

Osmotic stress sensitizes naturally resistant cells to TNF- α -induced apoptosis

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Abstract

Most cells are naturally resistant to TNF- α -induced cell death and become sensitized when NF- κ B transactivation is blocked or in the presence of protein synthesis inhibitors that prevent the expression of anti-apoptotic genes. In this report we analyzed the role of osmotic stress on TNF- α -induced cell death. We found that it sensitizes the naturally resistant HeLa cells to TNF- α -induced apoptosis, with the involvement of an increase in the activity of several kinases, the inhibition of Bcl-2 expression, and a late increase on NF- κ B activation. Cell death occurs regardless of the enhanced NF- κ B activity, whose inhibition produces an increase in apoptosis. The inhibition of p38 kinase, also involved in NF- κ B activation, significantly increases the effect of osmotic stress on TNF- α -induced cell death.

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Abbreviations: TNF- α , tumor necrosis factor- α ; TRADD, TNF receptor-associated death domain; TRAF2 TNF receptor-associated factor 2; RIP, receptor interacting protein; FADD Fas-associated death domain; JNK, *c-jun* amino terminal kinases; NIK, NF- κ B-inducing kinases; I κ B, inhibitor of NF- κ B; MAPKs, MAP kinases; Erk1/2, extracellular regulated kinases; SAPKs, stress activated protein kinases; SZ, sulfasalazine; GADD, growth arrest and DNA damage; ROI, reactive oxygen intermediates; Mn-SOD, Mn-superoxide dismutase.

Introduction

Tumor necrosis factor- α (TNF- α) is an inflammatory cytokine that elicits a wide range of biological responses, apoptosis being one of them.^{1,2} These responses occur after TNF- α -induced trimerization of cell surface receptors p55 and p75. Although most of the biological activities of TNF- α appear to be transduced by p55, many can also be mediated by p75,^{2,3} although this receptor is a poorer inducer of apoptosis.

TNF- α -induced assembling of the TNF receptor recruits other proteins,^{4,5} including the TNF receptor-associated death domain (TRADD), the TNF receptor-associated factor 2 (TRAF2) and the receptor-interacting protein (RIP) to p55 cytoplasmic death domain.^{6–8} TRADD acts as an adapter that recruits the downstream transducers Fas-associated death domain-containing protein (FADD) and activates the apoptotic caspase 8. In turn, TRAF2 has been implicated in the activation of two distinct pathways that leads to: the activation of AP-1 via the *c-Jun* amino terminal kinases (JNKs) and, together with RIP, NF- κ B activation, via the NF- κ B-inducing kinase (NIK).^{9,10}

The cellular resistance to TNF- α has been attributed to a molecular mechanism by which TNF- α exerts on the target cell two different signals. One of them is the activation of caspases that leads to cell death, while the other, that requires the *nov*o synthesis of proteins, involves a protective pathway where NF- κ B regulated anti-apoptotic genes such as c-IAP1, c-IAP2, TRAF1 and TRAF2 are induced.^{9,11–17} The induction of these protective genes could be the reason that explains why most cells carrying p55, that are resistant become sensitized to TNF- α -induced death with protein or RNA synthesis inhibitors, such as cycloheximide or actinomycin D.¹⁸ These observations are in agreement with other experiments showing that cells naturally resistant to TNF- α , transfected with an expression vector for the inhibitor of NF- κ B (I κ B), become sensitized and undergo apoptosis.^{14,17}

Although most evidences support an anti-apoptotic role for NF- κ B,^{11–17} p53 and *c-myc*, are target genes of this transcription factor, both of which may act as pro-apoptotic.^{19,20} Interestingly, NF- κ B can either induce or repress apoptosis in CD4⁺CD8⁺ thymocytes, depending upon the nature of the activator.²¹ In addition, its role in apoptosis may be dependent of the cell type, probably due to cross talk with other signals. The precise molecular mechanisms that determine the switch from an anti- to a pro-apoptotic role have not been clearly defined. A similar observation can be applied to AP-1, as some authors have reported no effect of AP-1 activation on TNF- α -induced cell death,⁹ while others give to AP-1 activation a cell type-specific anti-apoptotic role.¹¹

The MAP kinases (MAPKs) were originally described as proteins activated by mitogens. Lately the family of MAPKs has expanded and a variety of stimuli, including non-

mitogenic ones, have been reported as MAPKs activators. Three major MAPK families have been discovered. Erk1/2, which responds mostly to signals that trigger cell proliferation;²² JNK, which leads to AP-1 activation, through the phosphorylation of *c-Jun*; and ATF-2 transcription factors^{23–25} and the p38 family.^{26,27} Four isoforms of p38 have been identified: p38 α (or simply p38), p38 β , p38 γ (ERK6) and p38 δ .²⁷

Although most reports link MAPKs to nuclear signaling, it seems clear that all of them phosphorylate both nuclear and cytoplasmic targets. The JNKs and p38s are activated by most environmental stresses (hence the name SAPKs for Stress Activated Protein Kinases) such as hyperosmolarity^{28,29} and some of them can also be activated by cytokines and TNF- α .^{9,26–30} All these kinases play critical roles as components of the signal transduction pathways that control cell differentiation, the cell cycle, and cell death.^{26–32} As observed with the transcription factors NF- κ B and AP-1, the role of MAP kinases in apoptosis is equally controversial. While there is a tendency to link Erk1/2 with cell growth and the SAPKs with the onset of apoptosis, both pro- and anti-apoptotic roles have been attributed to the members of the MAPK family.^{27,32}

Accordingly, in this work, we analyzed the activity of these kinases as well as the role of NF- κ B activation on sensitivity to TNF- α -induced-cell death in HeLa cells, which are naturally resistant to this cytokine. We found that osmotic stress induces a two-phase NF- κ B activity, involving a late induction of this activity, while the early NF- κ B activity induced by TNF- α is inhibited. Regardless of triggering a late anti-apoptotic signal, it sensitizes them to TNF- α -induced apoptosis by a mechanism that might involve the activation of several kinases and inhibition of the anti-apoptotic gene *bcl-2*.

