## NATURAL PRODUCT COMMUNICATIONS

An International Journal for Communications and Reviews Covering all Aspects of Natural Products Research



NPC-SILAE: Special Issue Volume 6. Issue 7. Pages 925-1054. 2011 ISSN 1934-578X (printed); ISSN 1555-9475 (online) www.naturalproduct.us



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# NPC Natural Product Communications

# Validation of the Ethnopharmacological Use of *Polygonum persicaria* for its Antifungal Properties

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#### Received: November 13<sup>th</sup>, 2010; Accepted: March 16<sup>th</sup>, 2011

*Polygonum* L. genus (Polygonaceae) is represented in Argentina by 21 species and some of them have been used in the traditional medicine of our country to treat affections related with fungal infections, such as skin ailments and vaginal diseases. With the aim of contributing to the correct ethnopharmacological use of this genus, in the present work we describe the antifungal properties of *P. persicaria* (species not studied up to now) and the bio-guided isolation of the main active compounds. Results showed that dichloromethane extracts was the most active with MICs (Minimun Inhibitory Concentrations) between  $31.2 - 1000 \mu g/mL$ , validating the ethnopharmacological use of *P. persicaria* to treat affections related with fungal infections in the Argentinean traditional medicine.

Keywords: Validation, Ethnopharmacological use, Polygonum, Antifungal activity.

Since the early 1980s, fungal infections have emerged as major causes of morbi-mortality, mainly among immunocompromised patients. The majority of deaths were associated with species of Candida, Aspergillus and Cryptococcus [1]. Instead, dermatophytes such as Trichophyton and Microsporum spp. produce superficial infections (tineas) which are usually not threatening but dramatically diminish the quality of life of human beings [2]. Although it appears to be an array of antifungal agents (polyenes, azoles, allylamines and the recent echinocandins) there are, in fact, few therapeutic options. Decreased susceptibilities of yeasts to the currently available antifungal agents [3] added to the increase in the number of reported cases of resistance [4], have led to a general consensus that new efforts for detecting novel antifungal entities remain a priority. In this context, the study of plants with history of ethnopharmacological use for ailments related to fungal infections, can serve two goals: validation of the use of traditional medicines and finding new leads [5].

*Polygonum* L. genus (Polygonaceae) is represented in Argentina by 21 species and some of them have been used to treat affections related with fungal infections, such as skin ailments and vaginal diseases [6]. Previous studies of this genus reported that *P. punctatum* possessed antifungal properties against yeasts and dermatophytes [7]. With the aim of contributing to the correct ethnopharmacological use of this genus, in a previous work we described the antifungal properties of *P. acuminatum* [8] and in this work we describe those of *P. persicaria* (species not studied up to now) and the bio-guided isolation



Figure 1: A) Sesquiterpene [polygodial (1) and isopolygodial (2)] and B) flavonoids [pinostrobin (3), flavokawin B (4) and cardamonin (5)] isolated from *P. persicaria* DCM extract.

of two sesquiterpene dialdehydes: polygodial (1), isopolygodial (2), and three flavonoids: pinostrobin (3), flavokawin B (4) and cardamonin (5) (Figure 1). Compounds 1 and 2 were previously isolated from *Drymis spp.* [9,10], *P. punctatum* [7] and *P. acuminatum* [8], while compounds 3-5 were previously isolated from *Boesenbergia pandurata*, *Myrica pensilvanica*, *P. ferrugineum* and *Piper spp* [11-13].

Compounds 1-5 were evaluated for their antifungal activities with the microbroth dilution assay recommended by the Clinical and Laboratory Standards Institutes (CLSI)

Species	Extract	Antifungal activity (MICs in µg/mL)									
		Ca	Sc	Cn	Afu	Afl	An	Mg	Tr	Tm	
P. persicaria Standards drugs	Hex	Ι	Ι	1000	Ι	Ι	Ι	1000	500	1000	
	DCM	1000	500	500	1000	1000	1000	125	62.5	31.2	
	EtOAc	1000	1000	1000	Ι	Ι	Ι	1000	500	1000	
	MeOH	Ι	Ι	Ι	Ι	Ι	Ι	Ι	Ι	Ι	
	Ketoconazole	0.50	0.50	0.25	0.12	0.50	0.25	0.04	0.02	0.02	
	Amphotericin	1.00	0.50	0.25	0.50	0.50	0.50	0.12	0.07	0.07	
	Terbinafine	-	-	-	-	-	-	0.04	0.01	0.04	

Table 1: Antifungal activity (MICs in µg/mL) of P. persicaria extracts.

Ca: Candida albicans ATCC 10231; Sc: Saccharomyces cerevisiae ATCC 9763; Cn: Cryptococcus neoformans ATCC 32264; Afu: Aspergillus fumigatus ATCC 26934; Afl: Aspergillus flavus ATCC 9170; An: Aspergillus niger ATCC 9029; Mg: Microsporum gypseum C 115; Tr: Trichophyton rubrum C 113; Tm: Trichophyton mentagrophytes ATCC 9972. I: Inactive = MIC > 1000  $\mu$ g/mL.

