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Effect of release rate and enantiomeric composition on response to pheromones of *Megaplatypus mutatus* (Chapuis) in poplar plantations of Argentina and Italy

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Abstract

Megaplatypus mutatus (= *Platypus sulcatus* Chapuis) is an Ambrosia beetle native to South America, which was recently introduced in Italy and its presence there is causing severe damage to the local poplar plantations. The male *M. mutatus* pheromone is composed of (S)-(+)-6-methyl-5-hepten-2-ol [(+)-sulcatol], 6-methyl-5-hepten-2-one (sulcatone) and 3-pentanol. A series of field trials testing dose, blend and enantiomer composition performed in Argentina and Italy evaluated attraction and found that the optimal release rate of pheromone components as baits in cross vane baited traps (CIPEIN-CV) was 6, 6 and 30 mg day⁻¹ of sulcatone, (+)-sulcatol and 3-pentanol, respectively. It was also determined that racemic sulcatol is as effective as the pure (+)-isomer for the purpose of beetle catch, due to the inert nature of the (–)-isomer allowing the usage of low cost racemic sulcatol instead of highly expensive (+)-sulcatol. The results of our work contribute to the development of pheromone-based local technologies with low environmental impact and low cost for control or monitoring of an important pest.

Keywords: *Megaplatypus mutatus*, pheromone, (+)-sulcatol, sulcatone, 3-pentanol, trap efficiency

(Accepted 14 February 2013)

Introduction

Megaplatypus mutatus (Chapuis) (Wood, 1993, 2007) is an ambrosia beetle native to South America that is a serious problem in poplar *Populus* sp. commercial plantations (Achinelli *et al.*, 2005; Alfaro *et al.*, 2007). It has recently been introduced in Italy (Tremblay *et al.*, 2000; Allegro & Della Beffa, 2001; Allegro & Griffo, 2008), raising concerns about its possible economic impact on poplar plantations.

Unlike most ambrosia beetles *M. mutatus* attacks only living trees, penetrating into the xylem of its host by boring large tunnels. This weakens the stem, making the tree more vulnerable to breakage during wind storms. Males excavate a tunnel through the bark and build a crown-shaped rim around the gallery entrance with boring dust (Santoro, 1962) from which they emit their sex pheromone blend in order to attract females (González Audino *et al.*, 2005). A similar behaviour has been reported for *Platypus apicalis* White and *P. gracilis* Broun in New Zealand (Milligan & Ytsma, 1988).

In previous studies we reported that volatile emissions from males are composed of (S)-(+)-6-methyl-5-hepten-2-ol ((+)-sulcatol), 6-methyl-5-hepten-2-one (sulcatone)

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50 (González Audino *et al.*, 2005) and 3-pentanol (Gatti Liguori
51 *et al.*, 2008). Individual tests of these compounds showed that
52 sulcatol and sulcatone elicited antennal responses in electro-
53 antennogram recordings and were attractive to beetles in
54 an olfactometer, however 3-pentanol did not elicit antennal
55 response in electroantennogram recordings (González Audino
56 *et al.*, 2005; Gatti Liguori *et al.*, 2008). In our previous field work
57 we tested the blend of sulcatol and sulcatone both released
58 from their own dispensers with release rates selected based on
59 the release rate of commercial bubblecaps (Funes *et al.*, 2009).
60 Also, in a single field test we compared sulcatone plus sulcatol
61 with the three components blend and found that 3-pentanol
62 increased catches (unpublished results).

