Accepted Manuscript

Otolith edge fingerprints as approach for stock identification of Genidens barbus

Esteban Avigliano, Barbara Maichak de Carvalho, Mathieu Leisen, Rurik Romero, Gonzalo Velasco, Marcelo Vianna, Fernando Barra, Alejandra Vanina Volpedo

PII: S0272-7714(17)30340-2

DOI: 10.1016/j.ecss.2017.06.008

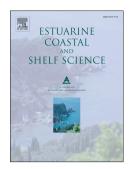
Reference: YECSS 5497

To appear in: Estuarine, Coastal and Shelf Science

Received Date: 3 April 2017
Revised Date: 1 June 2017
Accepted Date: 5 June 2017

Please cite this article as: Avigliano, E., Maichak de Carvalho, B., Leisen, M., Romero, R., Velasco, G., Vianna, M., Barra, F., Volpedo, A.V., Otolith edge fingerprints as approach for stock identification of *Genidens barbus, Estuarine, Coastal and Shelf Science* (2017), doi: 10.1016/j.ecss.2017.06.008.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



- 1 Otolith edge fingerprints as approach for stock identification of Genidens barbus.
- 2 Esteban Avigliano^{1*}, Barbara Maichak de Carvalho², Mathieu Leisen³, Rurik Romero³, Gonzalo
- 3 Velasco⁴, Marcelo Vianna⁵, Fernando Barra³, Alejandra Vanina Volpedo^{1,6}

4

- 5 ¹CONICET- Universidad de Buenos Aires. Instituto de Investigaciones en Producción Animal
- 6 (INPA). Buenos Aires, Argentina. Facultad de Ciencias Veterinarias, Universidad de Buenos Aires
- 7 (UBA), Av. Chorroarín 280 (C1427CWO), Ciudad Autónoma de Buenos Aires, Argentina.
- 8 ²Programa de Pós Graduação em Zoologia, Departamento de Zoologia UFPR, Centro
- 9 Politécnico (19.020), Jardim das Américas, Curitiba, Paraná, Brazil.
- 10 ³Departamento de Geología y Centro de Excelencia en Geotermia de Los Andes (CEGA),
- 11 Universidad de Chile, Plaza Ercilla 803 (8370450), Santiago, Chile.
- ⁴Instituto de Oceanografia, Universidade Federal do Rio Grande (FURG), Av. Italia, Km 8
- 13 (96203-900), Rio Grande, RS, Brazil.
- ⁵Laboratório de Biologia e Tecnologia Pesqueira, Instituto de Biologia, Universidade do Rio de
- 15 Janeiro (UFRJ), Av. Carlos Chagas Filho 373 B1 (21941-599), RJ, Brazil.
- 16 ⁶Centro de Estudios Transdisciplinarios del Agua (CETA-UBA), Facultad de Ciencias
- 17 Veterinarias, Universidad de Buenos Aires (UBA), Av. Chorroarín 280 (C1427CWO), Ciudad
- 18 Autónoma de Buenos Aires, Argentina.
- 19 corresponding author: *estebanavigliano@conicet.gov.ar, Tel/Fax: +54-11-45248484

20

21	Abstract
22	The purpose of this paper is to assess the use of multi-elemental otolith fingerprints as a tool to
23	delimit catfish Genidens barbus fish stocks in four estuaries from the southwestern Atlantic
24	Ocean. Barium:Calcium (Ca), Magnesium:Ca, Manganese:Ca, Sodium:Ca and Strontium:Ca
25	ratios in the otolith edge were determined by LA-ICPMS. PERMANOVA analysis reveal
26	significant differences in the multi-element signatures among estuaries (p=0.0001-0.002).
27	Reclassification rates of quadratic discriminant analysis are high, averaging 89.9 % (78-100%).
28	The new data presented here show that the otolith chemistry is a potential tool for stock
29	identification, and indicates the presence of at least four stocks which should probably be handled
30	independently.
31	
32	Keywords: Catfish; multi-elemental fingerprints; stock delimitation; South-western Atlantic
33	estuaries
34	

