

Ticks (Acari: Ixodidae) identified from prey–predator interactions via fecal analysis of Brazilian wild carnivores

Thiago F. Martins · Thaís R. Diniz-Reis · Gustavo S. Libardi ·
Alexandre R. Percequillo · Luciano M. Verdade · Eliana R. Matushima ·
Marcelo B. Labruna

Received: 1 September 2014 / Accepted: 8 February 2015
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Abstract Between July 2008 and May 2010, we conducted a trophic study on 12 Brazilian wild carnivore species through their faecal analysis in a silvicultural landscape at Angatuba municipality, southern São Paulo state. Predator faeces was identified by morphology, predator hair, and surrounding tracks; prey remnants within faeces were used for morphological identification of the prey. Among the recovered ectoparasites, there were 89 specimens of six tick species in 21 (4.0 %) out of 523 analysed samples. Ticks were identified to species level, based on external morphological characters, as following: adults of *Amblyomma ovale* and *Amblyomma sculptum*; nymphs of *Amblyomma brasiliense*, *Amblyomma calcaratum*, *Amblyomma dubitatum*, *A. ovale*, and *Ixodes schulzei*; and larvae of *Amblyomma* sp. and *Ixodes* sp. Generally, the recovered immature ticks were associated

T. F. Martins (✉) · M. B. Labruna
Departamento de Medicina Veterinária Preventiva e Saúde Animal, Faculdade de Medicina Veterinária e Zootecnia, Universidade de São Paulo, São Paulo, SP 05508-270, Brazil
e-mail: thiagodogo@hotmail.com

M. B. Labruna
e-mail: labruna@usp.br

T. R. Diniz-Reis · L. M. Verdade
Centro de Energia Nuclear na Agricultura, Universidade de São Paulo, Piracicaba, SP, Brazil
e-mail: thaisdinizreis@gmail.com

L. M. Verdade
e-mail: lmverdade@usp.br

G. S. Libardi · A. R. Percequillo
Escola Superior de Agricultura “Luiz de Queiroz”, Universidade de São Paulo, Piracicaba, SP, Brazil
e-mail: gslibardi@gmail.com

A. R. Percequillo
e-mail: percequillo@usp.br

E. R. Matushima
Departamento de Patologia, Faculdade de Medicina Veterinária e Zootecnia, Universidade de São Paulo, São Paulo, SP, Brazil
e-mail: ermatush@usp.br

with consumed prey (small birds or small mammals), whereas adults were associated with the predator itself, ingested during its self-grooming. Our data show that faeces is an additional information source on ticks in Brazil and which may provide information on ectoparasite-predator-prey interactions.

Keywords Ectoparasite · Ixodidae · Faeces · Carnivora · Brazil

Introduction

Ticks are obligate ectoparasites of amphibians, reptiles, birds, and mammals. They are important vectors of pathogenic virus, bacteria, protozoa, and nematodes to animals and humans; therefore ticks are of major medical and veterinary concern worldwide (Jongejan and Uilenberg 2004). The Brazilian tick fauna is currently composed of 66 species—45 Ixodidae and 21 Argasidae (Martins et al. 2014; Nava et al. 2014). At least 21 of those, mostly within the genus *Amblyomma* (15 species), have been reported on wild Carnivora hosts native to Brazil (Labruna et al. 2005). Brazilian native Carnivora are divided into five families: Felidae (eight species), Canidae (six species), Mustelidae (six species), Procyonidae (four species), and Mephitidae (two species) (Paglia et al. 2012). Although tick species that parasitize domestic animals have been well studied, knowledge on the natural history of ticks parasitizing wildlife is still incipient for most of the South American tick fauna.

Collecting ticks directly from hosts, is time and money consuming, and laborious for elusive and low density hosts, such as wild carnivores. Rosalino et al. (2007) reported an alternative approach based on the analysis of host faecal samples collected in the environment. Whereas such analysis is a common tool in studies on carnivores' diet, including identification of prey species (Reynolds and Aebischer 1991; Crawshaw Júnior 2006), the study of Rosalino et al. (2007) successfully applied the scat analysis of two European wild carnivores (*Vulpes vulpes* and *Genetta genetta*) for determination of host-tick interactions, with 276 ticks found inside 940 scat samples. In this context, the present study aimed at reporting the occurrence of different tick species that were identified at prey-predator interactions through faecal analysis of some free-ranging wild carnivores in Brazil.

