CALCIUM TRIGGERED ARRHYTHMIAS AND THE GOLDILOCKS' PRINCIPLE

Carlos A. Valverde¹, Alejandra Cely Ortiz¹, Luis A. Gonano¹, Elena C. Lascano², Alicia Mattiazzi^{1*}

¹ Centro de Investigaciones Cardiovasculares, CCT-La Plata-CONICET, Facultad de Cs. Médicas, UNLP, La Plata, Argentina.

² Instituto de Medicina Translacional, Transplante y Bioingeniería, Universidad Favaloro, (CONICET), Buenos Aires, Argentina.

* Correspondence to: aliciamattiazzi@gmail.com

ABSTRACT

Beyond the importance of calcium cycling on cardiac contractile function, calcium can also affect membrane currents and lead to arrhythmias.

In this revision we will discuss how the balance between release and reuptake of calcium from and to the sarcoplasmic reticulum affects the probability of arrhythmias.

A mathematical human cardiomyocyte model that can simulate increasing RyR2 open probabilities (Po) along with high or low SERCA2A mediated reuptake velocities showed that arrhythmias occur in an intermediate physiological zone for both RyR2 Po and reuptake velocity. These results allow to make a parallelism with the "goldilocks and the tree bears" story in which she is looking to drink a cup of porridge with an intermediate optimal temperature.

Along the text we explain how the Goldilocks principle is fulfilled in different well-known experimental situations.

Keywords: Heart, Arrhythmias, Calcium, Sarcoplasmic Reticulum, RyR2, PLN

RESUMEN

Más allá del papel que el ciclado de calcio cumple en la función contráctil del corazón, el manejo de calcio en los cardiomiocitos es capaz de afectar corrientes de membrana y, de este modo, las alteraciones en el manejo de calcio son capaces de generar arritmias cardíacas. En esta revisión discutiremos los efectos que el balance entre liberación y recaptura de calcio desde y hacia el retículo sarcoplasmático (RS) tiene sobre la probabilidad de gatillar arritmias cardíacas. Mediante un modelo matemático de miocito cardiaco humano, que simula probabilidades de apertura (Po) crecientes de los receptores de rianodina (RyR2) en asociación con aumentos y disminuciones de la velocidad de secuestro de Ca²⁺ por la Ca-ATPasa del RS (SERCA2a), se determinó que las arritmias ocurren con más probabilidad en una zona intermedia (zona óptima para las arritmias) tanto de Po como de la velocidad de secuestro de Ca²⁺ están libres de arritmias. Estos resultados permiten establecer un paralelismo con la historia de "Ricitos de oro y los tres osos" y su búsqueda de una taza de avena con temperatura intermedia como aquella óptima para ser bebida.

A lo largo del texto explicaremos cómo este principio se cumple en situaciones experimentales conocidas.

Palabras Clave: Corazón, Arritmias, Calcio, Retículo Sarcoplasmático, RyR2, PLN

Introduction

Mechanical dysfunction and arrhythmias are a leading cause of morbidity and mortality worldwide [1,2], and it is now well established that a large fraction of ventricular arrhythmias is initiated at the cellular level by focal triggered mechanisms such as abnormal spontaneous Ca^{2+} discharges (Ca^{2+} sparks) from the sarcoplasmic reticulum (SR). This excessive Ca^{2+} release during diastole propagates as regenerative Ca^{2+} waves that travel through the cytosol and activate inward membrane currents, mainly the electrogenic Na^+/Ca^{2+} exchanger (NCX) working in the forward mode [3-5].

Abnormal spontaneous Ca^{2+} discharges from the SR occur under conditions in which SR Ca^{2+} load exceeds a threshold that is largely determined by the state of the ryanodine receptors (RyR2). It is known, for instance, that RyR2 point mutations render the channel more prone to spontaneous SR Ca^{2+} release during adrenergic stimulation. Patients with this inherited anomaly exhibit catecholaminergic polymorphic ventricular tachycardia, a known cause of sudden cardiac death [6]. We and others have described that RyR2 phosphorylation by Ca^{2+} –calmodulin-dependent protein kinase II (CaMKII) at the Ser2814 site is associated with SR Ca^{2+} leak and arrhythmogenesis in cardiac pathologies of different etiologies [7-12]. These results define the crucial role of altered RyR2 activity on triggered arrhythmias.

In contrast, the effect of increasing SR Ca^{2+} uptake on cardiac triggered events is not clear and there is concern on whether the increase in SR Ca^{2+} uptake, which has been shown to be a useful therapy to revert depressed cardiac contractility in human and experimental heart failure [13], is protective against Ca^{2+} triggered arrhythmias or exacerbates them. Indeed, either the increase or decrease of SR Ca^{2+} uptake has led to contradictory results [14-21].

