

A NEW ANISAKID SPECIES PARASITIZING THE IMPERIAL CORMORANT *PHALACROCORAX ATRICEPS* FROM THE NORTH PATAGONIAN COAST, ARGENTINA

Lucas E. Garbin, Julia I. Diaz, Florencia Cremonte*, and Graciela T. Navone

Centro de Estudios Parasitológicos y de Vectores, Calle 2 #584. 1900 La Plata, Argentina. e-mail: lgarbin@cepave.edu.ar

ABSTRACT: The anisakid species *Contracaecum chubutensis* n. sp. was found parasitizing the imperial cormorant *Phalacrocorax atriceps* at 2 different localities, Bahía Bustamante ($45^{\circ}11'S$, $66^{\circ}30'W$) and Puerto Madryn ($42^{\circ}47'S$, $65^{\circ}02'W$) on the Patagonian coast. Morphometrical analysis and further studies of adult specimens of *C. chubutensis* n. sp. and fourth-stage larvae of *Contracaecum* sp. were done using light and scanning electron microscopy. The possession and disposition of 7 pairs of tail papillae differentiate this new species from *C. osculatum*, *C. radiatum*, *C. plagiaticum*, and *C. mirounga*. Double-sized spicules separate it from *C. variegatum* and *C. pelagicum*. Moreover, the paracloacal papillae disposition and tail shape differentiate this new species from *C. travassosi*. The presence of bifurcated interlabia separate it from *C. ogmorrhini* and *C. margolisi*. Three well-marked lip notches, a sharp tail shape, and the of the transversal paracloacal papillae disposition distinguishes *C. chubutensis* n. sp. from *C. magnipapillatum* and *C. septentrionale*. The species here described can be differentiated from *C. caballeroi* by its bifurcated interlabia, longer spicules, and for possessing more precloacal papillae pairs. It can be separated from *C. rudolphii* by a thinner interlabia at the base, deeper lip notches, a sharper tail, the disposition of transversal paracloacal papillae, and greater distance between both papillae of the first subventral distal papillae pair. In addition, fourth-stage larvae recovered from the same host and localities were identified and described as *Contracaecum* sp. Prevalence of adult parasites was 66.7 %, mean intensity was 4.0, and mean abundance was 7.1.

Species of *Contracaecum* Raillet and Henry, 1912 are associated with fresh and brackish water, as well as marine systems; the definitive hosts are piscivorous birds and mammals (Anderson, 2000). Garbin, Navone, et al. (2007) summarized the records of the genus in Argentina and redescribed *Contracaecum pelagicum* Johnston and Mawson, 1942 in *Spheniscus magellanicus* Foster (Sphenisciformes: Spheniscidae) and *Thalassarche melanophris* Temminck (Procellariiformes: Diomedidae) at Península Valdés. *Contracaecum* species parasitizing cormorants on the southwestern Atlantic coast include *C. travassosi* Gutiérrez, 1943, which was described from *Phalacrocorax atriceps* (=*P. albiventer*) Lesson (Pelecaniformes: Phalacrocoracidae) from Península Valdés, Argentinean seacoast, and *C. caballeroi* Bravo Hollis, 1939 in *P. brasiliensis* (=*P. olivaceus*) from the Uruguayan coast (Gutiérrez, 1943; Lent and Freitas, 1948). Later, Malacalza et al. (1998) found *Contracaecum* sp. in regurgitated pellets of *P. atriceps* (=*P. albiventer*) from Punta León, Chubut Province, Argentina.

The imperial cormorant *P. atriceps* King (both color morphs “*atriceps*” and “*albiventer*,” sensu Rasmussen, 1994) is one of several that breed along the southern coast of South America (Punta et al., 1993; Malacalza and Bertellotti, 2001; Punta et al., 2003). It is a relatively abundant seabird on the Argentine coasts and southern islands (Frere et al., 2005), but is distributed throughout the Southern Hemisphere south of 45° latitude, reaching the South African islands and New Zealand (Behn et al., 1955; del Hoyo et al., 1996). Although information on its diet is scarce, fishes and marine invertebrates are thought to be

the main prey items (Espitalier-Noel et al., 1987; Gosztongyi and Kuba, 1998).

The aim of the present paper is to describe adults of a new species of *Contracaecum* and fourth-stage larvae of *Contracaecum* sp. parasitizing the imperial cormorant *P. atriceps* from 2 different breeding sites in Chubut Province, Argentina. Morphological details observed using scanning electron microscopy (SEM). In addition, data regarding prevalence, mean intensity, and abundance of infection of adults are provided.

MATERIAL AND METHODS

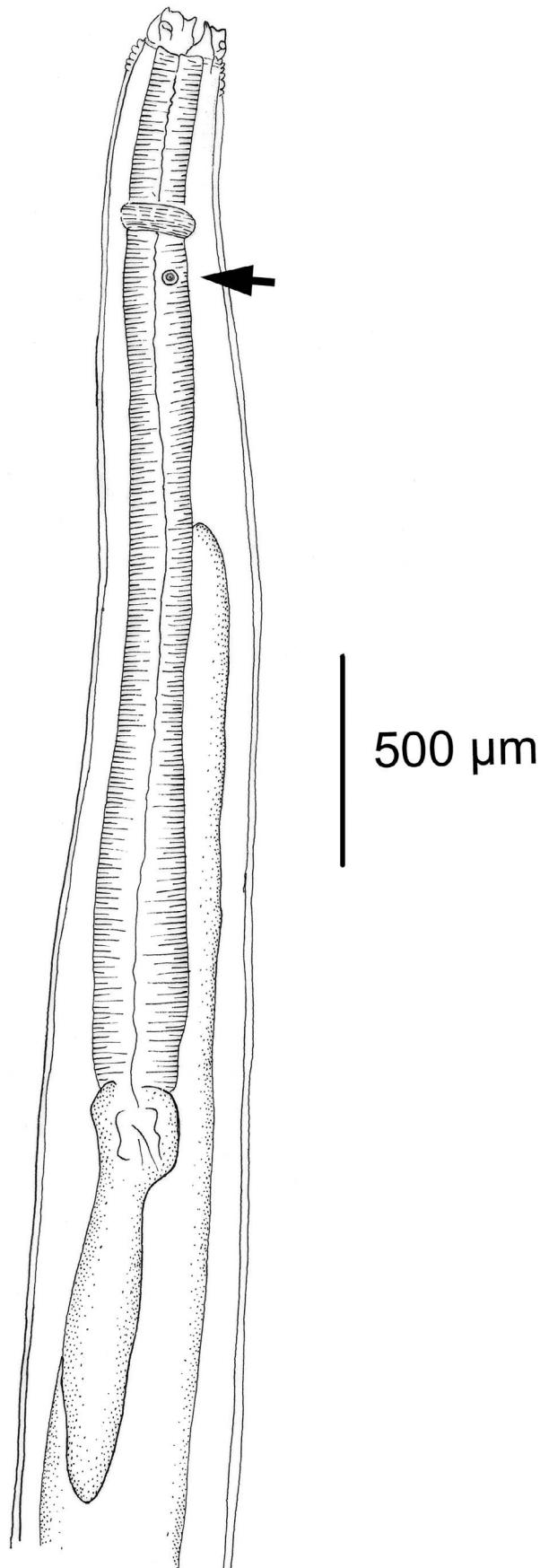
At irregular intervals from 1999 to 2006, 12 dead imperial cormorants were collected from nesting sites on Bahía Bustamante ($n = 10$; $45^{\circ}11'S$, $66^{\circ}30'W$) and Puerto Madryn ($n = 2$; $42^{\circ}47'S$, $65^{\circ}02'W$), Chubut Province, Argentina. All birds, which were in good condition, were dissected and the digestive tracts fixed in 10% formalin. In addition, regurgitated pellets from imperial cormorants from the first locality were collected and fixed in 10% formalin. In the laboratory, viscera and pellets were examined using stereomicroscopy; nematodes were removed and stored in cold 70% ethanol. Twenty adult nematodes (10 males and 10 females) and 10 fourth-stage larvae from Bahía Bustamante, and 8 adults (5 males and 3 females) and 10 fourth-stage larvae from Puerto Madryn, were cleared in lactophenol and examined by light microscopy. Drawings were made with the aid of a drawing tube.