63 Other platypodids (Renwick *et al.*, 1977; Shore & Mc Lean,
64 1983) and scolytids (Byrne *et al.*, 1974; Borden & Mc Lean,
65 1979; Fletchmann & Berisford, 2003) are also attracted to
66 sulcatol but not to sulcatone. 3-Pentanol has not been found in
67 other platypodids but it is part of the pheromonal blend of
68 *Metamasius hemipteris* (Oliv.) (Perez *et al.*, 1995)

69 Pheromone-baited traps could be used in detection and
70 management of low level populations of *M. mutatus* in infested
71 plantations. Past research has focused on optimal trap design
72 (Gatti *et al.*, 2007; Funes *et al.*, 2009) and the influence of
73 translucent vs. black colour (González Audino *et al.*, 2011) but
74 not on optimal pheromone ratio and release rate. Considering
75 that non-natural ratios or isomers can make the attractant less
76 active than natural sources (Borden, 1990), our goal was to test
77 different pheromone release rates and component ratios in
78 order to optimize blend proportions for field attraction.

79 In addition, since the commercial racemic mixture of
80 (±)-sulcatol is significantly less expensive than the pure
81 (+)-isomer, it made sense from an economic perspective to
82 evaluate whether attraction to the racemic mixture of
83 (±)-sulcatol is similar to the attraction to the natural isomer
84 (+)-sulcatol. A series of field trials testing pheromone dose,
85 blend and enantiomer composition were performed in
86 Argentina and Italy to evaluate attraction in the field.

87 Materials and methods

88 Field trials were conducted to test total pheromone release
89 rates (Experiment 1), blend ratios (Experiments 2 and 3) and
90 enantiomeric composition of sulcatol (Experiment 4).

91 Pheromone dispensers

92 Analytical grade (98%) sulcatone, (±)-sulcatol, and
93 3-pentanol (Aldrich, Saint Louis, MO, USA) were added to
94 reservoir-type dispensers.

95 The dispensers were polyethylene bags made with a
96 non-permeable side (high-density polyethylene of 80 µm;
97 PBB Polisor, Buenos Aires, Argentina) and a semi-permeable
98 side (low-density polyethylene of 40 µm), or polyethylene bags
99 with two semi-permeable sides (low-density polyethylene of
100 40 µm), and/or glass vials with polyethylene semipermeable
101 caps (table 1).

102 The bags were filled with 0.5–2 ml of pheromone
103 component and carefully sealed with heat. They were each
104 loaded with only a single compound, and two or three dis-
105 pensers were placed in a single trap to make up blends.
106 Release rates were measured in the laboratory, as weight loss
107 per day in a wind tunnel (27–28°C, 0.5 m s⁻¹) (Funes *et al.*,
108 2009, 2011). These dispensers had a constant release rate (zero-
109 order kinetics) until total consumption as is usual for

Table 1. Effective release area and daily release rate (mg per day ±SE) of pheromone lures containing (+)-sulcatol, (±)-sulcatol, sulcatone and 3-pentanol at 27–28°C and 0.5 m s⁻¹ in a laboratory wind tunnel (*n* = 3).

Pheromone	Dispenser	Effective release area (cm ²) (dimensions)	Release rate (mg per day) ±SE
(+)–Sulcatol or (±)–sulcatol	Polyethylene bag	16 (4 × 4 cm)	11.3 ± 0.5
	Polyethylene bag	8 (4 × 2 cm)	6.0 ± 0.3
	Polyethylene bag	30 (3 × 10 cm)	26.3 ± 0.8
Sulcatone	Polyethylene bag	20 (5 × 4 cm)	57.7 ± 5.5
	Glass vial with polyethylene cap	1.2	13.4 ± 0.8
3-Pentanol	Glass vial with polyethylene cap	0.3	6.7 ± 0.6
	Polyethylene bag	20	29.8 ± 1.8
	Glass vial with polyethylene cap	0.26	1.9 ± 0.1

diffusion-controlled membrane-moderated reservoir systems (Tojo, 1985), and also as we showed in our previous work (Funes *et al.*, 2009, 2011). Pheromone dispensers were suspended inside traps.

To obtain different release rates we modified the sizes of the semipermeable surfaces, according to the correlation between pheromone release rate and permeable surface of the dispensers previously studied (Funes *et al.*, 2011).