35 1. Introduction

36	Among the major fisheries in South America that have strongly declined in recent decades is the
37	catfish Genidens barbus (Lacépède 1803), a species of commercial importance that is distributed
38	in subtropical and temperate zones of the southwestern Atlantic Ocean (López and Bellisio, 1965;
39	Velasco et al., 2007). The biology of catfish is highly complex, due to the presence of different
40	migratory patterns such as cyclic semi-amphidromy, amphidromy, anadromy and freshwater
41	residency (Avigliano et al., 2017b, 2015a). Reproductive events occur in spring and summer in
42	relatively low salinity environments (rivers and estuaries) (Araújo, 1988; Reis, 1986a, 1986b).
43	Parental care by males is observed in this species, where eggs or juveniles are carried in the
44	oropharingeal cavity up to 3 months from the mating area to the external estuary (moderate-high
45	salinity) (Reis, 1986a). Recently, the species was included in the Red List of endangered species
46	in Brazil and several studies are being carried out for the recovery of the fisheries (Avigliano and
47	Volpedo, 2016; Di Dario et al., 2015).
48	The identification of fish stocks is a prerequisite to study the dynamics and structure of the
49	fishery management units. In this respect, the efficiency of the management of a given fishery
50	depends on the correct delimitation of stocks (Cadrin et al., 2013). Several methodologies have
51	been historically used for stock delimitation as meristic, morphometric landmark, parasites, fatty
52	acid profiles, allozymes, mitochondrial DNA, external an internal tags, otolith morphometry and
53	microchemistry (Cadrin et al., 2013).
54	Multi-elemental fingerprints in the otolith edge has contributed to stock identification of several
55	fish species in the world (Avigliano and Volpedo, 2016; Campana, 2013; Tanner et al., 2015).
56	Fish otoliths are apposition structures composed of calcium carbonate deposited in a protein
57	matrix, with small quantities of certain elements such as Ba, Li, Mg, Mn, Na, Sr, and Zn
58	(Campana et al., 1997). These trace elements are acquired by an individual fish during the

59	ontogenetic period and preserved within the otolith microstructure. Hence, multi-elemental
60	fingerprints in the edge may reflect geographic groups or stocks (Campana, 2013; Tanner et al.,
61	2015).
62	The trace element composition of catfish lapillus otolith has proved to be a useful tool as a
63	natural tag and has allowed the identification of nursery areas, population structure and life
64	history (Avigliano et al., 2017b, 2016, 2015a, 2015b). Recently, Avigliano et al. (2015b) have
65	suggested the presence of two fish stocks south of the species known distribution. However, the
66	delimitation of fish stocks remains poorly constrained in the rest of the species distribution area.
67	The purpose of this study was to evaluate the multi-elemental fingerprints of the lapillus otolith
68	edge as a tool to delimit G. barbus fish stocks in the southwestern Atlantic Ocean. Here, we
69	determined the Ba:Ca, Mg:Ca, Mn:Ca, Na:Ca and Sr:Ca ratios of otolith edge by LA-ICPMS in
70	catfish caught in four different estuaries (Guanabara Bay, Paranaguá Bay, Patos Lagoon in
71	Brazil, and Plata River estuary in Argentina). Multi-elemental otolith fingerprints were compared
72	among fish collected from each estuary in order to establish patterns in the data that may be used
73	to identify and evaluate the presence of stocks. This information is important for the sustainable
74	exploitation of the resource, the development of assessment and management models.

75

76

2. Materials and Methods

77 2.1 Sample collection and preparation

Adult catfish were caught in Guanabara Bay (GB), Paranaguá Bay (PB), Patos Lagoon (PL), and Plata River estuary (PR) (Fig. 1) with gillnets, hooks and longlines between November 2010 and May 2015. The total fish length (in cm) was recorded and both *lapilli* otoliths were removed and rinsed with ultrapure water (18.2 MΩ/cm) (Millipore, São Paulo, Brazil).

82	Otoliths (N=46) were weighed using an analytical balance (Sartorius AG ED 2242, Göttingen,
83	Germany), washed with ultrapure water and dried. The left otolith of each pair was embedded in
84	epoxy resin and sectioned transversely through the core to a thickness of 700 µm using a Buehler
85	Isomet low speed saw (Hong Kong, China) equipped with twin diamond edge blade. Only fishes
86	between 8-12 years were used (randomly selected) for analysis to avoid possible effects caused
87	by the age of the fish on the data interpretation (Avigliano et al., 2017a). Mean age±standard
88	deviation (in years) were 10.3±1.75, 10.2±1.56, 8.64±0.92 and 8.73±1.27 for PR (N=15), PL
89	(N=9), PB (N=11) and GB (N=11). Annual periodicity of ring formation was validated by Reis
90	(1986a). Mean total length±standard deviation and range (in cm) were 63.2±7.05 (52.0-74.2),
91	60.7±6.32 (54.7–71.2), 64.9±6.02 (58.0–81.9), 59.2±9.16 (45.9–75.4) for PR, PL, PB and GB.