Some specimens were dried by the critical point method, then observed and photographed using a SEM (JSV 6063 LV; Jeol, Akishima City, Tokyo, Japan). Unless otherwise stated, measurements are given in millimeters; those of paratypes (including holotype and allotype) are expressed by the mean followed by the range in parentheses. Spatial location of papillae was described according to Fagerholm (1990), except for distal papillae, which were identified as subventral and laterodorsal.

Prevalence (P), mean intensity (MI), and mean abundance (A) followed Bush et al. (1997) and were calculated separately for adult and larval infections from Bahía Bustamante. Holotype, allotype, and paratypes specimens were deposited in the Helminthological Collection of Museo de La Plata (CHMLP).

Received 27 June 2007; revised 11 October 2007; accepted 11 December 2007.

* Centro Nacional Patagónico, Boulevard Brown 2825, U9120ACF Puerto Madryn, Argentina.



***Contracaecum chubutensis* n. sp.**

(Figs. 1–10; Table I)

Description

General morphology (20 adult specimens from *P. atriceps* from Bahía Bustamante): Entire body transversally striated (Figs. 2, 3, 6, 7). Conspicuous cephalic collar, with deep concentric anteriorly directed free edges and narrow V-shaped lateral region without striations (Figs. 3–6). Lips with 3 apical notches, central notch deeper and larger than lateral notches (Figs. 3–6); with 2 auricles, each with processes placed obliquely to body's longitudinal axis, with a sensory pit at distal end at level of apical notches (Figs. 3–6, see arrows). Large double lip papillae, 2 on the dorsal lip and 1 on each ventrolateral lip (Figs. 3, 4, 6). Three triangle-shaped bifurcated interlabia shorter than lips (Figs. 2–6). Inconspicuous button-shaped deirids (Figs. 1, 2, see arrows).

Male (holotype): Body length 25.92. Maximum body width 0.83. Distance from anterior end to nerve ring and deirids 0.58 and 0.66, respectively. Esophagus length 3.62; intestinal caecum length 2.41; ventriculus length 0.30; ventricular appendix length 0.80 (Fig. 1). Spicules of equal length reaching almost half of body. Spicules length 10.82 (Fig. 9). Tail length 0.19 (Fig. 10). Caudal extremity conical, bearing 39 pairs of precloacal papillae. Pts zone including 3 pairs of precloacal papillae (Fig. 7). Seven pairs of postcloacal papillae; 2 large paracloacal pairs situated aside cloaca, in transversal position to body axis, 2 subventral pairs, 2 sublateral pairs, and 1 pair of phasmids among latter ones (Figs. 7, 10). Distal spicule tip extended and pointed length of free distal end longer than spicule width (0.057 vs. 0.024). Spicule wings slope distally toward shaft and inserted at different points (Figs. 8, 10) (male paratypes, see Table I).

Female (allotype): Body length 29.20. Maximum body width 0.99. Distance from anterior end to nerve ring and deirids 0.54 and 0.63 respectively. Esophagus length 3.12; intestinal caecum length 2.08; ventriculus length 0.25; ventricular appendix length 0.66. Vulva in anterior half of body. Distance from anterior end to vulva 8.72. Tail length 0.40. Embryonated egg ($n = 10$) diameter 0.069 (0.065–0.074) (female paratypes, see Table I continued).

Taxonomic summary

Type host: *Phalacrocorax atriceps* King (Aves: Phalacrocoracidae).

Type locality: Bahía Bustamante (45°11'S, 66°30'W), Chubut Province, Argentina.

Other localities: Puerto Madryn (42°47'S, 65°02'W), Chubut Province, Argentina.

Infection site: Stomach.

Prevalence: 60% (6/10) from Bahía Bustamante.

Mean intensity: 4.0.

Mean abundance: 2.67.

Specimens deposited: Holotype (male) (5748 CHMLP), allotype (female) (5749 CHMLP), and 18 paratypes (5750 CHMLP); Helminthological Collection of Museo de La Plata, La Plata, Buenos Aires, Argentina.