We believe that a possible explanation to these conflicting results may rest, at least in part, in the opposite effects inherent to the augmented cytosolic SR Ca²⁺ uptake (Figure 1) i.e., increasing the rate of SR Ca²⁺ uptake would reduce cytosolic Ca²⁺ overload and the risk of cardiac arrhythmias, but would necessarily increase SR Ca²⁺ content, favoring RyR2 Ca²⁺ sensitization, improving diastolic SR Ca²⁺ leak and the risk of Ca²⁺ waves. This situation might be exacerbated if the increase in SR Ca²⁺ uptake coexists with an increase in the open probability of the RyR2, as that produced by CaMKII-dependent phosphorylation of the Ser2814 site [7, 12, 22].

Of note, SR Ca²⁺ uptake and release are highly regulated processes. The activity of SERCA2a and RyR2 open probability (Po) are dependent on a complex regulation that includes SR Ca²⁺ load and different proteins interacting with either SERCA2a, RyR2 or both (See for review [23-25]. Among these regulatory proteins we will mention phospholamban (PLN) not only because of its relevant action on SERCA2a activity but also because some of the experiments to be described here use genetic modified mice with PLN ablation (PLNKO mice, (26)). Under dephosphorylated conditions, PLN tonically inhibits SERCA2a. PLN phosphorylation or ablation, relieves this inhibition increasing SERCA2a activity and SR Ca²⁺ load [26-28].

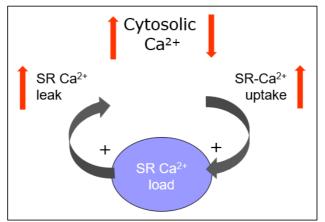


Figure 1. Opposite effects of increasing SR Ca^{2+} uptake on cytosolic Ca^{2+} . The increase in SR Ca^{2+} reuptake diminishes cytosolic Ca^{2+} and increases SR Ca^{2+} leak.

Triggered arrhythmias and the relationship between SR Ca²⁺ uptake and leak

By using genetic altered mice and the myocyte mathematical model of Negroni- Lascano [22, 29-31], we previously showed that triggered arrhythmias are highly dependent on the relationship between SR Ca²⁺ uptake, given by the activity of SERCA2a and SR Ca²⁺ leak, given by the Po of RyR2. In these experiments, we were able to define an arrhythmogenic zone (arrhythmogenic island) and a non-arrhythmogenic area (in blue), surrounding "the island", constituted by different combinations of RyR2 Po (conductance in the model) and SR Ca²⁺ uptake.

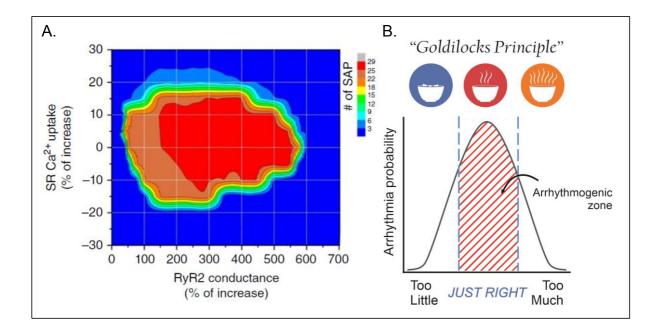


Figure 2. A. The interplay between SR Ca^{2+} uptake and leak define the occurrence of arrhythmias. **B.** The Goldilocks principle. Too much uptake (with respect to the leak) or too much leak with respect to the uptake), decreases the probability of arrhythmias.

As it is shown in the figure, very low or very high SR Ca^{2+} uptake or leak (blue zone), is unable to evoke triggered arrhythmias. Instead, maximal arrhythmia probability occurs in the middle zone of the relationship, which reminds the classic Goldilocks principle by analogy to the children's story "The Three Bears" by Robert Southey [32]. In this tale, a young girl named Goldilocks tastes three different bowls of porridge and finds she prefers porridge that is neither too hot nor too cold, but has just the right temperature. In our case, the red zone of the island constitutes the area with the just right combination of SR Ca^{2+} uptake and leak that produces arrhythmias.