92

93

2.2 Determination of elements by LA-ICP-MS and data analysis

Elemental concentrations in otolith sections were measured using a 193nm ArF laser ablation 94 system (Photon Machines Analyte G2) coupled to an ICP-MS iCapQ ThermoFisher at the 95 96 Andean Geothermal Center of Excellence (CEGA), Universidad de Chile, Santiago, Chile. The abundances of isotopes ²³Na, ²⁴Mg, ⁴³Ca, ⁵⁵Mn, ⁸⁸Sr, ¹³⁸Ba was determined by laser ablation 97 on fifty µm line-scans performed on the outermost 300 microns of the otolith edge, which 98 represents approximately the last year of life. NIST SRM 612 silicate glass reference material 99 was used as an external standard (Jochum et al., 2011), whereas the USGS synthetic calcium 100 carbonate MACS-3 (Jochum et al., 2012) and silicate glass NIST SRM 610 were analyzed as 101 102 secondary standards. The two external standards measurements allowed us to calculate the 103 precision and the accuracy of the analysis. Precision was less than 10% for all elements analyzed. Accuracy was less than 1%, except for Mg (~13%). This higher value for Mg can be explained by 104 the uncertainty of the Mg concentration in the NIST SRM 612 standard (Jochum et al., 2011). 105

106	Prior to recording transect measurements; the otolith surface was pre-ablated using a spot size of
107	$85~\mu m$ and a scan speed of $30~\mu m/s.$ Then, the ablation scans were performed using a $50~\mu m$ spot
108	size, a scan speed of 10 μ m/s, an energy density of 5 mJ/cm ² and a repetition rate of 10 Hz.
109	Element concentrations are expressed as molar ratios (element:Ca = mmol/mol). Limit of
110	detection (LOD) was calculated from the standard deviation of the blank and normalized to Ca
111	(0.0011, 0.00075, 8.99, 0.067 and 0.028 mmol/mol for Sr:Ca, Ba:Ca, Na:Ca, Mg:Ca and Mn:Ca).
112	Uncertainties were determined for each the analyzed elements (i.e., Na: 5.8%; Ba: 8%; Sr: 8%;
113	Mn: 8.5%; and Mg: 13.1%).
114	Nonparametric statistics were used to compare the elemental ratios between sampling sites
115	because the ratios did not fit the normal distribution and homogeneity of variance (Shapiro-Wilk,
116	p < 0.05; Levene, $p < 0.05$) even after logarithmic transformation. To ensure that differences in
117	fish age did not confound spatial patterns in elemental composition, the effect of age on the
118	elemental ratios was examined using analysis of covariance (ANCOVA) with age as co-variate
119	(Campana, 2013; Longmore et al., 2010). Only the Sr:Ca ratio showed a correlation with age (p <
120	0.05). This age effect was corrected by subtracting the common slope (b = 0.11) in ANCOVA
121	(Campana, 2013; Longmore et al., 2010).
122	Univariate and multivariate statistics were used to evaluate the presence of different stocks
123	(Campana, 2013). Each ratio was compared among sampling sites using Kruskal-Wallis. Because
124	the data did not fit the multi-dimensional non-normality assumptions; permutational multivariate
125	analysis of variance (PERMANOVA) was used to evaluate geographical differences in the multi-
126	elemental fingerprints of the otolith edge. After testing the multicollinearity between variables,
127	quadratic discriminant function analysis (QDA) was used to assess the ability of elemental ratios
128	to sort fish into specific catch areas. The calculation of the expected prior probability
129	classification was based on sample sizes and group numbers (White and Ruttenberg, 2007). A

130	randomization test was used to determine if the classification success rate was significantly
131	different from random data (White and Ruttenberg, 2007). Statistical tests were performed using
132	the Ginkgo 1.7 and SPSS 19 programs.
133	

