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New Progresses in the Enantioselective Synthesis and Biological Properties of Carbocyclic Nucleosides

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Abstract: The recent advances in the chemistry of carbocyclic nucleosides focused on different synthetic approaches that lead to optically pure products as well as a comprehensive overview of their biological properties are discussed. In the latter aspect, molecular recognition of enzymes of pharmacological importance such as: reverse transcriptase, adenosine deaminase, thymidine kinase, DNA cytosine-C5 methyl transferase, S-adenosylhomocysteine hydrolase, etc are considered. The role of conformation and puckering of the glycon moiety in modulating the biological activity and also the use of carbanucleosides as building blocks to prepare oligonucleotides are carefully illustrated.

INTRODUCTION

The term carbanucleoside or carbocyclic nucleoside refers to a nucleoside analogue in which a methylene group has replaced the oxygen atom of the furanose ring. These analogues have potent metabolic stability because they are unaffected by phosphorylases and hydrolases that cleave the glycosidic bond of natural nucleosides. On the other hand, the pharmacological actions of conventional nucleosides have been extensively studied, especially as antiviral and antitumor agents. Interestingly, the same enzymes that recognize normal nucleosides displaying a wide range of biological properties, also recognize their carba-analogues. Enantioselective synthesis of these modified nucleosides has received much attention in the last years motivated by the urgent need of having effective antiviral agents especially against HIV. In effect, the lack of therapeutically valuable drugs against viral infections gave an enormous impetus to the search for new synthetic protocols that lead to optically pure nucleoside analogues. In addition, the methodology for the preparation of carba-C nucleosides is also included. In this case, a C-C bond instead of a N-C bond connects the heterocyclic base to the pseudosugar moiety. At the present time, several review articles have appeared covering not only the synthetic aspects of the chemistry of carbocyclic nucleosides [1-7] but also the important pharmacological properties they exhibit [8–9].

Different synthetic approaches have been developed for the synthesis of carbanucleosides, including the critical steps the preparation of the pseudosugar and the incorporation of the heterocyclic base onto the carbocyclic ring (linear or convergent approach). The linear strategy, that is step-bystep base construction, will not be covered in this article. It

Solid Phase Synthesis of Carbocyclic Nucleosides

In order to overcome the problems associated to the Mitsunobu reaction, solid phase syntheses of different carbanucleosides have been attempted [14, 15]. Chu et al. have developed a solid phase strategy for synthesis of L-2'deoxynucleosides of general structure 7 employing a Mitsunobu reaction to incorporate all the nucleic bases. The required synthetic intermediate 3 to achieve this approach was prepared from D-ribose (1) in 15 steps via the known carbocyclic ring 2. Therefore, with the use of a p-nitrophenyl carbonate resin 4 [16], the purification problems associated to this reaction were overcome and also the reaction yields were significantly improved. In the case of pyrimidines, the regioselectivity of this reaction $(N^1$ - vs O^2 - alkylated pyrimidines) was enhanced with the exemption of the cytosine derivative where the O^2 -alkylated cytosine was isolated as the major product. This effect is more noticeable in the preparation of purine derivatives where the undesired N^7 -alkylated products were not detected [15] (Scheme 1).

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deserves to mention among the convergent methods those that are suitable for a specific transformation such as the base nucleophilic attack to an epoxide group; a nucleophilic attack of a suitable base on a sulfur or selenium-activated double bond; a palladium catalyzed alkylation; and base addition to a Michael receptor, and those in which a S_N2 attack on an activated hydroxyl group is involved. In this case the Mitsunobu-type reaction arises as the main protocol to introduce a nucleic base into a carbocyclic ring [10-12]. Nevertheless, this Mitsunobu type reaction presents some disadvantages regarding the competition between N-9 versus N-7 alkylation in the case of purines and N-alkylation versus O-alkylation when employed N-1-protected pyrimidines. In addition, the major drawback of this reaction is the elimination of associated products formed in the course of the reaction: triphenylphosphine oxide and the respective dialkyl hydrazinedicarboxylate. The use of basic phosphines to facilitate the removal of the corresponding oxidized product has not been widely accepted [13].

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Solid phase synthesis of "unnatural" carbanucleosides

Structures of the corresponding heterocyclic bases (B'H):

Scheme 1. Reagents and Conditions: (a) 3 (3 eq.), DMAP (1 eq.), DIPEA (10 eq.), CH₂Cl₂, 40 °C, 2 days; (b) PPTs, n-BuOH/1,2-dichloroethane, 60 °C; (c) B'H (8a-e), DEAD, Ph₃P, DMF/1,4-dioxane or DMF, rt; (d) K₂CO₃, THF/MeOH, rt 50% for 7a, 100% for 7b, 97% for 7c, 55% for 7d, 62% for 7d (all yields correspond to four steps from 3).

Palladium-catalyzed allylic substitution of an allylic ester with either a purine or pyrimidine base is another useful coupling method for the direct introduction of a heterocyclic base onto the corresponding pseudosugar [17–19]. In this reaction, the yield of the reaction can be drastically impaired by formation of the N^7 -alkylated product. By the use of this

approach, Crimmins et al. have recently reported the solid phase synthesis of abacavir (9) and other closely related compounds to 9 and to carbovir ((-)-10), in which the N^9 -alkylated product was obtained as a single compound in all cases. This particular behavior can be rationalized taking into account that the presence of the resin provokes an

Scheme 2. Reagents and Conditions: (a) 10 mol % Pd₂(dba)₃, 0.4 equiv. PPh₃, 6.0 equiv. 1,2,2,6,6-pentamethylpiperidine, 1.5 equiv. 2-amino-6-chloropurine, THF-DMSO (1:1), 45 °C, 16 h; (b) cyclopropylamine, EtⁱPr₂N, BuOH, 80 °C, 4 h; (c) 5% TFA, CH₂Cl₂.

enhancement of the steric effect in the trajectory of the base in its nucleophilic attack (Scheme 2).

Asymmetric Synthesis of Carbocyclic Nucleosides via Aldol/ring-closing Metathesis Reaction

Abacavir (9) [20] and carbovir ((-)-10) [21] are synthetic carbanucleosides, which exhibit similar inhibitory activity than AZT against viral reverse transcriptase of the Human Immunodeficiency Virus (HIV). It deserves to point out that abacavir is a recently FDA-approved anti-HIV drug with less toxic effects than any other nucleoside analogues. These

drugs and additional structurally related carbocyclic nucleosides have been enartioselectively prepared via an asymmetric aldol/ring-closing metathesis [22]. Thus, employing three powerful reactions, from the synthetic point of view, these carbanucleosides were synthesized in an elegant and efficient way. The reactions involved in this strategy are: (a) an asymmetric aldol addition employing Evans chiral auxiliaries to define the relative and the absolute stereochemistry [23, 24]; (b) a ruthenium-catalyzed ring-closing metathesis (RCM) to form the carbocyclic pseudosugar [25]; (c) palladium-catalyzed allylation reaction to incorporate the nucleic base onto the ring [17–19]. The last reaction is the key step in this commonly used

Scheme 3. Reagents and Conditions: (a) *n*-BuLi, THF, -78 °C, pivaloyl 4-pentenoate, 99%; (b) TiCl₄, (-)-sparteine, CH₂Cl₂, acrolein, 82%; (c) CH₂Cl₂, Cl₂(Cy₃P)₂Ru=CHPh, 25 °C, 97%; (d) LiBH₄, THF, MeOH, 78%; (e) Ac₂O, CH₂Cl₂, DMAP, Et₃N, 90%; (f) 2-amino-6-chloropurine, Pd(PPh₃)₄, NaH, THF-DMSO (1:1), 45 °C (g) 2-amino-6-cyclopropyl-aminopurine, Pd(PPh₃)₄, NaH, THF-DMSO (1:1), 40 °C, 62%; (h) NaOH, H₂O, 68%; (i) NaOH, H₂O.

convergent method for the preparation of carbocyclic nucleosides [1]. Therefore, the oxazolidinone 16 was used as chiral auxiliary. Compound 16 was acylated by treatment with 4-pentenoic pivalic mixed anhydride to give 17, which on reaction with crotonaldehyde afforded the syn aldol product 18. This compound was the substrate for the RCM reaction to give rise to carbocyclic 19. Finally the third critical step was the coupling of the base employing a π -allylic palladium rearrangement as it is outlined in Scheme 3.