Trap setting

The traps used were Lindgren multiple funnel traps (eight funnels) (Lindgren, 1983), CIPEIN – CV cross vane (Funes *et al.*, 2009) and Mastrap[®] (Isagro S.R.L. Milan, Italy) cross vane traps. Lindgren is a multiple funnel trap made of eight black funnels of diameter 18.5 cm (Contech, BC, Canada). Total height of the trap is 1.5 m. CIPEIN-CV is a cross-vane trap of two black acrylic panels in a cross arrangement above a funnel. The diameter of the funnel is 20 cm, the total height 70 cm. Mastrap is L[®] A version (Isagro S.R.L., Milán, Italia) is a grey cross-vane trap, with effective surface 630 cm². Total height of the trap is 15 cm and width 28 cm.

Baited and empty control traps were hung from trees with their top at 1.8 m above ground level and were set 30–40 m apart in parallel lines separated by 45–50 m throughout each treatment plot. A previous detailed survey of the plantation showed that the distribution of *M. mutatus* galleries was homogeneous in the field (unpublished data) and consequently allowed us to use setup traps in random distribution. In addition, *M. mutatus* is a primary pest attacking vigorous trees, which also ruled out the selection of plots where the heterogeneity of soil characteristics causes different types of growth rates between trees in the same plot. Five identical control traps were set up for each experiment.

Traps were repositioned randomly and dispensers were checked and replaced before complete pheromone emission every one or two weeks, depending on access to the field site.

Field trial locations

The experiment to determine the influence of total pheromone release rate (Experiment 1) was performed between

Table 2. Release rate (mg per day \pm SE) of pheromone lures containing (+)-sulcatol sulcatone, or 3-pentanol used in field experiments (release rates at 27–28°C and 0.5 m s⁻¹ in a laboratory wind tunnel) and mean \pm SE number per trap per week of *M. mutatus* captured with different release rates and ratios of (+)-sulcatol, sulcatone and 3-pentanol. Different letters within each experiment indicate significant differences ($P < 0.05$). Controls: unbaited traps. There were no insects caught in any of the control traps.

Expt.	Treatment	Release rate (mg d ⁻¹)			Mean catch females \pm SE
		(+)-Sulcatol	Sulcatone	3-Pentanol	
1	1	11.3	6.7		6.33 \pm 1.5 ^b
	2	22.6	13.4		1.33 \pm 2.3 ^a
	Control				0
2	3	11.3	6.7	29.8	1.82 \pm 0.35 ^a
	4	6	6.7	29.8	2.75 \pm 0.48 ^a
	5	11.3	6.7	1.9	1.39 \pm 0.27 ^b
	6	6	6.7	1.9	1.37 \pm 0.28 ^b
	Control				0
3	7	6	57.7	29.8	1.66 \pm 0.23 ^a
	8	6	6.7	29.8	1.33 \pm 0.17 ^a
	Control	–			0

148 10 October 2009 and 9 March 2010, in a commercial poplar
149 (*Populus deltoides*) plantation located in Alberti, Province of
150 Buenos Aires, Argentina (35°10'S, 60°29'W) at an elevation of
151 50 m above sea level. The experimental area consisted of 8 ha of
152 a 10 year-old plantation, with a tree density of 1111 trees ha⁻¹
153 (3 m \times 3 m spacing) and a mean diameter at breast height
154 (DBH) of 23.2 cm \pm 0.5 (SE).

