



Hepatitis C virus genotype 4 in Southern and Central Spain does not come from recent foreign migration waves

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Title Page

Title: Hepatitis C virus genotype 4 in Southern and Central Spain does not come from recent foreign migration waves.

Authors: Di Lello Federico A.¹, Neukam Karin¹, Parra-Sanchez Manuel¹, Plaza Zulema², Soriano Vicente², Cifuentes Celia¹, Mira José A.¹, Poveda Eva², Pineda Juan A.^{1*}

Affiliations: ¹Unit of Infectious Diseases and Microbiology, Hospital Universitario de Valme, Seville, Spain. ²Department of Infectious Diseases, Hospital Carlos III, Madrid, Spain

***Corresponding author:** Dr. Juan A. Pineda. Unit of Infectious Diseases and Microbiology. Hospital Universitario de Valme. Avenida de Bellavista s/n. 41014 Seville. Spain. Phone: +34955015684. E-mail: japineda@telefonica.net.

Short Running Title: Hepatitis C Virus genotype 4 diversity in Spain

Abstract

Hepatitis C virus genotype 4 (HCV-4) is highly prevalent in Spain, but the information on the molecular characterization of HCV-4 in this region is scarce. Due to this, the molecular characteristics and the evolution of HCV-4 infection in Seville were analyzed (Southern Spain) and compared them with samples from Madrid. HCV genotype was determined by LIPA 2.0 assay and confirmed by sequence analysis of NS5B. Phylogenetic tree was estimated by MEGA 5.10. Bayesian coalescent-based methods were used to estimate the substitution rate and the age of the most recent common ancestor (MRCA). In the phylogenetic analysis of 50 NS5B HCV-4 from Seville and 11 from Madrid, two clusters were distinguished: The first cluster (HCV-4a) included 48% of the sequences from Seville and 9% of sequences from Madrid. The second cluster included the remaining sequences belonging to HCV-4d. The mean estimated substitution rate was 2.39×10^{-3} for HCV-4a and 1.81×10^{-3} for HCV-4d for Seville and 2.32×10^{-3} for HCV-4d from Madrid. The date for MRCA was estimated to be around 1981 to 1984 for HCV-4 from Seville. The dates for MRCA were dated before the recent flow of immigration in Spain. Therefore, the results presented in this study argues against the possibility of a foreign introduction of the HCV-4 from other regions with high prevalence, at least during the last two, decades in which there was a great flow of immigrants. Additionally, an unusual high prevalence of subtype 4a was observed in Seville.

Keywords: Hepatitis C virus; diversity; phylogeny; Beast; evolution.

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Introduction

Hepatitis C virus genotype 4 (HCV-4) is considered a difficult-to-treat genotype in patients infected with HCV and patients infected with HCV and HIV, because the rate of sustained virologic response (SVR) to pegylated interferon plus ribavirin is poorer than that for HCV genotypes 2 or 3 (Torriani *et al.*, 2004; Carrat *et al.*, 2004; Pineda *et al.*, 2007; Nuñez *et al.*, 2007; Mira *et al.*, 2012). Recent studies carried out in Europe have reported an increase in the prevalence of HCV-4 (Medrano *et al.*, 2011). Thus, in some Mediterranean countries the rates of prevalence for this genotype are between 10 to 24% (Morice *et al.*, 2001; Van Asten *et al.*, 2004; Nicot *et al.*, 2005; Payan *et al.*, 2005; Ansaldi *et al.*, 2005; Fernandez-Arcas *et al.*, 2006; Ramos *et al.*, 2007; Esteban *et al.*, 2008). Similar figures are observed in Spain, depending on the geographical areas and the risk group the analyzed patients belong to. Accordingly, rates from 10.2 to 18% have been reported in Southern Spain (Fernandez-Arcas *et al.*, 2006; Cifuenes *et al.*, 2012), 8.5 to 20% in Central Spain (Pérez-Olmeda *et al.*, 2002; Medrano *et al.*, 2011; Poveda *et al.*, 2012) and 8.2 to 19% in Northern Spain (Rubio *et al.*, 2001; Ramos-Sanchez *et al.*, 2003; Martinez *et al.*, 2005).

Most cases of HCV-4a infection in Europe have been attributed to the immigration from Africa (more precisely Egypt) and Middle East (de Bruijne *et al.*, 2009; Eriksen *et al.*, 2010). Egypt is one of the countries with highest prevalence of HCV infection (15%). HCV-4a accounts for 90% of infections in Egypt (Egyptian Ministry of Health, 2007; Khattab *et al.*, 2011). However, recent studies have also reported a high prevalence of HCV-4d, which has European origin (de Bruijne *et al.*, 2009; Eriksen *et al.*, 2010) and that it is spreading throughout European countries mainly in relation to illicit intravenous drug using (Fernandez-Arcas *et al.*, 2006; de Bruijne *et al.*, 2009; Eriksen *et al.*, 2010).