Synthesis of Carbovir via Ring-expansion Reaction of Cyclobutanones

An interesting new approach for the synthesis of carbanucleosides is the preparation of the unnatural isomers of carbovir ((+)-10) and aristeromycin ((+)-23) described by Hegedus [26]. For this purpose, a ring-expansion reaction of cyclobutanones was employed for the synthesis of the carbocyclic ring. The required chiral α -alkyl- α -alkoxycyclobutanone 26 is prepared via a photolytic reaction of the chromium carbene complex 24 with the Evans-type chiral enecarbamate 25 [27, 28]. Once the cyclobutanone is at hand, ring expansion is achieved by treatment with lanthanide triflates used as selective and mild Lewis acids

[26]. The ring expansion proceeds with high regioselectivity favoring the desired β -substituted cyclopentanone 27 as indicated in Scheme 4. In this example the base is also coupled by the palladium-catalyzed coupling reaction to produce the corresponding carbocyclic nucleosides precursors 30 and 31, respectively.

This π -allyl coupling method mediated with Pd (0) has been recently used by Smith *et al.* to prepare famciclovir (33) [29]. As illustrated in Scheme 5, the isopropyliden keto derivative (34) is reacted with vinyl magnesium bromide and the resulting alkoxy intermediate was trapped with methyl chloroformate to afford 35, which is coupled with 2-amino-6-chloropurine to give the advanced carbanucleoside precursor 36. Employing 1,2-bis(diphenylphosphino) ethane as a ligand and tris(dibenzylideneacetone) dipalladium(0) as palladium source, the ratio of N^9/N^7 alkylation product is 35:1. Cleavage of the protecting isopropylidene group afforded 37 that is further transformed into the desired drug 33 by treatment with acetic anhydride.

Preparation of Carbovir Analogues

Bearing in mind the potential usefulness of carbovir as antiviral drug, several closely related compounds has been

Scheme 4. Reagents and Conditions: (a) hv, CH_2Cl_2 , 76%; (b) $Me_3S(O)I$, NaH, $Sc(OTf_3)$, DMF; (c) Li_2CO_3 , 74%; (d) H_2 (80 psi), $[Rh(COD)dppb]BF_4$, DMF, 77%; (e) LDA, THF, 0 °C; (f) DIBAL, THF, -78 °C, 50%; (g) $CICO_2Et$, py, CH_2Cl_2 , 87%; (h) $Pd(PPh_3)_4$, 2-amino-6-chloropurine, DMF, -40 °C \rightarrow 10 °C, 63%; (i) BCl_3 , CH_2Cl_2 , -78 °C \rightarrow 0 °C, then MeOH, 83%; (j) 0.5% aq. NaOH, 100 °C, 3.5 h, 72%; (k) $Pd(PPh_3)_4$, adenine, DMF, rt, 90 min, 65%; (l) BCl_3 , CH_2Cl_2 , -78 °C \rightarrow 0 °C, then MeOH, 64%; (m) OsO₄, DMF, -15 °C, 36 h, 43%.

OCO₂Me
$$\begin{array}{c}
OCO2Me \\
A_{2}N \\
A_{3}
\end{array}$$

$$\begin{array}{c}
OCO2Me \\
A_{2}N \\
A_{3}N \\
A_{3}
\end{array}$$

$$\begin{array}{c}
OCO2Me \\
A_{3}N \\
A_{3}N \\
A_{3}N \\
A_{3}$$

$$\begin{array}{c}
OCO2Me \\
A_{3}N \\
A_{3}N \\
A_{3}$$

$$\begin{array}{c}
OCO2Me \\
A_{3}N \\
A_{$$

Scheme 5. Reagents and Conditions: (a) CH₂=CHMgBr, THF, then ClCO₂Me, 73%; (b) Pd₂(dba)₃, dppe, Cs₂CO₃, DMF, 80 °C, 7.5 h, 72%; (c) i. H₂, Pd/C, Et₃N, THF, ii. MeOH/HCl, 93%; (d) Ac₂O, DMAP, Et₃N, MDC, 80%.

HO
Ogle

$$A$$
OR
 A
OR

Scheme 6. Reagents and Conditions: (a) i. Hg(OAc)₂, NaBH₄, or ii. β -glucosidase, NaBH₄, 90%; (b) isobutyric anhydride, py, 0 °C \rightarrow 25 °C, 90 min, 90%; (c) Ac₂O, py, 1 h, 99%; (d) i. N_6 -benzoyl-adenine, MeCN, TMSCl, HMDS, ii. CF₃(CF₂)₃Si(CH₃)₃, CH₂Cl₂, reflux, 8 h, 20%; (e) DIBAL, CH₂Cl₂, -78 °C, 95%.

designed and prepared. In fact, Bianco et al. have recently depicted the enantioselective synthesis of aucubovir II (38) [30] employing the Vorbrüggen methodology for introducing the base onto the carbocyclic ring as key step [31, 32]. In this case, aucubin 1 (39) was employed as chiral starting material (Scheme 6). There is no information about the antiviral activity of aucubovir II.

Roberts et al. have described a very interesting example in the preparation of di- and tri-phosphorylated fluor-

containing derivatives of carbovir [33] keeping the base as guanine based on the modified triphosphorylated drugs 44 and 45, which proved to be potent inhibitors of HIV-reverse transcriptase [34]. These kinds of nucleotides are able to liberate phosphonoacetic acids. In this case, carbanucleotides (46-48) act as prodrugs [33] releasing compounds 49-51. These drugs have been designed not only to evaluate the nucleotides per se but also on the basis that phosphonoacetic acids are non-competitive inhibitors of viral DNA polymerases (herpes simplex virus, cytomegalovirus, HIV)

Scheme 7. Reagents and Conditions: (a) morpholine, DCC, ¹BuOH-H₂O, reflux, 4 h; (b) 46, 47 or 48 (NBu₃, salt), DMSO, 14-39 h, rt, 50% for 49, 32% for 50, 13% for 51.

[35]. Therefore, a synergistic effect would be anticipated because these metabolites acting as DNA polymerase inhibitors would combine their action with the original reverse transcriptase blocking effect of the nucleosides. Unfortunately these derivatives are almost devoid of activity against HIV reverse transcriptase [33]. The corresponding tributylammonium salts of these phosphonoacetic acids (46-48) are coupled with morpholidate 53, which in turn was prepared from the known phosphonate 52 [36] (Scheme 7).

Cyclopropane Derivatives

Tsuji et al. have designed and prepared several carbocyclic nucleosides structurally related to acyclovir (54), ganciclovir (55) and penciclovir (56), as well as the

carbanucleoside analogues of oxetanocin G (57), and cyclobut-G (58) [37]. For the design of these drugs, the relative position and special alignment of both 3' and 5' hydroxyl groups of 2'-deoxyribose ring were considered [37]. A cyclopropane group, an epoxide ring, or a double bond were employed to impart a certain degree of rigidity required to orientate the hydroxyl groups for molecular recognition. The designed compounds are presented in Scheme 8. At the beginning, only racemic syntheses were carried out, but once the biological evaluation was at hand, the enantioselective preparation of the more prospective compounds was developed. From the synthesized carbanucleosides, (\pm)-59 resulted to be very effective against HSV-1 (IC₅₀ 0.046 μ g/mL) and 20 times more potent than acyclovir (IC₅₀ 0.81 μ g/mL) under the same assays

Scheme 8. Reagents and Conditions: (a) TBDMSCl, Im, DMF, 0 °C, 56%; (b) i. OsO4, NMO, THF-H₂O, rt, 84 h, ii. NaIO₄, rt, 16 h, 81%; (c) NaBH₄, MeOH, 0 °C, 30 min, 87%; (d) BzCl, py, 0 °C, 30 min, 97% for **65**, 30% for **67**; (e) aq. HCl, MeOH, rt, 40 min, 100%; (f) ClTs, 4-DMAP, CH₂Cl₂, 0 °C, 1 h, 84%; (g) 2-amino-6-(benzyloxy)purine, K₂CO₃, 18-crown-6, DMF, 60 °C, 2 h, 61%; (h) i. MeONa/MeOH, 40 °C, 30 min, ii. 1 N HCl, 50 °C, 30 min, 81%; (i) EtONa/EtOH, diethyl malonate, 75 °C, 20 h, 70%; (j) NaBH₄, EtOH, rt, 2 h, 69%; (k) 2,2-dimethoxypropane, *p*-TsOH, DMF, rt, 12 h, 49%; (l) LiBH₄, THF, reflux, 12 h, 64%; (m) NaH, DMF, BnBr, rt, 14 h, 90%; (n) HCl, THF, 0 °C, 30 min, 86%; (o) BzCl, py, CHCl₃, 0 °C, 12 h, 68%; (p) H₂, 10% Pd/C, EtOH-EtOAc, 1 atm, 3 days, 99%.

conditions, while the pure enantiomer (-)-59 presented an IC_{50} value of 0.020 µg/mL, that is, 2-fold more potent than the racemic mixture [37]. The enantioselective synthesis of (-)-59 was achieved starting from the available cyclopropane lactone 71, which in turn, is obtained by condensation of diethyl malonate and (R)-(-)-epichlorhydrin (70) according to Pirrung et al. [38] (Scheme 8).(+)-59, is prepared in the same way from (S)-(+)-epichlorhydrin. This unusual activity can be explained by the introduction of a conformational restriction modulated by the cyclopropane ring, which force the hydroxyl groups to adapt a somewhat defined spatial orientation that is required for a better interaction with the involved enzymes. All of these cyclopropyl-containing nucleosides are not completely rigid, they have preferential hydroxyl groups orientation, in other words, they have such a flexibility that can interact either with thymidine kinases or viral DNA polymerases in an efficient way leading to potent antiviral agents [37]. Epoxides derivative like (±)-60 is moderately active against HSV-1. This low activity may be attributed to ring-opening under the assays conditions.

