



# REVISTA ARGENTINA DE MICROBIOLOGÍA

[www.elsevier.es/ram](http://www.elsevier.es/ram)



## BRIEF REPORT

# Fist report on fumagillin production by *Aspergillus fumigatus sensu stricto* gliotoxigenic strains recovered from raw cow milk and clinical samples in Argentina

Verónica A. Alonso<sup>a,c</sup>, Marina A. Velasco Manini<sup>b</sup>, Gabriela A. Pena<sup>a,c</sup>,  
Lilia R. Cavaglié<sup>a,c,\*</sup>

<sup>a</sup> Departamento de Microbiología e Inmunología, Facultad de Ciencias Exactas, Físico-Químicas y Naturales, Universidad Nacional de Río Cuarto, Río Cuarto, Córdoba, Argentina

<sup>b</sup> Fellow of Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Argentina

<sup>c</sup> Member of Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Argentina

Received 10 June 2020; accepted 4 March 2022

## KEYWORDS

Fumagillin;  
*Aspergillus fumigatus*;  
Raw cow milk strains;  
Clinical strains

**Abstract** In Argentina there are no reports on *Aspergillus fumigatus* fumagillin-producing strains. In this study we describe the isolation and mycotoxin production capacity of ten *A. fumigatus* strains isolated from farm and clinical samples. Farm strains were isolated from milk samples taken from dairy cows in Córdoba province, some of which were associated with subclinical mastitis. A culture medium was defined to optimize fumagillin production and a detection method was developed by HPLC chromatography. It is known that in addition to the host immune status, strain virulence is a fundamental characteristic that will determine its pathogenicity and, in this sense, fumagillin is considered to be among the virulence factors. In the present work, all the strains tested for the production of fumagillin were able to synthesize it, highlighting that the strain *A. fumigatus* RC2243, from a milk sample from a cow with clinical mastitis, was the most productive. The existence of fumagillin-producing strains represents a potential risk of mycotoxins being transferred to raw milk, constituting a public health risk.

© 2022 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## PALABRAS CLAVE

Fumagillin;  
*Aspergillus fumigatus*;

Primer reporte en Argentina sobre la producción de fumagilina por cepas de *Aspergillus fumigatus sensu stricto* productoras de gliotoxina, aisladas de leche cruda y de casos clínicos

**Resumen** En Argentina no existen reportes sobre cepas de *Aspergillus fumigatus* productoras de fumagilina. En este trabajo se describe el aislamiento y la producción de dicha micotoxina

\* Corresponding author.

E-mail address: [lcavaglié@exa.unrc.edu.ar](mailto:lcavaglié@exa.unrc.edu.ar) (L.R. Cavaglié).

<https://doi.org/10.1016/j.ram.2022.03.003>

0325-7541/© 2022 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Cepas aisladas de leche cruda;  
Cepas clínicas

por 10 cepas, provenientes del medioambiente rural y aisladas de muestras clínicas. Las cepas de origen rural fueron aisladas de vacas lecheras en tambores de la provincia de Córdoba, y algunas de esas cepas se asociaron a casos de mastitis subclínica. Se definió la composición de un medio de cultivo para optimizar la producción de fumagilina y se desarrolló un método de cromatografía HPLC para su determinación. Es conocido que, además del estado inmunitario del huésped, la virulencia de la cepa es una de las características fundamentales que determinan su potencial patogénico y, en este sentido, la fumagilina es considerada un factor de virulencia. En el presente trabajo todas las cepas estudiadas fueron capaces de sintetizarla y la cepa *A. fumigatus* RC2243, proveniente de leche de una vaca con mastitis subclínica, se destacó como la cepa más productora. La existencia de cepas productoras de fumagillina representa un riesgo potencial por el pasaje de dicha micotoxina a la leche, lo cual constituye un problema para la salud pública.

© 2022 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

*Aspergillus fumigatus* is a worldwide distributed saprophytic fungus and an opportunistic pathogen for human and animals. This fungus is of great importance to public health because it is capable of producing tremorgenic mycotoxins and inducing neurological syndromes in contaminated food handlers, both in maize silage and other contaminated animal feeds as well as in rural workers<sup>5,8</sup>. *Aspergillus fumigatus* pathogenicity depends not only on the host immune status but also on the virulence of the fungal strain, which is not given by a single factor of essential virulence, but is under polygenic control. It is known that *A. fumigatus* produces several immunosuppressive mycotoxins including gliotoxin, fumagillin, helvolic acid, fumitoxins, fumiclavines A and C, fumitremorgins, verruculogen, among others. All these mycotoxins inhibit the function of leukocytes in terms of migration, superoxide production and fungicidal activity<sup>9</sup>. Among them, fumagillin has the ability to inhibit endothelial cell proliferation acting as an anti-angiogenic factor, and suppress neutrophil function<sup>2</sup>. Our research group has demonstrated that there were no differences between animal and human isolated strains from aspergillosis cases<sup>8</sup>. In this way, they may become pathogenic to cows and also to farm handlers. In Argentina, there are currently no reports on the production capacity of fumagillin in native strains. The presence of *A. fumigatus sensu stricto* able to produce other mycotoxins such as fumagillin in raw cow milk could be a very important risk factor since milk and its by-products are destined for human consumption, therefore the aim of this study was to describe the fumagillin production ability of isolated Argentinean *A. fumigatus sensu stricto* strains from raw cow milk and clinical samples.