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4 67 Currently, there are almost 6 million of foreign-born residents in Spain. This numbers
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6 68 correspond to 12.1% of the total population (Instituto Nacional de Estadística 2012). Of
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8 69 these, 3.3 million (7% of the total population) were born outside the European Union,
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10 70 including a large proportion of immigrants from Africa (Instituto Nacional de Estadística
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12 71 2012), where genotype 4a is highly prevalent. The rest of the 2.4 million (5.1% of the
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14 72 total population) were born in another European Union member state (Instituto Nacional
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16 73 de Estadística 2012), where there are also several countries with high prevalence of
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18 74 genotype 4d. Consequently, due to the elevated immigration rate from endemic areas to
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20 75 Spain, it is important to assess the molecular epidemiology and evolution of HCV-4
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22 76 infection in this area, which was the purpose of this study. To extend the evolutionary
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24 77 analysis, the coalescence-based population genetic method was used in order to
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26 78 estimate the origin and diversification of HCV-4 in Spain.
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80 **Methods**

82 *Design and study population*

83 From November 1992 to November 2011, a total of 1523 patients with positive serology
84 for HCV were seen at the Infectious Diseases Units of a tertiary care center in Seville,
85 Spain. HCV genotype by LIPA was available in 970 (63.7%) of these patients. In this
86 retrospective cross-sectional study, those 150 (15.5%) patients infected with HCV or
87 patients infected with HCV and HIV who bore HCV genotype 4 were included. A
88 phylogenetic analysis of NS5B partial gene sequences obtained from HCV-4 was
89 performed in 50 patients selected randomly from de above population. On the other
90 hand, 11 patients infected with HCV-4 and HIV, belonging to a cohort of 424 subjects,
91 including 85 patients infected with HCV-4 and followed in the Department of Infectious
92 Diseases of a tertiary care center in Madrid, were also included in the phylogenic
93 analysis, in order to compare with sequences from Seville.

95 *Laboratory determinations*

96 HCV genotype was determined using a RT-PCR hybridization assay (Versant HCV
97 Genotype 2.0 LIPA; Siemens, Tarrytown, NY, USA).
98 The HCV NS5B partial region (positions 7816-8218 according to reference HCV-1a
99 sequence AF009606) was amplified using specific primers. For both HCV-4 subtypes,
100 the primers used were: POLEF4 (5'-GGA TCR GAG GAY GTM GTR TG- 3') and
101 POLER4 (5'TGT GAT AAA TGT CYC CCC CG 3') as outer primers and POLIF4 (5'-
102 CTG CCM ATY ARC CCC CTG AG- 3') and POLIR4 (5'-GGC AAT GGA GTG AGT
103 YTG- 3') as inner primers. PCR amplicons were directly sequenced in both senses using
104 the ABI PRISM 3100 Genetic Analyser using the ABI PRISM Rhodamine Terminator
105 reaction kit (Applied Biosystems, Foster City, CA, USA).

106 *Phylogenetic analysis*

107 NS5B sequences were subjected to alignment with CLUSTALX v2.0 software (Larkin *et*
108 *al.*, 2007). A Maximum Likelihood (ML) phylogenetic tree were estimated by MEGA 5.10
109 (Kumar *et al.*, 2008). Evolutionary models were inferred according to the Akaike
110 Information Criterion (AIC) statistics obtained with the jModeltest program (Posada and
111 Crandall 1998). The bootstrap resampling test with 10,000 replicates was performed for
112 the NJ tree and 1,000 for ML to assess the robustness of the tree arrangement. All HCV-
113 4 sequences available in GenBank and that corresponded to the amplified region in this
114 work were included in the phylogenetic analysis. D90208 (HCV-1b), AF009606 (HCV-
115 1a), AY746460 (HCV-2a) and D17763 (HCV-3a) were added as out-groups.

117 *Molecular evolutionary rate*

118 Bayesian coalescent-based methods were used to estimate the substitution rate. The
119 estimates of the rate of nucleotide substitutions per site per year (s/s/y) were obtained by
120 means of the Bayesian Markov Chain Monte Carlo (MCMC) techniques implemented in
121 the BEAST v1.7.2 program (Drummond *et al.*, 2007). Both strict and relaxed
122 (uncorrelated lognormal and uncorrelated exponential) molecular clocks were enforced
123 to estimate the rate of nucleotide substitution and the age of the most recent common
124 ancestor (MRCA). Five demographic models were applied as coalescent priors: constant
125 population size, exponential growth, expansional growth, logistic growth and Bayesian
126 skyline plot. These analyses were performed using the general time reversible
127 substitution model with gamma-distributed rates across sites and a proportion of sites
128 assumed to be invariable (GTR+G+I). The best-fitted model analyzed in this study was
129 selected with the Akaike Information Criterion (AIC) by using Modeltest Version 3.06
130 (Posada *et al.*, 1998). The length and number of MCMC were chosen, so that the
131 effective sample sizes (ESS) were above 100, indicating that the parameter space was

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132 explored sufficiently. The convergence of the parameters to a stationary distribution was
133 assessed with the TRACER v1.5 program (Rambaut *et al.*, 2011) and the statistical
134 uncertainties were summarized from the 95% highest probability density (HPD) intervals.
135 Model comparisons were performed by a Bayes Factor (BF) analysis (Suchard *et al.*,
136 2001).
137
138 Frequencies were compared using the chi-square test or the Fisher's test. The Student's
139 *t*-test and the Mann-Whitney U were used for comparing continuous variables between
140 two groups. The statistical analysis was carried out using the SPSS statistical software
141 package release 19.0 (IBM SPSS Inc, Chicago, IL, USA).