155 The experiments to determine the influence of pheromone
156 release rates and ratios (Experiments 2 and 3) were con-
157 ducted between 30 October 2009 and 17 December 2010
158 (Experiment 2) and from 8 January to 4 May 2010 (Experi-
159 ment 3). The field site was a commercial poplar (*Populus*
160 *deltoides*, Australiano clone I29/60) plantation located in
161 Morse, Junín, Buenos Aires, Argentina (S) 34°43'56.3", (W)
162 60°51'11.5") at an elevation of 59 m above sea level. The field
163 plot was a 12 ha 11 year-old plantation with a tree density of
164 625 trees ha⁻¹ (4 m \times 4 m spacing) and a mean DBH of 32.6 cm,
165 \pm 0.2 (SE). The experiment to study the influence of pure vs.
166 racemic sulcatol (Experiment 4) was conducted in Caserta,
167 Campania Region, Italy, during the 2008 season between
168 19 May and 19 September at a poplar plantation located in
169 Falciano del Massico (41°09'07"N, 13°57'54.3"E and 38 m
170 above sea level). The plantation (*Populus \times euroamericana*
171 (Dode) Guinier Louisa Avanzo clone) consisted of 1 ha of
172 11 year-old trees at a density of 494 trees ha⁻¹ (4.5 \times 4.5 m
173 spacing) and a mean DBH of 26.6 \pm 1.0 cm (SE).

174 The pest is thought to have been introduced into Italy
175 recently from Argentina (Tremblay *et al.*, 2000).

176 Pheromone treatments

177 Experiment 1 investigated the influence of total pheromone
178 release rate. We tested two release rates of (+)-sulcatol plus
179 sulcatone in a proportion of 2:1 (table 2, Experiment 1, Treat-
180 ments 1 and 2): one lower based on our previous results (Funes
181 *et al.*, 2009) and the other doubling the rate. Five replicates
182 were performed for each treatment and five identical traps
183 without lures were considered control. Treatments were
184 rotated weekly or every two weeks according to the possibility
185 of accessing the field.

186 Experiment 2 on the influence of (+)-sulcatol and
187 3-pentanol release rates in the pheromone blend, tested two
188 release rates for sulcatol and 3-pentanol respectively, holding

the sulcatone release rate constant. The component 3-pentanol,
189 was tested at two concentrations that differed by a factor of
190 16 (table 2, Experiment 2, Treatments 3–6); the component
191 (+)-sulcatol was tested in 6 and 11 mg day⁻¹ according to the
192 results of Experiment 1, and also based on optimal release rate
193 information reported for another Ambrosia beetle (Liu *et al.*,
194 1989). The release rate of sulcatone was chosen according to
195 previous studies (Funes *et al.*, 2009, 2011). There were five
196 replicates per treatment and five identical traps without lures
197 were used as blank controls.

198 Experiment 3 investigated the influence of sulcatone re-
199 lease rate in pheromone blend. Based on the results of treat-
200 ments 3–6, we held sulcatol and 3-pentanol release rates
201 constant and varied the release rate for sulcatone (table 2,
202 Experiment 3, Treatments 7 and 8). There were six replicates
203 per treatment. Five identical traps without lure were used as
204 blank controls.

205 Experiment 4 determined the influence of enantiomeric
206 composition of sulcatol in pheromone blend. Traps were
207 baited with sulcatone (6 mg per day), 3-pentanol (29.8 mg
208 per day), and either (+)-sulcatol (6.6 mg per day) or racemic
209 sulcatol (11.2 mg per day). The release rate of racemic sulcatol
210 was twice that of (+)-sulcatol so the total amount of (+)-isomer
211 released was the same in both treatments. Fifteen replicates
212 were set for each treatment and five traps without lures were
213 used as blank controls.

214 We recorded the number of *M. mutatus* adults captured per
215 trap per week during each period for Experiments 1–3 and
216 females per trap per day for Experiment 4. Differences be-
217 tween treatments were analysed by one way analysis of
218 variance (ANOVA) followed by Duncañs multiple range
219 (Experiments 2–4) or χ^2 test (Experiment 1). Data were tested
220 for homoscedasticity by Levene's test along with Shapiro-
221 Wilks test for normality and no transformations of data were
222 necessary. In Experiment 1, the traps used were Lindgren
223 multiple funnel traps. In Experiments 2 and 3, the traps used
224 were CIPEIN – CV cross vane. In Experiment 4, the traps used
225 were Mastrap traps.