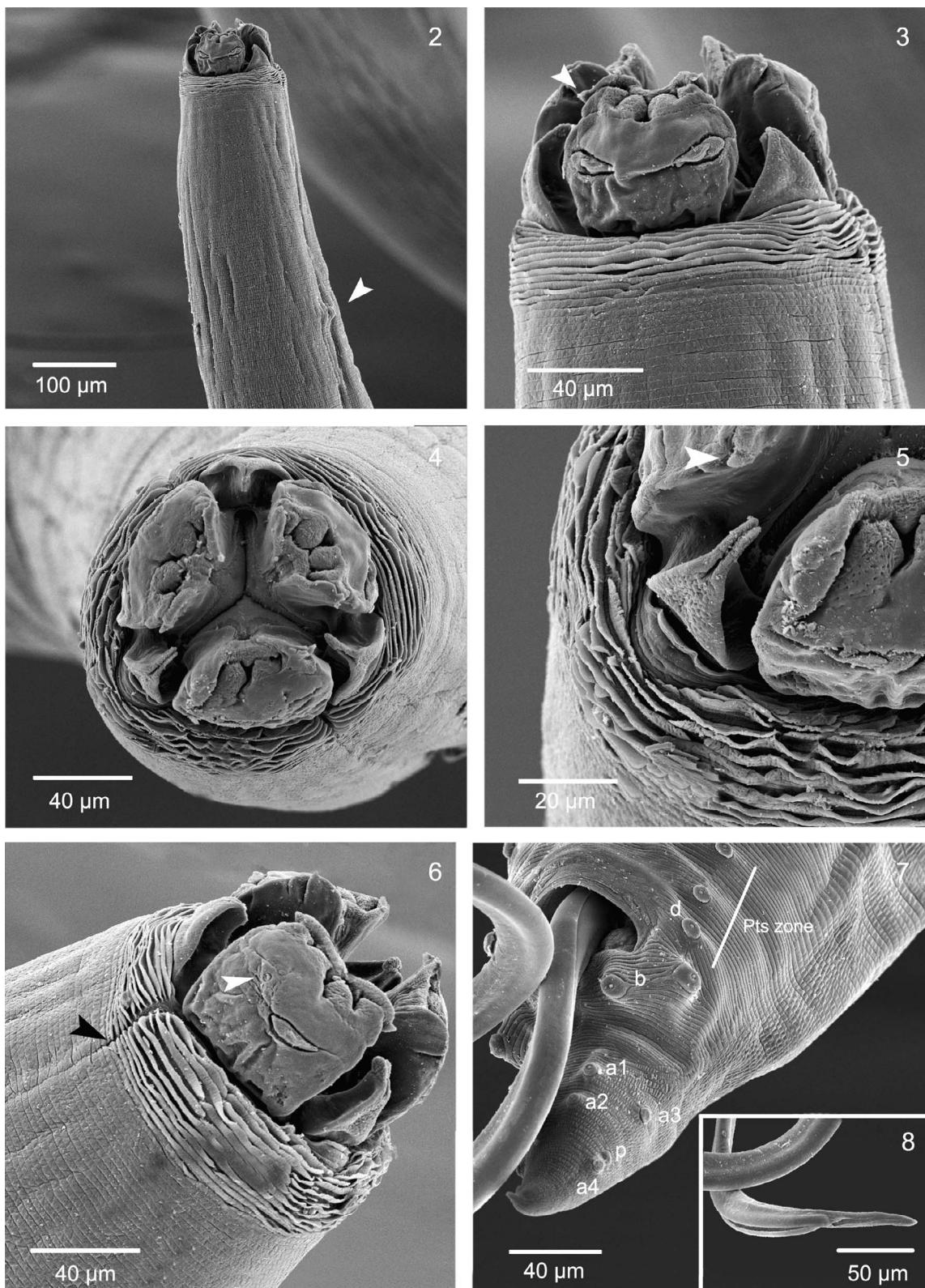
Remarks

Contracaecum chubutensis n. sp. can be differentiated from other *Contracaecum* species parasitizing sea mammals that overlap their geographic distribution in the southern Atlantic Ocean with that of Phalacrocoracidae, such as *C. osculatum* (Rudolphi, 1802), *C. radiatum* (Linstow, 1907), *C. ogmorrhini* s. s. Johnston and Mawson, 1941, *C. mirounga* Nikolskii, 1974, and *C. margolisi* Mattiucci et al., 2003, by its bifid interlabia (Baylis, 1936; Fagerholm, 1988; Fagerholm et al., 1996; Mattiucci et al., 2003).

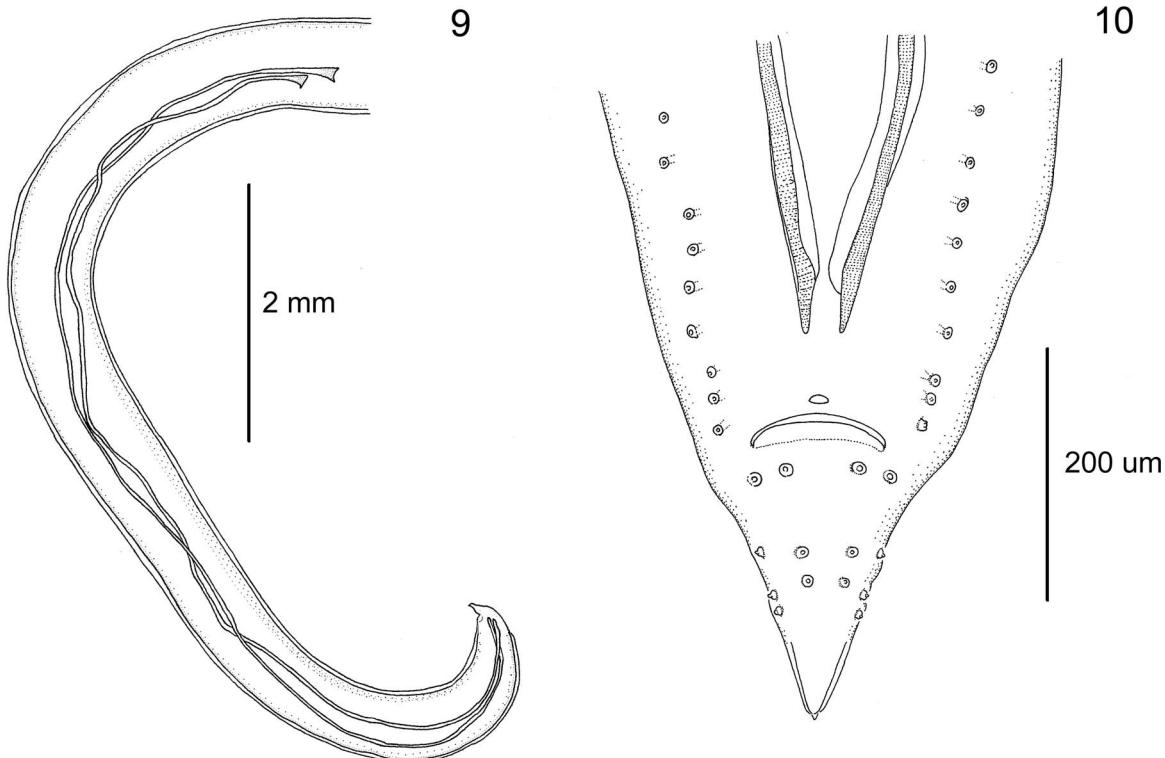
From the *Contracaecum* species that parasitize seabirds, *C. variegatum* Rudolphi, 1809 from *Gavia stellata* (Pontoppidan) (Gaviidae), *C. pelagicum* from *Spheniscus magellanicus* Foster (Spheniscidae), *C.*

←

FIGURE 1. *Contracaecum chubutensis* n. sp. from *P. atriceps* from the Chubut province coast. Anterior end, lateroventral view, nerve ring, esophagus, intestinal caecum, ventriculus, ventricular appendix, and deirid (arrow).