143 *Ethical aspects*

144 The study was designed and performed according to the Helsinki declaration and was
145 approved by the Ethic Committee of the participating Hospitals.

147 Results

148

149 *Characteristics of the analyzed population*

150 One hundred and fifty patients bore HCV-4 in the population from Seville. All patients
151 were born in Spain. The median age (Q1-Q3) was 37.6 (32-44) years. Among these
152 patients, 270 (48.2%) were infected with HCV and 279 (49.8%) were infected with HCV
153 and HIV. With regard to the way of transmission, 434 (77.5%) were injecting drug users
154 (IDU), in 66 (11.8%) the infections mode was unknown, in 39 (7%) blood transfusion and
155 in 3 (0.5%) patients HCV infection was considered to have been acquired by sexual
156 intercourse. Characteristics of the patients infected with HCV-4, separated by those with
157 sequenced (N=50) and no sequenced (N=100) samples, are shown in Table 1. On the
158 other hand, the median age (Q1-Q3) for patients from Madrid was 39 (37-42) years. All
159 patients from Madrid were infected with HCV and HIV.

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161 *Phylogenetic analysis and subtype distribution of HCV-4*

162 Fifty partial NS5B gene [position 7816 to 8227 according to HCV-1a-H77 AF009606,
163 (412bp)] from patients infected with HCV-4 from Seville and 11 from Madrid were
164 sequenced. The best-fit model of DNA evolution for the analyzed data was GTR + I + Γ .
165 Phylogenetic analyses (Figure 1) distinguished clearly two groups of sequences, each
166 forming a well-defined cluster. The first cluster (bootstrap=99) comprised 24 out of 50
167 (48%) sequences from Seville and 1 out of 11 (9%) from Madrid, belonging to the HCV-
168 4a subtype and included also samples from Egypt [four of them forming a separate
169 cluster (bootstrap=100)], Spain, France, Ireland and United States. The second cluster
170 (bootstrap=99) comprised the remaining 26 (52%) sequences from Seville and 10 out of
171 11 (91%) from Madrid belonging to the HCV-4d subtype and incorporated samples from

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other regions of Spain (Barcelona and Albacete), France, Canada and United States. Altogether, the subtypes determined by sequencing of NS5B were not in agreement with those obtained by Versant HCV Genotype 2.0 LIPA in 27 out of 48 (56.3%) samples infected with HCV-4. The frequency of HIV-coinfection and the way of HCV infection was not significantly different between patients with subtype 4a and those with subtype 4d. Thus, 14 (58.3%) subjects with subtype 4a and 12 (46.2%) with subtype 4d were HIV-infected ($p=0.282$). Twenty-one (91.7%) patients with subtype 4a vs. 19 (73.1%) harbouring subtype 4d were IDU ($p=0.089$).

Molecular evolutionary rate

In samples from Seville, the BF analysis favored the relaxed uncorrelated exponential molecular clock and the exponential population size over the other models for the partial NS5B analyzed herein. However, the relaxed uncorrelated lognormal molecular clock was used since is the top-performer and the estimates are very consistent across all the models compared. Lengths of MCMC of 320 million for HCV genotypes 4a and 4d from Seville were needed to reach values of ESS above 100. On the other side, for the sequences from Madrid, the BF analysis favored the relaxed uncorrelated lognormal molecular clock and the constant population size. The MRCA date and the mean estimated substitution rate for every group of sequences are showed in table 2.

Since viral recombination could affect coalescent analyses (Schierup and Hein, 2000), the SimPlot program was used to discard this possibility. No recombination was observed and absence of recombination events was confirmed by bootscanning, using a putative recombinant sequence as a query (Worobey, 2001).

Discussion:

In this study, it has been found that the distribution of HCV-4 in Seville, Southern Spain, shows a low diversity, with only two subtypes, HCV-4a and 4d, and almost without the formation of internal clusters. In contrast with the rest of Spain, as happened in the samples from Madrid analyzed in this study, there was a high prevalence of HCV-4a in Seville. The time for the MRCA of the strains from Seville was between 1981 and 1984, that is, just before the big increase of foreign immigration in Spain.

The HCV-4 subtype distribution observed in this study in Southern Spain is different to that reported previously in other regions of Spain, where subtype 4d is very predominant (Ramos-Sanchez *et al.*, 2003; Martinez *et al.*, 2005; Fernandez-Arcas *et al.*, 2006). However, in Seville, a prevalence of subtypes similar to that reported in other countries of Western Europe, such as France, Greece or Cyprus, was found (Morice *et al.*, 2001; Nicot *et al.*, 2005; Katsoulidou *et al.*, 2006; de Bruijne *et al.*, 2009; Demetriou *et al.*, 2009). In previous studies, the presence of HCV-4a in Europe was associated strongly to immigration from Egypt or Middle East (Morice *et al.*, 2001; Rubio *et al.*, 2001; Nicot *et al.*, 2005; Demetriou *et al.*, 2009). This is not the case observed in Seville, since all the patients included in this study were native from Spain and, as the phylogenetic analysis suggested, seems to be not related with the Egyptian samples.