Materials and methods

Between July 2008 and May 2010, we conducted a study on the diet of 12 Brazilian wild carnivores in a silvicultural landscape [Fazenda Três Lagoas (23°22'0''S; 48°28'0''W, 3242 ha) and Fazenda Arca (23°20'0''S; 48°27'30''W, 1123 ha)] in the municipality of Angatuba, state of São Paulo, southeastern Brazil. The landscape of this region is a mosaic of native vegetation remnants (Cerrado savannah and semideciduous forest) merged with an agro-forestry matrix, dominated by exotic pastures and *Eucalyptus* plantations (Athayde 2012). Its climate is subtropical (Cwa in Köppen climate classification) with mean temperatures ranging from 17 to 22 °C along the year (EECFI, 23°10'00''S, 48°40'00''O).

Carnivore faecal samples were collected through all landscape elements with the aid of a motorcycle and identified to the most precise taxonomic level by morphology (Kruuk 1978; Lloyd 1980; Chame 2003), predator hair structures (Teerink 1991; Quadros and Monteiro-Filho 2006) and surrounding footprints (Becker and Dalponte 1991). In the

laboratory, faecal samples were washed through a 1 mm sieve, oven-dried, and non-digested prey remnants (e.g., feathers, hairs, bones, teeth and scales) and ticks were separated under a stereomicroscope. Prey remnants were identified to the possible lowest taxonomic level, with the help of experts and by comparison with reference collections and/or material in the following zoological museums: “Museu de Zoologia da Universidade de São Paulo” (MZUSP) and “Coleção de Mamíferos do Laboratório de Zoologia de Vertebrados da Escola Superior de Agricultura Luiz de Queiroz” (LZV), both of the University of São Paulo. The nomenclature used for the identification of prey and predators followed Wilson and Reeder (2005) and Trigo et al. (2013). Ticks were preserved in 70 % alcohol and identified to species (nymphs and adults) or to genus level (larvae) following Onofrio et al. (2006), Barros-Battesti et al. (2007), Martins et al. (2010) and Nava et al. (2014). Collected ticks were deposited in the tick collection “Coleção Nacional de Carrapatos” (CNC) of the University of São Paulo under accession numbers CNC- 2188-2207, 2867. Tick-predator-prey interactions were identified by the presence of each individual tick in predator faeces with consumed prey (bird, reptile and mammal).

Results

A total of 523 faecal samples of 12 species of free-ranging wild carnivores were collected, as follows: Canidae—*Chrysocyon brachyurus* (150 samples), *Cerdocyon thous* (62), *Lycalopex* sp. (20); Felidae—*Puma concolor* (21), *Puma yagouaroundi* (22), *Leopardus pardalis* (36), *Leopardus wiedii* (43), *Leopardus guttulus* (26), *Leopardus* sp. (74); Procyonidae—*Nasua nasua* (41), *Procyon cancrivorus* (13); Mustelidae—*Eira barbara* (10), *Galictis cuja* (5). A total of 89 ticks (seven adults, 14 nymphs and 68 larvae) were found in 21 faecal samples (4.0 %) of eight carnivore species, mainly from felids (Table 1), and were identified as follows: *Amblyomma ovale* (three males, three females, and five nymphs), *Amblyomma dubitatum* (four nymphs), *Amblyomma calcaratum* (two nymphs), *Ixodes schulzei* (two nymphs), *Amblyomma sculptum* (one female), *Amblyomma brasiliense* (one nymph), *Amblyomma* sp. (67 larvae), and *Ixodes* sp. (one larva). Prey remnants were identified to each of three vertebrate groups: birds, reptilian, or mammals. Whereas birds could not be assigned to any lower taxonomic level, reptilia could be assigned to the Squamata order, and most of the mammals were assigned to family, tribe, genus or species level. The description of tick-predator-prey species interactions is presented in Table 1.