227 Results

228 Table 2 shows the mean number of female *M. mutatus*
229 caught per trap per week for Experiments 1–3 using different

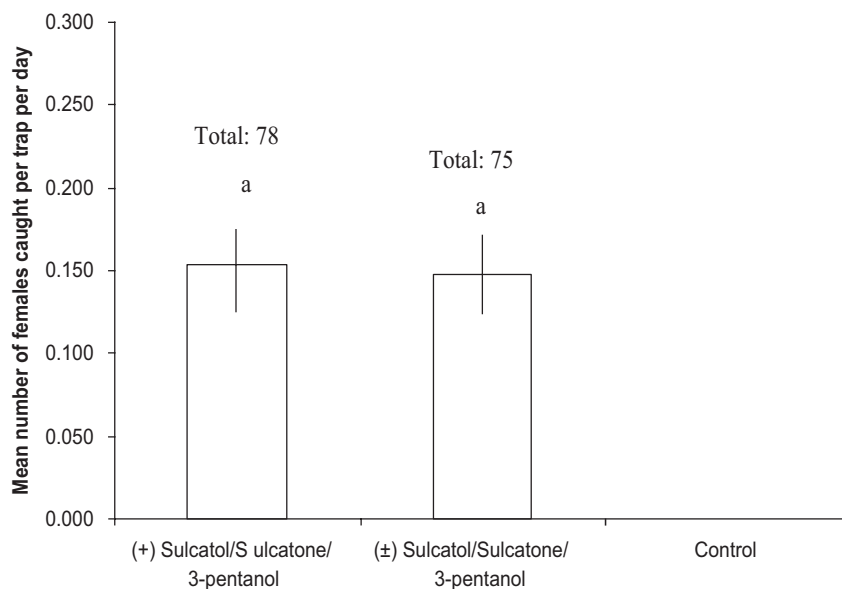


Fig. 1. Experiment 4. The mean number of females of *M. mutatus* captured per trap per day in traps baited with sulcatone (6 mg d^{-1}), 3-pentanol (29.8 mg d^{-1}) and either (\pm)-sulcatol 11.2 mg d^{-1} or (+)-sulcatol 6.6 mg d^{-1} . Caserta, Italy, 2008. Treatments did not differ significantly ($P=0.89$). Controls: unbaited traps. There were no insects caught in any of the control traps.

230 pheromone release rates. No insects were caught in the
231 unbaited control traps for any of the experiments.

232 In Experiment 1 on the influence of overall pheromone
233 release rate, significantly more insects were captured in traps
234 baited with (+)-sulcatol and sulcatone at a total release rate
235 of 18 mg per day compared with traps baited with double
236 the release rate (36 mg per day) (χ^2 : 17.9, $df=2,57$, $P=0.0367$)
237 (table 2).

238 In Experiment 2 on the influence of (+)-sulcatol and
239 3-pentanol release rates in pheromone blend with sulcatone
240 at fixed rate, (+)-sulcatol released at 6 mg day^{-1} (table 2, exp. 2,
241 treatments 4 and 6) and 11 mg day^{-1} (+)-sulcatol (table 2,
242 exp. 2, treatments 3 and 5) had the same attractive effect when
243 combined with both high and low release rate of 3-pentanol
244 ($F=2.75$, $dF=4111$, $P=0.26$ and $F=1.8$, $dF=3112$, $P=0.39$,
245 respectively)

246 However, 3-pentanol released at 29.8 mg d^{-1} (table 2,
247 exp. 2, treatments 3 and 4) was more attractive than 1.9 mg
248 day^{-1} (table 2, exp. 2, treatments 5 and 6), when combined
249 with (+)-sulcatol released at 6 or 12 mg day^{-1} , $P=0.045$ and
250 $P=0.022$, respectively).