Among these cyclopropyl-containing carbanucleosides, Ortuño *et al.* have reported an enantioselective synthesis of a new series of this family of compounds, in which the base, in the case of thymine, is bonded to the cyclopropyl group instead of to the methylene group as Tsuji's nucleosides to produce 80 [39]. These compounds were prepared according to Scheme 9 employing (–)-(Z)-2,3-methanohomoserine (81) as a chiral starting material [39]. This linear approach is very efficient and occurs with high stereoselectivity with 16% overall yield for the thymidine derivative.

It deserves to mention another synthetic methodology to prepare *cis,cis*-trisubstituted cyclopropyl carbocyclic nucleosides with the base also linked to the cyclopropyl ring [40]. These compounds were designed taking into account that the base closeness to the carbocyclic ring would favour the interactions with the different involved enzymes.

Scheme 9. Reagents and Conditions: (a) LiBH₄, 94%; (b) TBDMSCl; (c) H₂, Pd(OH)₂, 83% two steps; (d) 3-methoxy-2-methylacrylic acid, (COCl)₂, AgOCN, 30%; (e) 0.2 M HCl, 68%.

However, these compounds are devoid of antitumor activity. The preparation of the adenosine and thymidine derivatives, compounds 86 and 87, respectively, is illustrated in Scheme 10. The critical step in this approach is the ring contraction of a cyclobutane ring in compound 91 to give rise to the desired cyclopropyl intermediate 92 in excellent yield [41].

Cyclohexane Derivatives

Analogues of anhydrohexitol nucleosides 102 represent very good examples of the potential utility of nucleosides analogues (Scheme 11). These drugs, developed by Herdewijn *et al.*, exhibit potent antiviral activity [42, 43], while the carbocyclic analogues of general formula 103 are devoid of antiviral activity [44]. This lack of antiviral action may be attributable to the fact that, in anhydronucleosides,

Scheme 10. Reagents and Conditions: (a) LiAlH₄, 96%; (b) NaH, BrBn, 94%; (c) NBS, H₂O, 76%; (d) NaOH, 100%; (e) Jones' reagent, -5 °C, 10 h, 71%; (f) i. ClCO₂Et, Et₃N, THF, -5 °C, 1 h, ii. NH₃/THF, 0 °C, 1 h, 82%; (g) PhI(OAc)₂, KOH, MeOH, rt, 2 h, 78%; (h) KOH, MeOH, reflux, 48 h, 83%; (i) 3-methoxy-2-methylacryloyl chloride, AgOCN, PhH, reflux, 20 h, 90%; (j) NH₄OH, MeOH, 85 °C, 24 h, 57%; (k) i. BCl₃, CH₂Cl₂, -78 °C, ii. MeOH, -78 °C, 60%; (l) 5-formamido-4,6-dichloropyrimidine, Et₃N, dioxane, reflux, 8 h, 50%; (m) i. diethoxymethylacetate, 120 °C, 22 h, ii. NH₄OH, MeOH, rt, 1 h, 58%; (n) NH₃/MeOH (saturated at -78 °C) 80 °C, 24 h, 73%; (o) BCl₃, CH₂Cl₂, -78 °C, ii. MeOH, -78 °C, 75%.

the base adapts an axial position (conformation ¹C₄), while in the carbocyclic nucleosides the base is equatorial orientated (conformation ⁴C₁) [45]. Moreover, NMR conformational studies indicate that carbocyclic nucleosides do not mimic quite well the characteristic 3'-endo conformation of conventional nucleosides, while the conformation found in anhydrohexitol nucleosides is almost similar to that found in normal nucleosides [45]. In spite of these results, a new series of carbanucleosides bearing an additional hydroxyl group was investigated [46]. In this case, an α-oriented hydroxyl group at the C-5' position in carbanucleosides of general formula 104 was designed, synthesized and evaluated against different viruses due to the presence of this new substituent might favor the conformation ¹C₄ similar to that found in anhydrohexitoles [45]. Neither of these drugs exhibited any inhibitory action against HSV-1, HSV-2, HIV-1, HIV-2, vaccinia virus, vesicular stomatitis virus, etc. According to the ¹H NMR analysis, the conformation ⁴C₁ with equatorial base is still preferred [45]. The enatioselective synthesis for this family of compounds employs (R)-(-)-carvone as chiral source. The

key steps for this approach are the regio- and stereoselective hydroboration reaction of the double bond of a 2methylencyclohexane-1,3-diol derivative (compound 114) to give 115, and the introduction of the base through a Mitsunobu-type reaction to afford the title compounds 121 and 124 (Scheme 11). Bearing in mind that the antiviral activity of nucleosides is attributed to complex interactions among a set of enzymes, Herdewijn et al. designed a somewhat flexible carbanucleoside analogue capable of interacting with all the enzymes involved. Employing this concept, and in order to have a low energetic barrier between both conformations, a double bond was introduced onto the carbocyclic ring of general formula 125 and 126. Molecular modelling studies and careful analysis of the proton NMR spectra confirm that the pseudoaxial position of the base corresponds to the preferred conformer, probably, due to the existence of overlapping between the π orbital (C5'-C6') and the antiligand bond C1'-N that favors the conformation ³H₂ over the ²H₃. In addition, the values of the energy barrier result to be 1.6 kJ/mol less than the cyclohexanes derivatives [45]. The synthetic strategy to prepare

Scheme 11. Reagents and Conditions: (a) H₂O₂,/NaOH, MeOH, 83%; (b) L-selectride, THF, -65 °C; (c) TBDMSCl, Im, DMF, 93%; (d) 1% OsO₄/NaIO₄, THF-H₂O, rt, 83%; (e) *m*-CPBA, CHCl₃, pH = 8, rt, 73%; (f) K₂CO₃, MeOH-H₂O, 73%; (g) TBDMSCl, Imidazole, DMF, 80%; (h) LiTMP/Et₂AlCl, PhH, 0 °C, 100%; (i) NaH, BnBr, TBAI, THF, rt; (j) 9-BBN, THF, rt, 62%; (k) TBAF (1 equiv.), THF, rt, 93%; (l) PhCH(OMe)₂, PTSA, dioxane, 90%; (m) TBAF, THF, 93%; (n) PhCO₂H, DEAD, PPh₃, dioxane; (o) K₂CO₃, MeOH, 72%; (p) adenine, DEAD, PPh₃, dioxane, rt, 93%; (q) Pd(OH)₂/C, cyclohexene, MeOH, reflux, 73%; (r) 6-chloro-2-aminopurine, DEAD, PPh₃, dioxane, rt, 37%; (s) CF₃CO₂H/H₂O (3:1), rt, 75%; (t) Pd(OH)₂/C, cyclohexene, MeOH, reflux, 63%.

cyclohexenyl carbanucleosides also employ (R)-(-)-carvone as chiral starting material and has several points of coincidence (Scheme 12). Once the synthetic intermediate 129 is available, this product is further transformed into the allylic alcohol 135, which is coupled either with adenine or 2-amino-6-chloropurine under Mitsunobu-type conditions to give the corresponding advanced carbanucleosides precursors

136 and 137, respectively, that after cleavage of the corresponding protecting groups affords adenosine and guanosine derivatives D-125 and D-126, respectively [46,47]. As can be observed in Scheme 12, both enantiomers of each nucleic base have been synthesized from the same starting material (D-A, L-A, D-G, L-G). Guanosine derivative D-126 results to be a potent and highly selective antiviral agent