Table 2: Antifungal activity (MICs in µg/mL) of compounds isolated from P. persicaria.

Compounds	Antifungal activity (MICs in µg/mL)									
-		Ca	Sc	Cn	Afu	Afl	An	Mg	Tr	Tm
1		3.90	15.6	7.8	250	250	250	62.5	7.8	7.8
2		250	125	125	250	250	250	62.5	62.5	31.2
3		250	250	250	125	125	250	62.5	62.5	62.5
4		Ι	Ι	250	250	125	250	125	125	125
5		250	250	250	250	250	250	62.5	15.6	15.6
Standards drugs	Ketoconazole	0.50	0.50	0.25	0.12	0.50	0.25	0.04	0.02	0.02
-	Amphotericin	1.00	0.50	0.25	0.50	0.50	0.50	0.12	0.07	0.07
	Terbinafine	-	-	-	-	-	-	0.04	0.01	0.04

Ca: Candida albicans ATCC 10231; Sc: Saccharomyces cerevisiae ATCC 9763; Cn: Cryptococcus neoformans ATCC 32264; Afu: Aspergillus fumigatus ATCC 26934; Afl: Aspergillus flavus ATCC 9170; An: Aspergillus niger ATCC 9029; Mg: Microsporum gypseum C 115; Tr: Trichophyton rubrum C 113; Tm: Trichophyton mentagrophytes ATCC 9972. I: inactive = MIC > 250  $\mu$ g/mL).

[14] and results are shown in Table 2. They were active against yeasts, *Aspergillus spp.* and dermatophytes with MICs between  $3.90 - 250 \mu g/mL$ 

As it can be observed in Table 2, the five compounds isolated from *P. persicaria*, drimanes as well as flavonoids, all showed antifungal activity. Among them, polygodial (1) showed the best activity against yeasts and dermatophytes with MICs between 3.9 to 62.5  $\mu$ g/mL and it was almost inactive against species of *Aspergillus* genus. Its epimer, isopolygodial (2), showed a lower antifungal activity (MICs between 31.2 to 250  $\mu$ g/mL), suggesting that the C-9 configuration plays an important role in the antifungal activity, as we have been found in a previous paper [8].

Regarding flavonoids 3-5, there is not a clear difference among the antifungal activities of them against yeasts and *Aspergillus* spp. Nevertheless, chalcone 5 showed a high antifungal activity against *T. rubrum* and *T. mentagrophytes* with MICs = 15.6 µg/mL, eight times higher than the activity showed by chalcone 4 against the same strains. This striking difference in activity against *Trichophyton* spp. could be attributed to the phenolic OH present in compound 5 which is absent in 4.

These results show that the antifungal activity of *P. persicaria* could be attributed to polygodial but it is clear that the rest of the isolated compounds could contribute to the antifungal behavior of this traditional used species. In addition, these results validate the ethnopharmacological use of *P. persicaria* to treat affections related to fungal infections in the Argentinean traditional medicine and add a new evidence that the ethnopharmacological approach is

useful in guiding the discovery of antifungal compounds against dermatophytes, as it was demonstrated in a recent survey among seven Latinaoamerican countries [15].

#### Experimental

Extracts preparation and compounds isolation: Air-dried aerial parts of each species (100 g) were powdered and successively macerated  $(3 \times 24 \text{ h each})$  with Hexane (Hex), dichloromethane (DCM), ethyl acetate (EtOAc) and methanol (MeOH) with mechanical stirring to obtain the corresponding extracts, after filtration and evaporation. Bioassay-guided fractionation of DCM extract allowed us to isolate the compounds responsible for the antifungal activity. 1.1 g of P. persicaria DCM extract were submitted to column chromatography using mixtures of Hex: AcOEt in increasing polarity as elution solvents. We obtained 10 fractions; three of them were actives (fractions 6-8). From 150 mg of fraction 6, by repeated column chromatography, we obtained 55 and 30 mg of compounds 1 and 2 respectively. From 170 mg of fraction 7, by repeated column chromatography, we obtained 50, 46 and 25 mg of compounds 3, 4 and 5 respectively. Additionally, from 70 mg of fraction 8, we obtained 10 mg of compound 5. All the compounds were characterized by UV-visible, IR, <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopy.

Antifungal assay: For the antifungal evaluation, strains from the American Type Culture Collection (ATCC, Rockville, MD, USA) and Centro de Referencia en Micología, CEREMIC [C, Faculty of Biochemical and Pharmaceutical Sciences, Suipacha 531 (2000)-Rosario, Argentina] were used: Candida albicans (Ca) ATCC 10231, Saccharomyces cerevisiae (Sc) ATCC 9763, Cryptococcus neoformans (Cn) ATCC 32264, Aspergillus flavus (Afl) ATCC 9170, Aspergillus fumigatus (Afu) ATTC 26934, Aspergillus niger (An) ATCC 9029, Trichophyton rubrum (Tr) C 110, Trichophyton mentagrophytes (Tm) ATCC 9972 and Microsporum gypseum (Mg) C 115. Strains were grown on Sabouraudchloramphenicol agar slants for 48 h at 30 °C, maintained on slopes of Sabouraud-dextrose agar (SDA, Oxoid) and subcultured every 15 days to prevent pleomorphic transformations. Inocula of cell or spore suspensions were obtained and quantified following reported procedures (CLSI).[14]