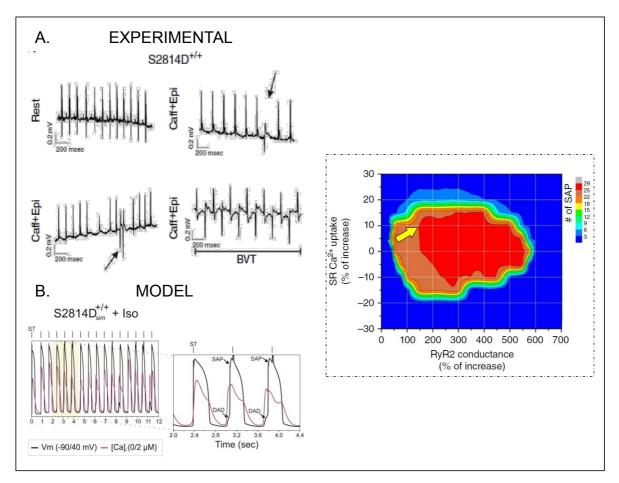


Figure 3. A. Representative ECG tracings in conscious S2814D+/+ knock-in mice at rest and after I.P. injection of caffeine/Adrenaline (Caff/Adr). Arrows in S2814D+/+ tracings indicate premature ventricular complexes (PVCs). Bidirectional ventricular tachycardia (BVT) was observed in the majority of S2814D+/+ mice after Caff/Adr challenge. **B.** The model reproduces the experimental results. Under conditions of stress (Isoproterenol (Iso) stimulation), there is an increase of spontaneous action potentials (SAP) originated from delayed afterdepolarizations (DADs). ST: Stimulus. Inset: The yellow arrow indicates the possible position in the graph of S2814D hearts under stress conditions. Experimental.

Figure 3A depicts the results obtained in a genetic modified mouse with an increased Po produced by constitutive pseudo-phosphorylation of RyR2 by CaMKII without SR Ca²⁺ uptake modifications (Ser2814D mice, [33]). Under resting conditions, there are no detectable arrhythmias. However, under stress conditions (Norepinephrine plus caffeine infusion), there is an increase in cardiac arrhythmias. **Figure 3B** shows that the mathematical model simulating the conditions of S2814D hearts, reproduces the experimental behavior of these hearts under stress conditions [22].

The behavior of Ser2814D hearts would follow the yellow arrow in the inset of Figure 3. Under control conditions the SR Ca^{2+} load is low, because of the increased SR Ca^{2+} leak of these myocytes without any increase in SR Ca^{2+} uptake. When submitted to stress (Adrenaline plus caffeine or Isoproterenol) there is an increase in SR Ca^{2+} uptake, SR Ca^{2+} load and SR Ca^{2+} leak that approximates S2814D myocytes to the red zone of the graph (high arrhythmia probability).

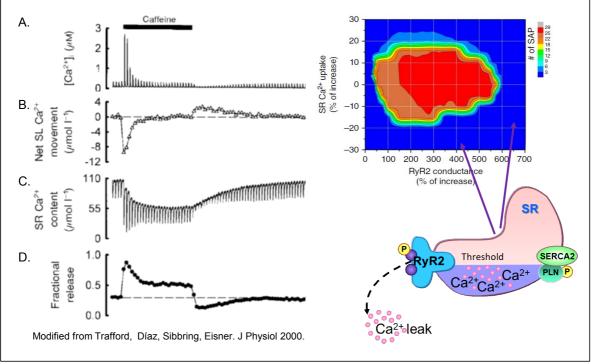


Figure 4. Potentiation of RyR2 enhances SR Ca²⁺ leak and diminishes SR Ca²⁺ content.

There are some "blue" zones in the graph of Figure 2 in which it is difficult to intuitively visualize why they represent an arrhythmia-free zone, despite the high increase in RyR2 conductance. The interpretation of the zone below "the island" is similar to that given for S2814D myocytes. Figure 4 illustrates what occurs experimentally when there is an increase in SR Ca²⁺ leak without any corresponding increase in SR Ca²⁺ uptake, able to reestablish SR Ca²⁺ load to the threshold for SR Ca²⁺ leak. The experiment from Eisner's group shows that administrating isolated myocytes low doses of caffeine is able to sensitize RyR2, producing an increase in the Ca²⁺ transient due to the

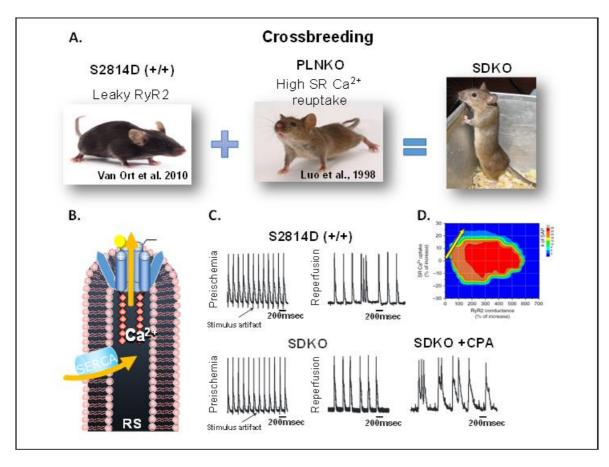


Figure 5.A. Mutants characteristics. **B.** Graphical representation of RS with an increase in both SR Ca^{2+} uptake and leak. **C.** Results obtained in S2814D and SDKO hearts ssubjected to a protocol of ischemia/reperfusion. **D.** Yellow arrow show possible trajectory from the non-arrhythmogenic to the arrhythmogenic area when S2814D mice were crossbred with PLNKO mice to produce SDKO animals.

increase in SR Ca²⁺ release, but which rapidly returns to control values as a result of the decrease in SR Ca²⁺ load [34]. However, the consequence of RyR2 sensitization can be seen: The SR was able to release the same amount of Ca²⁺, despite the decrease in SR Ca²⁺ load. Thus, RyR2 sensitization, although unable to evoke a persistent increase in Ca²⁺ transient amplitude, produces a persistent increase in fractional Ca²⁺ release, i.e., the Ca²⁺ released at a given SR Ca²⁺ load [31].