134

3. Results

Considering all sampling sites, element:Ca levels (mean±standard deviation and range) were 135 0.007 ± 0.01 (0.001-0.03), 0.04 ± 0.03 (0.02-0.2), 0.002 ± 0.003 (0.0003-0.02), 9.9 ± 1.8 (7.0-15.1), 136 4.6±0.9 (2.0-6.1) mmol/mol for Ba:Ca, Mg:Ca, Mn:Ca, Na:Ca and Sr:Ca, respectively (Fig. 2). 137 Ba:Ca ratio was found to be high in PR and PB and low for PL (p=0.0001-0.04). The otolith edge 138 Mg:Ca ratio was high for PR, intermediate for PL and low for GB (P<0.02). Na:Ca ratio was 139 140 higher for PR, low for PB and GB, and intermediate for PL (P<0.04). Sr:Ca was high for PB and low for PL, and intermediate for PR and GB. No significant differences (p>0.05) were found 141 between sites for Mn:Ca ratios. 142 Multivariate methods were highly effective at detecting different otolith fingerprints among sites, 143 144 indicating the existence of four stocks. Specifically, PERMANOVA analysis revealed significant differences in the multi-element signatures of the otolith edge between all sampling sites 145 (p=0.0001-0.002). Reclassification rates of QDA were generally high, averaging 89.9% (Table 146 1). Percentages of correctly classified individuals were 100% for PR, 78% for PL, 82% for PB 147 and 100% GB. These values are significantly different from random data (prior probabilities for 148 groups: 0.32 for PR, 0.20 for PL, 0.24 for PB and 0.24 for GB) (randomization test: p< 0.05). 149 150 Hence, the multi-elemental signatures appear to be a powerful tool to discriminate populations.

151

152

4. Discussion

153	According to Campana (2013), the presence of different fingerprints among groups of fish with
154	comparable age implies different life histories and that the otolith elemental composition can be
155	used as a stock delimitation indicator. Univariate analysis showed significant elemental variations
156	between some estuaries, with the sole exception of the Mn:Ca ratio. These variations were
157	maximized with the multivariate analysis of PERMANOVA and QDA. Significant differences
158	and high classification percentages were observed for every studied site. The two multivariate
159	methods show that the otolith edge chemical signatures are an efficient approach to discriminate
160	catfish stocks among the studied estuaries. Our results also indicate the presence of at least four
161	fish stocks for the species associated with each estuary. In addition, the reclassification rates
162	obtained for PL and PB (≥78%) suggest a relatively low connectivity among nearby estuaries,
163	which is consistent with the chemical signature of the core of adult specimens (Avigliano et al.,
164	2016). However, reclassification rates of 100% have been obtained for the extremes of the
165	distribution (PR and GB populations), indicating that these groups tend to be closed. The
166	presence of relatively closed populations could be linked to the homing behavior of the species
167	suggested by Avigliano et al. (2016).
168	Previous works have suggested the existence of different fish stocks between PL and PR using
169	the Sr:Ca, Ba:Ca and Mg:Ca ratios (Avigliano et al., 2015b). Nevertheless, these authors have
170	used the micromilling technique averaging the last four years of life. In the present work, the LA-
171	ICPMS technique and the possibility to measure several additional trace elements allowed us to
172	increase the percentage of PL classification from 63.6% to 78%, while the percentage obtained
173	for PR was not modified (100%) (Avigliano et al., 2015b).
174	Avigliano et al. (2017b, 2015a) have reported different migratory patterns for the species. The
175	most common is the amphidromous types (amphidromy and semi-amphidromy), although
176	freshwater specimens can reside in PL. Variability in migratory behaviors should be taken into

account for future studies of stock identification, as the inclusion of resident specimens could 177 affect the classification percentages when studying the connectivity of migrating individuals. 178 The incorporation of elements in the otolith is species-dependent and could also be influenced by 179 environmental (salinity, temperature) (Bouchard et al., 2015; Brown and Severin, 2009; Elsdon 180 and Gillanders, 2003; Martin et al., 2004), genetic (Barnes and Gillanders, 2013) and 181 physiological factors (growth rates, metabolic changes) (Kalish, 1991; Radtke and Shafer, 1992; 182 183 Sturrock et al., 2014). The specific factors involved in the incorporation of elements in catfish G. barbus remains largely unknown, as are the possible differential effects among populations. The 184 determination of these factors and their impacts could contribute to a better stock identification 185 and assessment of connectivity between different sites or environments. The fish age can also be 186 a factor, however in this study only fishes between 8-12 years were selected for analysis, and 187 hence the influence of age can be dismissed. So far, only the Sr:Ca and Ba:Ca ratios of catfish 188 189 otolith have been related to factors undoubtedly affect the characteristics of the water and could have printed distinctive salinity (Avigliano et al., 2017b, 2015a, 2015b). The relationship 190 between Sr:Ca and Ba:Ca with salinity has been useful to define the range of habitat of the 191 species (from freshwater to ocean), as well as to describe different migratory patterns (freshwater 192 residence or migration between freshwater, estuary and ocean) (Avigliano et al., 2017b, 2015a). 193 The study area includes four estuaries distributed in a tropical, subtropical and temperate region 194 and there is a decreasing temperature gradient from north-south direction. Estuaries had different 195 climatic and topographic features, depths, salinity ranges, oceanographic patterns and 196 hydrographic dynamics (Avigliano et al., 2016). These signatures in the otoliths, which explains 197 the multi-elemental differences found in this paper. Moreover, it is possible that different 198 oceanographic patterns or climatic regimes (El Niño occurrences) have affected the level of 199