FIGURES 2–8. *Contracaecum chubutensis* n. sp. from *P. atriceps* from the Chubut province coast. (2) Anterior end details, deirid (arrow); (3) anterior end, laterodorsal view, cephalic collar, dorsal lip, interlabia, cephalic lip papillae, lip auricle tip (arrow); (4) anterior end, apical view, lips, bifid interlabia, cephalic lip papillae, auricles, and lip notches; (5) interlabium details, lip notches, lip auricle tip (arrow), and excretory pore; (6) anterior end, laterodorsal view, ventrolateral lip, double cephalic lip papilla, amphid (white arrow) cephalic collar, and V-shaped lateral region without striations (black arrow); (7) posterior male end, postcloacal papillae distribution: distal subventral papillae [a1–a2], distal sublateral papillae [a3–a4], post paracloacal papilla pair [b], proximal precloacal papillae [d], phasmid [p]; (8) distal spicule end, lateroventral view.



FIGURES 9, 10. *Contracaecum chubutensis* n. sp. from *P. atriceps* from the Chubut province coast. (9) Posterior male extremity, whole spicules; (10) Posterior male extremity, distal spicule end, cloaca, precloacal papillae, postcloacal papillae.

eudyptulae Johnston and Mawson, 1944 from *Eudyptula minor* (Forster) (Spheniscidae), and *C. plagiaticum* Lent and Freitas, 1948 from *Ncticorax nycticorax naevius* (Bodd.) (Ardeidae) are similar to this new species in having bifurcated interlabia (Johnston and Mawson, 1944; Fagerholm et al., 1996). However, *C. variegatum* possesses a longer interlabium, thinner at base with a more conspicuous furrow, more than a double number of precloacal papillae, shorter spicules (4.40–4.86 vs. 5.88–12.60), and, therefore, larger body-to-spicule length ratio (4.00–6.50 vs. 2.18–3.13) (Fagerholm et al., 1996). *Contracaecum pelagicum* can be differentiated from *C. chubutensis* n. sp. mainly for their shallower lip notches, rounded lips, prominent auricle and lip papillae, shorter spicules (3.07–5.07 vs. 5.88–12.60), and greater body-to-spicule length ratio (3.40–5.10 vs. 2.18–3.13) (Garbin, Navone, et al., 2007). *Contracaecum eudyptulae* has shorter intestinal caecum, fewer precloacal and postcloacal papillae pairs (12 and 6, respectively), and an oblique paracloacal papillae disposition (Johnston and Mawson, 1944). *Contracaecum plagiaticum* has shorter spicules (2.32–3.49 vs. 5.88–12.60), greater body-to-spicule length ratio (4.80–5.40 vs. 2.18–3.13), and 8 postcloacal papillae pairs (1 more subventral papillae pair) (Lent and Freitas, 1948). *Contracaecum magnipapillatum* (= *C. magnicollare* Johnston and Mawson, 1941) from *Anous minutus* Chapin (Laridae) and *C. septentrionale* Kreis, 1955 from *P. aristotelis* (Linnaeus) (Phalacrocoracidae) share some morphological features with the new species such as the number and spatial distribution pattern of postcloacal papillae. However, they differ from each other for the absence of bifurcated interlabia (Johnston and Mawson, 1941; Gutierrez, 1943; Kreis, 1955; Hartwich, 1964; Fagerholm et al., 1996).

Among *Contracaecum* species parasitizing species of Phalacrocoracidae from the southeastern South American coast, *C. travassosi* from *P. atriceps* (= *P. albiventer*) can be differentiated from *C. chubutensis* n. sp. by its blunt tail shape with a constriction at the proximal subventral papillae level, paracloacal papillae disposition oblique 45° to the body longitudinal axis, and shallow lip notches (Gutierrez, 1943). *Contracaecum caballeroi*, from the Neotropic cormorant *P. brasiliensis*, can be distinguished from the new species by its lack of bifurcated interlabia, much shorter spicules (0.90–1.09 vs. 5.88–12.60), and larger

body-to-spicule length ratio (24.74–26.98 vs. 2.18–3.13) (Lent and Freitas, 1948). *Contracaecum rudolphii* Hartwich, 1964 s.l. from *P. carbo* is the most similar species to *C. chubutensis* n. sp. However, it can be distinguished from the new species by its rounded lips with more exaggerated papillae, blunt tail shape with a constriction at the proximal subventral papillae level, oblique and closer paracloacal papillae disposition, first distal subventral papillae pair more separate each other (42–50 μm vs. 37 μm), and shorter and blunter free distal spicule end (≈30 μm vs. 58 μm) (Hartwich, 1964; Barus et al., 2000; Abollo et al., 2001; Amato et al., 2006).

Contracaecum sp. (Figs. 11–14; Table II)

Description

Fourth-stage larvae (L4) (10 specimens from Bahía Bustamante): Cuticle transversally striated, specially marked at anterior extremity and forming a developed cephalic collar (Figs. 12, 13). Oral opening with 3 well-developed lips with conspicuous winglike processes on their anterior corners (Figs. 12, 13). Dorsal lip bearing 2 internal papillae and 2 small cephalic papillae (Fig. 12). Each ventrolateral lip possesses a single internal papilla (Fig. 12). Excretory pore opening right beneath the ventrolateral lips (Fig. 12, see arrow). Deirids nonconspicuous, located just posterior to the nerve ring (Fig. 11, see arrow). Ventriculus small and subspherical. Intestinal caecum dorsal to esophagus, longer than ventricular appendix (Fig. 11). Conical striated tail (Fig. 14).

Taxonomic summary

Infection sites: Esophagus and stomach.

Prevalence: 80% (8/10).

Mean intensity: 8.5.

Abundance: 7.1.

Specimens deposited: Paratypes (5751 CHMLP); Helminthological Collection of Museo de La Plata, La Plata, Buenos Aires, Argentina.

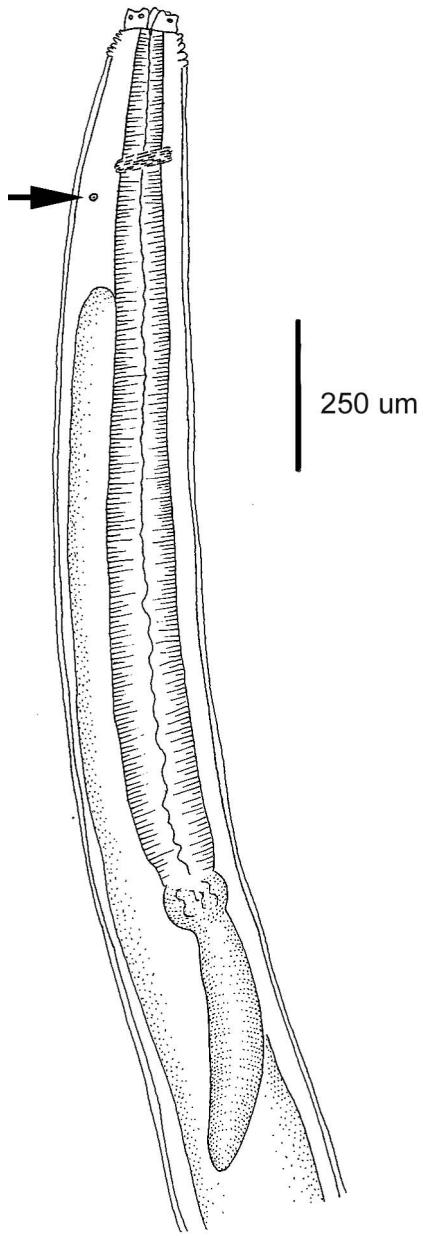


FIGURE 11. *Contracaecum* sp. from *P. atriceps* from the Chubut province coast. Fourth-stage larvae (L4): anterior end, esophagus, intestinal caecum, ventriculus, ventricular appendix, and deirid (arrow).