251 Catches with treatments 3–6 of Experiment 2 were shown
252 to be independent (according to multiple effects ANOVA),
253 as there were no interactions between them ($P=0.322$). This
254 fact allowed us to use the results obtained for sulcatol and
255 3-pentanol in Experiment 3.

256 In Experiment 3, there was no significant difference in number
257 of beetles captured in traps baited with sulcatone released
258 at 6.7 and 57.7 mg day^{-1} (table 2, exp. 3, treatments 5 and 6,
259 $P=0.24$), indicating that both low and high sulcatone release
260 rates have similar attractiveness for capturing *M. mutatus*
261 females. No insects were caught in the unbaited control traps
262 and zero catches values were not included.

263 In Experiment 4 on the influence of enantiomeric compo-
264 sition of sulcatol in pheromone blend, the number of beetles
265 captured in traps baited with racemic sulcatol released at

12 mg day^{-1} was not significantly different from the number
266 captured with the (+)-isomer with release rate of 6 mg day^{-1}
267 (fig. 1, $P=0.89$). The presence of the (–)-sulcatol isomer did not
268 cause an inhibitory behavioural response when combined
269 with the bioactive isomer component.
270

Discussion

271
272 In these experiments, traps baited with pheromones caught
273 significant numbers of female *M. mutatus* and no beetles were
274 caught in unbaited traps, confirming previous results on attrac-
275 tiveness of the proposed pheromone components (Funes *et al.*,
276 2009). Doubling the release rate of the two component blends
277 of sulcatol and sulcatone from 18 mg d^{-1} to 36 mg d^{-1} resulted
278 in a drop of $>80\%$ in the trap catch (Experiment 1), whereas
279 increasing the release rate of sulcatone caused no effect
280 (Experiment 3). This might suggest that the beetles are highly
281 sensitive to the release rate of sulcatol, but relatively insen-
282 sitive to the release rate of sulcatone. This fact correlates with
283 the fact that sulcatone is less active than sulcatol in individual
284 laboratory bioassays and that sulcatol, but not sulcatone, is an
285 active pheromone component of other ambrosia beetles.

286 The two (+)-sulcatol and sulcatone rates captured a similar
287 number of insects indicating that it is not necessary to increase
288 costs by using higher release rates. However, the higher
289 release rate of 3-pentanol tested (30 mg per day) demonstrated
290 higher efficacy than the lower one (2 mg per day). These results
291 suggest that the optimal release ratio for the three pheromone
292 components is 6 , 6 and 30 mg d^{-1} (ratio 1:1:5) of (+)-sulcatol,
293 sulcatone and 3-pentanol, respectively. These release rates are
294 much larger than the amounts produced by male beetles,
295 which are in the range of micrograms per day per beetle (Gatti
296 Liguori *et al.*, 2011). However, the beetles produced (+)-sulca-
297 tol and sulcatone at similar rates (Gatti Liguori *et al.*, 2011), as
298 found in the optimal blend of synthetic compounds proposed
299 above. Liu *et al.* (1989) found that mg d^{-1} doses of sulcatol

300 seem to be optimal for catching *Gnathotricus sulcatus*, with
301 1.5 mg day⁻¹ more efficiency than lower (0.5 mg d⁻¹) and
302 higher release rates (5 and 10 mg d⁻¹).

303 The racemic sulcatol mixture is as effective as the
304 (+)-isomer for capturing *M. mutatus*, suggesting that the
305 (-)-isomer has no discernable biological activity, either posi-
306 tive or negative. Also, Hoover *et al.* (2000) found that response
307 of *Tripodendrum lineatum* to lineatin is independent of the
308 enantiomeric ratio, and depends only on the content of (+)
309 lineatin and the (-) isomer is inactive. This is important
310 because it enables the use of low cost racemic sulcatol instead
311 of (+)-sulcatol. In this case, the release rates should be 12, 6
312 and 30 (ratio 2:1:5) mg per day of (±)-sulcatol, sulcatone and
313 3-pentanol, respectively.