The blue area above the "arrhythmogenic island" is more difficult to explain. In this case the increase in SR Ca^{2+} leak associated with an increase in SR Ca^{2+} uptake can hardly be visualized as a non-arrhythmogenic combination. To explore this paradox, we performed experiments in genetic modified mice obtained by crossbreeding S2814D mice with mice with PLN ablation mice (SDKO mice) (Figure 5A).

This provides an animal model with an increase in both SR Ca²⁺ uptake and leak (Figure 5B). Figure 5C shows the results obtained in S2814D hearts subjected to a protocol of ischemia/reperfusion. Immediately after ischemia, it is possible to observe the classic reperfusion arrhythmias found in S2814D hearts. PLN ablation avoids reperfusion arrhythmias in SDKO hearts. As a proof of concept, decreasing the activity of SERCA2a by perfusion of the hearts with the SERCA2a inhibitor cyclopiazonic acid (CPA) at the time of reperfusion, evokes again reperfusion arrhythmias. Figure 5D shows a possible trajectory of S2814D from the non-arrhythmogenic to the arrhythmogenic area when these mice were crossbred with PLNKO mice to produce SDKO animals.

Which is the mechanism by which PLN ablation avoids the arrhythmias that usually occur under stress conditions in mice with an increase in Po? We mentioned above that an increase in SR Ca²⁺ uptake increases SR Ca²⁺ load and SR Ca²⁺ leak (Figure 1). Is this mechanism absent in SDKO hearts? To test this idea, we measured Ca²⁺ sparks in SDKO hearts subjected to either a stress [22] or to the

ischemia/reperfusion protocol [12]. Figure 6 A is a representative example obtained using confocal microscopy in the intact heart, revealing that during reperfusion there is an increase in SR Ca^{2+} leak (Ca^{2+} sparks) in SDKO when compared with S2814D hearts.

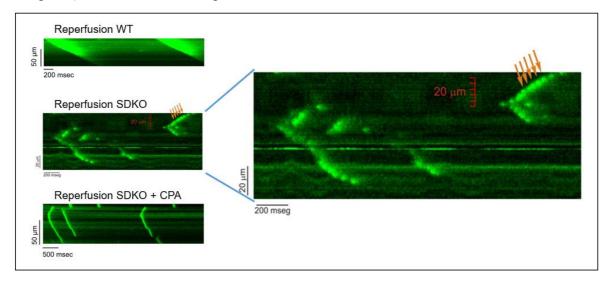


Figure 6. Protocol and typical examples of line scan recordings of epicardial Ca^{2+} waves obtained at the beginning of reperfusion in intact WT and SDKO hearts. Protocol and typical recordings of line scan before and after the addition of the SERCA2a inhibitor CPA in SDKO hearts. Red arrows indicate fragmented SR Ca^{2+} waves.

The question is then, why does this increase in SR Ca^{2+} leak fail to produce reperfusion arrhythmias as expected? As explained above, Ca^{2+} triggered arrhythmias occur because excess of Ca^{2+} leak from the SR, "travels" as Ca^{2+} waves in the cytosol and is extruded through the electrogenic NCX, producing a membrane depolarization that, if high enough, may lead to an ectopic beat. Figure 6B shows that whereas there are Ca²⁺ waves during reperfusion in S2814D hearts, in SDKO heart Ca²⁺ waves are aborted and converted into non-arrhythmogenic Ca^{2+} events, termed mini waves. Abortion of Ca²⁺ waves in these mice may occur due to the fact that the increase in SR Ca²⁺ uptake produced by PLN ablation, increases cytosolic Ca²⁺buffer [34], that would hamper the traveling of the wave as a whole. Interestingly, this mechanism was also observed to be present in cardiac myocytes treated with Istaroxime, a cardiotonic steroid that is under clinical evaluation to treat heart failure [35]. Istaroxime, which combines Na+/K+ ATPase inhibition with SERCA-mediated uptake acceleration, increases SR Ca²⁺ load but brakes Ca²⁺ wave propagation leading to non- arrhythmogenic mini-wave occurrence [36]. In contrast, recent experiments show that the dual-action (CaMKII-dependent Ca^{2+} leak and CaMKII-independent Ca²⁺-uptake) of aging -induced increase in stress kinase JNK2 (c-Jun N-terminal kinase2) enhances atrial fibrillation vulnerability [37]. In this case, the increase in cytosolic Ca²⁺ buffer capacity produced by the enhanced SR Ca²⁺-uptake seems not to be sufficient to break cytosolic Ca^{2+} waves. Instead, the increased SR Ca^{2+} uptake favors arrhythmia events by keeping SR Ca^{2+} load above the threshold necessary to enhance SR Ca^{2+} leak. The comparison of the arrhythmic island obtained with mathematical model with the Goldilocks principle, may sound counterintuitive. In contrast to the Three Beer Story, the right combination of