Scheme 12. Reagents and Conditions: (a) n-BuLi, TMP, Et₂AlCl, PhH-PhMe (1:1), 0 °C, 3 h, 71%; (b) 9-BBN, THF, 0 °C, then H₂O₂, 2 N NaOH, 74%; (c) TBDMSCl, Imidazole, DMF, rt, 70%; (d) MsCl, Et₃N, CH₂Cl₂, 0 °C, 92% for 131, 98% for 141; (e) 10% Pd/C, HCO₂NH₄, MeOH, reflux, 76%; (f) MnO₂, CH₂Cl₂, rt 48% and 47% recovery starting material; (g) NaBH₄, CeCl₃•7H₂O, MeOH, 0 °C → rt, 91 % for 135, 75% for 144; (h) adenine, DEAD, PPh3, dioxane, rt, 66% for 136, 40% for 145, 2-amino-6-chloropurine, DEAD, PPh3, dioxane, rt, for 137 and 146; (i) TFA/H2O, rt, 78% for D-125, 46% from 135 for D-126, K2CO3, MeOH, rt for L-125, 58% from 144 for 146 followed by NH₃/MeOH, 80 °C, 2 days 73% for L-126; (j) Bz₂O, DMAP, CH₂Cl₂, 0 °C, 98%; (k) TBAF (1 equiv), THF, rt, 74%; (l) TBAF, THF, rt, 86%; (m) PDC, CH2Cl2, rt, 48%.

with effectiveness similar to the commonly employed drugs for the treatment of herpes infection, acyclovir and ganciclovir. The selectivity presented for these drugs make them good candidates for in vivo assays. It deserves to pointing out that both enantiomers exhibit almost the same potency as antiviral agents. Docking experiments with HSV-1 thymidine kinase (TK) shows that both enantiomers fit well in the active site when the conformation is flipped to that with a pseudo equatorial oriented base moiety. This constitutes an interesting example because the enzyme interacting with same efficacy at the active site can recognize both enantiomers. These results are in agreement with previous studies, in which inversion of configuration occurs when hexitol nucleosides are co-crystallized with HSV-1 TK. Very recently, Herdewijn et al. have reported that incorporation of cyclohexenyl carbanucleosides into oligonucleotides produces a remarkable heteroduplex stabilization [48]. Certainly, incorporation of one, two, or three unit of p-125 in the DNA strand produces an increment of Tm in +1.1, +1.6, and +5.2 °C, respectively. Interestingly, the stabilization effect of anhydrohexitol 102 (B = adenine) is two-fold higher than **D-125** [48]. However,

the oligoadenilate-poly(dT) complex is significantly more stable than the corresponding anhydrohexitol oligoadenilate-poly(dT) complex. As cyclohexenyl carbanucleosides have structural similarities compared with natural furanosyl nucleic acids, they produce similar conformational changes than conventional nucleosides. In addition, since cyclohexenyl carbocyclic nucleosides combine the advantages of stabilizing heteroduplex complexes, of being stable in serum, and of activating RnaseH, these properties make these compounds good candidates for antisense evaluation in cellular systems. It is interesting to notice that 102 (B = adenine) is recognized by a number of DNA polymerases, especially by the B-family, however, in any case no more than two consecutive triphosphate derivative can be incorporate [49].

Cyclopentenyl Carbanucleosides

A very motivating novel class of nucleosides analogues are apio and thioapio dideoxydidehydronucleosides derivatives [50]. Although these drugs cannot be considered

Scheme 13. Reagents and Conditions: (a) H_2 , 10% Pd/C, EtOAc, 99%; (b) LiHMDS, THF, -78 °C, then TMSCl, rt; (c) PhSeBr, THF, -78 °C, 77% from 151; (d) i. DIBAL, PhMe, -78 °C, ii. Ac₂O, py, rt, 85%; (e) NH₄SO₄ (cat), HMDS, 140-150 °C \rightarrow rt, then HMDS removal, 1,2-dichloroethane, 5 °C, TMSOTf, 90% for 155a, 63% for 155b, 62% for 155c, 69% for 155d, 91% for 155e, 68% for 155f; (f) CH₂Cl₂, 30% H₂O₂, rt, then TBAF, THF, 0 °C, 84% for 149a, 44% for 149b, 44% for 149c, 90% for 149d, 90% for 149e, 45% for 149f; (g) 1 N NaOH, 1 h, then (CH₃)₂SO₄, DMSO, 1 h, 89%; (h) MsCl, py, rt; (i) KSAc, DMF, rt, 7 h, 86% from 156; (j) NaOH, H₂O, EtOH, 50 °C, 2.5 h, then DCC, DMAP, CH₂Cl₂, 30 min, 70%.

Scheme 14. Reagents and conditions: (a) DIBAL-H, PhMe, -78 °C, 81%; (b) CH₂=CHMgBr, THF, -78 °C, 100%; (c) TPAP, NMNO, CH₂Cl₂, 0 °C, 66%; (d) DIBAL-H, PhMe, -78 °C, 84%; (e) 60% NaH, DMSO, THF, PPh₃CH₃Br, 0 °C \rightarrow rt, 88%; (f) Grubbs catalyst, CHCl₃, rt, 2 h, 89%; (g) MnO₂, CH₂Cl₂, rt, 5 h, 83%; (h) Ph₃PCH₃Br, NaH, DMSO, THF, rt, overnight, 72%; (i) (COCl)₂, DMSO, CH₂Cl₂, -78 °C, 1 h, then NEt₃ \rightarrow rt; (j) CH₂=CHMgBr, THF, -78 °C, 1 h, 63%; (k) Grubbs catalyst, CHCl₃, rt, 2 h, 77%; (l) MnO₂, CH₂Cl₂, rt, 15 h, 81%.

as carbanucleosides, the fact that the oxygen atom of the furanose ring moves to the C-2 position make these drugs not recognisable by adenosine deaminase, phosphorilases and hydrolases, behaviour found in carbanucleosides. These drugs were designed on the basis of the antiviral activities shown by 2',3'-dideoxycytidine (ddC, 147) and 2',3'dideoxy-2',3'-didehydrothymidine (d4T, 148) by moving the oxygen atom and the double bond to form nucleosides analogues of general formula 149. These compound were synthesized according to Scheme 13 starting from the known α, β-unsaturated lactone 150 [51]. All the 149 series is devoid of activity against HIV-1, HSV-1 and HSV-2, but the fluorouracil derivative (149c) is a potent anti-HCMV agent at the low micromolar level, while the rest of apio derivatives are moderately potent drugs against HCMV [50]. The thioapio nucleosides are not active against HCMV.

A very useful synthetic intermediate (compound 159) is now available for the preparation of D- and L-cyclopentenyl carbocyclic nucleosides of pharmacological importance including the naturally occurring neplanocin A (160) and aristeromycin (23) [52]. Both enantiomers of 159 can be prepared from the same starting material, D-isoascorbic acid (161) as depicted in Scheme 14. 161 is easily transformed into the isopropyliden lactone 162 according to published procedures [53]. The levo-isomer (-)-159 leads to D-carbocyclic nucleosides while the enantiomer (+)-159 leads to the L-series [52].

Conformationally Constrained Carbanucleosides

Of special interest is the sugar conformation / biological activity studies made in a variety of carbocyclic nucleosides. In solution, the ribose unit exits in a dynamic equilibrium between the Northern-type geometry and the facing Southern-type geometry according to the concept of the pseudorotational cycle [54]. To the contrary, only one of these conformers is found in the crystalline structure, and only the Northern or Southern conformer is solely responsible for molecular recognition as well because only one form is present in the drug-enzyme complex. However, as the ribose ring is bendable and the conformation in solution may be unlike than that found in the solid state, any attempt to correlate sugar conformation with biological action would be flawed unless the crystal and the solution conformation are the same. A cyclopropane or epoxy ring can confer such rigidity to the sugar moiety that the equilibrium $N \Leftrightarrow S$ is not observed indicating that the solution conformation is virtually identical to that found in the crystal [55]. However, the conformational restriction induced by an epoxy group at the C-2', C-3' positions fixes the sugar conformation far away from the Northern or Southern geometry that is required for biological activity. In carbocyclic nucleosides, the elimination of the oxygen atom of the furanose ring brings about a significant change in terms of stereoelectronic effects. Therefore, the anomeric effect as well as significant gauche interactions between this

Scheme 15. Reagents and conditions: (a) 6-chloropurine, PPh₃, DEAD, THF, rt, 1 h; (b) AcOH, 50 °C, 24 h, 40% from 173; (c) *m*-CPBA, CH₂Cl₂, 0 °C \rightarrow rt, 10 days, (d) NH₃/MeOH, 70 °C, 5 h, 75% for 177, 63% for 179; (e) H₂, 10% Pd/C, MeOH, 3 atm, 88% for 172, 72% for 180.