Results

Stress by hyperosmolarity sensitizes HeLa cells to TNF- α -induced apoptosis

It was previously shown that cells naturally resistant to TNF- α -induced cell death become sensitive upon previous treatment with inhibitors of *de novo* protein synthesis, either at the transcriptional or translational level, as cycloheximide or actinomycin D¹⁶ as shown in Figure 1A. Those chemicals are well known for their inhibitory actions, but they induce as well a variety of biochemical changes in the cell, including an increase in the enzymatic activity of some stress activated protein kinases or SAPKs. In order to analyze if additional pathways, not related to an inhibition of *de novo* synthesis or NF- κ B suppression could be involved on TNF- α -induced apoptosis we investigated the role of osmotic stress. For this purpose, HeLa cells were stimulated with TNF- α at different doses and 100 mM of NaCl for different periods of time and then cell survival was determined (Figure 1A, B, C). Although the treatment with NaCl alone has no effect on cell survival, it sensitizes cells to the apoptotic effect of TNF- α in a dose-dependent fashion (Figure 1A). Similar results were obtained with sorbitol 300 mM plus TNF- α (data not shown).

TNF- α may induce cell death by two different pathways, depending of the target cell, such as necrosis or apoptosis. The integrity of the cellular membrane was not lost by stimulation with high doses of TNF- α plus NaCl, as we determined by acridine orange staining (Figure 1C). In addition, the typical internucleosomal DNA fragmentation by this treatment is shown (Figure 1C).

In some models where TNF- α induces apoptosis, it triggers caspase 8 activation and Bcl-2 has a poor protective effect.³³ In addition, there are enough evidences that support an anti-apoptotic role for Bcl-2 in the pathways that involve caspase 9 activation by mitochondrial products, like cytochrome *c*.^{34,35} In order to determine whether this pathway may be involved in this model of osmotic stress sensitization to TNF- α -induced cell death, we analyzed the levels of Bcl-2 and Bax in HeLa cells under different treatments. As shown in Figure 2A, Bcl-2 was significantly inhibited by treatment with NaCl plus TNF- α , while the same dose of NaCl or TNF- α alone had not effect. In addition, the levels of the pro-apoptotic protein Bax, were not affected by any treatment (Figure 2B). As these results may be indicating the participation of the mitochondrial pathway in the cell death induced by NaCl plus TNF- α , we then analyzed the activity of caspase 9. Given that activation of caspases is associated with the proteolytic processing of this enzyme, we examined its cleavage by Western blot. Figure 2B shows a slow increase of the cleavage product of 37 kDa and a reduced amount of not cleaved substrate at 6 h after NaCl plus TNF- α treatment. In view that TNF- α may induce apoptosis through the activation of caspase 8 we also analyzed the activation of this caspase. As shown in Figure 2B, there is not difference on the levels of the procaspase 8 between the different treatments. Similar results were obtained for both caspases at 3 h and 12 h post treatment. The absence of caspase 8 activity probably points out that caspase 9 activation occurs in a pathway that is independent of caspase 8. These data, may suggest that hyperosmolarity sensitizes the naturally resistant cells to TNF- α -induced cell death in an apoptotic way that involves a mitochondrial pathway.

Hyperosmolarity and addition of TNF- α activate several kinases

There are several evidences that indicate the participation of MAPKs in the control of cell cycle and survival.^{25–30} In addition, it is well known that stress and TNF- α addition activate several of these kinases.^{9,10,26–29,32}

In order to determine if a particularly different pattern of MAPK induction by a simultaneous treatment with NaCl plus TNF- α could be related to the induction of apoptosis, we performed kinase assays. We measured the levels of JNK, p38 and ERK6 activity on cell lysates from cultures stimulated with NaCl, TNF- α and NaCl plus TNF- α as indicated. As expected, the treatment with NaCl alone produced a stimulation of all the three kinases (Figure 3). In agreement with previous works, TNF- α -induced not only JNK and p38,^{9,10,31} but also ERK6 (Figure 3). The