release and uptake is far from producing a pleasant effect as that evoked by the warm bowl of porridge. In this case, the right combination leads to unwanted results, like the enhanced possibility of threatening arrhythmias. Conversely, too much or too little, gives rise to a good result. This is not the case of some related phenomena. For instance, based on different previous works [38-40], Liu et al., [41] suggested a Goldilocks behavior in the relationship between Ca^{2+} release restitution (CRR) and arrhythmias: An increase in the velocity of CRR, predisposes to triggered arrhythmias but precludes Ca^{2+} alternans [40], whereas a slow recovery of CRR can increase the risk of alternans, i.e. recovery that is either too fast or too slow may be detrimental, whereas a rate of recovery between the

potentially dangerous extremes may be ideal. This relationship seems to hold true under a variety of conditions that exclusively affect CRR. However, our results showed that when the enhancement of CRR achieved by increasing SR Ca²⁺ uptake, as is the case of PLN ablation is high enough, triggered arrhythmias are prevented [12].

In summary, the results indicate that the arrhythmogenic effect of increasing RyR2 Po can only persist if there is an associated increase in SR Ca^{2+} uptake that allows SR Ca^{2+} load to be maintained above the necessary level (threshold) that produces leak. However, if the increase in SR Ca^{2+} uptake is high enough, it might prevent arrhythmias by aborting Ca^{2+} waves.

Perspectives

Based on the Goldilocks'principle, we can consider that two therapeutic strategies could reduce Ca^{2+} triggered arrhythmias. The first is the pharmacological reduction of RyR2 opening, with several compounds accumulating evidence about their capacity to prevent waves occurrence [42-45]. However, concerns about their "alternogenic" potential have been raised [46]. The second approach is to enhance SR Ca^{2+} uptake velocity to brake waves propagation and reduce their arrhytmogenic potential. Interestingly, it could also prevent alternans due to an acceleration of CRR. Drugs such as istaroxime, that circumvent the Goldilocks principle regarding SR Ca^{2+} release and reuptake [36], would be useful to treat heart failure, increasing contractility without the risk of triggering arrhythmias. Nevertheless, further studies are necessary to analyze the relationship not only between intracellular Ca^{2+} handling and arrhythmias, but also associated with CRR and alternans, to establish the best balance of Ca^{2+} management where electrical abnormalities are prevented.

References

- [1] Cleland JG, Chattopadhyay S, Khand A, Houghton T & Kaye GC. Prevalence and incidence of arrhythmias and sudden death in heart failure. *Heart Fail Rev* 2002; 7: 229–242.
- [2] Mozaffarian D, Anker SD, Anand I, Linker DT, Sullivan MD, Cleland JG, Carson PE, Maggioni AP, Mann DL, Pitt B et al. Prediction of mode of death in heart failure: the Seattle Heart Failure Model. *Circulation* 2007; 116: 392–398.
- [3] **Pogwizd SM & Bers DM.** Cellular basis of triggered arrhythmias in heart failure. *Trends Cardiovasc Med.* 2004; 14, 61–66.
- [4] Laurita KR & Rosenbaum DS. Mechanisms and potential therapeutic targets for ventricular arrhythmias associated with impaired cardiac calcium cycling. *J Mol Cell Cardiol*. 2008; 44: 31–43.
- [5] Luo M & Anderson ME. Mechanisms of altered Ca²⁺ handling in heart failure. *Circ Res.* 2013; 113: 690–708.
- [6] Liu N & Priori SG. Disruption of calcium homeostasis and arrhythmogenesis induced by mutations in the cardiac ryanodine receptor and calsequestrin. *Cardiovasc Res* 2008; 77: 293–301.
- [7] Ai X, Curran JW, Shannon TR, Bers DM & Pogwizd SM. Ca²⁺/calmodulin-dependent protein kinasemodulates cardiac ryanodine receptor phosphorylation and sarcoplasmic reticulum Ca²⁺ leak in heart failure. *Circ Res* 2005; 97: 1314–1322.
- [8] Said M, Becerra R, Palomeque J, Rinaldi G, Kaetzel MA, Diaz-Sylvester PL, Copello JA, Dedman JR, Mundina-Weilenmann C, Vittone L et al. Increased intracellular Ca²⁺ and SR Ca²⁺ load contribute to arrhythmias after acidosis in rat heart. Role of Ca²⁺/calmodulin-dependent protein kinase II. *Am J Physiol Heart Circ Physiol* 2008; 295: H1669–H1683.
- [9] Said M, Becerra R, Valverde CA, Kaetzel MA, Dedman JR, Mundina-Weilenmann C, Wehrens XH, Vittone L & Mattiazzi A. Calcium-calmodulin dependent protein kinase II (CaMKII): a main signal responsible for early reperfusion arrhythmias. J Mol Cell Cardiol 2011; 51: 936–944.

