







http://pubs.acs.org/journal/acsodf

Mini-Review

Evolution of the Synthesis of Remdesivir. Classical Approaches and **Most Recent Advances**

Didier F. Vargas, Enrique L. Larghi,* and Teodoro S. Kaufman*

Metrics & More



ACCESS

Cite This: https://doi.org/10.1021/acsomega.1c03082



Optimization of OPG reagents, reactions **PGO** TMSCN PG= Bn, TBDMS Remdesivir (GS-5734)

ABSTRACT: The broad-spectrum antiviral Remdesivir, a monophosphate nucleoside analogue prodrug (ProTide), was repurposed. In May 2020, it received emergency approval by the FDA, being the first drug approved to fight the new coronavirus (COVID-19) disease which targets the virus directly. The main synthetic strategies toward Remdesivir, and their relevant modifications, are presented and discussed, to provide a panoramic view of the state-of-the-art and the more important advances in this field. Recent progress, proposed improvements, and uses of novel technologies for the synthetic sequence are also detailed.

R= CI, 4NO₂C₆H₄O, F₅C₆H₄O

1. INTRODUCTION

In 2012, Cho et al. discovered that C-nucleotide mimics containing 1'-substituted 4-aza-7,9-dideazaadenosine are analogues of ATP with potent activity against RNA viruses. The concomitant outbreak of the Ebola epidemics of 2014 prompted the development of new specific antivirals.²

The incorporation of phosphate groups into drugs is problematic, due to their high metabolic lability and charged nature, which hinder membrane crossing. Therefore, they are synthesized as prodrugs, such as phosphoramidates, where metabolically labile protecting groups replace the acidic oxygens of the phosphate. This results in compounds which are uncharged, more lipophilic, and less prone to phosphoesterase-mediated hydrolysis; they can be converted into the bioactive forms inside the cells.

Remdesivir (1, Scheme 1), a phosphoramidate prodrug nucleotide (ProTide), is a result of this approach. Developed by Gilead Sciences Inc. as GS-5734 since 2009 to fight the hepatitis C virus, the drug was later claimed as active against the Ebola virus and others, including those causing SARS and MERS, as well as against the Hendra, Junin, Lassa, Nipah, Marburg, Zika, and respiratory syncytial viruses.

In vitro, 1 blocks virus infection at low micromolar concentration with high selectivity index. These bioactivity data and the preexistence of safety and clinical data eased FDA approval of 1 as a repurposed drug for treatment of severe COVID-19 cases, being the first approved treatment that directly targets the virus.

Scheme 1. Retrosynthetic Analysis of Remdesivir (1) Intermediate 3 (GS-441524), and Bioactive Form 4 (GS-443902)

Article Recommendations

June 11, 2021 Received: Accepted: July 14, 2021



The pharmaceutical use of 1 instead of compound 3 improves the potency and efficacy of the treatment. The ProTide 1 can enter the cells (passive diffusion or via the nucleoside transporter), while the permeabilities of 3 and its monophosphate derivative GS-704277 (3a) are much lower. Inside the cells, 1 undergoes an esterase-mediated *in vivo* bioactivation, ultimately leading to the monophosphate 3a. Phosphate loss under the action of phosphatase or nucleosidase gives the adenosine analogue GS-441524 (3), which has shown *in vivo* efficacy in a veterinary setting. In the cells, the monophosphate 3a (a species which avoids the rate-limiting first phosphorylation step) can also be in equilibrium with the nucleoside 3.

Then, a kinase-mediated phosphorylation yields the active form, the triphosphate 4 (GS-443902). The latter confuses the RNA-dependent viral RNA polymerase, decreasing viral RNA production; when incorporated into viral RNA, it compromises UTP uptake in the first transcription, causing chain termination in the second transcription.