314 Two factors contribute to the promising success of
315 pheromones for monitoring or controlling this species: the
316 components of the pheromone blend of male *M. mutatus* are
317 inexpensive, and traps can be easily made with low cost plastic
318 translucent materials that perform better than commercial
319 traps (González Audino *et al.*, 2011). However, other factors
320 influencing the final cost of pheromone-based programmes
321 like deploying and maintaining large arrays of traps should
322 also be considered. The results of our work contribute to the
323 development of pheromone-based local technologies with low
324 environmental impact and low cost for control or monitoring
325 of an important pest.

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References

334
335 Achinelli, F.G., Liljerström, G., Aparicio, A., Delgado, M.,
336 Jouanny, M. & Mastrandrea, C. (2005) Daños por taladrillo
337 (*Megaplatypus mutatus* (= *Platypus sulcatus*)) en plantaciones
338 de álamo (*Populus* spp.) de Alberti, Buenos Aires: análisis
339 preliminar de la magnitud y distribución de fustes quebra-
340 dos. *Revista Asociación Forestal Argentina* **59**, 8–11 (In Spanish).
341 Alfaro, R., Humble, L.M., Gonzalez Audino, P., Villaverde, R. &
342 Allegro, G. (2007) The threat of the ambrosia beetle
343 *Megaplatypus mutatus* (Chapuis) [= *Platypus mutatus* Chapuis]
344 to world poplar resources. *Forestry* **80**, 471–479.
345 Allegro, G. & Della Beffa, G. (2001) Un nuovo problema ento-
346 mologico per la pioppicoltura Italiana: *Platypus mutatus*
347 Chapuis (Coleoptera: Platypodidae). *Sherwood Foreste ed*
348 *alberi oggi* **66**, 31–34 (In Italian).
349 Allegro, G. & Griffo, R. (2008) I rischi di diffusione di
350 *Megaplatypus mutatus*. *L'Informatore Agrario* **13**, 73–76.
351 Borden, J. (1990) Use of Semiochemicals to manage coniferous tree
352 pests in Western Canada. pp. 281–316 in Ridgway, R.L.,
353 Silverstein, R.M. & Inscoc, M.N. (Eds) *Behavior Modifying*
354 *Chemicals for Insect Management. Applications of Pheromones*
355 *and Other Attractants*.
356 Borden, J.H. & Mc Lean, J.A. (1979) Secondary attraction in
357 *Gnathotricus retusus* and cross attraction of *G. sulcatus*
358 (Coleoptera: Scolytidae). *Journal of Chemical Ecology* **5**, 79–88.