In the sequence analysis of NS5B, which is the reference method for HCV genotyping, a perfect concordance with LIPA at genotype level was observed, but discrepant results were observed in a very high proportion of samples at subtype level. Direct sequencing of the NS5B region is preferable for the precise identification of HCV subtypes (Simmonds *et al.*, 2005). The inability of LIPA to identify the correct subtype has been

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223 reported by other studies, especially, regarding subtypes 1a and 1b, but also for
224 genotype 4 subtypes (Chen *et al.*, 2002; Nolte *et al.*, 2003; Zekri *et al.*, 2005). In this
225 work, more than one half of discordant subtypes between both methods for HCV-4
226 samples from Seville and Madrid were found. This is a critical point, as differences
227 regarding the response to therapy reported for subtypes HCV-1a and HCV-1b (Rallon *et*
228 *al.*, 2012) could also happen for different subtypes of HCV-4.

229

230 During the last two decades, Spain has been a transit place or the final destination for a
231 very important number of immigrants from other areas of the world with elevated
232 prevalence's of HCV. In spite of this fact, the infection with both subtype 4a and 4d were
233 dated between 1981 and 1984 in samples from Seville, just before the big increase of
234 immigration in Spain, which began in the middle of the nineties (Instituto Nacional de
235 Estadística 2012). As the MRCA represents the time of the diversification process of all
236 sequences included in the analysis, in the case of HCV-4 sequences from Seville (a
237 non-monophyletic group), this would imply the time at which currently circulating virus in
238 Seville started diversifying. This epidemiological information is important because the
239 evolutionary reconstructions indicate that several HCV-4a and HCV-4d strains with a
240 common ancestor originating in the early 1980 entered in Seville. This would not imply
241 that the diversification has occurred in Seville, but the polyphyletic nature of the group
242 would suggest multiple viral introductions instead. All these results suggest that the high
243 prevalence of HCV-4 found in Seville is not due to the large immigration received in the
244 last 10 to 15 years. Furthermore, the countries that contributed most to the immigrant
245 community in the recent years, such as Morocco, Romania, Germany, UK and some
246 Latin American countries, have a low prevalence of HCV-4 infection (Fay *et al.*, 2006;
247 Quarleri *et al.*, 2007; Brant *et al.*, 2010; Mora *et al.*, 2010; Sultana *et al.*, 2011; Brahim *et*
248 *al.*, 2012). Consequently, there was not a potential risk for a growth in the prevalence of

249 HCV-4 due to this issue. Nonetheless, periodical surveillance studies of the prevalence
250 of HCV-4 infection should be considered in the context of patients infected with HCV and
251 HIV and, principally, among IDUs.

252

253 This study has a limitation. The sequences of all HCV-4 infected patients could not be
254 analyzed. However, the analyzed samples were obtained at random and significant
255 differences between patients with and without sequenced virus were not observed
256 (Table 1). Moreover, the sample size was large enough to make a good molecular
257 characterization of distribution of HCV-4 subtypes and the estimation of MRCA in this
258 area.

259

260 In summary, the substitution rate calculated for HCV-4 is between the published
261 previously range (Di Lello et al, 2012) and the date for MRCA for samples from Seville
262 was dated before the big increasing of immigration in Spain. Accordingly, the results
263 presented in this study argue against the possibility of a foreign introduction of the HCV-
264 4 from other regions with high prevalence, at least during the two last decades in which
265 there was a great influx of people. Additionally, an unusual high prevalence of subtype
266 4a was observed in Seville (Southern Spain), but not in Madrid (Central Spain).

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Reference:

- Ansaldi F, Bruzzone B, Salamaso S, Rota MC, Durando P, Gasparini R, Icardi G. 2005
Different seroprevalence and molecular epidemiology pattern of hepatitis C virus
infection in Italy. *J Med Virol* 76: 327-332.
- Brahim I, Akil A, Mtairag el M, Pouillot R, Malki AE, Nadir S, Alaoui R, Njouom R, Pineau
P, Ezzikouri S, Benjelloun S. 2012. Morocco underwent a drift of circulating hepatitis C
virus subtypes in recent decades. *Arch Virol* 157: 515-520.
- Brant LJ, Ramsay ME, Tweed E, Hale A, Hurrelle M, Klapper P, Ngui SL; Sentinel
Surveillance of Hepatitis Testing Group. 2010. Planning for the healthcare burden of
hepatitis C infection: Hepatitis C genotypes identified in England, 2002-2007. *J Clin Virol*
48: 115-119.
- Carrat F, Bani-Sadr F, Pol S, Rosenthal E, Lunel-Fabiani F, Benzekri A, Morand P,
Goujard C, Pialoux G, Piroth L, Salmon-Céron D, Degott C, Cacoub P, Perronne C;
ANRS HCO2 RIBAVIC Study Team. 2004. Pegylated interferon alfa-2b, plus ribavirin,
for chronic hepatitis C in HIV-infected patients. A randomized controlled trial. *JAMA* 292:
2839-2848.
- Chen Z, Weck KE. 2002. Hepatitis C virus genotyping: interrogation of the 5'
untranslated region cannot accurately distinguish genotypes 1a and 1b. *J Clin Microbiol*
40: 3127-3134.
- Cifuentes C, Mira JA, Vargas J, Neukam K, Escassi C, García-Rey S, Gilabert I,
González-Monclova M, Bernal S, Pineda JA. 2012. Prevalence of hepatitis virus