- [10] Chelu MG, Sarma S, Sood S,Wang S, van Oort RJ, Skapura DG, Li N, Santonastasi M, Muller FU, Schmitz W et al. Calmodulin kinase II-mediated sarcoplasmic reticulum Ca²⁺ leak promotes atrial fibrillation in mice. *J Clin Invest* 2009; 119: 1940–1951.
- [11] Gonano LA, Sepulveda M, Rico Y, Kaetzel M, Valverde CA, Dedman J, Mattiazzi A & Vila Petroff M (2011). Calcium-calmodulin kinase II mediates digitalis induced arrhythmias. *Circ Arrhythm Electrophysiol* 2011; 4: 947–957.
- [12] Valverde CA, Mazzocchi G, Di Carlo MN, Ciocci Pardo A, Salas N, Ragone MI, Felice JI, Cely Ortiz, A, Consolini AE et al. Ablation of PLN rescues reperfusion arrhythmias but exacerbates myocardium infarction in hearts with Ca²⁺/calmodulin kinase II constitutive phosphorylation of ryanodine receptors. *Cardiovasc. Res.* 2019; 115:556-569.
- [13] Hajjar RJ, Zsebo K, Deckelbaum L, Thompson C, Rudy J, Yaroshinsky A, Ly H, Kawase Y, Wagner K, Borow K et al. Design of a phase 1/2 trial of intracoronary administration of AAV1/SERCA2a in patients with heart failure. *J Card Fail* 2008; 14: 355–367.
- [14] Lukyanenko V, Subramanian S, Gyorke I, Wiesner TF & Gyorke S. The role of luminal Ca²⁺ in the generation of Ca²⁺ waves in rat ventricular myocytes. *J Physiol* 1999; 518: 173–186.
- [15] Davia K, Bernobich E, Ranu HK, delMonte F, Terracciano CM, MacLeod KT, Adamson DL, Chaudhri B, Hajjar RJ & Harding SE. SERCA2A overexpression decreases the incidence of aftercontractions in adult rabbit ventricular myocytes. *J Mol Cell Cardiol* 2001; 33: 1005–1015.
- [16] **del Monte F, Lebeche D, Guerrero JL, Tsuji T, Doye AA, Gwathmey JK & Hajjar RJ.** Abrogation of ventricular arrhythmias in a model of ischemia and reperfusion by targeting myocardial calcium cycling. *Proc Natl Acad Sci* USA 2004; 101: 5622–5627.
- [17] Landgraf G, Gellerich FN & Wussling MH. Inhibitors of SERCA and mitochondrial Cauniporter decrease velocity of calcium waves in rat cardiomyocytes. *Mol Cell Biochem* 2004; 256– 257: 379–386.
- [18] Stokke MK, Hougen K, Sjaastad I, Louch WE, Briston SJ, Enger UH, Andersson KB, Christensen G, Eisner DA, Sejersted OM & Trafford AW. Reduced SERCA2 abundance decreases the propensity for Ca²⁺ wave development in ventricular myocytes. *Cardiovasc Res* 2010; 86: 63–71.
- [19] Prunier F, Kawase Y, Gianni D, Scapin C, Danik SB, Ellinor PT, Hajjar RJ & Del Monte F. Prevention of ventricular arrhythmias with sarcoplasmic reticulum Ca²⁺ ATPase pump overexpression in a porcine model of ischemia reperfusion. *Circulation* 2008; 118: 614–624.
- [20] Bai Y, Jones PP, Guo J, Zhong X, Clark RB, Zhou Q, Wang R, Vallmitjana A, Benitez R, Hove-Madsen L et al. Phospholamban knockout breaks arrhythmogenic Ca²⁺ waves and suppresses catecholaminergic polymorphic ventricular tachycardia in mice. *Circ Res* 2013; 113: 517–526.
- [21] Liu GS, Morales A, Vafiadaki E, Lam CK, Cai WF, Haghighi K, Adly G, Hershberger RE & Kranias EG. A novel human R25C-hospholamban mutation is associated with super-inhibition of calcium cycling and ventricular arrhythmia. *Cardiovasc Res* 2015; 107: 164–174.
- [22] Mazzocchi G, Sommese L, Palomeque J, Felice JI, Di Carlo MN, Fainstein D, Gonzalez P, Contreras P, Skapura D, McCauley MD et al. Phospholamban ablation rescues the enhanced propensity to arrhythmias of mice with CaMKII-constitutive phosphorylation of RyR2-S2814 site. *J Physiol* (Lond). 2016; 594: 3005-3030.
- [23] **Kranias EG and Hajjar RJ.** Modulation of cardiac contractility by the phospholamban/SERCA2a regulatome. *Circ. Res.* 2012; 110: 1646–1660.
- [24] Landstrom AP, Dobrev D, Wehrens XHT. Calcium Signaling and Cardiac Arrhythmias. *Circ Res.* 2017;120:1969-1993