oxygen atom and any electronegative substituent at C-2' or C-3' is lacking. The naturally occurring carbocyclic nucleoside neplanocin C (172) [56-59], is a good prototype of a conformationally locked nucleoside analogue. This compound, isolated from Ampullariela regularis, is a minor component of the neplanocin family of antibiotics, and it is built on a [3.1.0]-bicyclic system, which allows it to exhibit the typical northern-type (N) conformation, specifically in the ₂E conformation that is very close to a pure 3T_2 (P = 0°) geometry as determined by the P value of the pseudorotational cycle (P value = 338.03° and v_{max} = 21.89°) from the solved X ray structure [59]. The structure of neplanocin C was taken as lead drug for the development of a huge number of conformationally restricted carbanucleosides of biological importance [60-71]. The enantioselective synthesis of this valuable carbocyclic nucleoside has been recently achieved [72, 73] employing cyclopentenol 173 as advanced synthetic intermediate, which in turn, is available from D-ribonolactone in seven steps [72]. The critical step for the synthesis of neplanocin C is the epoxidation reaction, in which it is required ten days of reaction time to produce the neplanocin C precursor 176 and its diasteromer 178 in a 1:1 ratio. Both of these isomers are easily separated by column chromatography. The unusual stereochemical course of the epoxidation reaction, where the Hembest rule is not applicable, can be explained bearing in mind that the base in the anti position in 175 is almost 6 kcal/mol more stable that the syn conformer. Then, the electrophilic epoxidizing agent can also coordinate with the N-3 rather than exclusively with the hydroxyl groups affording equal amounts of 176 and 178. The other critical point is the remarkable stability of the epoxy functionality under methanolic ammonia at high temperature to produce 177. This fact had already been described in simpler and closely related carbanucleosides built in a [3.1.0]oxabicyclo systems [74]. As it was mentioned before several carbanucleosides has been synthesized on the basis of

neplanocin C structure. Among them, it is worth pointing out the following conformationally rigid carbocyclic nucleosides: (a) the adenosine derivative of 2',3'dideoxycarbanucleosides locked in the N geometry (181) is moderately active against HIV [60, 61]. The 5'-triphosphate of conformationally restricted carbocyclic analogues of AZT locked in the Northern conformation (182) is exclusively responsible for reverse transcriptase (RT) inhibition [62], while the 5'-triphosphate of its isomer rigid in the S geometry 183 ((S)-methanocarba-AZT) is devoid of activity against RT [62]. In addition, neither of these two rigid AZT analogues exhibit anti-HIV activity as such, and as the S geometry is the conformation needed for molecular recognition by kinases, it is reasonable that 182 is not efficiently phosphorylated, but it is very surprising that 183, in spite of having the required S conformation (specifically (3E) conformation), is not capable of incorporation of phosphate either. This behavior might be attribute to the fact that the rigidity of this prodrug forces the thymine base to a syn orientation destabilizing the enzyme-substrate complex. These results explain why relatively flexible nucleosides prodrugs like AZT can be phsophorylated and then adopt the required N conformation to inhibit RT. (c) (N)methanocarbathymidine (184) is an extremely potent drug against herpes simplex virus 1 and 2 (HSV1 and HSV2) and even more potent than acyclovir, a well known antiviral agent that is employed at the present time for treatment of herpetic infections [63, 64]. In addition, kinetics and crystal structure of 184 interacting with thymidine kinase from Herpes simplex virus type-1 has been conducted [75] demonstrating that the antiviral activity may be attributed to thymidine salvage pathway [75]. methanocarbathymidine and (S)-methanocarbathymidine (185) also shows potential to be used as building block in antisense chemotherapy, in fact, when 184 replaces thymidine in DNA/RNA heteroduplexes an increment of thermodynamic stability is observed, while 185, which is

Fig. (1).

locked in the S geometry, produces an opposite destabilizing effect [65, 66]. Adenosine derivative (N)-methanocarba-2'-desoxyadenosine (186) is a substrate of adenosine deaminase (ADA), the enzyme responsible for catalyzing irreversible deamination of adenosine to inosine, this compound is deaminated 100 times faster than its antipodal rigid conformer 187, (S)-methanocarba-2'-deoxyadenosine [67] (Figure 1). ADA is a crucial enzyme in the purine metabolic pathway that appears at extremely high levels in tumoral lymphocytes, whereas it is not well expressed in several inherited immunodeficient diseases. In a rigorous study of preferential conformation for catalysis it is described that the N-type nucleosides form a stable complex with ADA [76].

Certainly, the N-locked carbanucleoside analogue 186 is deaminated 100-fold faster than the S-type analogue 187 [77]. One attractive application of the concept of lock conformation is the preparation of DNA duplexes bearing conformationally constrained abasic sugars to investigate the mechanism of base flipping by *Hha*I DNA (cytosine C5)-methyltransferase [78]. In conclusion, some of the Northern analogues prove to be extremely potent antiviral agents while the Southern derivatives exhibit vanishing inhibitory action.

A novel synthetic approach for access to bicyclo[3.1.0]hexane carbocyclic nucleosides that employs

Scheme 16. Reagents and conditions: (a) LDA, THF, -78 °C, CH₂=CHCHO; (b) ¹BuPh₂SiCl, CH₂Cl₂, 53% from ethyl acetoacetate; (c) TsN₃, Et₃N, MeCN, 99%; (d) CuSO₄, cyclohexane, Δ, 61%; (e) NaBH₄, MeOH, rt; (f) LiAlH₄, Et₂O, 79% from 184; (g) BzCl; (h) n-Bu₄NF, THF; (i) adenosine deaminase, 80%.

an intramolecular cyclopropanation as key step has been recently reported [79]. In this strategy, a carbene generated from an appropriate β -ketoester gives rise with high diastereoselectivity to the desired carbocyclic ring in an elegant and efficient way. The natural enantiomers of the adenosine and guanosine are obtained with the use of ADA to discriminate the "natural" carbanucleosides. Therefore, (\pm)-199 is irreversible deaminated to give the corresponding guanosine derivative (+)-200, while (\pm)-196 to the inosine derivative (-)-197 plus the unnatural adenoside (+)-198. (-)-

197 is converted back into carbocyclic (-)-196 by known procedures (Scheme 16) [79]. With the use of this intramolecular carbene insertion on a double bond is also possible to access to the rigid S conformers [80]. As is illustrated in Scheme 17, this reaction provides the required bicyclo[3.1.0]hexane ring that is further transformed into the cyclopentylamine precursor. The allylic alcohol 201 is employed as starting material. The critical synthetic step in this synthetic approach are the Claisen rearrangement reaction on 202 via the intermediate 203 to produce 204, the

Scheme 17. Reagents and conditions: (a) PMBOCH₂COOH, DCC, DMAP, CH₂Cl₂, 99%; (b) i. LDA, THF, ii. TMSCl, Et₃N, 91%; (c) i. carbonyldiimidazole, THF, ii. LiCH₂CO₂CH₃, THF, 90%; (d) TsN₃, Et₃N, MeCN, 95%; (e) cupric acetylacetonate, cyclohexane, 52%; (f) NaBH₄, MeOH/CH₂Cl₂, (2:1), 91%; (h) CuSO₄, H₂SO₄ (cat), Me₂CO, 93%; (i) 1.0 N NaOH, MeOH, 100%; (j) i. (PhO)₂PON₃, Et₃N, PhH, ii. 2.0 N NaOH, THF, 64%.

Scheme 18. Reagents and conditions: (a) *p*-TsOH, Me₂CO, 56 °C, 2 h, 57%; (b) TPAP, NMO, 4 Å molecular sieves, CH₂Cl₂, rt, 100%; (c) CH₃P(Ph)₃Br, *n*-BuLi, THF, 0 °C, 30 min, 90%; (d) BH₃.THF, NaBO₃-H₂O, 99%; (e) BnBr, NaH; (f) 1.0 N HCl, MeOH-THF, reflux, 88%; (g) NEt₃, SOCl₂, 0 °C, 5 min, 80%; (h) adenine, NaH, DMF, 18-crown-6, 50%; (i) CS₂, ICH₃, NaH; (j) Et₃B, n-Bu₃SnH; (k) Pd black, HCO₂H, 61%; (l) NaN₃, DMF, 100 °C, 10 h, 81%.