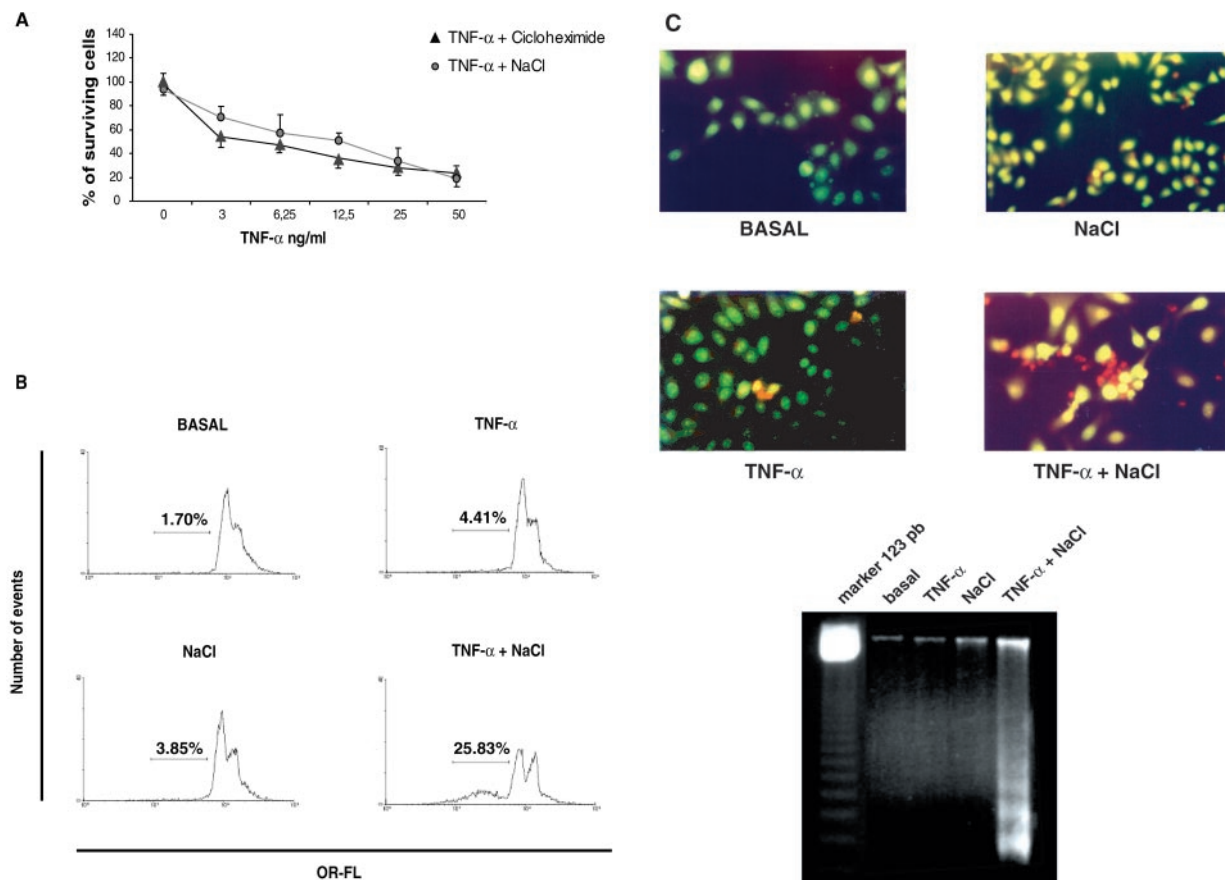


Figure 1 NaCl sensitizes HeLa cells to TNF- α -induced cell death. **(A)** Cells were treated with TNF- α alone or in the presence of 0.5 μ g/ml cycloheximide or 100 mM NaCl as indicated by 18 h. Absorbance of stained surviving cells was determined at 570 nm. Each value corresponds to the average of three wells \pm S.D., where the percentage of surviving cells was determined respect to the basal absorbance (0.531 ± 0.095). The treatment with NaCl or TNF- α (50 ng/ml) alone have no effect on the cell number: absorbance 0.478 ± 0.075 and 0.504 ± 0.060 respectively. **(B)** HeLa cells were treated with 10 ng/ml TNF- α , 100 mM NaCl, or both. After 12 h of treatment cells were harvested, nuclei were stained with propidium iodide and analyzed by flow cytometry. The number above the marker in the FACS histogram represents the percentage of sub-diploid particles. **(C)** HeLa cells were treated with 100 mM NaCl, 10 ng/ml TNF- α , or both for 18 h. Cells were stained with ethidium bromide and acridine orange. Apoptotic cells (red stained) show condensation respect to the green viable cells. The treatment with NaCl did not produce any change in the morphology of the cells, being large green cells, similar to the treatment with TNF- α or basal conditions. DNA laddering was visualized in an agarose gel electrophoresis stained with ethidium bromide. Similar results were obtained from three independent experiments

treatment with NaCl plus TNF- α , both in a dose whose combination induces apoptosis (Figure 1B), caused a significant increase of the three kinase activities if compared with each one of these stimulus alone (Figure 3). The observed changes on kinase activities was not due to an increase on the levels of kinase synthesis by the treatment with NaCl, TNF- α or both simultaneously, as was confirmed by Western blots for the three kinases (Figure 3, lower panel).

These results indicate that nevertheless, being NaCl and TNF- α two types of stimulus, rely different concerning their biological role, receptors and transduction signals, yet both of them may activate at least the same three kinases. In addition, although each one of these stimuli is unable to induce apoptosis, the target cell interprets the simultaneous stimulation as a death signal. According with our results, this death signal probably correlates with a high increase or a new balance of kinase activities where p38, ERK6 and JNK are hyper-activated.

Osmotic stress and TNF- α induce the activation of NF- κ B

It was previously shown that several stress signals activate the NF- κ B pathway, and different kinases, including p38 kinase, have been involved in the control of its activity.^{30,36} In addition, there are strong evidences that support an anti-apoptotic role for this transcription factor.^{11–15} Since in our experimental conditions, the simultaneous stimulation with two non-apoptotic signals, results in cell death, we analyzed the activity of NF- κ B under osmotic stress, TNF- α and the simultaneous treatment with both stimuli. Transactivation was determined at 12 h, being that the time that cell death may be detected by microscopy and flow cytometry and earlier, at 6 h post stimulation, when cells look viable. Figure 4A shows that at 12 h, in both, the osmotic stress or, as expected, TNF- α treatment, the transcriptional activity of NF- κ B is significantly increased with respect to the basal levels. In addition, even as the simultaneous treatment with both stimuli produces an