1
2
3 302 infection markers in HIV-infected patients in Southern Spain. *Enferm Infecc Microbiol*
4
5 303 *Clin* 30: 452-457.
6
7 304
8
9
10 305 de Bruijne J, Schinkel J, Prins M, Koekkoek SM, Aronson SJ, van Ballegooijen MW,
11
12 306 Reesink HW, Molenkamp R, van de Laar TJ. 2009. Emergence of hepatitis C virus
13
14 307 genotype 4: phylogenetic analysis reveals three distinct epidemiological profiles. *J Clin*
15
16 308 *Microbiol* 47:3832-3838.
17
18 309
19
20 310 Demetriou VL, van de Vijver DAMC, The Cyprus HCV Network, and Kostrikis LG. 2009.
21
22 311 Molecular Epidemiology of Hepatitis C Infection in Cyprus: Evidence of Polyphyletic
23
24 312 Infection. *J Med Virol* 81:238–248.
25
26 313
27
28 314 Di Lello FA, Macias J, Plaza Z, García-Rey S, Soriano V, Cifuentes C, González M del
29
30 315 M, Parra-Sánchez M, Labarga P, Recio E, Poveda E, Pineda JA. 2012. No influence of
31
32 316 antiretroviral therapy on the mutation rate of the HCV NS5B polymerase in HIV/HCV-
33
34 317 coinfecting patients. *Antiviral Res* 2012; 95: 67-71.
35
36 318
37
38 319 Drummond AJ, Rambaut A. 2007. “BEAST”: Bayesian evolutionary analysis by sampling
39
40 320 trees. *BMC Evo Biol* 7: 214.
41
42 321
43
44 322 Egyptian Ministry of Health. Egyptian Ministry of Health Annual Report: 2007. Available
45
46 323 at <http://www.mohp.gov.eg/Main.asp> (accessed 6 July 2012).
47
48 324
49
50 325 Eriksen MB, Jørgensen LB, Krarup H, Laursen AL, Christensen PB, Møller A,
51
52 326 Schlichting P, Kuiken C, Bukh J, Weis N; DANHEP Group. 2010. Molecular and
53
54
55
56
57
58
59
60

- 327 epidemiological profiles of hepatitis C virus genotype 4 in Denmark. *J Med Virol* 82:
328 1869-1877.
329
330 Esteban JI, Sauleda S, Quer J. 2008. The changing epidemiology of hepatitis C virus
331 infection in Europe. *J Hepatol* 48: 148-162.
332
333 Fay O, Rey J, Vladimirovsky S. 2006. Epidemiology of HCV infection in Argentina. *Acta*
334 *Gastroenterol Latinoam* 36 Suppl 1:S10-12.
335
336 Fernández-Arcas N, López-Siles J, Trapero S, Ferraro A, Ibáñez A, Orihuela F,
337 Maldonado J, Alonso A. 2006. High prevalence of hepatitis C virus subtypes 4c and 4d
338 in Malaga (Spain): phylogenetic and epidemiological analyses. *J Med Virol* 78: 1429-
339 1435.
340
341 Katsoulidou A, Sypsa V, Tassopoulos NC, Boletis J, Karafoulidou A, Ketikoglou I,
342 Tsantoulas D, Vafiadi I, Hatzis G, Skoutelis A, Akriviadis E, Vasiliadis T, Kitis G,
343 Magiorkinis G, Hatzakis A. 2006. Molecular epidemiology of hepatitis C virus (HCV) in
344 Greece: temporal trends in HCV genotype-specific incidence and molecular
345 characterization of genotype 4 isolates. *J Viral Hepat* 13: 19-27.
346
347 Khattab MA, Ferenci P, Hadziyannis SJ, Colombo M, Manns MP, Almasio PL, Esteban
348 R, Abdo AA, Harrison SA, Ibrahim N, Cacoub P, Eslam M, Lee SS. 2011. Management
349 of hepatitis C virus genotype 4: Recommendations of an international expert panel. *J*
350 *Hepatol* 54: 1250-1262.

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352 Kumar S, Nei M, Dudley J, Tamura K. 2008. MEGA: A biologist-centric software for
353 evolutionary analysis of DNA and protein sequences. *Brief Bioinform* 9: 299-306.

354 Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H,
355 Valentin F, Wallace IM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG. 2007.
356 CLUSTAL W and CLUSTAL X version 2.0. *Bioinformatics* 23: 2947-2948.