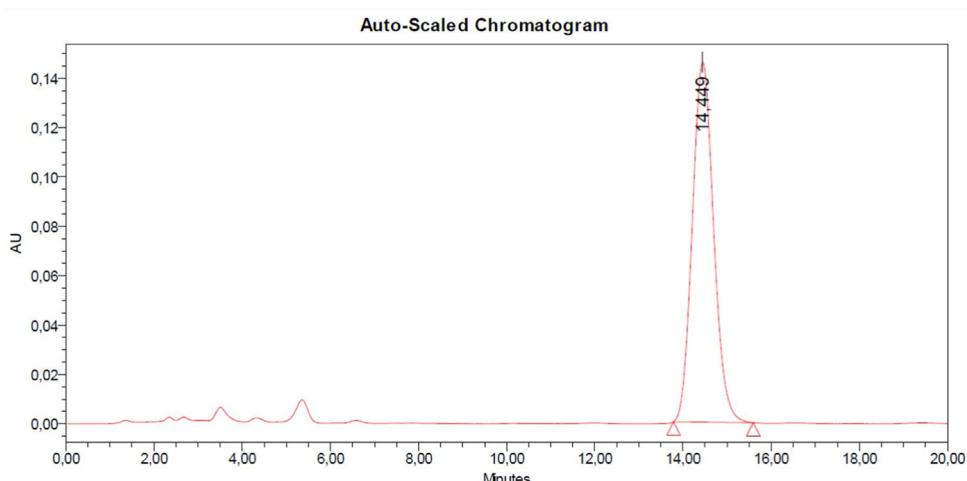
Strains correspond to a broad sampling conducted between March and September 2017; the animals sampled came from 44 dairy establishments located in Córdoba province, in the central region of Argentina. Each one of these establishments owned between 100 and 250 milking cows with daily average milk production of 10–20 l/cow. Every dairy establishment was visited once during the morning milking. In each herd, between 20% and 40% of the milking animals (Holando-Argentino breed) were randomly

sampled in accordance with the instructions recommended by the National Mastitis Council (2004)<sup>6</sup>. A total of 901 individual milking cows were sampled. The udders were disinfected with ethanol 70%, dried with individual paper towel and the first jet of milk discarded. As a result of this sampling, several fungal strains were isolated and identified. Seven of them, corresponding to *A. fumigatus* isolates were deposited in Universidad Nacional de Río Cuarto, Córdoba, Argentina Collection Center (RC). Three strains isolated from human pulmonary aspergillosis were also added to the collection. All the strains were confirmed as *A. fumigatus sensu stricto* species by morphological and molecular characterization, by sequencing the benA gene of selected strains and by a maximum parsimony analysis<sup>8</sup>.

Strains were subcultured on malt extract agar (MEA) and incubated at 25 °C for 7 days in darkness. Three agar plugs of 6 mm diameter (approx. 85 mm<sup>2</sup> of surface) were removed from different points of the colony to represent its variability, then weighed and introduced into a 250 ml Erlenmeyer flask containing 50 ml of YES medium (sucrose 150 g/l, yeast extract 20 g/l modified with 0.5 g/l of SO<sub>4</sub>Mg·7H<sub>2</sub>O, in distilled water) at 25 °C for 7 days.

The selected base medium for fumagillin production was developed by Peterson and Goldstein (1957) (succharose 30 g/l, starch 15 g/l, NaNO<sub>3</sub> 3 g/l, K<sub>2</sub>HPO<sub>4</sub> 1 g/l, MgSO<sub>4</sub>·7H<sub>2</sub>O 0.5 g/l, KCl 0.5 g/l, FeSO<sub>4</sub>·7H<sub>2</sub>O 0.01 g/l) modified by the addition of carboxymethylcellulose (CMC) 10 g/l to obtain small pellets<sup>10</sup>. Cultures were grown at 30 °C for 6 days in an orbital shaker at 220 rpm (Heidolph Rotamax 120, Heidolph Instruments GmbH, Schwabach, Germany). A 5 ml aliquot of broth was transferred to centrifuge tubes, with 5 ml of methanol; the extraction process was carried out at 30 °C and 220 rpm for 3 h. The obtained suspension was centrifuged at 3000 × g for 10 min, supernatant was filtered through a membrane filter (0.45 µm) and dried under nitrogen stream. The extracts were resuspended in 1 ml of acetonitrile for HPLC analysis.