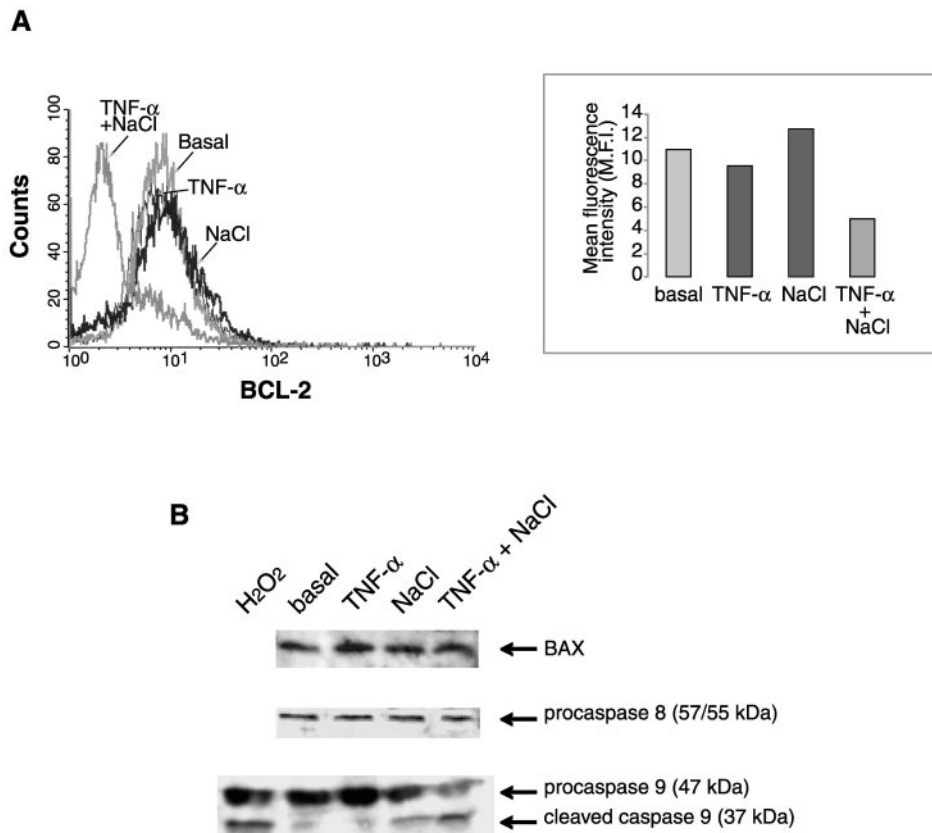


Figure 2 Analysis of caspases and expression of pro- and anti-apoptotic genes. **(A)** HeLa cells were treated as in 1 C and cells were permeabilized, incubated with anti-Bcl-2 (Santa Cruz) and labeled with anti-mouse IgG-FITC. Cells were analyzed by flow cytometry. **(B)** HeLa cells were stimulated by 6 h, and the specific proteins and cleavage products were detected by Western blot using specific antibodies (Santa Cruz). Similar results were obtained for caspase 8 and 9 at 3 and 12 h. As a positive control for caspase 9 activation, cells were stimulated by 4 h with 0.2% H₂O₂

apoptotic signal, it induces NF- κ B activity and there is not a synergism between both signals.

Early, at 6 h post stimulation the pattern of stimulation looks different. The transcriptional activity of NF- κ B is significantly increased under TNF- α treatment, but surprisingly NaCl has a poor stimulatory effect respect to the basal levels. In addition, the simultaneous treatment with both stimuli produces a significant inhibition on NF- κ B activity respect to the cultures stimulated with TNF- α alone (Figure 4C).

Although both stimuli produce an increase on NF- κ B activity at one late phase in the route to apoptosis these results, clearly demonstrate that each one of these inductors have a different kinetics, that probably involves a specific particular cascade leading to NF- κ B activation. Furthermore, some additional inhibitory signal for NF- κ B activation is triggered early by hyperosmotic stress.

It was previously reported that p38 activity is required for the NF- κ B activity induced by TNF- α .³⁰ In order to analyze the requirement of this kinase for each one of the routes to NF- κ B activation used by TNF- α , NaCl or both simultaneously, we analyzed the effect of the specific p38 inhibitor SB203580. Our results indicate that the addition of the specific p38 inhibitor, in a dose that is clearly effective

(Figure 4B) has an inhibitory effect on NF- κ B transactivation induced by every treatment (Figure 4A,C). These results demonstrate that some different pathways that lead to the increase on NF- κ B activation are used by osmotic stress, TNF- α or both, simultaneously. However, this kinase plays an important role on regulating NF- κ B transactivation, which is independent of the type of stimuli or its kinetics.

Inhibition of NF- κ B or p38 increases the NaCl sensitization to TNF- α -induced cell death

According to our results, NF- κ B is activated by osmotic stress plus TNF- α through a pathway that involves at least p38. Therefore, we wanted to determine the role of NF- κ B and the p38 pathway in this model of apoptosis. For this purpose, the cells were stimulated with the specific inhibitor of NF- κ B, sulfasalazine, in a dose that is not cytotoxic (Figure 5A) and inhibits NF- κ B activity (Figure 5B) and then stimulated with NaCl 100 mM plus TNF- α . Flow cytometry measurements were performed and as shown in Figure 5A, the addition of sulfasalazine, at a dose that has not basal effect, increases the cell death induced by the simultaneous treatment with NaCl plus TNF- α with respect to the cultures with the same stimulation, but without the inhibitor. These results clearly