Discussion

Among the six tick species found in wild carnivore faecal samples in the present study, *A. ovale* was the most abundant, with a total of six adults and five nymphs. This species was reported as the most common tick species on wild carnivores in most regions of Brazil (Labruna et al. 2005). With very few exceptions (Guglielmone et al. 2003), only the adult stage of *A. ovale* is found feeding on carnivores (Labruna et al. 2005). On the other hand, small rodents are preferred hosts for *A. ovale* larva and nymphs (Jones et al. 1972; Martins et al. 2012; Saraiva et al. 2012; Szabó et al. 2012). Thus, it is reasonable to speculate that the six *A. ovale* adults of the present study are associated with ingestion during self-cleaning behaviour (grooming) of carnivore hosts (Felidae, Canidae and Mustelidae), whereas the five nymphs are associated with prey, since these nymphs were found together with a variety of small rodent remnants in the faecal samples (Table 1). The two nymphs of

Table 1 Ticks and prey remnants found in faecal samples of free-ranging wild carnivores collected in a silvicultural landscape in Angatuba, southeastern Brazil

Predators	Preys identified in predators' faecal samples	No. Ticks (L: larvae; N: nymphs; M: males; F: females)
Felidae		
<i>Puma concolor</i>	Squamata; small mammal	2L <i>Amblyomma</i> sp.; 1N <i>Amblyomma brasiliense</i> ; 1M, 1F <i>A. ovale</i>
<i>Puma yagouaroundi</i>	Squamata; small mammal	1L <i>Amblyomma</i> sp.
<i>Leopardus pardalis</i>	<i>Calomys</i> sp.; <i>Oligoryzomys</i> sp.	1N <i>Amblyomma ovale</i>
<i>L. pardalis</i>	Small bird; <i>Akodon</i> sp.; <i>Nectomys</i> sp.; <i>Oligoryzomys</i> sp.	1N <i>Ixodes schulzei</i>
<i>L. pardalis</i>	<i>Akodon</i> sp.; <i>Calomys</i> sp.; <i>Oligoryzomys</i> sp.; small rodent	3L <i>Amblyomma</i> sp.; 1N <i>A. ovale</i>
<i>L. pardalis</i>	Reptilia; Didelphidae; <i>Akodon</i> sp.; <i>Calomys</i> sp.; <i>Oligoryzomys</i> sp.; <i>Necromys lasiurus</i>	1L <i>Ixodes</i> sp.
<i>L. pardalis</i>	Small bird; <i>Akodon</i> sp.; <i>Oligoryzomys</i> sp.	1N <i>Amblyomma calcaratum</i>
<i>L. pardalis</i>	Small bird; <i>Necromys lasiurus</i>	1L <i>Amblyomma</i> sp.
<i>L. pardalis</i>	Small mammal	2L <i>Amblyomma</i> sp.
<i>Leopardus wiedii</i>	Didelphidae; <i>Calomys</i> sp.; <i>Oligoryzomys</i> sp.	1F <i>Amblyomma sculptum</i>
<i>Leopardus guttulus</i>	Squamata; <i>Akodon</i> sp.	1M <i>A. ovale</i>
<i>L. guttulus</i>	Squamata; small bird; Didelphidae; small rodent	5L <i>Amblyomma</i> sp.
<i>L. guttulus</i>	Small bird; <i>Akodon</i> sp.; <i>Oligoryzomys</i> sp.	35L <i>Amblyomma</i> sp.
<i>Leopardus</i> sp.	Small bird; Didelphidae; <i>Akodon</i> sp.; <i>Calomys</i> sp.; <i>N. lasiurus</i> ; <i>Oligoryzomys</i> sp.	16L <i>Amblyomma</i> sp.
<i>Leopardus</i> sp.	Squamata; <i>Akodon</i> sp.; small rodent	1N <i>A. ovale</i>
<i>Leopardus</i> sp.	Small bird; Didelphidae; <i>Akodon</i> sp.; <i>N. lasiurus</i> ; <i>Oligoryzomys</i> sp.; small rodent	1L <i>Amblyomma</i> sp.; 1N <i>A. calcaratum</i> ; 1N <i>I. schulzei</i>
<i>Leopardus</i> sp.	Squamata; small bird; <i>Akodon</i> sp.; <i>Calomys</i> sp.; <i>N. lasiurus</i> ; small rodent	1L <i>Amblyomma</i> sp.
Canidae		
<i>Cerdocyon thous</i>	Small bird; Didelphidae; <i>N. lasiurus</i> ; <i>Nectomys</i> sp.; Akodontini	1M, 1F <i>A. ovale</i>
<i>C. thous</i>	Small bird; <i>Oligoryzomys</i> sp.	1N <i>A. ovale</i>
<i>Chrysocyon brachyurus</i>	Squamata; small bird; <i>Calomys</i> sp.; <i>Oligoryzomys</i> sp.; <i>Hydrochoerus hydrochaeris</i>	1N <i>A. ovale</i> ; 4N <i>Amblyomma dubitatum</i>
Mustelidae		
<i>Eira barbara</i>	Small mammal	1F <i>A. ovale</i>