200	closed populations, the metapopulations or the connectivity (Bakun and Broad, 2003; Mann,
201	1993; Selkoe et al., 2007).
202	On the other hand, it has been reported that there could be inter-annual variation in the
203	concentration of some elements of the otolith, probably due to temporary climatic variations,
204	therefore future stock evaluations should be performed by limiting the age and cohorts (Avigliano
205	et al., 2017a).
206	Microchemistry of the otoliths is a useful tool to delimitate stocks and is of potential interest for
207	catfish because it requires fewer resources in comparison to other methodologies such as external
208	tags. Methods such as the geometric morphometry of the body are not recommended because fish
209	presents a relative flaccidity out of the water making difficult the positioning of landmarks.
210	However, a holistic approach is recommended, and it is advisable to apply other approaches such
211	as genetics, otolith morphometry or parasites (Avigliano and Volpedo, 2016; Begg and Waldman,
212	1999; Cadrin et al., 2013).
213	Considering the results obtained, we propose the use of Ba:Ca, Mg:Ca, Mn:Ca, Na:Ca and Sr:Ca
214	ratios to delimit stocks. The Sr:Ca and Ba:Ca ratios are essential because they are directly related
215	to the salinity which varies between the different estuaries (Avigliano et al., 2017b). In addition,
216	other elemental ratios such as the Li:Ca ratio are of potential use to discriminate between
217	different catfish groups (Avigliano et al., 2016).
218	Even though the relative low sampled size used in this work could be a limitation, the results
219	indicate that the methodology used is effective to delimit fish stocks. In addition, the presence of
220	at least four fish stocks suggests that they should probably be handled independently. Therefore,
221	it is recommended to include the determination of multi-elemental signatures in future otoliths
222	studies. This work is a baseline for future projects with a view to the correct delimitation of
223	management units and the subsequent administration of the resource.

224	
225	5. Acknowledgments
226	Authors are indebted to CONICYT-Fondequip instrumentation grant EQM120098, CONICET
227	(PIP 112-20120100543CO), ANPCyT (PICT 2015-1823), Universidad de Buenos Aires
228	(UBACYT 20020150100052BA), Universidade Federal do Rio Grande (FURG), Centros
229	Asociados para el Fortalecimiento de Posgrados Program (CAFP BA 43/13 Brazil/Argentina),
230	Long Term Ecological Program-CNPq (PELD403809/2012-6) and FAPERJ (E26/110.114/2013),
231	E26/ 112.636/2012) and CNPQ (141267/2015-1) for financial support. We also wish to
232	acknowledge the anonymous reviewers for their constructive comments, which helped us to
233	improve the manuscript
234	
235	6. References
236	Araújo, F.G., 1988. Distribuição, abundância relativa e movimentos sazonais de bagres marinhos
236237	Araújo, F.G., 1988. Distribuição, abundância relativa e movimentos sazonais de bagres marinhos (Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5,
237	(Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5,
237 238	(Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5, 509–543. doi:10.1590/S0101-81751988000400002
237238239	(Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5, 509–543. doi:10.1590/S0101-81751988000400002 Avigliano, E., Carvalho, B., Velasco, G., Tripodi, P., Vianna, M., Volpedo, A.V., 2016. Nursery
237238239240	(Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5, 509–543. doi:10.1590/S0101-81751988000400002 Avigliano, E., Carvalho, B., Velasco, G., Tripodi, P., Vianna, M., Volpedo, A.V., 2016. Nursery areas and connectivity of the adults anadromous catfish (<i>Genidens barbus</i>) revealed by
237238239240241	(Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5, 509–543. doi:10.1590/S0101-81751988000400002 Avigliano, E., Carvalho, B., Velasco, G., Tripodi, P., Vianna, M., Volpedo, A.V., 2016. Nursery areas and connectivity of the adults anadromous catfish (<i>Genidens barbus</i>) revealed by otolith core microchemistry in the southwestern Atlantic Ocean. Mar. Freshw. Res.
237238239240241242	(Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5, 509–543. doi:10.1590/S0101-81751988000400002 Avigliano, E., Carvalho, B., Velasco, G., Tripodi, P., Vianna, M., Volpedo, A.V., 2016. Nursery areas and connectivity of the adults anadromous catfish (<i>Genidens barbus</i>) revealed by otolith core microchemistry in the southwestern Atlantic Ocean. Mar. Freshw. Res. 10.1071/MF16058. doi:10.1071/MF16058
237238239240241242243	(Siluriformes, Ariidae) no estuário da Lagoa dos Patos (RS), Brasil. Rev. Bras. Zool. 5, 509–543. doi:10.1590/S0101-81751988000400002 Avigliano, E., Carvalho, B., Velasco, G., Tripodi, P., Vianna, M., Volpedo, A.V., 2016. Nursery areas and connectivity of the adults anadromous catfish (<i>Genidens barbus</i>) revealed by otolith core microchemistry in the southwestern Atlantic Ocean. Mar. Freshw. Res. 10.1071/MF16058. doi:10.1071/MF16058 Avigliano, E., Carvalho, B., Velasco, G., Tripodi, P., Volpedo, A.V., 2017a. Inter-annual