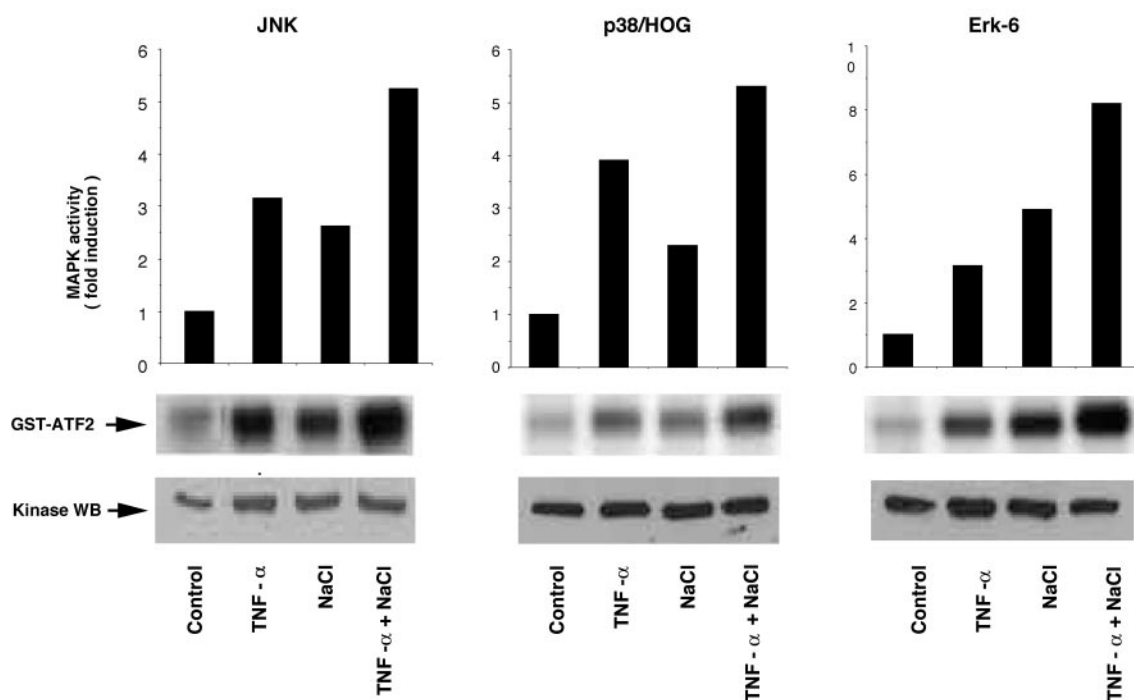


Figure 3 Activation of stress activated protein kinases in HeLa cells. HeLa cells were stimulated with 10 ng/ml TNF- α , 100 mM NaCl or both during 20 min. Then the cells were harvested and the endogenous kinases were immunoprecipitated with the aid of a specific antibody for JNK or p38. In the case of Erk-6, a plasmid expressing the corresponding cDNA expressed as a fusion protein with the HA epitope was transfected. The activity was quantified and presented as fold induction relative to the unstimulated controls. The panel in the middle shows the bands corresponding to phosphorylated substrates and corresponds to a representative experiment. The Western blots in the lower panel show the amount of total kinase present in the assays. Similar results were obtained in three independent experiments

demonstrate that NF- κ B is a protective signal, which is induced later under apoptotic conditions such as osmotic stress plus TNF- α , but is not sufficient to avoid cell death.

We demonstrate in this work that inhibition of p38 activity down-regulates NF- κ B transactivation induced by NaCl, TNF- α or both simultaneously (Figure 4A, C). Therefore we wanted to analyze the role of this kinase in the pathway to apoptosis. The inhibition of p38 by treatment with SB203580 strongly enhances the osmotic stress sensitization to TNF- α -induced cell death (Figure 5A). In order to determine if the increase on NaCl plus TNF- α -induced cell death by blockade of p38 activity is a consequence of a decreased NF- κ B transactivation, we analyzed the effect of Rel-A. The cells were transfected with an expression vector for Rel-A in a dose that clearly induces NF- κ B activity (Figure 5B) or the empty vector as a control, and then stimulated as shown in Figure 5A. From our results, it is evident that although Rel-A totally reverts the increase on sub-diploid cells, due to the inhibitory effect of sulfasalazine on the activation of NF- κ B by NaCl plus TNF- α , it partially reverts the increase on cell death produced by p38 inhibition. In addition, although sulfasalazine does not have a significant inhibitory effect on NF- κ B transactivation under Rel-A constitutively expressed (Figure 5B), this activity is significantly inhibited by SB203580 (Figure 5B). From these data, we conclude that p38 is a protective signal. Although we cannot exclude some additional anti-apoptotic pathway, p38 could be acting through NF- κ B activation, which is not

totally dependent on the levels of available Rel-A, but probably involves some downstream signal related to NF- κ B transactivation.

Discussion

Although most cells are resistant to TNF- α -induced cell death, there are several ways to turn them sensitive, and all of them involve the inhibition of protective NF- κ B target genes.^{11–17} In this work, we report a new model of TNF- α -induced apoptosis by sensitization with osmotic stress. In our model, an early inhibition of NF- κ B activity is involved, but also, cell death is induced in spite of the increase of the late NF- κ B activation by these signals. Although these results could be probably suggesting that NF- κ B has a pro-apoptotic role in this model, our results clearly show that blocking NF- κ B activation increases the sensitivity to apoptosis. Thus, we may conclude that in these specific conditions that lead the cells to apoptosis, a two-phase response is taking place. This transcription factor is induced in a later phase as a defensive response, but its action is not strong enough to prevent cell death, while its early inhibition by NaCl is probably a critical point that leads the cells to apoptosis under TNF- α stimulation.

The loss of cell volume is a fundamental feature of apoptosis.^{37–41} Under hypotonic stress cells can re-adjust their volume after transient osmotic swelling which is accomplished mainly by KCl efflux.⁴⁰ There are enough