357

358 Martinez SM, Delgado MB, Castroagudín JF, Aguilera A. High Rate of Infection with
359 Hepatitis C Virus Genotype 4 in a District of Galicia, Spain. *J Clin Microbiol* 2005; p.
360 5403-5404.

361

362 Medrano J, Resino S, Vispo E, Madejón A, Labarga P, Tuma P, Martín-Carbonero L,
363 Barreiro P, Rodriguez-Novoa S, Jiménez-Nacher I, Soriano V. 2011. Hepatitis C Virus
364 (HCV) treatment uptake and changes in the prevalences of HCV genotypes in HIV/HCV
365 coinfectd patients. *J viral Hepat* 18: 325-330.

366

367 Mira JA, Rivero A, de Los Santos-Gil I, López-Cortés LF, Girón-González JA, Márquez
368 M, Merino D, del Mar Vilorio M, Téllez F, Ríos-Villegas MJ, Omar M, Rivero-Juárez A,
369 Macías J, Pineda JA; Grupo HEPAVIR de la Sociedad Andaluza de Enfermedades
370 Infecciosas (SAEI). 2012. Hepatitis C virus genotype 4 responds better to pegylated
371 interferon plus ribavirin than genotype 1 in hiv-infected patients. *AIDS* 26: 1721-1724.

372

373 Mora MV, Romano CM, Gomes-Gouvêa MS, Gutiérrez MF, Carrilho FJ, Pinho JR. 2010.
374 Molecular characterization, distribution, and dynamics of hepatitis C virus genotypes in
375 blood donors in Colombia. *J Med Virol* 82: 1889-1898.

376

- 377 Morice Y, Roulot D, Grando V, Stirnemann J, Gault E, Jeantils V, Bentata M, Jarrousse
378 B, Lortholary O, Pallier C, Dény P. 2001. Phylogenetic analyses confirm the high
379 prevalence of hepatitis C virus (HCV) type 4 in the Seine-Saint-Denis district (France)
380 and indicate seven different HCV-4 subtypes linked to two different epidemiological
381 patterns. *J Gen Virol* 82: 1001-1012
- 382
- 383 Nicot F, Legrand-Abravanel F, Sandres-Saune K, Boulestin A, Dubois M, Alric L, Vinel
384 JP, Pasquier C, Izopet J. 2005. Heterogeneity of hepatitis C virus genotype 4 strains
385 circulating in south-western France. *J Gen Virol* 86: 107-114.
- 386
- 387 Nolte FS, Green AM, Fiebelkorn KR, Caliendo AM, Sturchio C, Grunwald A, Healy M.
388 2003. Clinical evaluation of two methods for genotyping hepatitis C virus based on
389 analysis of the 5' noncoding region. *J Clin Microbiol* 41: 1558-1564.
- 390
- 391 Núñez M, Miralles C, Berdún MA, Losada E, Aguirrebengoa K, Ocampo A, Arazo P,
392 Cervantes M, de Los Santos I, San Joaquín I, Echeverría S, Galindo MJ, Asensi V,
393 Barreiro P, Sola J, Hernandez-Burruezo JJ, Guardiola JM, Romero M, García-
394 Samaniego J, Soriano V; PRESCO Study Group. 2007. Role of weight-based ribavirin
395 dosing and extended duration of therapy in chronic hepatitis C in HIV-infected patients:
396 the PRESCO trial. *AIDS Res Hum Retroviruses* 23: 972-982.
- 397
- 398 Payan C, Roudot-Thoraval F, Marcellin P, Bled N, Duverlie G, Fouchard-Hubert I,
399 Trimoulet P, Couzigou P, Cointe D, Chaput C, Henquell C, Abergel A, Pawlotsky JM,
400 Hezode C, Coudé M, Blanchi A, Alain S, Loustaud-Ratti V, Chevallier P, Trepo C,
401 Gerolami V, Portal I, Halfon P, Bourlière M, Bogard M, Plouvier E, Laffont C, Agius G,
402 Silvain C, Brodard V, Thieffn G, Buffet-Janvresse C, Riachi G, Grattard F, Bourlet T,

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53
54
55
56
57
58
59
60

403 Stoll-Keller F, Doffoel M, Izopet J, Barange K, Martinot-Peignoux M, Branger M,
404 Rosenberg A, Sogni P, Chaix ML, Pol S, Thibault V, Opolon P, Charrois A, Serfaty L,
405 Fouqueray B, Grange JD, Lefrère JJ, Lunel-Fabiani F. 2005. Changing of hepatitis C
406 virus genotype patterns in France at the beginning of the third millennium: the GEMHEP
407 Geno CII Study. *J Viral Hepat* 12: 405-413.
408
409 Pérez-Olmeda M, Ríos P, Núñez M, García-Samaniego J, Romero M, Soriano V. 2002.
410 Virological characteristics of hepatitis C virus infection in HIV-infected individuals with
411 chronic hepatitis C: implications for treatment. *AIDS* 16: 493-495.
412
413 Pineda JA, Mira JA, de los Santos-Gil I, Valera-Bestard B, Rivero A, Merino D, Girón-
414 González JA, Ríos-Villegas MJ, González-Serrano M, Collado A, García-García JA,
415 Carrillo-Gómez R, López-Cortés LF, Gómez-Mateos J. 2007. Influence of concomitant
416 antiretroviral therapy on the rate of sustained virological response to pegylated interferon
417 plus ribavirin in hepatitis C virus/HIV-coinfected patients. *J Antimicrob Chemother* 60:
418 1347-1354.
419
420 Plaza Z, Soriano V, Gonzalez MM, Di Lello FA, Macias J, Labarga P, Pineda JA, Poveda
421 E. 2011. Impact of antiretroviral therapy on the variability of the HCV NS5B polymerase
422 in HIV-HCV co-infected patients. *J Antimicrob Chemother* 66: 2838-2842.
423
424 Posada D, Crandall KA. 1998. Modeltest: testing the model of DNA substitution.
425 *Bioinformatics* 14: 817-818.
426