For obtaining the calibration curve, 1 mg pure fumagillin (Sigma USA) was dissolved in 1 ml acetonitrile HPLC grade (Sintorgan S.A.). A stock solution of fumagillin 10 µg/m



**Figure 1** Chromatogram obtained with the injection of 20 µl of a 10 µg/ml solution of fumagillin.

**Table 1** Fumagillin concentration produced by *A. fumigatus* strains in Peterson-Goldstein modified medium.

Origin	Strain	Fumagillin ( $\mu\text{g} \cdot \text{ml}^{-1}$ )
Cow milk	RC2210	9.23
	RC2213	9.23
	RC2229	9.97
	RC2240	5.4
	RC2243	16.79
	RC2249	5.47
	RC2251	4.2
Human aspergillosis	RC2257*	8.93
	RC2261*	12.74
	RC2263*	7.97

and the working solutions of 100, 50, 20, 10, and 5 ng/ml were prepared. A Waters e2695 HPLC was used, with a 20 µl loading loop and equipped with a diode array detector Waters 2998; separations were carried out on a C18 Phenomenex Luna RP C18 column (150 mm × 4.6 mm, 5 µm particle size, Phenomenex Inc, CA, USA) at 25 °C. An isocratic mode was used, as the mobile phase, acetonitrile (Sintorgan S.A., Argentina) and water (Sintorgan S.A., Argentina) with 1% acetic acid were used in a 50/50 v/v ratio. The flow for the mobile phase was 1 ml/min and the wavelength used was 351 nm. Retention time was 15 min. The fumagillin concentrations were determined from the areas obtained as a function of the calibration curve. All the strains investigated were able to produce fumagillin at levels between 4.20 and 16.79 µg/ml (Table 1). A chromatogram of the fumagillin peak corresponding to the injection of 20 µl of pure standard (10 µg/ml) can be observed at 14.5 min (Fig. 1). The analyzed strains from raw cow milk samples showed a variable production capacity of fumagillin. The strain *A. fumigatus* RC2243, from a cow milk sample with clinical mastitis, showed the highest production capacity among all the tested strains. Strains from healthy cow samples yielded intermediate values between 9.23 and 9.97 µg/ml, while the lowest level

was shown in *A. fumigatus* RC2251 strain, isolated from a subclinical mastitis sample. With the purpose of comparing the toxic potential of strains from different origins, three strains of *A. fumigatus sensu stricto* from human pulmonary aspergillosis were included in the study. Among these strains, one of them had the highest production capacity (12.74 µg/ml) while the others had moderate fumagillin production capacity. Pellegrino et al. (2013)<sup>7</sup> conducted a mycological survey of the milk samples from which the strains of the current study were isolated and found that *A. fumigatus* was the second more frequently isolated species of *Aspergillus* after *A. flavus*. Moreover, they demonstrated the gliotoxinogenic ability of the strains and its association with high somatic cell counts in milk samples. The results obtained in the present work constitute a first step in the knowledge of the production capacity of fumagillin from *A. fumigatus sensu stricto* gliotoxin producer strains obtained from animal and human samples in Argentina. Boudra and Morgavi (2005)<sup>1</sup> analyzed the toxicogenic capacity of *A. fumigatus* strains in different natural feed substrates (wheat, corn, barley, triticale, rye grass, orchard grass, and tall fescue); fumagillin production levels up to 25.9 µg/ml were obtained after six days of development in a medium other than that used in the present study (minimal essential medium Eagle supplemented with fetal bovine serum and glucose). In another work, the growth and production of fumagillin by *A. fumigatus* NRRL 2436 was measured in Peterson-Goldstein medium modified with carboxymethylcellulose and similar fumagillin levels to ours were obtained (average of 13.4 mg/l)<sup>10</sup>. The medium used in the present work was useful to perform a screening among strains in order to identify those with a capacity to produce fumagillin. This knowledge is relevant due to the well-known antiangiogenic effects of fumagillin that reduce the proliferation of endothelial cells and blood vessel formation<sup>4</sup> probably related to the process of *A. fumigatus* pathogenesis in invasive aspergillosis. Guruceaga et al. (2018)<sup>3</sup> showed the gene-expression and metabolite bioactivity of the fumagillin/pseurotin gene cluster during invasive lung aspergillosis after intranasal exposure to *A. fumigatus* conidia using a murine model. They concluded