247

248	and freshwater residency, typical behaviors in <i>Genidens barbus</i> inferred by otolith
249	chemistry. Fish. Res. 193, 184–194. doi:10.1016/j.fishres.2017.04.011
250	Avigliano, E., Velasco, G., Volpedo, A. V, 2015a. Assessing the use of two southwestern
251	Atlantic estuaries by different life cycle stages of the anadromous catfish Genidens barbus
252	(Lacépède, 1803) as revealed by Sr:Ca and Ba:Ca ratios in otoliths. J. Appl. Ichthyol. 31,
253	740–743. doi:10.1111/jai.12766
254	Avigliano, E., Velasco, G., Volpedo, A. V, 2015b. Use of lapillus otolith microchemistry as an
255	indicator of the habitat of Genidens barbus from different estuarine environments in the
256	southwestern Atlantic Ocean. Environ. Biol. Fishes 98, 1623–1632. doi:10.1007/s10641-
257	015-0387-3
258	Avigliano, E., Volpedo, A. V., 2016. A review of the application of otolith microchemistry
259	toward the study of Latin American fishes. Rev. Fish. Sci. Aquac. 24, 369-384.
260	doi:10.1080/23308249.2016.1202189
261	Bakun, A., Broad, K., 2003. Environmental "loopholes" and fish population dynamics:
262	Comparative pattern recognition with focus on El Niño effects in the Pacific, in: Fisheries
263	Oceanography. pp. 458–473. doi:10.1046/j.1365-2419.2003.00258.x
264	Barnes, T.C., Gillanders, B.M., 2013. Combined effects of extrinsic and intrinsic factors on
265	otolith chemistry: Implications for environmental reconstructions. Can. J. Fish. Aquat. Sci.
266	70, 1159–1166. doi:10.1139/cjfas-2012-0442
267	Begg, G.A., Waldman, J.R., 1999. An holistic approach to fish stock identification. Fish. Res. 43
268	35–44. doi:DOI: 10.1016/S0165-7836(99)00065-X
269	Bouchard, C., Thorrold, S.R., Fortier, L., 2015. Spatial segregation, dispersion and migration in
270	early stages of polar cod Boreogadus saida revealed by otolith chemistry. Mar. Biol. 162,
271	855–868. doi:10.1007/s00227-015-2629-5

