
226. Hansen KE, Jones AN, Lindstrom MJ, *et al.* (2010) Do proton
1823 pump inhibitors decrease calcium absorption? *J Bone Miner*
1824 *Res* **25**, 2786–2789.
227. Wahl TO, Gobuty AH, Lukert BP, *et al.* (1981) Long-term
1825 anticonvulsant therapy and intestinal calcium absorption.
1826 *Clin Pharmacol Ther* **30**, 506–512.
228. von Borstel Smith M, Crofoot K, Rodriguez-Proteau R, *et al.*
1827 (2007) Effects of phenytoin and carbamazepine on calcium
1828 transport in Caco-2 cells. *Toxicol In Vitro* **21**, 855–862.
229. Charoenphandhu N, Teerapornpuntakit J, Lapmanee S, *et al.*
1829 (2012) Duodenal calcium transporter mRNA expression
1830 in stressed male rats treated with diazepam, fluoxetine,
1831 reboxetine, or venlafaxine. *Mol Cell Biochem* **369**, 87–94.
230. Kim S, An BS, Yang H, *et al.* (2013) Effects of octylphenol
1832 and bisphenol A on the expression of calcium transport genes
1833 in the mouse duodenum and kidney during pregnancy.
1834 *Toxicology* **303**, 99–106.
231. Alevizaki CC, Ikkos DG & Singhelakis P (1973) Progressive
1835 decrease of true intestinal calcium absorption with age in
1836 normal man. *J Nucl Med* **14**, 760–762.
232. Chen C & Kalu DN (1999) Strain differences in bone density
1837 and calcium metabolism between C3H/HeJ and C57BL/6J mice.
1838 *Bone* **25**, 413–420.
233. Wu L, Martin BR, Braun MM, *et al.* (2010) Calcium requirements
1839 and metabolism in Chinese-American boys and girls. *J Bone*
1840 *Miner Res* **25**, 1842–1849.
234. Weaver CM, McCabe LD, McCabe GP, *et al.* (2008) Vitamin D
1841 status and calcium metabolism in adolescent black and
1842 white girls on a range of controlled calcium intakes. *J Clin*
1843 *Endocrinol Metab* **93**, 3907–3914.
235. Bryant RJ, Wastney ME, Martin BR, *et al.* (2003) Racial
1844 differences in bone turnover and calcium metabolism in
1845 adolescent females. *J Clin Endocrinol Metab* **88**, 1043–1047.
236. Braun M, Palacios C, Wigertz K, *et al.* (2007) Racial differences
1846 in skeletal calcium retention in adolescent girls with varied
1847 controlled calcium intakes. *Am J Clin Nutr* **85**, 1657–1663.
237. Replogle RA, Li Q, Wang L, *et al.* (2014) Gene-by-diet
1848 interactions influence calcium absorption and bone density
1849 in mice. *J Bone Miner Res* **29**, 657–665.
238. Reid IR (2014) Should we prescribe calcium supplements for
1850 osteoporosis prevention? *J Bone Metab* **21**, 21–28.