- 427 Poveda E, Vispo E, Barreiro P, de Mendoza C, Labarga P, Fernández-Montero JV,
428 Martín-Carbonero L, Soriano V. 2012. Predicted effect of direct acting antivirals in the
429 current HIV-HCV-coinfected population in Spain. *Antivir Ther.* 2012;17:571-575.
430
- 431 Quarleri JF, Bolcic FM, Bouzas MB, Laufer N, Gómez Carrillo M, Mammana L, Kaufman
432 S, Pérez H, Cahn P, Salomon H. 2007. HCV Genotype distribution among HIV co-
433 infected individuals in Argentina: relationship with host and viral factors. *Acta*
434 *Gastroenterol Latinoam* 37:76-83.
435
- 436 Rallon NI, Pineda JA, Soriano V, Neukam K, Vispo E, Rivero A. 2012. Differences in
437 virological response to peginterferon- plus ribavirin in HIV-positive patients coinfected
438 with HCV subtypes 1a or 1b. *J Acquir Immune Defic Syndr* 60: 117-123.
439
- 440 Rambaut A, Drummond AJ. Tracer v1.5, Available from <http://beast.bio.ed.ac.uk/Tracer>
441 (Accessed at October 20, 2012).
442
- 443 Ramos B, Núñez M, Toro C, Sheldon J, García-Samaniego J, Ríos P, Soriano V. 2007.
444 Changes in the distribution of hepatitis C virus (HCV) genotypes over time in Spain
445 according to HIV serostatus: Implications for HCV therapy in HCV/HIV-coinfected
446 patients. *J Infect* 54: 173-179.
447
- 448 Ramos-Sánchez MC, Torío-Cabezón R, Mazón-Ramos MA, Martín-Gil FJ, del Alamo M.
449 2003. Hepatitis C virus genotype 4 in a North-west Spain district. *J Clin Virol* 28: 223-
450 224.
451

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9
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42
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46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

452 Rubio M, Rubio C, Nogués A, Manonelles A. 2001. Hepatitis C virus genotypes. Study of
453 302 patients coinfectd by the human immunodeficiency virus. Med Clin (Barc). 116:
454 650-651.
455
456 Schierup MH, and Hein J. 2000. Consequences of recombination on traditional
457 phylogenetic analysis. Genetics 156:879-879.
458
459 Sección prensa / Notas de prensa publicadas <http://www.ine.es/prensa/np710.pdf>.
460 (accessed 27 July 2012)
461
462 Simmonds P, Bukh J, Combet C, Deleage G, Enomoto N, Feinstone S, Halfon P,
463 Inchauspé G, Kuiken C, Maertens G, Mizokami M, Murphy DG, Okamoto H, Pawlotsky
464 JM, Penin F, Sablon E, Shin-I T, Stuyver LJ, Thiel HJ, Viazov S, Weiner AJ, Widell A.
465 2005. Consensus Proposals for a Unified System of Nomenclature of Hepatitis C Virus
466 Genotypes. Hepatology 42: 962-973.
467
468 Suchard MA, Weiss RE, Sinsheimer JS. 2001. Bayesian selection of continuous time
469 Markov chain evolutionary models. Mol Biol Evol 18: 1001-1013.
470
471 Sultana C, Oprisan G, Szmal C, Vagu C, Temereanca A, Dinu S, Teleman MD, Ruta S.
472 2011. Molecular Epidemiology of Hepatitis C Virus Strains from Romania. J
473 Gastrointestin Liver Dis 20: 261-266.
474
475 Torriani FJ, Rodríguez-Torres M, Rockstroh JK, Lissen E, González-García J, Lazzarin
476 A, Carosi G, Sasadeusz J, Katlama C, Montaner J, Sette H Jr, Passe S, De Pamphilis J,
477 Duff F, Schrenk UM, Dieterich DT; APRICOT Study Group. 2004. Peginterferon alfa-2a