Scheme 19. Reagents and conditions: (a) $Hg(OAc)_2$, AcOH, 20 °C, 12 h; (b) $NaBH(OMe)_3$, CH_2Cl_2 , 20 °C, 24 h, 53% from (+)-226 and from (-)-226; (c) $LiAlH_4$, THF, reflux, 5 h, 87% for (+)-228 and for (-)-228; (d) $(COCl)_2$, DMSO, NEt_3 , CH_2Cl_2 , -50 °C \rightarrow 20 °C, 30 min; (e) $Ph_3P=CHCO_2Et$, EtOH, $PhCO_2H$ (1%), 0 °C \rightarrow 20 °C, 24 h, 75% from 230.

incorporation of two nitrogen atoms by a diazo transfer reaction, and the thermolysis of 206 to produce the rigid bicyclo[3.1.0]hexane template 207, which after three synthetic step is converted into the cyclopentyl amine 211 that following a linear strategy leads to the adenosine derivative 212 locked in the Southern conformation. Another interesting example of conformationally locked carbanucleosides in the N geometry is the preparation of ring-expanded oxetanocin analogues [81,82]. The synthetic strategy followed for the preparation of these carbanucleosides employs a regioselective opening reaction of the cyclic sulfite 220 as key step. The synthesis begins

with the readily available cyclopentanyl alcohol 213, which in turn is prepared from 173 [65]. Then, on reaction either with adenine or azide ion, 220 is transformed into the desired carbocyclic nucleoside precursor 221 or the cyclopentylazide 223 with high diastereoselectivity. The hydroxymethyl group at the C-3' results to be a bad substituent in terms of the antiviral activity [81].

Miscellaneous

Very recently, a simple procedure for the preparation of precursors of both enantiomers of 5'-homo-aristeromycin has

Scheme 20. Reagents and conditions: (a) i. DIBAL-H, Et₂0, -78 °C, 10 min, ii. H₂NCH₂CO₂Et.HCI, MeOH, 2 h; (b) i. C1CO₂Et, DBU, CH₂Cl₂, 2 h, ii. NaOMe, MeOH, 40 min, 41% from 235; (c) i. BzN=C=S, CH₂Cl₂, ii. MeI, DBN, iii. NH₃/MeOH, 90 °C, 16 h, 57%; (d) CF₃CO₂H/H₂O (2: 1, v/v), 50 °C, 3 h, 68% for 231, 68% for 232, 60% for 233; (e) HC(=NH)NH₂.HOAc, EtOH, reflux, 4 days, 81%; (f) HCl (c), MeOH, rt, 2 h, 72%; (g) MsCl, py, rt, 4 h, 76%; (h) Te, Et₃BHLi, THF, 50 °C, 7 h, 65%.

been reported [83]. In fact, (-)-(1R,2R,3S,4R)- and (+)-(1S,2S,3R,4S)-4-hydroxyethylcyclopentane-1,2,3-triol (compounds (-)-228 and (+)-228, respectively), are useful template for the synthesis of unnatural and natural aristeromycin [83]. The advances intermediates are straightforwardly obtained from D-ribose [84] in three steps and D-arabinose in two steps [85] (Scheme 19).

A very promising field in the design of carbanucleosides is the preparation of carbocyclic C-nucleosides, in which the nitrogen atom of the bases bonded to the pseudosugar ring is replaced by a carbon atom [86,87]. Compounds 231-234 emerge as attractive members of this family of drugs. The preparation of this particular class of carbanuclesides is outlined in Scheme 20 starting from the intermediate 235 that is readily prepared from D-ribonolactone [87]. Unfortunately, these drugs are not effective against HIV. Another interesting carbanucleoside is the fluor-containing unnatural isomer of 2',3'-dideoxyadenosine 243. L-

(1'S,3'S)-9-[3-Fluoro-3-(hydroxymethyl)cyclopentan-1-yl] adenine is synthesized in 14 steps from 2,3-isopropyliden-D-glyceraldehyde as chiral source [88] via the known advanced intermediate 244 [89]. The antiviral action of this drug is not reported [88] (Scheme 21).

A novel synthetic approach to access to carbocyclic rings has been developed by Jacobson *et al.* [90] starting from the synthetic intermediate **257**, which is straightforwardly prepared from D-ribonolactone [91]. The critical step in this novel methodology to prepare the well-known ketone **261** is a ring closure metathesis reaction on **259** to produce the cyclopentenyl alcohol **260** as a major product and its diastereoisomer **173** (Scheme 22). Ketone **261** was further transformed into the ring-constrained cyclopentenyl alcohol **213** in order to obtain conformationally rigid carbanucleosides such as **262**. This compound is able to inhibit nucleoside transport by the equilibrative transporters at the low nanomolar level [90]. Conformationally

Scheme 21. Reagents and conditions: (a) LiAlH₄, THF, -40 °C $\rightarrow -35$ °C, 90 min, 95%; (b) NaH, THF, 0 °C \rightarrow rt, 1 h, then BnBr, TBAI, 0 °C to rt, overnight, 74%; (c) O₃, MeOH, -78 °C, 45 min, then Me₂S, 0 °C, 2 h, 50%; (d) (EtO)₂P(O)CH₂CO₂Et/NaHMDS, THF, -78 °C, 1 h, 80%; (e) H₂, 10% Pd/C, cyclohexane, rt, 24 h, 95%; (f) MsCl, Py, CH₂Cl₂, 0 °C \rightarrow rt, 24 h, 85%; (g) NaH, THF, reflux, 16 h, 75%; (h) NaOH/H₂O, EtOH, rt, 5 h, then AcOH, 0 °C, 81%; (i) Pb(OAc)₄, CCl₄, hv, reflux, 15 min, then I₂, CCl₄, hv, reflux, 2 h, 82%; (j) NaHCO₃, 15% (v/v) H₂O/ HMPA, 100 °C, 16 h, 40%; (k) 6-chloropurine, PPh₃, DEAD, rt, 6 h, 80%; (l) NH₃/MeOH, 100 °C, 2.5 h, 83%; (m) TBAF, THF, rt, 30 min, 98%.

Scheme 22. Reagents and conditions: (a) (COĈl)₂, DMSO, THF, -78 °C, then TEA, rt; (b) PPh₃CH₃Br, n-BuLi, THF; (c) TBAF, CH₃CN; (d) (COCl)₂, DMSO, THF, -78 °C, then TEA, rt; (e) CH₂=CHBr, THF, -78 °C, (f) Grubbs catalyst, CH₂Cl₂; (g) MnO₂, CHCl₃.

Scheme 23. Reagents and conditions: (a) MPM-Br/NaH, THF, rt, 6 h, 86%; (b) 0.8% H₂SO₄, MeOH, rt, 10 h, 75%; (c) MeSO₂C1, Et₃N, DMAP, CH₂Cl₂, rt, 1 h; (d) NaI, EtCOMe, reflux, 8 h, 64%; (c) Grubbs catalyst, CH₂Cl₂, rt, 6 h, 80%; (f) 0.4% H₂SO₄, dioxane, reflux, 2 h; (g) NaIO₄, CH₂Cl₂, SiO₂, rt, 1 h, 69%; (h) NaBH₄, MeOH, rt, 1 h, 70%; (i) Im-CO-Im, PhH, reflux, 4 h, 69%; (j) 6-aminopurine, Pd(PPh₃)₄, DMSO, THF (1:1), 45 °C, 2 h, 54%; (k) DDQ, CH₃CN/H₂O, rt, 2 h, 72%.

Scheme 24. Reagents and conditions: (a) 4-methoxy-α-toluenethiol, K₂CO₃, THF, 99%;(b) BH₃.THF, 60%; (c) DIAD, PPh₃, 6-chloropurine, THF; (d) NH₃, MeOH, 110 °C, 71%; (e) TFA, PhOH, reflux, 75%.

constrained adenosine derivatives have been designed as ligands at adenosine receptors (A₁, A_{2A}, A_{2B}, and A₃) and nucleotide receptors (P2) [92, 93].

An elegant synthetic approach for the preparation of a carbovir hydroxylated analogue (263) has been recently published [94]. This compound was synthesized starting from 1,2;5,6-di-O-isopropylidene-α-D-glucofuranose via the intermediate 264 [95]. In this case, a RCM reaction was

employed as a key step in order to form the carbocyclic ring. The preparation of **263** is illustrated in Scheme 23.