- [25] Federico M, Valverde CA, Mattiazzi A, Palomeque J. Unbalance Between Sarcoplasmic Reticulum Ca2 + Uptake and Release: A First Step Toward Ca²⁺ Triggered Arrhythmias and Cardiac Damage. *Front Physiol.* 2020; 10:1630, 1-22.
- [26] Luo W, Grupp IL, Harrer J, Ponniah S, Grupp G, Duffy JJ, Doetschman T & Kranias EG. Targeted ablation of the phospholamban gene is associated with markedly enhanced myocardial contractility and loss of beta-agonist stimulation. *Circ Res* 1994; 75: 401–409
- [27] Chu G, Ferguson DG, Edes I, Kiss E, Sato Y & Kranias EG. Phospholamban ablation and compensatory responses in the mammalian heart. *Ann NY Acad Sci* 1998; 853: 49–62.
- [28] Mundiña-Weilenmann C, Vittone L, Ortale M, Chiappe de Cingolani G, Mattiazzi A. Immunodetection of phosphorylation sites gives new insights into the mechanisms underlying phospholamban phosphorylation in the intact heart. *J. Biol. Chem.* 1996; 271: 33561-67.
- [29] Lascano EC, Said M, Vittone L, Mattiazzi A, Mundiña-Weilenmann C, Negroni JA. Role of CaMKII in post acidosis arrhythmias: a simulation study using a human myocyte model. *J Mol Cell Cardiol* 2013; 60: 172–183.
- [30] Lascano E, Mattiazzi A, Negroni JA. Mathematical Models: A precious tool for Research Physiology. *Physiol Mini Reviews*. 2015; 8: 12-22.
- [31] Lascano E, Negroni J, Vila-Petroff M, Mattiazzi A. The impact of RyR2 potentiation on myocardial function. *Am J Physiol (Heart Circ Physiol.)* 2017; 312:H1105-H1109.
- [32] Southey R. The Story of the Three Bears. In The Doctor & Co. 1837; 7 vol.
- [33] van Oort RJ, McCauley MD, Dixit SS, Pereira L, Yang Y, Respress JL, Wang Q, De Almeida AC, Skapura DG, Anderson ME et al. Ryanodine receptor phosphorylation by calcium/calmodulin-dependent protein kinase II promotes life-threatening ventricular arrhythmias in mice with heart failure. *Circulation* 2010; 122: 2669–2679.
- [34] Trafford AW, Díaz ME, Sibbring GC, Eisner DA. Modulation of CICR has no maintained effect on systolic Ca2: simultaneous measurements of sarcoplasmic reticulum and sarcolemmal Ca2 fluxes in rat ventricular myocytes. J Physiol. 2000; 522: 259–270.
- [35] Carubelli V, Zhang Y, Metra M, Lombardi C, Felker GM, Filippatos G, O'Connor CM, Teerlink JR, Simmons P, Segal R et al. Istaroxime ADHF Trial Group. Treatment with 24-hour istaroxime infusion in patients hospitalised for acute heart failure: a randomised, placebo-controlled trial. *Eur J Heart Fail*. 2020 22:1684-1693.
- [36] Racioppi MF, Burgos JI, Morell M, Gonano LA, Vila Petroff M. Cellular Mechanisms Underlying the Low Cardiotoxicity of Istaroxime. *J Am Heart Assoc.* 2021; 10: e018833.
- [37] Yan JJ, Bare D, DeSantiago J, Zhao W, Mei Y, Chen Z, Ginsburg K, Solaro JR, Wolska B, Bers DM et al. JNK2, a newly identified SERCA2 enhancer, augmenting [Ca²⁺]SR uptake and shifting an arrhythmic leak-load relationship. *Circ Res.* 2021; 128: 455–470
- [38] Xie LH, Sato D, Garfinkel A, Qu Z, Weiss JN. Intracellular Ca²⁺ alternans: coordinated regulation by sarcoplasmic reticulum release, uptake, and leak. *Biophys J* 2008; 95: 3100–10.
- [39] **Restrepo JG, Weiss JN, Karma A.** Calsequestrin-mediated mechanism for cellular calcium transient alternans. *Biophys J* 2008; 95: 3767–89.
- [40] Kornyeyev D, Petrosky AD, Zepeda B, Ferreiro M, Knollmann B, Escobar AL. Calsequestrin deletion shortens the refractoriness of Ca²⁺ release and reduces rate dependent Ca²⁺-alternans in intact mouse hearts. *J Mol Cell Cardiol* 2012 52:21-31.
- [41] Liu OZ, Lederer WJ, Sobie EA. Does the Goldilocks Principle apply to calcium release restitution in heart cells? *J Mol Cell Cardiol*, 2012; 52:3-6.
- [42] Kryshtal DO, Blackwell DJ, Egly CL, Smith AN, Batiste SB, Johnston JN, R Laver DR, Knollmann BC. RYR2 Channel Inhibition Is the Principal Mechanism of Flecainide Action in CPVT Circ Res. 2021; 128:321-331.