A. calcaratum found in the faeces of two felids of the genus *Leopardus* are also likely to be associated with a prey item (in these cases, small birds; Table 1), because immature stages of *A. calcaratum* feed primarily on passerine birds (Jones et al. 1972; Labruna et al. 2007; Ogrzewalska et al. 2010, 2014; Sanches et al. 2013). Similarly, the findings of two nymphs of *I. schulzei* along with remnants of small rodents (including a *Nectomys* sp.; Table 1) also indicate that these ticks were attached to ingested prey, because nymphs of *I. schulzei* are known to feed exclusively on small rodents, especially of the genus *Nectomys* (Labruna et al. 2003; Guglielmone et al. 2011; Saraiva et al. 2012; Onofrio et al. 2013). The four nymphs of *A. dubitatum* found in a *C. brachyurus* faecal sample were probably associated with the consumed capybara (*Hydrochoerus hydrochaeris*), the primarily host of all parasitic stages of *A. dubitatum* or, to a much lesser extent, with the consumed small rodents (Table 1), which have been found infested by immature stages of *A. dubitatum* (Nava et al. 2010; Débarbora et al. 2014). Finally, the *A. sculptum* female and the *A. brasiliense* nymph could be related either with predator or prey, since these ticks are known to have a broad host range, including small to large mammals (Aragão 1936; Szabó et al. 2013; Nava et al. 2014).

The present study corroborates a previous report from Portugal (Rosalino et al. 2007), by adding faecal samples as suitable source of valuable information for the study of ticks, being a complementary method for the traditional ones that require capture of live wild animals, especially elusive carnivores. On the other hand, this approach needs to be carefully carried on because faecal samples could be misidentified among close-related species, and in some cases, it could be impossible to determine the host of the ingested tick, namely the predator or the plurality of consumed preys.

Our results also provide epidemiological support for the life cycle of the protozoan *Hepatozoon canis* in Brazil, which is thought to be transmitted to carnivores through the ingestion of *H. canis*-infected *A. ovale* ticks (Forlano et al. 2005; Rubini et al. 2009). Many other tick-borne pathogens, such as *Rickettsia rickettsii* (the agent of Rocky Mountain spotted fever), are transmitted to vertebrate hosts during the parasitism of infected ticks (Parola et al. 2013). Because it was shown in the laboratory that domestic dogs can acquire *R. rickettsii* infection through the ingestion of infected rodents (Moreira and Magalhães 1935), our results support the assumption that predator carnivores are exposed to a broad array of tick-borne agents, which include not only those agents transmitted by carnivore ticks, but also to a variety of agents primarily associated with prey and their ticks (Widmer et al. 2011). As demonstrated here, faecal analysis might be a cheap and unique tool for tick investigations, especially because it provides simultaneous information on ectoparasite-predator-prey interaction, what cannot be obtained by traditional direct-host collection. For this reason, this methodology should be further applied and refined on future sanitary and ecological studies.

Acknowledgments This study was supported by Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) and Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq). Luciano M. Verdade holds a CNPq Productivity Scholarship (309468/2011-6).

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