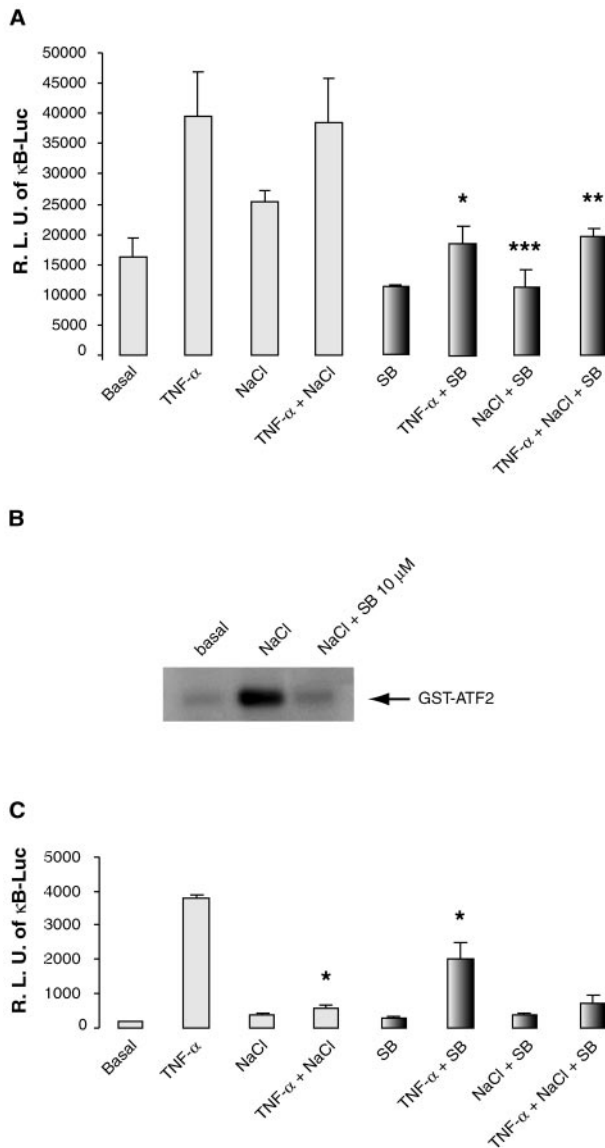


Figure 4 p38 MAP kinase has a role in the NF- κ B activity induced by TNF- α and NaCl. **(A)** HeLa cells were transfected with 0.5 μ g κ B-Luc reporter plasmid, 0.5 μ g of RSV- β -Gal and 2 μ g of empty vector, and stimulated by 12 h with 10 ng/ml TNF- α , 100 mM NaCl and 10 μ M SB203580 as indicated. Relative luminescence units are the average of triplicate \pm S.D. and were normalized with the corresponding β -galactosidase values. **(B)** HeLa cells were treated with 100 mM NaCl in the presence or the absence of 10 μ M SB203580 for 30 min and p38 MAPK induced phosphorylation of GST-ATF2 was assayed as described in Materials and Methods. **(C)** As in (A) but the cells were stimulated by 6 h. For all conditions, at least $P < 0.05$ (ANOVA with Fisher's test). *Respect to TNF- α , **respect to TNF- α +NaCl, ***respect to NaCl

evidences supporting the idea that the efflux of ions associated with cell shrinkage, particularly K^+ , plays a critical role in the cell death program³⁷ and is a necessary event for DNA degradation.⁴¹ In addition, it was previously shown that ions in high concentration, as 150 mM may act as repressors of apoptosis in living cells.⁴¹ In this work, although TNF- α , like other inducers of apoptosis may cause a reduction on cell volume and the loss of ions,⁴⁰ this should be prevented under hyperosmotic condition such as

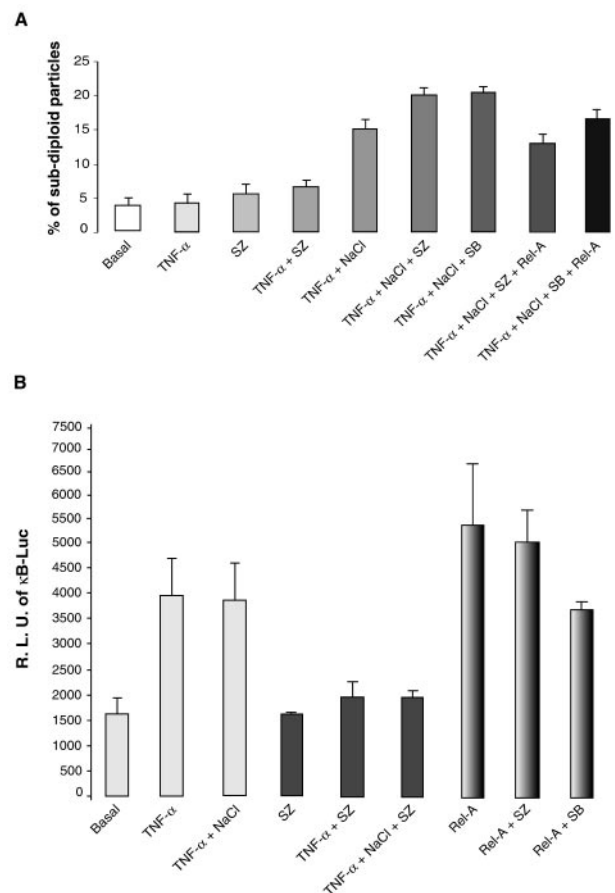


Figure 5 Effect of the p38 MAPK inhibitor SB203580 and sulfasalazine in apoptosis induced by TNF- α in cells sensitized with NaCl. **(A)** HeLa cells were transfected by using lipofectamine plus (GIBCO BRL) with 2 μ g of CMV-RelA or 2 μ g of empty vector and 0.5 μ g RSV- β -Gal, and after 48 h of culture without lipofectamine, they were treated with 100 mM NaCl and 10 ng/ml TNF- α in the presence or not of 10 μ M SB203580 or 200 μ M sulfasalazine (SZ), as indicated. After 12 h of treatment, cells were harvested, stained with propidium iodide, and analysed by flow cytometry. The transfection efficiency (70%) was determined by X-gal staining. The diagram bars corresponds to the average \pm S.D. of triplicates. **(B)** In order to confirm that 200 μ M of sulfasalazine is an effective dose for inhibition of NF- κ B activity, HeLa cells were co-transfected with 0.5 μ g (B-Luc reporter plasmid, 2 μ g of CMV-RelA or empty vector, and 0.5 μ g RSV- β -Gal for normalization. Cells were stimulated with 10 ng/ml TNF- α in the presence or not of 200 μ M of sulfasalazine by 24 h. The diagram bars correspond to the average of triplicate wells \pm S.D. Similar results were obtained in three independent experiments

100 mM of NaCl. In our work, both high 100 mM NaCl and 300 mM sorbitol sensitize the naturally resistant HeLa cells to TNF- α -induced cell death. Taken together all these observations and our own results, we can hypothesize that in the model of hyperosmotic sensitization to TNF- α -induced cell death, the loss of intracellular ions may be not a necessary event.

There are evidence that hyperosmolarity (150 mM NaCl) causes growth arrest of murine kidney cells, which involves the induction of the growth arrest and DNA damage-inducible (GADD) gene products.⁴² In this model, ERK1/2 and p38 are activated, but while p38 stimulates GADD induction, ERK1/2 is inhibitory. It was previously shown that

TNF- α , as well as hyperosmotic stress, induce the activity of several kinases.^{9,10,26,30,32} In this regard, it has been proposed that the balance between the intracellular levels of ceramide and sphingosine-1-phosphate and their regulatory effects on JNK and ERK pathway respectively, could be determining the fate of the cell.³²

According with this idea, a combination of both stimuli, such as TNF- α plus NaCl, could be producing a new balance in the levels of kinase activity probably involved in cell death. In our study all of the kinases induced by each one of the stimuli show a similar pattern of activation being maximal with the combination of the osmotic stress and TNF- α treatment. Although the increased activity of some of these pathways could be playing a role in the control of TNF- α -induced death/survival response, we cannot discard that additional signaling mechanisms, not related to these cascades could be involved in the switch. Previous reports show the participation of p38 on NF- κ B activation,^{30,43} but the precise mechanism by which this occurs is not so clear at present and might be dependent on the cell context. In addition, some evidence supports the requirement of p38 activity on TNF- α -induced NF- κ B transactivation.³⁰ Moreover, according to our data, the requirement of this kinase seems to be critical for NF- κ B activation and independent of the kinetics of each one of the stimulus.