Minimum Inhibitory Concentration (MIC) of each extract or compound was determined by using broth microdilution Natural Product Communications Vol. 6 (7) 2011 933

techniques according to the guidelines of CLSI for yeasts: document M27-A2 and for filamentous fungi, M38A. For the assay, stock solutions of extracts or pure compounds (100  $\mu$ L) were two-fold diluted with the culture medium. A volumen of 100  $\mu$ L of inoculum suspension [adjusted to 1–5 × 10<sup>4</sup> cells/spores as Colony Forming Units (CFU/mL)] was added to each well with the exception of the sterility control where sterile water was added to the well instead. Ketoconazole (Sigma Chem. Co., St. Louis, MO), Terbinafine (Novartis) and Amphotericin B (Sigma) were used as positive controls.

Acknowledgments - CONICET, ANPCyT, UNR, ERASMUS MUNDUS, UNIBO.

#### References

- [1] Pfaller M, Diekema D. (2007) Epidemiology of invasive candidiasis: a persistent public health problem. *Clinical Microbiology Reviews*, 20, 133-163
- [2] Weitzman I, Summerbell R. (1995) The dermatophytes. *Clinical Microbiology Reviews*, 8, 240-259
- Hsueh P, Lau Y, Chuang Y, Wan J, Huang W, Shyr J, Yan J, Yu K, Wu J, Ko W, Yang Y, Liu Y, Teng L, Liu Ch, Luh K. (2005) Antifungal susceptibilities of clinical isolates of *Candida* species, *Cryptococcus neoformans*, and *Aspergillus* species from Taiwan: Surveillance of multicenter antimicrobial resistance on Taiwan program data from 2003. *Antimicrobial Agents and Chemotherapy*, 49, 512-517
- [4] White T, Holleman S, Dy F, Mirels L, Stevens D. (2002) Resistance mechanisms in clinical isolates of *Candida albicans*. *Antimicrobial Agents and Chemotherapy*, 46, 1704-1713
- [5] Verpoorte R. (2000) Pharmacognosy in the new millennium: leadfinding and biotechnology. *Journal of Pharmacy and Pharmacology*, 52, 253-262
- [6] Del Vitto L, Petenatti E, Petenatti M. (2003) Materia Medica Vegetal. Plantas medicinales nativas y exóticas empleadas en fitomedicinas, homeopáticos, galénicos. *Servicio Técnico Herbario UNSL*, San Luis, pp. 1-63
- [7] De Almeida Alves T, Lacerda Ribeiro F, Kloos H, Zani C. (2001) Polygodial, the fungitoxic component from the Brazilian medicinal plant *Polygonum punctatum. Memórias do Instituto Oswaldo Cruz*, 96, 831-833
- [8] Derita M, Leiva M, Zacchino S. (2009) Influence of plant part, season of collection and content of the main active constituent, on the antifungal properties of *Polygonum acuminatum* Kunth. *Journal of Ethnopharmacology*, 124, 377-383
- [9] Cechinel Filho V, Schlemper V, Santos A, Pinheiro T, Yunes R, Mendes G, Calixto J, Delle Monache F. (**1998**) Isolation and identification of active compounds from *Drymis winteri* barks. *Journal of Ethnopharmacology*, **62**, 223-227
- [10] Muñoz-Concha D, Vogel H, Yunes R, Razmilic I, Bresciani L, Malheiros A. (2007) Presence of polygodial and drimenol in *Drymis* populations from Chile. *Biochemical Systematics and Ecology*, 35, 434-438
- [11] Hodgetts K. (2001) Approaches to 2-substituted chroman-4-ones: synthesis of (-)-pinostrobin. *Tetrahedron Letters*, 42, 3763-3766.
- [12] Burke B, Nair M. (1986) Phenylpropene, benzoic acid and flavonoids derivatives from fruits of Jamaican *Piper* species. *Phytochemistry*, 25, 1427-1430
- [13] López S, González Sierra M, Gattuso S, Furlán R, Zacchino S. (2006) An unusual homoisoflavanone and a structurally-related dihydrochalcone from *Polygonum ferrugineum*. *Phytochemistry*, **67**, 2152-2157
- [14] CLSI (Clinical and Laboratory Standards Institute). (2002) Methods M 27-A2, Vol. 22 (15): 1-29 and M 38-A, Vol. 22 (16): 1-27. Wayne Ed.
- [15] Svetaz L, Zuljan F, Derita M, Petenatti E, Tamayo G, Cáceres A, Cechinel Filho V, Giménez A, Pinzón R, Zacchino S, Gupta M. (2010) Value of the ethnomedical information for the discovery of plants with antifungal properties. A survey among seven Latin American countries. *Journal of Ethnopharmacology*, 127, 137-158

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