that fumagillin contributed to tissue damage during invasive aspergillosis, and allowed *A. fumigatus* progress by the lungs through angioinvasion and disruption of the lung parenchymal structure. Furthermore, it has been shown that fumagillin inhibits neutrophil function contributing to the reduction of the local immune response<sup>2</sup>. These facts confirm or increase the risk posed by the workers' exposure to fumagillin contamination. Although fumagillin toxicity to humans is important because it poses a direct risk to the consumer, this mycotoxin is on the list of products approved for use in beekeeping in several countries including Argentina, according to SENASA regulations (Cert. No. 97.117) for the treatment of nosema disease (*Nosema apis* and *Nosema ceranae*) and is commonly used. In the present work, all the strains tested for the production of fumagillin were able to synthesize it, highlighting that the strain *A. fumigatus* RC2243, from a milk sample of a cow with clinical mastitis, was the most productive. It should be noted that the strains isolated and studied in this work came from the animal environment, specifically the dairy farm environment, all of which are able to produce at least two of the most important mycotoxins that have been identified in *A. fumigatus*. This increases the possibility of being transferred to milk and polluting the environment in which they develop, constituting a public health risk. It is necessary to conduct specific trials for investigating the efficiency of different mycotoxin adsorbents and biodegradation products against several toxins that are mainly detected in forage and avoid its transfer to the human food chain.

## Conflict of interest

The authors declare that they have no conflicts of interest.

## Acknowledgements

This work was supported by grants from FONCyT-PICT 2033/15.

## References

1. Boudra H, Morgavi DP. Mycotoxin risk evaluation in feeds contaminated by *Aspergillus fumigatus*. *Anim Feed Sci Technol*. 2005;120:113–23.
2. Fallon JP, Reeves EP, Kavanagh K. Inhibition of neutrophil function following exposure to the *Aspergillus fumigatus* toxin fumagillin. *J Med Microbiol*. 2010;59:625–33, <http://dx.doi.org/10.1099/jmm.0.018192-0>.
3. Guruceaga X, Ezpeleta G, Mayayo E, Sueiro-Olivares M, Abad-Díaz-De-Cerio A, Aguirre Urízar JM, Hong GL, Wiemann P, Bok JW, Filler SG, Keller NP, Hernando FL, Ramirez-Garcia A, Rementeria A. A possible role for fumagillin in cellular damage during host infection by *Aspergillus fumigatus*. *Virulence*. 2018;9:1548–61.
4. Takayuki K, Uehara T, Osawa M, Fukumoto H, Mine S, Ueda K, Hasegawa H, Katano H. Fumagillin, a potent angiogenesis inhibitor, induces Kaposi sarcoma-associated herpesvirus replication in primary effusion lymphoma cells. *Biochem Biophys Res Commun*. 2015;463:1267–72.
5. Keller L, Keller M, Monge MP, Pereyra CM, Alonso VA, Cavagliari LR. Gliotoxin contamination in and pre- and postfermented corn, sorghum and wet brewer's grains silage in São Paulo and Rio de Janeiro State, Brazil. *J Appl Microbiol*. 2012;112:865–73.
6. National Mastitis Council. Microbiological procedures for the diagnosis of bovine udder infection and determination of milk quality. 4th ed. Madison: National Mastitis Council Publications; 2004.
7. Pellegrino M, Alonso V, Vissio C, Larriestra A, Chiacchiera S, Bogni C, Cavagliari L. Gliotoxinogenic *Aspergillus fumigatus* in the dairy herd environment. *Mycotoxin Res*. 2013;29:71–8.
8. Pena G, Alonso V, Velasco Manini M, Pellegrino M, Cavagliari L. Molecular characterization of *Aspergillus fumigatus* isolated from raw cow milk in Argentina: molecular typing of *A. fumigatus* from raw cow milk. *Int J Food Microbiol*. 2018;275:1–7.
9. Rementeria A, Lopez-Molina N, Ludwig A, Vivanco AB, Belen A, Bikandi J, Pontón J, Garaizar J. Genes and molecules involved in *Aspergillus fumigatus* virulence. *Rev Iberoam de Micol*. 2005;22:1–23.
10. Yang W, Kim WS, Fang A, Demain AL. Carbon and nitrogen source nutrition of fumagillin biosynthesis by *Aspergillus fumigatus*. *Curr Microbiol*. 2003;46:275–9.