In other way, we showed that inhibition of p38 kinase significantly increases the cell death induced by osmotic stress and TNF- α . Thus, in the new balance of kinases induced by osmotic stress and TNF- α , p38 plays a protective role that is not strong enough to prevent apoptosis. Interestingly, according to our results, the effect of p38 inhibition cannot be totally reverted by the over-expression of Rel-A. This could be compatible with the inhibitory effect of SB203580 on NF- κ B transactivation, even under a constitutive Rel-A over-expression (Figure 5B). These results could probably indicate that the anti-apoptotic role of p38 is mediated by NF- κ B, but in a further step downstream of I κ B proteolysis and not totally dependent of the levels of the active molecules of transcription factor. However, we should not discard that p38 plays its protective role not only by enhancing NF- κ B activity, but also, some additional protective signals, not related to this transcription factor could be triggered.

Besides the activation of caspases, it was previously reported that the cytotoxic effects of TNF- α are due in part to its ability to induce the generation of reactive oxygen intermediates (ROI) and oxidative damage,^{44,45} that would be the principal pathway for death in cells naturally sensitive to TNF- α . Overexpression of Mn-superoxide dismutase (Mn-SOD) has been shown to be a major pathway by which cells can protect themselves against cytotoxic effects of TNF- α .⁴⁵ Mitochondria are the major source of these species, which are increased after loss of the electrochemical gradient across the inner membrane.⁴⁵ According with this knowledge, it has been suggested that in TNF- α resistant cell lines, mitochondrial damage is prevented by an increase in Mn-SOD.⁴⁴ In this regard, perhaps, any signal that impairs the integrity of the inner mitochondrial membrane, or inhibits either, anti-oxidants or the mitochondrial protective signals, like Bcl-2 could,

probably sensitize cells to TNF- α -induced cell death. How hyperosmolarity could be affecting some of these mechanisms remains to be determined.

It is well known that TNF- α activates caspase 8 but there are signals that directly affect the mitochondria which leads to caspase 9 activation without the requirement of caspase 8 activity^{34,35} and Bcl-2 has a regulatory role on preventing this effect.³⁵ According to our results, the levels of Bcl-2 are inhibited by the apoptotic signal elicited by osmotic stress plus TNF- α , while Bax levels are not affected. In addition, caspase 9 seems to be activated by all the different stimulus, and a slow increase is observed under the apoptotic treatment of NaCl plus TNF- α . In these conditions, a diminished ratio of Bcl-2/Bax as occurs under NaCl plus TNF- α treatment, suggest that in this particular model the mitochondrial pathway could play an important role on signaling for cell death, where inhibition of the protective Bcl-2, plus the early activation of NF- κ B could be critical.

Finally, we show that a not apoptotic stimulus as the hyperosmotic stress may sensitize to TNF- α -induced cell death in cells that are naturally resistant to this cytokine, which is the case for most cells in the organism. Since almost all the tissues, in normal and inflammatory response are exposed to both TNF- α and changes in the osmolarity of physiological fluids, these results could be an important contribution to the knowledge of some physiological or pathological conditions involving hypertension, high levels of inflammatory cytokines in plasma and tissue damage.

Materials and methods

Cells and reagents

Human cervix carcinoma HeLa cells were grown in Dulbecco's modified Eagle's medium (DMEM) (Gibco Laboratories, Grand Island, NY, USA) supplemented with 10% fetal calf serum (FCS) (Gibco, Paisley, UK), penicillin (100 units/ml) and streptomycin (100 mg/ml). Cells were maintained at 37°C in a humidified atmosphere with 5% CO₂. Unless stated, reagents were obtained from Sigma Chemical Co (St. Louis, MO, USA), Pharmacia (Uppsala, Sweden) or Calbiochem.

Determination of NF- κ B activity

The NF- κ B activity determination was performed as previously described.¹⁵ Briefly, cells were cultured in 6-well plates at a density of 2.5×10^4 cells/well in DMEM without serum or antibiotics and transiently transfected with a total of 3 μ g of DNA (including 0.5 μ g of κ B-Luc reporter 0.5 μ g of constitutive RSV- β -Gal control vector and 2 μ g of CMV-Rel-A or empty vector) using lipofectamine or lipofectamine plus (Gibco BRL). After 5 h (or 3 h when using lipofectamine plus), transfection medium was replaced by DMEM 10% serum. Cells were stimulated with human TNF- α (Calbiochem) at a concentration of 10 ng/ml, NaCl 100 mM, the NF- κ B inhibitor sulfasalazine (SZ) 200 μ M which inhibits NF- κ B activation^{15,47} and 10 μ M p38 inhibitor SB203580, as indicated, and cultured for 6 h or 12 h prior harvesting. Cellular extracts for luciferase and β -galactosidase assays were prepared, and assays were performed using the appropriate substrates following the manufacturer's protocols (Promega corp.). For experiments of surviving, cells transfected with the expression vector for Rel-A or empty vector, by

using lipofectamine plus were replated after 24 h and stimulated 24 h later.