Remarks

The presence of an intestinal caecum and ventricular appendix and the position of the excretory pore right beneath the ventrolateral lips allow the inclusion of these larvae as a species of *Contracaecum*. The lips are well developed, but interlabia are lacking, which usually characterizes fourth-stage larvae (Berland, 1989; Fagerholm, 1990; Fagerholm et al., 1996). To date, there are few descriptions of *Contracaecum* spp. fourth-stage larvae in birds. *Contracaecum magnipapillatum* fourth-stage larvae found by Fagerholm et al. (1996) in *Anous minutus* possess large winglike expanded auricles. Fourth-stage larvae of *C. rudolphi* from *P. carbo* (Linnaeus) have an interlabium, a feature not usually present in this larval stage (Barus et al., 2000); perhaps these specimens might belong to immature adult specimens (Garbin, Navone, et al., 2007). *Contracaecum osculatum* fourth-stage larvae do not have auricle expansions, which differentiate them from larval specimens de-

scribed in this work. The present fourth-stage larvae seem to be similar to those described by Garbin, Navone, et al. (2007) from *S. magellanicus*. They agree with almost all of the morphometrical features except the ventricular appendix length and ventriculus plus ventricular appendix length, greater in the former, and the excretory gland distance to the anterior end, greater in the latter. According to the similarities among larvae of this genus, it is impossible to assign present specimens to a given species without experimental infections or molecular genetic analysis.

DISCUSSION

Differences mentioned above allow for the separation of *C. chubutensis* n. sp. from *C. rudolphi*, *C. travassosi*, and *C. caballeroi*, which are the closest related species. Thus, differences such as the postcloacal spatial papillae distribution, tail shape, mouth, and spicules are strong enough to discriminate these species from *C. chubutensis* n. sp.

There are several studies dealing with the diet of *P. atriceps* diet along the Chubut coast. *Engraulis anchoita* and nototheniid fishes constitute some of the most frequent *P. atriceps*' prey items, whereas *Odontesthes* spp., *Merluccius hubbsi*, and *Eleginops maclovinus* are consumed in smaller quantities (Punta et al., 1993; Malacalza et al., 1994; Punta et al., 2003; Gosztonyi and Kuba, 1998). Some of these fish species were analyzed for parasites in the same locality, e.g., *M. hubbsi*, *E. anchoita*, and *Odontesthes* spp., and *Contracaecum* third-stage larvae were found (Sardella and Timi, 1996; Timi et al., 2001; Sardella and Timi, 2004; Garbin, Navone et al., 2007; C. Carballo, unpubl. obs.). However, studies based on collections and inspection of these nototheniid prey items at the foraging area and prospection for the endoparasite could confirm the *C. chubutensis* n. sp. transmission.

Molecular genetic studies of nematodes, such as those carried out by Nascetti et al. (1993) and Mattiucci et al. (1997) on the *Anisakis simplex* complex, and other authors on *Contracaecum* spp. (Mattiucci et al., 2002, 2003; Li et al., 2005; Mattiucci et al., 2007) would help to establish the specific relationship between third-stage larvae of *Contracaecum* species found in prey of *P. atriceps*, and those of fourth-stage larval *Contracaecum* and adults found in the digestive tract of this bird and other host species studied in the same area, i.e., *S. magellanicus*, *P. brasiliensis*, and *T. melanophris*.

Mean intensity and abundance of infection by *Contracaecum* spp. from different *Phalacrocorax* spp. in the region are higher in comparison to those from the present work (Torres et al., 2000; Amato et al., 2006). Low mean intensity and abundance values of infection could be related to both the regurgitation habit of this bird and to the low mean intensity and prevalence of *Contracaecum* larvae in the prey items consumed by *Phalacrocorax atriceps*. In this sense Malacalza et al. (1994) suggested that the regurgitation of *P. atriceps* (= *P. albiventer*) would be a depuration mechanism since high nematode intensities were found in regurgitated pellets. In addition, Garbin, Diaz, et al. (2007) observed low mean intensity (2.81) and abundance (1.47) of *Contracaecum* spp. third-stage larvae parasitizing *E. anchoita* from the Chubut province sea coast.

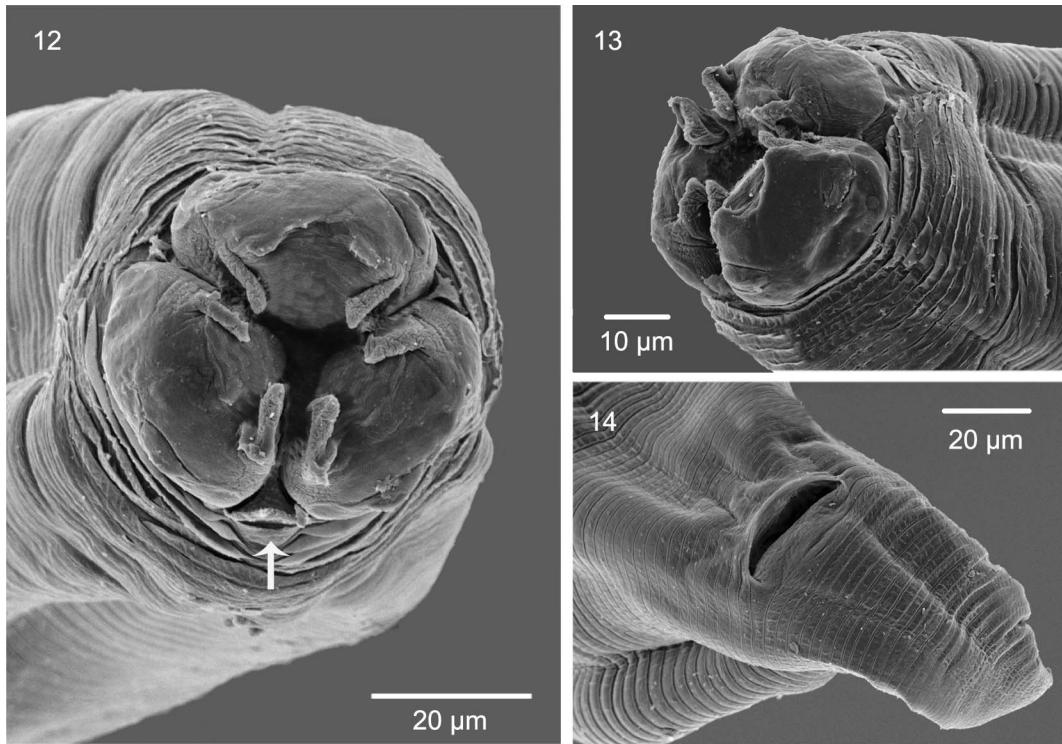
Future molecular studies could be carried out to confirm the validity of *C. chubutensis* n. sp., considering that most of the nominal species of anisakid nematodes now comprise several sibling species that cannot be recognized by only morphological analysis (Mattiucci et al., 2002; Li et al., 2005; D'Amelio et al., 2007; Mattiucci et al., 2008).