5'-Norcarbanucleosides are found to have important biological properties, in which 5'-nor-aristeromycin (271) arises as the main member of this family of nucleoside analogues. For instance, the enantiomer of this compound ((+)-271) exhibits potent action against hepatitis B virus [96]. On the basis of these results, several analogues of this product have been prepared [97]. The enantiomer analogue of

5'-nor-aristeromycin, in which the C-4' is lacking, proves to be 10-fold more potent than (+)-271 against HBV [97]. It is an interesting example the preparation of a mercapto analogue of both enantiomers of 5'-nor-aristeromycin ((-)-272 and (+)-272, respectively [98]. This compounds are prepared from either (-)-159 or (+)-159 to afford the natural or unnatural isomer, respectively. Only (-)-272 was moderately active against vaccinia virus [98] (Scheme 24).

CONCLUSIONS

The essential idea for the design and synthesis of carbocyclic nucleosides is the production of a metabolically stable glycosidic bond not affected by phosphorylases and hydrolases. Moreover, structural conformational variations due to the lack of gauche and anomeric effects should be avoid to a minimum in order to have recognizable nucleosides analogues.

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REFERENCES

- [1] Crimmins, M. T. Tetrahedron 1998, 54, 9229-9272.
- [2] Borthwick, A. D.; Biggadike, K. Tetrahedron 1992, 48, 571-623.
- [3] Agrofolio, L.; Suhas, E.; Farese, A.; Condom, R.; Challand, S. R.; Earl, R. A.; Guedj, R. Tetrahedron 1994, 50, 10611-10670.
- [4] Ichikawa, E.; Kato, K. Curr. Med. Chem. 2001, 8, 385–423.
- [5] Ferrero, M.; Gotor, V. Chem. Rev. 2000, 100, 4319-4347
- [6] Meldgaard, M.; Wengel, J. J. Chem. Soc. Perkin Trans. 2000, 1, 3539-3554.
- [7] Zhu, X.-F. Nucleosides Nucleotides & Nucleic Acids 2000, 19, 651-690.
- [8] Marquez, V. E.; Lim, M. -I. Med. Res. Rev. 1986, 6, 1-40.
- [9] Marquez, V. E. In Advances in Antiviral Drug Design; De Clercq, E. Ed.; JAI Press Inc.: Greenwich, CT, 1996; Vol 2, pp 89-146.
- [10] Mitsunobu, O. Synthesis 1981, 1-28.
- [11] Jenny, T. F.; Previsani, N.; Brenner, S. A. Tetrahedron Lett. 1991, 32, 7029-7032.
- [12] Jenny, T. F.; Horlacher, J.; Previsani, N.; Brenner, S. A. Helv. Chim. Acta 1992, 75, 1944-1954.
- [13] von Itzstein, M.; Mocerino, M. Synth. Commun. 1990, 20, 2049.

- [14] Choo, H.; Chong, Y.; Chu, C. K. Org. Lett. 2001, 3, 1471–1473.
- [15] Crimmins, M. T.; Zuercher, W. J. Org. Lett. 2000, 2, 1065-1067.
- [16] Dixit, D. M.; Leznoff, C. C. J. Chem. Soc. Chem. Commun. 1977, 798–799.
- [17] Trost, B. M.; Kuo, G. -H.; Benneche, T. J. Am. Chem. Soc. 1988, 110, 621-622.
- [18] Trost, B. M.; Li, L.; Guile, S. D. J. Am. Chem. Soc. 1992, 114, 8745-8747.
- [19] Trost, B. M.; Madsen, R.; Guile, S. D.; Brown, B. J. Am. Chem. Soc. 2000, 122, 5947-5956.
- [20] Foster, R. H.; Faulds, D. Drugs 1998, 55, 729-736.
- [21] Vince, R.; Hua, M. J. Med. Chem. 1990, 33, 17-21.
- [22] Crimmins, M. T.; King, B. W.; Zuercher, W. J.; Choy, A. L. J. Org. Chem. 2000, 65, 8499-8509.
- [23] Evans, D. A.; Bartroli, J.; Shih, T. L. J. Am. Chem. Soc. 1981, 103, 2127-2129.
- [24] Evans, D.; Rieger, D. L; Bilodeau, M., T.; Urpi, F. J. Am. Chem. Soc. 1991, 113, 1047-1049.
- [25] Trnka, T. M.; Grubbs, R. H. Acc. Chem. Res. 2001, 34, 18–21.
- [26] Brown, B.; Hegedus., L. S. J. Org. Chem. 2000, 65, 1865– 1872.
- [27] Riches, A. G.; Wernersbach, L. A.; Hegedus, L. S. J. Org. Chem. 1998, 63, 4691-4696.
- [28] Hegedus, L. S.; Bates, R. W.; Söderberg, B. C. J. Am. Chem. Soc. 1991, 113, 923-927.
- [29] Freer, R.; Geen, G. R.; Ramsay, T. W.; Share, A. C.; Slater, G. R.; Smith, N. M. Tetrahedron 2000, 56, 4589-4595.
- [30] Bianco, A.; Celletti, L.; Mazzei, R. A.; Umani, F. Eur. J. Org. Chem. 2001, 1331-1334.
- [31] Vorbrüggen, H.; Höfle, G. Chem. Ber. 1981, 114, 1256– 1258.
- [32] Vorbrüggen, H.; Bennua, B. Chem. Ber. 1981, 114, 1279– 1286.
- [33] Hamilton, C. J.; Roberts, S. M. J. Chem. Soc. Perkin Trans. 1 1999, 1051-1056.
- [34] Hamilton, C. J.; Roberts, S. M.; Shipitsin, A. Chem. Commun. 1998, 1087-1088.
- [35] Chrisp, P.; Clissod, S. Drugs 1991, 41, 102.
- [36] Merlo, V.; Roberts, S. M.; Storer, R.; Bethell, R. J. Chem. Soc. Perkin Trans. 1 1994, 1477.
- [37] Sekiyama, T; Hatsuya, S.; Tanaka, Y.; Uchiyama, M.; Ono, N.; Iwayama, S.; Oikawa, M.; Suzuki, K.; Okunishi, M.; Tsuji, T. J. Med. Chem. 1998, 41, 1284–1298.

- [38] Pirrung, M. C.; Dunlap, S. E.; Trinks, U. P. Helv. Chim. Acta 1989, 72, 1301–1310.
- [39] Rifé, J.; Ortuño, R. M. Org. Lett. 1999, 1, 1221-1223.
- [40] Gauvry, N; Huet, F. Tetrahedron 1999, 55, 1321-1328.
- [41] Mévellec, L.; Huet, F. Tetrahedron Lett. 1995, 36, 7441–7444.
- [42] Verheggen, I.; Van Aerschot, A.; Toppet, S.; Snoeck, R.; Jannsen, G.; Balzarini, J.; De Clercq, E; Herdewijn, P. J. Med. Chem. 1993, 36, 2033–2040.
- [43] Verheggen, I.; Van Aerschot, A.; Van Meervelt, L.; Rozenski, J.; Wiebe, L.; Snoeck, R.; Andrei, G.; Balzarini, J.; Claes, P.; De Clercq, E; Herdewijn, P. J. Med. Chem. 1995, 38, 826–835.
- [44] Maurinsh, Y.; Schraml, J.; De Winter, H.; Blaton, N.; Peeters, O.; Lescrinier, E.; Rozenski, J.; Van Aerschot, A.; De Clercq, E; Busson, R.; Herdewijn, P. J. Org. Chem. 1997, 62, 2861-2871.
- [45] Wang, J.; Herdewijn, P. J. Org. Chem. 1999, 64, 7820–7827.
- [46] Wang, J.; Busson, R.; Blaton, N.; Rozenski, J.; Herdewijn, P. J. Org. Chem. 1998, 63, 3051-3058.
- [47] Wang, J.; Froeyen, M.; Hendrix, C.; Andrei, G.; Snoeck, R.; De Clercq, E; Herdewijn, P. J. Med. Chem. 2000, 43, 736-745.
- [48] Wang, J.; Verbeure, B.; Luyten, I.; Lescrinier, E.; Froeyen, M.; Hendrix, C.; Rosemeyer, H.; Seela, F.; Van Aerschot, A.; Herdewijn, P. J. Am. Chem. Soc. 2000, 122, 8595-8602.
- [49] Vastmans, K.; Pochet, S.; Peys, A.; Kerremans, L.; Van Aerschot, A.; Hendrix, C.; Marlière, P.; Herdewijn, P. *Biochemistry* **2000**, *39*, 12757–12765.
- [50] Jeong, L. S.; Kim, H. O.; Moon, H. R.; Hong, J. H.; Yoo, S. J.; Choi, W. J.; Chun, M. W.; Lee, C. -K. J. Med. Chem. 2001, 44, 806-813.
- [51] Jeong, L. S.; Lee, Y. A.; Moon, H. R.; Chun, M. W. Nucleosides Nucleotides 1998, 17, 1473-1478.
- [52] Choi, W. J.; Park, J. G.; Yoo, S. J.; Kim, H. O.; Moon, H. R.; Chun, M. W.; Jung, Y. H.; Jeong, L. S. J. Org. Chem. 2001, 66, 6490-6494.
- [53] Cohen, N.; Banner, B. L.; Laurenzano, A. J.; Carozza, L. Org. Synth. 1984, 63, 127-139.
- [54] Saenger, W. Principles of Nucleic Acid Structure; Springer-Verlag: New York, 1984; pp 51-104.
- [55] Koole, L. H.; Neidle, S.; Crawford, M. D.; Krayevski, A. A.; Gurskaya, G. V.; Sandström, A.; Wu, J.-C.; Tong, W.; Chattopadhyaya, J. J. Org. Chem. 1991, 56, 6884–6892.
- [56] Yaginuma, S.; Muto, N.; Tsujino, M.; Sudate, Y.; Hayashi, M.; Otani, M. J. Antibiot. 1981, 34, 359–366.
 (b) Isono, K. J. Antibiot. 1988, 41, 1711–1739.
- [57] De Clercq, E. Antimicrob. Agents Chemother. 1985, 28, 84-89.