- [43] Zhou Q, Xiao J, Jiang D, Wang R, Vembaiyan K, Wang A, Smith CD, Xie C, Chen W, Zhang J et al. Back TG, Chen SR. Carvedilol and its new analogs suppress arrhythmogenic store overload-induced Ca²⁺ release. *Nat Med.* 2011; 10; 17:1003-9.
- [44] **Maxwell JT, Domeier TL, Blatter LA.** Dantrolene prevents arrhythmogenic Ca²⁺ release in heart failure. *Am J Physiol Heart Circ Physiol*. 2012 Feb 15; 302(4):H953-63.
- [45] Gonano LA, Sepulveda M, Morell M, Toteff T, Racioppi MF, Lascano E, Negroni J, Fernandez Ruocco MJ, Medei E, Neiman G et al. Non- beta-blocking carvedilol analog, VK- II-86, prevents ouabain- induced cardiotoxicity. *Circ J.* 2018; 83:41–51.
- [46] Zhong X, Sun B, Vallmitjana A, Mi T, Guo W, Ni M, Wang R, Guo A, Duff HJ, Gillis AM et al. Suppression of ryanodine receptor function prolongs Ca²⁺ release refractoriness and promotes cardiac alternans in intact hearts. *Biochem J*. 2016 Nov 1; 473(21):3951-3964.

ABOUT AUTHORS



Dr. Carlos A. Valverde obtained his degree in Biochemist in 2001 in La Plata, Argentina. He is actually working as Investigator of CONICET and as adjunct Professor of Physiology and Biophysics at the School of Medicine in La Plata (National University of La Plata). His main studies are focused on the role of CaMKII in ischemia/reperfusion injury and in calcium-triggered arrhythmias. He has published 25 papers on these and other topics related to cardiovascular pathophysiology. Most of their studies are done in intact perfused mouse hearts loaded with fluorescent dyes in order to study calcium dynamics and/or electrical properties.



Mg. Alejandra Cely-Ortiz is a biomedical engineer from the Manuela Beltrán University of Bogota- Colombia. She obtained her Master's degree in Biomedical Research from the National University of La Plata. She is doctoral fellow in Biological sciences area of UNLP-CONICET, Argentina. She currently studies the interplay between cardiac sarcoplasmic reticulum calcium leak and reuptake on cardiac damage during ischemia/reperfusion injury. She has two articles published, one as the first author and she has participated in 12 congresses and received special mention in 2 oral presentations.



Dr. Luis Gonano is an assistant researcher at the Centro de Investigaciones Cardiovasculares, CONICET-UNLP, Argentina and lecturer of human physiology at UNLP, Argentina. His research focuses on cardiac calcium handling and related mechanisms that trigger cardiac arrhythmia or contractile failure. Luis developed his career under the mentorship of Drs Vila Petroff and Mattiazzi and was postdoctoral fellow at the University of Otago, NZ working in the group of Dr. Peter Jones. Luis is currently treasurer of the Argentinean society of physiology (SAFIS) and works to grow his research group in La Plata in collaboration with local, regional and international researchers that share passion about cardiovascular physiology.



Dr. Elena Catalina Lascano Bachelor in Biological Sciences, PhD from the University of Buenos Aires. Independent Researcher and Associate Professor at Favaloro University Former Associate Professor at the Biomedical Engineering Institute, School of Engineering, University of Buenos Aires. Former Director of the Department of Biology at the Favaloro University. Author of 65 papers, and 165 congress presentations Has received awards from the National Academy of Medicine, the Argentine Society of Cardiology, the Argentine Federation of Cardiology, the Florencio Fiorini Foundation and MAFRE Foundation.



Dr. Alicia Mattiazzi is a consultant professor at the Faculty of Medicine, University of La Plata, and emeritus superior researcher from CONICET. She works at the Cardiovascular Research Center, Faculty of Medicine, University of La Plata. Author of more than 130 papers, her main interest focuses on calcium handling and mishandling, and calcium and CaMKII-induced arrhythmias and cardiac damage