TABLE I. Morphometrical data of *Contracaecum chubutensis* n. sp. adults from *P. atriceps* at 2 different Argentinean coastal sites.

	<i>C. rudolphi</i> Hartwich, 1964	<i>C. caballeroi</i> Bravo Hollis, 1939	<i>C. travassosi</i> Gutiérrez, 1943	<i>C. chubutensis</i> n. sp.	<i>C. chubutensis</i> n. sp.
References	Hartwich (1964), Barus (2000), Abollo (2001)	Gutiérrez (1948)	Gutiérrez (1943)	Present paper	Present paper
Type host	<i>P. carbo</i> , <i>Phalacrocorax</i> spp.	<i>Anhinga anhinga</i>	<i>P. atriceps</i>	<i>P. atriceps</i>	<i>P. atriceps</i>
Other hosts	<i>Phalacrocoraciidae</i> , Charadriiformes, Ciconiiformes	<i>P. brasiliensis</i>	—	—	—
Localities	Cosmopolitan	México, Uruguay	Chubut Province, Argentina	Bahía Bustamante, Chubut Province, Argentina	Puerto Madryn, Chubut Province, Argentina
<i>Males</i> (n)					
Body length	39	3	no data	10	5
Maximum body width	12.10–33.90	24.29–26.97	16.10–25.40	14.32–38.58	11.60–28.51
Nerve ring (dae)	0.24–0.95	0.53–0.64	0.70–1.10	0.43–0.98	0.65 (0.58–0.74)
Deirids (dae)	—	0.43–0.45	0.45–0.64	0.52 (0.36–0.60)	0.47 (0.33–0.55)
Esophagus length	—	0.44–0.48	—	0.64 (0.46–0.84)	0.57 (0.43–0.66)
Intestinal caecum length	2.03–4.26	3.18–3.48	2.80–4.10	3.39 (2.32–4.50)	2.83 (2.08–3.93)
Ventriculus length	1.53–3.68	2.71–3.01	1.90–3.20	2.25 (1.50–2.76)	2.17 (1.58–3.04)
Ventricular appendix length	—	0.10–0.10	—	0.23 (0.13–0.30)	0.18 (0.15–0.21)
Spicule length	—	0.51–0.61	0.74–1.30	0.67 (0.46–0.80)	0.64 (0.48–0.75)
Tail length	4.05–9.98	0.90–1.09	7.70–11.10	9.95 (6.40–12.60)	8.43 (5.34–10.35)
PrPP	0.14–0.24	0.13–0.15	0.19–0.26	0.20 (0.17–0.26)	0.20 (0.15–0.25)
BL/MBW	27–43	40	26–30	35–43	35–36
BL/EL	52.3 (29.4–98.1)	42.14–45.83†	23.00–23.09†	33.19 (27.68–39.32)	29.76 (19.97–46.66)
BL/TI	8.0 (5.25–10.8)	7.64–7.75†	5.75–6.19†	7.33 (6.15–9.27)	6.74 (4.87–7.54)
EL/ICL	131.8 (74.1–197.2)	179.80–186.84†	84.74–97.69†	124 (83.74–214.32)	94.5 (72.34–113.13)
EL/NAL	1.3 (1.11–1.54)	1.16–1.17†	1.28–1.49†	1.52 (1.44–1.63)	1.30 (1.26–1.36)
BL/SL	3.25 (1.82–4.25)	5.70–6.23†	3.15–3.78†	4.90 (3.49–6.02)	4.78 (4.20–5.20)
<i>Females</i> (n)					
Body length	36	—	No data	10	2
Maximum body width	10.10–57.60	—	22.7–31.5	29.60 (21.98–35.33)	17.97 (15.30–20.64)
Nerve ring (dae)	0.29–1.51	—	1.10–1.50	0.90 (0.61–1.27)	0.75 (0.61–0.89)
Deirids (dae)	—	—	0.59–0.73	0.56 (0.48–0.62)	0.63 (0.52–0.74)
Esophagus length	—	—	—	0.69 (0.58–0.80)	0.66 (0.55–0.76)
Intestinal caecum length	1.62–5.48	1.74–4.80	3.50–4.80	3.05 (1.19–4.28)	2.74 (2.36–3.12)
Ventriculus length	1.28–4.12	—	2.70–4.20	1.91 (1.08–2.93)	1.88 (1.60–2.16)
Ventricular appendix length	—	—	0.89–1.60	0.24 (0.16–0.26)	0.18 (0.16–0.21)
Vulva (dae)	5.12–17.7	—	—	0.74 (0.66–0.94)	0.63 (0.58–0.68)
Tail length	0.19–0.63	—	18.60–21.00	9.57 (8.32–11.56)	7.63 (6.54–8.72)
Embrionated egg	0.059–0.073	—	0.43–0.54	0.41 (0.30–0.65)	0.36 (0.30–0.41)
			0.068	0.070 (0.06–0.07)	0.068

* dae = distance from anterior end; PrPP = Precloacal papillae pairs; BL/MBW = Body length/Maximum body width ratio; BL/EL = Body length/Esophagus length ratio; EL/ICL = Esophagus length/Intestinal caecum length ratio; EL/NAL = Esophagus length/Ventricular appendix length ratio; BL/SL = Body length/Tail length ratio;

† = ratios calculate with maximum and minimum values.



FIGURES 12–14. *Contracaecum* sp. from *P. atriceps* from the Chubut province coast. Fourth-stage larvae (L4): (12) anterior end, apical view, lips with wing-like processes, internal and cephalic lip papillae, excretory pore (arrow), cephalic collar; (13) anterior end, laterodorsal view, lips with wing-like processes, internal and cephalic dorsal lip papillae, cephalic collar; (14) posterior extremity, anus, and striations.