1
2
3 478 plus ribavirin for chronic hepatitis C virus infection in HIV-infected patients. N Engl J Med
4
5 479 351: 438-450.
6
7 480
8
9
10 481 Van Asten L, Verhaest I, Lamzira S, Hernandez-Aguado I, Zangerle R, Rezza G, Broers
11
12 482 B, Robertson JR, Brettle RP, McMenemy J, Prins M, Cochrane A, Simmonds P,
13
14 483 Coutinho RA, Bruisten S; European and Italian Seroconverter Studies. 2004. Spread of
15
16 484 Hepatitis C Virus among European Injection Drug Users Infected with HIV: A
17
18 485 Phylogenetic Analysis. J infect Dis 189: 292-302.
19
20 486
21
22
23 487 Worobey M A novel approach to detecting and measuring recombination: new insights
24
25 488 into evolution in viruses, bacteria and mitochondria. Mol Biol Evol 2001; 18:1425-1434.
26
27 489
28
29 490 Zekri AR, El-Din HM, Bahnassy AA, El-Shehabi AM, El-Leethy H, Omar A, Khaled HM.
30
31 491 2005. TRUGENE sequencing versus INNO-LiPA for sub-genotyping of HCV genotype-4.
32
33 492 J Med Virol 75: 412-20.
34
35
36 493
37
38
39
40
41
42
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44
45
46
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Legend Figure:

Figure 1: Phylogenetic tree of 50 HCV-4 sequences from Seville (412 nt) and 11 from Madrid. The numbers at each node correspond to bootstrap values from a ML tree obtained with 1,000 replicates (bootstrap values lower than 70 are not shown). HCV-4 sequences from France (27), Ireland (1), USA (8), Canada (2), Spain (3), Egypt (11) and Portugal (1) were included in the phylogenetic analysis. D90208 (HCV-1b), AF009606 (HCV-1a), AY746460 (HCV-2a) and D17763 (HCV-3a) were added as out-groups. Full circles indicated samples from Seville and Empty circle indicated samples from Madrid. The end of each sample name corresponds to the HCV subtype.

Table 1: Characteristics of the patients with chronic hepatitis C genotype 4 (n=150) separated by those with sequenced and no sequenced virus.

Parameter	No Sequenced (N=100)	Sequenced (N=50)	P
Age (years)*	37 (31.7-42.8)	38 (34.8-44.1)	0.256
Male gender no. (%)	87 (87)	39 (78)	0.156
HIV Positive no. (%)	61 (61)	26 (52)	0.412
Subtype by LIPA no. (%)			
A	2 (2)	3 (6)	
A/B	1 (1)	0 (0)	
A/C/D	28 (28)	8 (16)	
B	2 (2)	2 (4)	
C/D	35 (35)	23 (46)	
E	1 (1)	1 (2)	
Undetermined	31 (31)	13 (26)	0.404
Transmission no. (%)			
IDU	87 (87)	39 (78)	
Unknown	10 (10)	6 (12)	
Sexual	0 (0)	2 (4)	
Transfusion	1 (1)	1 (2)	0.242

*Median (quartile 1-quartile 3).

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Table 2: Estimates of the MRCA for the NS5B gene by Bayesian coalescent.

N° of NS5B Sequences	Place	HCV-4 Subtype	MRCA (HPD 95%)	Mean Substitution Rate (HPD 95%)
24	Seville	4a	1981 (1947 / 1999)	1.32×10^{-3} (2.81×10^{-4} - 2.63×10^{-3})
26	Seville	4d	1984 (1940 / 1999)	1.77×10^{-3} (2.16×10^{-4} - 3.24×10^{-3})
10	Madrid	4d	1988 (1966 / 2002)	2.72×10^{-3} (5.91×10^{-4} - 4.82×10^{-3})

MRCA: Most Recent Common Ancestor

HPD 95%: Highest probability density 95%.

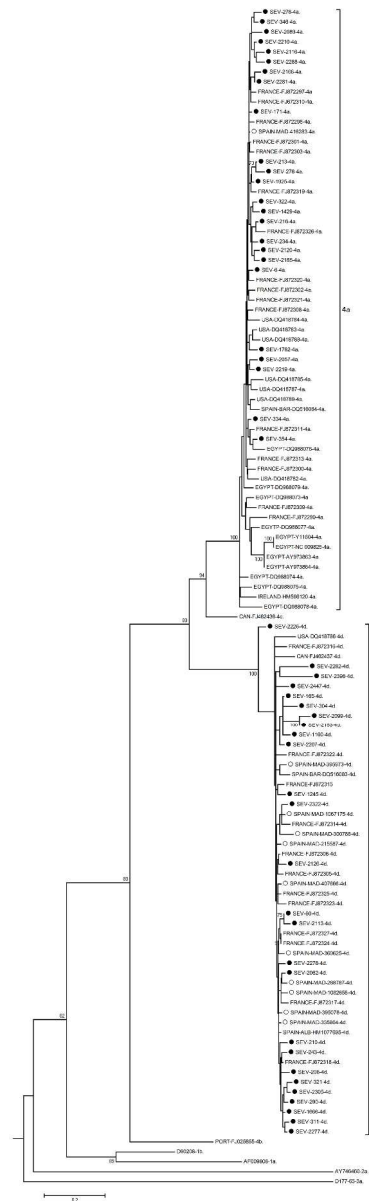


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