Byrne, K.J., Swigar, A.A., Silverstein, R.M., Borden, J.H. &
Stokkink, E. (1974) Sulcatol: population aggregation phero-
mone in the Scolytid beetle, *Gnathotricus sulcatus*. *Journal of*
Insect Physiology **20**, 1895–1900.
Fletchmann, C.A.H. & Berisford, C.W. (2003) Identification of
sulcatol, a potential pheromone of the ambrosia beetle
Gnathotricus materiarus (Col., Scolytidae). *Journal of Applied*
Entomology **127**, 189–194.
Funes, H., Zerba, E. & González-Audino, P. (2009) Comparison of
three types of traps baited with sexual pheromones for
Ambrosia beetle *Megaplatypus mutatus* in poplar plantations.
Journal of Economic Entomology **102**, 1546–1550.
Funes, H., Griffo, R., Zerba, E. & Gonzalez-Audino, P. (2011)
Mating disruption of the ambrosia beetle *Megaplatypus mu-*
tatus in poplar and hazelnut plantations using reservoir
systems for pheromones. *Entomologia Experimentalis et*
Applicata **139**, 226–234.
Gatti Liguori, P., Zerba, E. & González Audino, P. (2007) New
trap for emergent *Megaplatypus mutatus*. *Canadian*
Entomologist **139**, 894–896.
Gatti Liguori, P., Zerba, E., Alzogaray, R. & González-Audino, P.
(2008) 3-Pentanol: a new attractant present in volatile emis-
sions from the Ambrosia beetle, *Megaplatypus mutatus*.
Journal of Chemical Ecology **34**, 1446–1451.
Gatti Liguori, P., Zerba, E. & González Audino, P. (2011)
Anatomical site of pheromone accumulation and temporal
pattern of pheromone emission in the ambrosia beetle,
Megaplatypus mutatus. *Physiological Entomology* **36**, 201–207.
González Audino, P., Villaverde, R., Alfaro, R. & Zerba, E.
(2005) Identification of volatile emissions from *Platypus*
mutatus (= *sulcatus*) (Coleoptera: Platypodidae) and their
behavioral activity. *Journal of Economic Entomology* **98**, 1506–
1509.
González-Audino, P., Gatti, P. & Zerba, E. (2011) Traslucent
pheromone traps increase trapping efficiency of ambrosia
beetle *Megaplatypus mutatus*. *Crop Protection* **30**, 745–747.
Hoover, S.E.R., Lindgren, B.S., Keeling, C.I. & Slessor, K.N.
(2000) Enantiomer preference of *Tripodendron lineatum* and
effect of pheromone dose and trap length on response to
lineatin-baited traps in interior British Columbia. *Journal of*
Chemical Ecology **26**, 667–677.
Lindgren, B.S. (1983) A multiple funnel trap for scolytid beetles
(Coleoptera). *Canadian Entomologist* **115**, 299–302.
Milligan, R.H. & Ytsma, G. (1988) Pheromone dissemination
by male *Platypus apicalis* White and *P. gracilis* Broun (Col.
Platypodidae). *Journal of Applied Entomology* **106**, 113–118.
Perez, A.L., Campos-Piedra, Y., Chinchilla, C.M.,
Oehlschlager, A.C., Gries, G., Gries, R., Castrillo, G.,
Giblin-Davis, R.M., Peña, J.E., Duncan, R.E., Gonzalez, L.
M., Pierce, H.D. Jr, McDonald, R. & Andrade, R. (1995)
Aggregation pheromones and host kairomones of the West
Indian sugarcane weevil, *Metamasius hemipterus sericeus*
(Oliv.) (Coleoptera: Curculionidae). *Journal of Chemical*
Ecology **23**, 869–888.
Renwick, J.A., Vite, J.P. & Billings, R.F. (1977) Aggregation
pheromones in the ambrosia Beetle *Platypus flavicornis*.
Naturwissenschaften **64**, 226.
Santoro, F.H. (1962) La copula en *Platypus sulcatus* Chapuis
(Coleoptera: Platypodidae). *Revista Investigaciones Forestales*
3, 25–27 (in Spanish).
Shore, T.L. & Mc Lean, J.A. (1983) Attraction of *Platypus wilsoni*
Swaine (Coleoptera: Platypodidae) to traps baited with sul-
catol, ethanol and alpha-pinene. *Canadian Forestry Service*
Research Notes **3**, 24–25.

- 423 **Tojo, K.** (1985) Intrinsic release rate from matrix-type drug 428
424 delivery systems. *Journal Pharmacology Science* **74**, 685–687. 429
- 425 **Tremblay, E., Espinosa, B., Mancini, D. & Caprio, G.** (2000) Un 430
426 coleottero proveniente dal Sudamerica minaccia i pioppi. 431
427 *L'Informatore Agrario* **56**, 89–90. 432
433
- Wood, S.L.** (1993) Revision of the genera of Platypodidae 428
(Coleoptera). *Great Basin Naturalist* **53**, 259–281. 429
- Wood, S.L.** (2007) *Bark and Ambrosia Beetles of South America* 430
(Coleoptera: Scolytidae). Provo, Utah, Brigham Young 431
University, M.L. Bean Life Science Museum. 432
433