272 Brown, R.J., Severin, K.P., 2009. Otolith chemistry analyses indicate that water Sr:Ca is the primary factor influencing otolith Sr:Ca for freshwater and diadromous fish but not for 273 marine fish. Can. J. Fish. Aquat. Sci. 66, 1790–1808. doi:10.1139/F09-112 274 Cadrin, S.X., Karr, L.A., Mariani, S., 2013. Stock identification methods: an overview, in: 275 Cadrin, S.X., Kerr, L.A., Mariani, S. (Eds.), Stock Identification Methods. Applications in 276 277 Fishery Science. pp. 3–6. Campana, S.E., 2013. Otolirh elemental as a natural marker of fish stocks, in: Cadrin, S.X., Kerr, 278 L.A., Mariani, S. (Eds.), Stock Identification Methods: Applications in Fishery Science: 279 Second Edition. pp. 227–245. doi:10.1016/B978-0-12-397003-9.00011-4 280 Campana, S.E., Thorrold, S.R., Jones, C.M., Gunther, D., Tubrett, M., Longerich, H., Jackson, S., 281 Halden, N.M., Kalish, J.M., Piccoli, P., de Pontual, H., Troadec, H., Panfili, J., Secor, D.H., 282 Severin, K.P., Sie, S.H., Thresher, R., Teesdale, W.J., Campbell, J.L., 1997. Comparison of 283 284 accuracy, precision, and sensitivity in elemental assays of fish otoliths using the electron microprobe, proton-induced X-ray emission, and laser ablation inductively coupled plasma 285 mass spectrometry. Can. J. Fish. Aquat. Sci. 54, 2068–2079. doi:10.1139/cjfas-54-9-2068 286 Di Dario, F. Di, Alves, C.B.M., Boos, H., Frédou, F.L., Lessa, R.P.T., Mincarone, M.M., 287 Pinheiro, M.A.A., Polaz, C.N.M., Reis, R.E., Rocha, L.A., Santana, F.M., Santos, R.A., 288 Santos, S.B., Vianna, M., Vieira, F., 2015. A better way forward for Brazil's fisheries. 289 290 Science (80-.). 363, 1079–1079. Elsdon, T.S., Gillanders, B.M., 2003. Relationship between water and otolith elemental 291 concentrations in juvenile black bream Acanthopagrus butcheri. Mar. Ecol. Prog. Ser. 260, 292 263-272. doi:10.3354/meps260263 293 Jochum, K.P., Scholz, D., Stoll, B., Weis, U., Wilson, S.A., Yang, Q., Schwalb, A., Börner, N., 294 Jacob, D.E., Andreae, M.O., 2012. Accurate trace element analysis of speleothems and 295

296 biogenic calcium carbonates by LA-ICP-MS. Chem. Geol. 318–319, 31–44. doi:10.1016/j.chemgeo.2012.05.009 297 Jochum, K.P., Weis, U., Stoll, B., Kuzmin, D., Yang, Q., Raczek, I., Jacob, D.E., Stracke, A., 298 Birbaum, K., Frick, D.A., Günther, D., Enzweiler, J., 2011. Determination of reference 299 values for NIST SRM 610-617 glasses following ISO guidelines. Geostand. Geoanalytical 300 Res. 35, 397–429. doi:10.1111/j.1751-908X.2011.00120.x 301 Kalish, J.M., 1991. Determinants of otolith chemistry: seasonal variation in the composition of 302 blood plasma, endolymph and otoliths of bearded rock cod *Pseudophycis barbatus*. Mar. 303 Ecol. Prog. Ser. 74, 137–159. doi:10.3354/meps075137 304 Longmore, C., Fogarty, K., Neat, F., Brophy, D., Trueman, C., Milton, A., Mariani, S., 2010. A 305 comparison of otolith microchemistry and otolith shape analysis for the study of spatial 306 variation in a deep-sea teleost, Coryphaenoides rupestris. Environ. Biol. Fishes 89, 591– 307 308 605. doi:10.1007/s10641-010-9674-1 López, R., Bellisio, N., 1965. Contribución al conocimiento del *Tachysurus barbus* (Lacepede), 309 bagre del mar argentino (Pisces. Ariidae), in: Anais Do Segundo Congreso Latino-310 Americano de Zoología. Sao Paulo, Brazil, pp. 145–153. 311 Mann, K.H., 1993. Physical oceanography, food chains, and fish stocks: a review. ICES J. Mar. 312 Sci. J. du Cons. doi:10.1006/jmsc.1993.1013 313 314 Martin, G.B., Thorrold, S.R., Jones, C.M., 2004. Temperature and salinity effects on strontium incorporation in otoliths of larval spot (Leiostomus xanthurus). Can. J. Fish. Aquat. Sci. 61, 315 34-42. doi:10.1139/F03-143 316 Radtke, R., Shafer, D., 1992. Environmental sensitivity of fish Otolith Microchemistry. Mar. 317 Freshw. Res. doi:10.1071/MF9920935 318 Reis, E.G., 1986a. Reproduction and feeding habits of the marine catfish, Netuma barba 319