ACKNOWLEDGMENTS

The authors wish to thank Dr. Flavio Quintana and Dr. Alexandra Sapoznikow who collected some of the birds and nematodes, and to the staff of the Servicio de Microscopía Electrónica de Barrido of the Museo de La Plata. The present work was funded by the Agencia Nacional de Promoción Científica y Tecnológica (ANPCyT) (PICT 11902), the Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) (PIP 2714/5653), and the Universidad Nacional de La Plata (N 393). All authors are members of the CONICET.

LITERATURE CITED

- ABOLLO, E., C. GESTAL, AND S. PASCUAL. 2001. Anisakid infection in the European shag *Phalacrocorax aristotelis aristotelis*. Journal of Helminthology **75**: 209–214.
AMATO, J. F. R., C. M. MONTEIRO, AND S. B. AMATO. 2006. *Contracaecum rudolphii* Hartwich (Nematoda, Anisakidae) from the Neotropical Cormorant, *Phalacrocorax brasilianus* (Gmelin) (Aves, Phalacrocoracidae) in Southern Brazil. Revista Brasileira de Zoologia **23**: 1284–1289.

TABLE II. Morphometrical data of *Contracaecum* sp. fourth-stage larvae in *P. atriceps* at two different Argentine coastal sites. Means are followed by the standard deviation.

	<i>Contracaecum</i> sp.	<i>Contracaecum</i> sp.
References	Present	Present
Type host	<i>P. atriceps</i>	<i>P. atriceps</i>
Locality	Bahía Bustamente, Chubut Province, Argentina	Puerto Madryn, Chubut Province, Argentina
Individuals (n)	10	10
Body length	5.67 ± 2.05 (3.22–7.88)	7.23 ± 1.28 (5.72–9.14)
Maximum body width	0.24 ± 0.08 (0.13–0.36)	0.25 ± 0.04 (0.18–0.32)
Anterior end to nerve ring	0.24 ± 0.05 (0.15–0.29)	0.28 ± 0.05 (0.21–0.36)
Deirids (dae)	0.30 ± 0.11 (0.17–0.40)	0.39 ± 0.11 (0.24–0.56)
Esophagus length	1.14 ± 0.44 (0.51–1.72)	1.17 ± 0.16 (0.93–1.43)
Intestinal caecum length	0.74 ± 0.35 (0.26–1.24)	0.69 ± 0.12 (0.56–0.90)
Ventricular appendix length	0.48 ± 0.11 (0.36–0.60)	0.45 ± 0.14 (0.40–0.52)
Ventriculus + ventricular appendix length	0.50 ± 0.14 (0.28–0.70)	0.55 ± 0.09 (0.45–0.70)
Excretory gland (dpe)	2.31 ± 0.22 (2.16–2.56)	2.57 ± 0.28 (2.35–2.98)

* dae = distance from anterior end; dpe = distance from posterior end.