Kinase assays

Kinase assays were performed as previously reported.²⁴ Briefly, HeLa cells were transfected with 0.5 μ g of pCDNA-HA-Erk-6 by the lipofectamine technique. Total amount of plasmid DNA was adjusted to 3 μ g per well using empty vector. Two days later transfected cells were kept for 2 h in serum-free medium. Then, cells were left untreated or stimulated with TNF- α 10 ng/ml, with NaCl 100 mM or both for 20 min, washed with PBS, and lysed at 4°C in a buffer containing 25 mM HEPES (pH 7.5), 0.3 M NaCl, 1.5 mM MgCl₂, 0.2 mM EDTA, 0.5 mM DTT, 1% Triton X-100, 0.5% sodium vanadate, 1 mM phenylmethylsulphonylfluoride, 20 μ g/ml leupeptin. The epitope tagged Erk-6 (or the endogenous kinases, for JNK and p38) was immunoprecipitated from the cleared lysates by incubation with the specific antibody (Covance and Santa Cruz respectively) for 2 h at 4°C. The immunocomplex was recovered with the aid of Gamma Bind Sepharose beads (Pharmacia), washed three times with PBS containing 1% NP-40 and 2 mM sodium vanadate, once with 100 mM Tris (pH 7.5), 0.5 M LiCl, and once with kinase reaction buffer (12.5 mM MOPS (pH 7.5), 12.5 mM β -glycerophosphate, 7.5 mM MgCl₂, 0.5 mM EGTA, 0.5 mM sodium fluoride, 0.5 mM sodium vanadate). The Erk-6 kinase activity present in the immunocomplex was determined by resuspension in 30 μ l of kinase reaction buffer containing 1 μ Ci of [γ -³²P]ATP per reaction and 20 μ M unlabeled ATP, using 1 μ g of GST-ATF2 fusion protein as a substrate, similarly to the protocol already described for JNK.²⁴ The same substrate was used to measure JNK or p38 kinase activity. After 30 min at 30°C, reactions were terminated by addition of 10 μ l of 5 \times Laemmli buffer. Samples were heated at 95°C for 5 min and analyzed by SDS electrophoresis on 12% polyacrylamide gels. Autoradiography was performed to detect the phosphorylated products.

Cytotoxicity and apoptosis assays

Cytotoxicity was determined as previously described.¹⁵ Briefly, HeLa cells were plated at 1.5×10^3 cells/well in 96-well microtiter plates and stimulated with different doses of h-TNF- α and NaCl 100 mM. After 18 h, cells were fixed with 70% methanol and stained with 0.5% crystal violet. The absorbance corresponding to the stained surviving cells was determined at 570 nm. The per cent of surviving cells was determined respect to the basal conditions (without any treatment).

In some experiments, cells at a density of 3×10^5 cells/well in six-well plates, were transiently transfected with an expression vector for Rel-A or the empty vector as indicated before, and then stimulated with different doses of h-TNF- α and NaCl 100 mM, in the absence or the presence of SZ or the p38 kinase inhibitor SB203580, for 12 h.

Cells with degraded DNA were determined by gating on those displaying sub-diploid (sub-G0/G1) DNA content. These studies were performed by propidium iodide. After the different treatments, 2×10^6 cells were washed twice on PBS, and incubated overnight in 50 μ g/ml propidium iodide solution (0.1% sodium citrate, 0.1% Triton X-100) at 4°C. Samples were measured in a Cytoron Absolute (Ortho Diagnostic Systems, New Brunswick, NJ, USA). In order to determine the number of sub-diploid cells, the data were analyzed with a WinMDI 2.8 software.

Some cultures were stained with acridine orange and ethidium bromide and the morphology of death and surviving cells was observed at fluorescent microscopy. Ethidium bromide only enters into non-viable cells and stains chromatin a dark orange color. Acridine orange penetrates in viable cells and turns green when it intercalates

with DNA. These experiments confirm, as previously shown¹⁵ that the cell death induced by TNF- α plus NaCl is apoptosis.

DNA laddering

The inter-nucleosomal fragmentation of DNA is a characteristic of apoptotic cells.^{11–15} In order to analyze this parameter in our model, each cell suspension (5×10^6 cells) was washed twice with PBS and the cell pellet was resuspended in 800 μ l of lysis buffer (0.5% Triton X-100; 5 mM Tris-HCl pH 7.5; 20 mM EDTA). After 30 min on ice, the lysate was centrifuged (14 000 r.p.m.) for 10 min at 4°C. The aqueous phase was precipitated with sodium acetate 30 mM and 0.7 volumes of isopropanol. The pellet was washed with 80% ethanol, dried and resuspended in 100 μ l TE (10 mM Tris-HCl, 1 mM EDTA, pH 8). Samples were treated with ribonuclease A (0.5 mg/ml) for 1 h at 37°C, then a phenol-chloroform/isoamil alcohol extraction was performed and the aqueous phase was precipitated with 2 volumes of ethanol. The nucleic acid pellet was resuspended in 40 μ l of buffer TE, and electrophoresis was carried out in a 2% agarose gel in TBE buffer. DNA was visualized by ethidium bromide staining. A 123 bp marker (Gibco BRL, Grand Island, NY, USA) was run in parallel lanes as an indicator of molecular weight.

Caspases and Bax analysis

The activity of caspases 8 and 9 and the levels of the pro-apoptotic Bax protein were analyzed by Western blot. Briefly, HeLa cells were stimulated or not with 100 mM NaCl, 10 ng/ml TNF- α or the combination of both by 3, 6, 12 and 16 h. Then, the cultures were washed with PBS, and lysed at 4°C in a buffer containing 25 mM HEPES (pH 7.5), 0.3 M NaCl, 1.5 mM MgCl₂, 0.2 mM EDTA, 0.5 mM DTT, 1% Triton X-100, 0.5% sodium vanadate, 1 mM phenylmethylsulphonylfluoride, 20 μ g/ml leupeptin. Cytosolic extracts were denatured in Laemmli loading buffer at 95°C for 5 min and separated in a 12% SDS-PAGE gel. The proteins were transferred to nitrocellulose membrane for immunoblotting. The membranes were incubated with the specific antibodies: a rabbit polyclonal antibody against human Casapase-8, a rabbit polyclonal antibody against Casapase-9 and a rabbit polyclonal antibody against Bax (all from Santa Cruz). Specific proteins were detected using ECL reagents.

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