- [58] De Clercq, E.; Bernaerts, R.; Bregstrom, D. E.; Robins, M. J.; Montgomery, J. A.; Holy, A. Antimicrob. Agents Chemother. 1986, 29, 482-487.
- [59] Kinoshita, K.; Yaginuma, S.; Hayashi, M.; Nakatsu, K. Nucleosides Nucleotides, 1985, 4, 661-668.
- [60] Rodriguez, J. B.; Marquez, V. E.; Nicklaus, M. C.; Barchi, J. J., Jr. Tetrahedron Lett. 1993, 34, 6233-6236.
- [61] Rodriguez, J. B.; Marquez, V. E.; Nicklaus, M. C.; Mitsuya, H.; Barchi, J. J. Jr. J. Med. Chem. 1994, 37, 3389-3399.
- [62] Marquez, V. E.; Ezzitouni, A.; Russ, P.; Siddiqui, M. A.; Ford, H. Jr.; Feldman, R. J.; Mitsuya, H.; George, C.; Barchi, J. J. Jr. J. Am. Chem. Soc. 1998, 120, 2780-2789.
- [63] Siddiqui, M. A.; Ford, H., Jr.; George, C.; Marquez, V. E. Nucleosides Nucleotides 1996, 15, 235-250.
- [64] Marquez, V. E.; Siddiqui, M. A.; Ezzitouni, A.; Russ, P.; Wang, J.; Wagner, R. W.; Mateucci, M. D. J. Med. Chem. 1996, 39, 3739-3747.
- [65] Altmann, K.-H.; Kesselring, R.; Francotte, E.; Rihs, G. *Tetrahedron Lett.* 1994, 35, 2331–2334.
- [66] Altmann, K.-H.; Imwinkelried, R.; Kesselring, R.; Rihs, G. Tetrahedron Lett. 1994, 35, 7265-7268.
- [67] Marquez, V. E.; Russ, P.; Alonso, R.; Siddiqui, M. A.; Shin, K. -J.; George, C.; Nicklaus, M. C.; Dai, F., Ford, H. Jr., Nucleosides Nucleotides 1999, 18, 521-530.
- [68] Jeong, L. S.; Marquez, V. E.; Yuan, C. -S.; Borchardt, R. T. Heterocycles 1995, 41, 2651-2656.
- [69] Ezzitouni, A.; Marquez, V. E. J. Chem. Soc. Perkin Trans. 1 1997, 1073–1078.
- [70] Jeong, L. S.; Buenger, G.; McCormack, J. J.; Cooney D. A.; Hao, Z.; Marquez, V. E. J. Med. Chem. 1998, 41, 2572– 2578.
- [71] Ezzitouni, A.; Russ, P.; Marquez, V. E. J. Org. Chem. 1997, 62, 4870-4873.
- [72] Comin, M. J.; Rodriguez, J. B. Tetrahedron 2000, 56, 4639-4649.
- [73] Rodriguez, J. B. Tetrahedron 1999, 55, 2157-2170.
- [74] Comin, M. J.; Pujol, C. A.; Damonte, E. B.; Rodriguez, J. B. *Nucleosides Nucleotides* 1999, 18, 2219–2231.
- [75] Prota, A.; Vogt, J.; Pilger, B.; Perozzo, R.; Wurth, C.; Marquez, V. E.; Russ, P.; Schultz, G. E.; Folkers, G.; Scapozza, L. Biochemistry 2000, 39, 9597-9603.
- [76] Ford, Jr., H.; Dai, F.; Mu, L.; Siddiqui, M. A.; Nicklaus, M. C.; Anderson, L.; Marquez, V. E.; Barchi, J. J. Jr. Biochemistry 2000, 39, 2581-2592.
- [77] Marquez, V.E.; Russ, P.; Alonso, R.; Siddiqui, M.A. Hernandez, S.; George, C.; Nicklaus, M.C.; Dai, F.; Ford, H. Jr. Helv. Chim. Acta 1999, 82, 2119–2129.
- [78] Wang, P.; Brank, A. S.; Banavali, N. K.; Nicklaus, M. C.; Marquez, V. E.; Christman, J. K.; MacKerell, A. D. Jr. J. Am. Chem. Soc. 2000, 122, 12422-12434.

- [79] Moon, H. R.; Ford, H. Jr.; Marquez, V. E. Org. Lett. 2000, 24, 3793–3796.
- [80] Shin, K. J.; Moon, H. R.; George, C.; Marquez, V. E. J. Org. Chem. 2000, 65, 2172–2178.
- [81] Moon, H. R.; Kim, H. O.; Chun, M. W.; Jeong, L. S.; Marquez, V. E. J. Org. Chem. 1999, 64, 4733-4741.
- [82] Jeong, L. S.; Marquez, V. E. Tetrahedron Lett. 1996, 37, 2353–2356.
- [83] Gallos, J. K.; Dellios, C. C.; Spata, E. E. Eur. J. Org. Chem. 2001, 79–82.
- [84] Gallos, J. K.; Koumbis, A. E.; Xiraphaki, V. P.; Dellios, C. C.; Coutouli-Argyropoulou, E. Tetrahedron 1999, 55, 15167-15180.
- [85] Hall, A.; Meldrum, K. P; Therond, P. R.; Wightman, R. H. Synlett 1997, 123-125.
- [86] Chun, B. K.; Song, G. X.; Chu, C. K. J. Org. Chem. 2001, 66, 4852–4858.
- [87] Chun, B. K.; Chu, C. K. Tetrahedron Lett. 1999, 40, 3309-3312.
- [88] Gumina, G.; Chong, Y.; Choi, Y.; Chu, C. K. Org. Lett. **2000**, 2, 1229–1231.
- [89] Hong, J. H.; Lee, K.; Choi, Y.; Chu, C. K. Tetrahedron Lett. 1998, 39, 3443-3446.

- [90] Lee, K.; Cass, C.; Jacobson, K. A. Org. Lett. 2001, 3, 597– 599.
- [91] Ohira, S.; Sawamoto, T.; Yamato, M. Tetrahedron Lett. 1995, 36, 1537-1540.
- [92] Jacobson, K. A.; Ji, X. -D.; Li, A. H.; Melman, N.; Siddiqui, M. A.; Shin, K. J.; Marquez, V. E.; Ravi, R. J. J. Med. Chem. 2000, 43, 2196-2203.
- [93] Nandanan, E.; Jang, S. Y.; Moro, S.; Kim, H. O.; Siddiqui, M. A.; Russ, P.; Marquez, V. E.; Busson, R.; Herdewijn, P.; Harden, T. K.; Boyer, J. L.; Jacobson, K. A. J. Med. Chem. 2000, 43, 829-842.
- [94] Gurjar, M. K.; Maheshwar, K. J. Org. Chem. 2001, 66, 7552-7554.
- [95] Patra, R.; Bar, N. C.; Roy, A.; Achari, B.; Mandal, S. B. Tetrahedron 1996, 52, 11265-11272.
- [96] Seley, K. L.; Schneller, S. W.; Korba, B. Nucleosides Nucleotides 1997, 16, 2095-2099.
- [97] Seley, K. L.; Schneller, S. W.; Korba, B. J. Med. Chem. 1998, 41, 2168-2170.
- [98] Das, S. R.; Schneller, S. W.; Balzarini, J.; De Clercq, E. Bioorg. Med. Chem. 2002, 10, 457-460