- ANDERSON, R. C. 2000. Nematode parasites of vertebrates. Their development and transmission, 2nd ed. CABI Publishing, New York, New York, 650 p.
- BARUS, V., K. NAGASAWA, F. TENORA, AND M. PROKES. 2000. The head end morphology of *Contracaecum rudolphii* with remarks on *C. himeu* and *C. umiu* (Nematoda, Anisakidae). Acta Universitat Agriculturae et Silviculturae Mendelianae Brunensis **58**: 69–76.
- BAYLIS, H. A. 1936. On the ascarids parasitic in seals, with special reference to the genus *Contracaecum*. Parasitology **29**: 121–130.
- BEHN F., J. D. GOODALL, A. W. JOHNSON, AND R. A. PHILIPPI. 1955. The geographic distribution of the blue-eyed shags, *Phalacrocorax albiventer* and *Phalacrocorax atriceps*. The Auk **72**: 6–13.
- BERLAND, B. 1989. Identification of larval nematodes from fish. In H. Moller (ed.). Nematode problems in north Atlantic fish. Report from a workshop in Kiel. 3–4 April 1989, International Council for the Exploration of the Sea, Kiel, Germany, p. 16–22.
- BUSH, A. O., K. D. LAFFERTY, J. M. LOTZ, AND A. W. SHOSTAK. 1997. Parasitology meets ecology on its own term: Margolis *et al.* revisited. Journal of Parasitology **83**: 575–583.
- D'AMELIO, S., K. D. MATHIOPoulos, C. P. SANTOS, O. N. PUGACHEV, S. C. WEBB, M. PICANÇO, AND L. PAGGI. 2000. Genetic markers in ribosomal DNA for the identification of members of the genus *Anisakis* (Nematoda: Ascaridoidea) defined by polymerase chain reaction-based restriction fragment length polymorphism. Journal of Parasitology **30**: 223–226.
- DEL HOYO J., A. ELLIOTT, AND J. SARGATAL. 1996. Handbook of the birds of the world. Vol. 3. Lynx, Barcelona, Spain, 821 p.
- ESPITALIER-NOEL, G., N. J. ADAMS, AND N. T. KLAGES. 1987. Diet of the Imperial Cormorant *Phalacrocorax atriceps* at sub-Antarctic Marion Island. Emu **88**: 43–46.
- FAGERHOLM, H. P. 1988. Patterns of caudal papillae in *Contracaecum osculatum* (Nematoda) and some related species from different regions of the world. International Journal for Parasitology **18**: 1039–1051.
- . 1990. Systematic position and delimitation of ascaroid nematode parasites of the genus *Contracaecum* with a note on the superfamily Ascaridoidea. Ph.D. Thesis. National Veterinary Institute, Helsinki, Finland, 27 p.
- , R. M. OVERSTREET, AND I. HUMPHREY-SMITH. 1996. *Contracaecum magnipapillatum* (Nematoda, Ascaridoidea): Resurrection and pathogenic effects of a common parasite from the proventriculus of *Anous minutus* from the Great Barrier Reef, with a note on *C. variegatum*. Helminthologia **33**: 195–207.
- FRERE, E., F. QUINTANA, AND P. GANDINI. 2005. Cormoranes de la costa patagónica: estado poblacional, ecología y conservación. El Hornero **20**: 35–52.
- GARBIN, L. E., J. I. DIAZ, C. CARBALLO, F. CREMONTE, AND G. T. NAVONE. 2007. Recruitment and ecological parameters of three Anisakid larvae (Nematoda: Anisakidae) parasites of *Engraulis anchoita* (Pisces: Engraulidae) from southwest Atlantic Ocean. ISFP VII Abstracts. Parassitologia **49**: 219.
- , G. T. NAVONE, J. I. DIAZ, AND F. CREMONTE. 2007. Further study of *Contracaecum pelagicum* (Nematoda: Anisakidae) in *Spheniscus magellanicus* (Aves: Spheniscidae) from Argentinean coasts. Journal of Parasitology **93**: 143–150.
- GOSZTONYI, A. E., AND L. KUBA. 1998. Fishes in the diet of the Imperial Cormorant *Phalacrocorax atriceps* at Punta Lobería (Chubut, Argentina). Marine Ornithology **26**: 59–61.
- GUTIÉRREZ, R. O. 1943. Sobre la morfología de una nueva especie de *Contracaecum* (Nematoda: Ascaridoidea). Revista Brasileira de Biología **3**: 159–172.
- HARTWICH, G. 1964. Die Typen Parasitischer Nematoden in der Helminthen-Sammlung des Zoologischen Museums in Berlin. I. Ascaridoidea. Mitteilungen aus dem Zoologischen Museum. Berlin **40**: 1–53.
- JOHNSTON, T. H., AND P. M. MAWSON. 1941. Some parasitic nematodes in the collection of the Australian Museum. Records of the Australian Museum **21**: 9–16.
- , AND —. 1944. Remarks on some parasitic nematodes from Australia and New Zealand. Transactions of the Royal Society of South Australia **68**: 60–66.
- KREIS, H. A. 1955. *Contracaecum septentrionale*, ein neuer parasit aus dem Kormoran; sein Lebenslauf, sowie Angaben über die Entwicklung der Anisakinae. Zeitschrift für Parasitenkunde **17**: 106–121.
- LENT, H., AND J. F. T. FREITAS. 1948. Uma coleção de nematódeos, parasitos de vertebrados do Museu de Historia Natural de Montevideo. Memorias do Instituto Oswaldo Cruz **46**: 1–71.
- LI, A. X., S. D'AMELIO, L. PAGGI, F. HE, R. B. GASSER, Z. R. LUN, E. ABOLLO, M. TURCHETTO, AND X. Q. ZHU. 2005. Genetic evidence for the existence of sibling species within *Contracaecum rudolphii* (Hartwich, 1964) and the validity of *Contracaecum septentrionale* (Kreis, 1955) (Nematoda: Anisakidae). Parasitology Research **96**: 361–366.
- MALACALZA, V., AND M. BERTELLOTTI. 2001. Cambios poblacionales de los Cormoranes (*Phalacrocorax*) en Punta Lobería, Patagonia Argentina. Ornitología Neotropical **11**: 83–86.
- , —, AND T. PORETTI. 1998. Presencia de nematodos (*Contracaecum* sp.) en “pellets” del cormorán real (*Phalacrocorax albiventer*) en Punta León, Chubut, Argentina. Naturalia Patagónica Ciencias Biológicas **6**: 29–34.
- , T. I. PORETTI, AND M. BERTELLOTTI. 1994. La dieta de *Phalacrocorax albiventer* en Punta León (Chubut, Argentina) durante la temporada reproductiva. Ornitología Neotropical **5**: 91–97.
- MATTIUCCI, S., R. CIANCHI, G. NASCETTI, L. PAGGI, N. SARDELLA, J. TIMI, S. C. WEBB, R. BASTIDA, D. RODRIGUEZ, AND L. BULLINI. 2003. Genetic evidence for two sibling species within *Contracaecum ogmorhini* Johnston and Mawson, 1941 (Nematoda: Anisakidae) from otariid seals of boreal and austral regions. Systematic Parasitology **54**: 13–23.
- , G. NASCETTI, R. CIANCHI, L. PAGGI, P. ARDUINO, L. MARGOLIS, J. BRATTEY, S. C. WEBB, S. D'AMELIO, P. ORECCHIA, AND L. BULLINI. 1997. Genetic and ecological data on the *Anisakis simplex* complex with evidence for a new species (Nematoda, Ascaridoidea, Anisakidae). Journal of Parasitology **83**: 401–416.
- , M. PAOLETTI, J. OLIVERO-VERBEL, R. BALDIRIS, B. ARROYO-SALGADO, L. GARBIN, G. NAVONE, AND G. NASCETTI. 2008. *Contracaecum bioccii* n. sp. from the brown pelican, *Pelecanus occidentalis* (L.) in Colombia (Nematoda: Anisakidae): genetic evidence and morphological description. Systematic Parasitology **69**: 101–121.
- , M. TURCHETTO, F. BRIGANTINI, AND G. NASCETTI. 2002. On the occurrence of the sibling species of *Contracaecum rudolphii* complex (Nematoda: Anisakidae) in cormorants (*Phalacrocorax carbo sinensis*) from Venetian and Caorle lagoons: Genetic markers and ecological studies. Parasitología **44**: 105.
- NASCETTI, G., R. CIANCHI, S. MATTIUCCI, S. D'AMELIO, P. ORECCHIA, L. PAGGI, J. BRATTEY, B. BERLAND, J. W. SMITH, AND L. BULLINI. 1993. Three sibling species within *Contracaecum osculatum* (Nematoda, Ascaridida, Ascaridoidea) from the Atlantic arctic-boreal region: Reproductive isolation and host preferences. International Journal for Parasitology **23**: 105–120.
- PUNTA, G. E., J. R. SARAVIA, AND P. YORIO. 1993. The diet and foraging behaviour of two Patagonian cormorants. Marine Ornithology **21**: 27–36.
- , P. YORIO, G. HERRERA, AND J. SARAVIA. 2003. Biología reproductiva de los cormoranes Imperial (*Phalacrocorax atriceps*) y Cuello Negro (*P. magellanicus*) en el Golfo San Jorge, Chubut, Argentina. Hornero **18**: 103–111.
- RASMUSSEN, P. C. 1994. Geographic variation in morphology and allozymes of South American imperial shags. The Auk **111**: 143–161.
- SARDELLA, N. H., AND J. T. TIMI. 1996. Parasite communities of *Merluccius hubbsi* from the Argentinian-Uruguayan common fishing zone. Fisheries Research **27**: 81–88.
- , AND —. 2004. Parasites of Argentine hake in the Argentine Sea: Population and infracommunity structure as evidence for host stock discrimination. Journal of Fish Biology **65**: 1472–1488.
- TIMI, J. T., N. H. SARDELLA, AND G. T. NAVONE. 2001. Parasitic nematodes of *Engraulis anchoita* (Pisces: Engraulidae) from the Argentine and Uruguayan Seas. Acta Parasitologica **46**: 186–193.
- TORRES, P. J., VALDIVIESO, R. SCHLATTER, A. MONTEFUSCO, J. REVENGA, F. MARIN, J. LAMILLA, AND G. RAMALLO. 2000. Infection by *Contracaecum rudolphii* (Nematoda: Anisakidae) in the Neotropic cormorant *Phalacrocorax brasiliensis*, and fishes from the estuary of the Valdivia River, Chile. Studies on Neotropical Fauna and Environment **35**: 101–108.