320	(Siluriformes, Ariidae), in the estuary of the Patos Lagoon (Brazil). Atlantica 8, 35–55.
321	Reis, E.G., 1986b. Age and growth of the marine catfish, Netuma barba (Siluriformes, Ariidae),
322	in the estuary of the Patos Lagoon (Brasil). Fish. Bull. 84, 679-686.
323	Selkoe, K.A., Vogel, A., Gaines, S.D., 2007. Effects of ephemeral circulation on recruitment and
324	connectivity of nearshore fish populations spanning Southern and Baja California. Mar.
325	Ecol. Prog. Ser. 351, 209–220. doi:10.3354/meps07157
326	Sturrock, A.M., Trueman, C.N., Milton, J.A., Waring, C.P., Cooper, M.J., Hunter, E., 2014.
327	Physiological influences can outweigh environmental signals in otolith microchemistry
328	research. Mar. Ecol. Prog. Ser. 500, 245–264. doi:10.3354/meps10699
329	Tanner, S.E., Reis-Santos, P., Cabral, H.N., 2015. Otolith chemistry in stock delineation: A brief
330	overview, current challenges and future prospects. Fish. Res. 173, 206-213.
331	doi:10.1016/j.fishres.2015.07.019
332	Velasco, G., Reis, E.G., Vieira, J.P., 2007. Calculating growth parameters of Genidens barbus
333	(Siluriformes, Ariidae) using length composition and age data. J. Appl. Ichthyol. 23, 64-69.
334	doi:10.1111/j.1439-0426.2006.00793.x
335	White, J., Ruttenberg, B., 2007. Discriminant function analysis in marine ecology: some
336	oversights and their solutions. Mar. Ecol. Prog. Ser. 329, 301–305. doi:10.3354/meps329301
337	
338	

339	Figure 1: Fishing sites of the <i>Genidens barbus</i> (arrows).
340	Figure 2: Mean \pm SE elemental ratio (mmol/mol) in otolith edge from different sampling
341	locations. Different letters indicate statistically significant differences.
342	
343	

Table 1: Confusion matrix of quadratic discriminant analyses based on multi-elemental signature

(Ba:Ca, Mg:Ca, Mn:Ca, Na:Ca and Sr:Ca) of otolith edge. PR, Plata River; PL, Patos Lagoon;

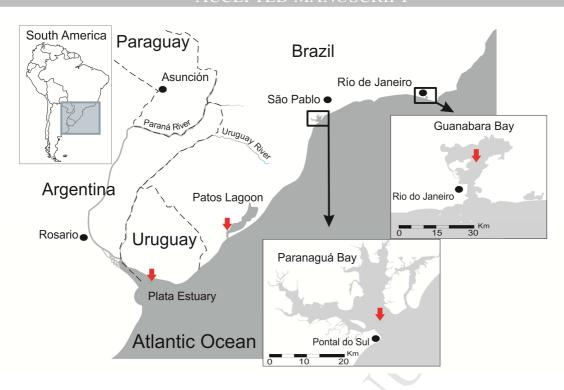
PB, Paranagua Bay and GB, Guanabara Bay. Percentage of correctly reclassified individuals are

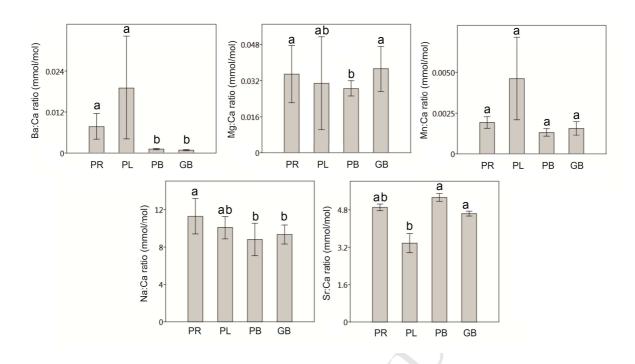
indicated in bold numbers.

348

	PR	PL	PB	GB
PR	100	0	0	0
PL	22	78	0	0
PB	0	0	82	18
GB	0	0	0	100

349





Otolith microchemisty is a potential tool for stock identification.

Results suggest the presence of at least 4 fish stocks of catfish.

High percentages of classification suggest low connectivity.

The stocks should be managed as separate groups.