Remdesivir has low aqueous solubility and suffers fast first pass clearance in the liver. Therefore, it is available in a parenteral dosage form (Veklury), which contains sodium β -cyclodextrin sulfobutyl ether. However, since the SARS-CoV-2 virus infects multiple tissues, its parenteral use ensures higher plasmatic drug levels, improving its exposure and distribution among the target tissues.

2. SYNTHESIS OF THE KEY FRAGMENTS OF REMDESIVIR

Retrosynthetically, 1 can be disconnected into an activated phosphoramidate (2) and an adenosine mimic (3), which in turn can be further retrosynthetically disassembled to unveil an activated pyrrolo[2,1-f][1,2,4]triazin-4-amine (6) along with a suitably protected D-ribono-1,4-lactone precursor (5) and a cyanide source (7).

2.1. Pyrrolo[2,1-f][1,2,4]triazine-4-amine Core. Klein et al. (Scheme 2) provided the first concise and efficient approach⁸ to this long known heterocycle (12). ⁹ 2-Formylpyrrole (8) was

Scheme 2

"Reagents and conditions: (a) 9, POCl₃, DMF, 20 °C, 1 h; (b) 8, NH₂OH, Ac₂O, Py, H₂O, 90 °C, 16 h (11, 93% from 8) or 9, HOSA, KOH, H₂O (10, 43%; 11, 37%) or 9, HOSA, NaHCO₃, H₂O, rt, 1 h (12, 92%); (c) NaH, ClNH₂ (71%); (d) HC(=NH)NH₂·AcOH, K₂CO₃, EtOH, reflux (66%) or HC(=NH)NH₂·AcOH, K₃PO₄, EtOH, reflux, 18 h (81%); (e) *t*-BuOC(O)NHNH₂, HCl, dioxane, 90°C (59%); (f) 1. ClSO₂NCO, MeCN, 5 °C, 45 min; 2. DMF, 0 °C, 45 min (77%); (g) 4 N HCl (8 equiv), dioxane, rt, 5 h (85%); (h) 14, DMF, -20°C, 1.5 h (90%); (i) NIS, 0 °C, 1 h (95%) or ICl, DMF -25 °C \rightarrow -10 °C, 3 h (90%) or 1. I₂, Py, EtOAc, 10 °C, 1 h; 2. H₂O₂, 20 °C, 10 h (85%).

treated with hydroxylamine-O-sulfonic acid (HOSA) in aqueous KOH to give 1-amino-2-pyrrolonitrile (10) in 43% yield, along with 37% yield of 2-pyrrolonitrile (11). Next, 10 was cyclocondensed with formamidine acetate in refluxing ethanol under mild basic conditions (K_2CO_3), furnishing 12 in 66% yield.

However, for the synthesis of Remdesivir, 12 was prepared from 2,5-dimethoxytetrahydrofuran (15) and *tert*-butyl carbazate in dioxane at 90 °C under HCl catalysis to give 16 (59% yield). This was exposed to chlorosulfonyl isocyanate in MeCN, resulting in 67% yield of nitrile 17 after reaction with DMF, which converted the *N*-chlorosulfonyl amide intermediate into a nitrile moiety.

Acid-mediated deprotection of 17 with HCl in dioxane gave an improved yield of 10 (85%),¹¹ which in turn delivered 12 in 81% yield in three crops, when the cyclocondensation with formamidine acetate was performed using potassium phosphate as base (31% overall yield from 15).

Finally, the heterocycle was selectively brominated with 1,3-dibromo-5,5-dimethylhydantoin (14) in DMF to give 13 in 90% yield. This heterocycle was employed for the first-generation synthesis of Remdesivir. Other Remdesivir generations and their modifications used 18, obtained in 95% yield by iodination of 12 with *N*-iodosuccinimide (NIS). Alternative procedures toward 18 employed ICl (90% yield) and molecular iodine (85% yield) as iodinating reagents.

Remdesivir sparked renewed interest in the synthesis of 12. In an improved alternative (39% overall yield), ¹⁷ the reaction of 8 and HOSA with NaHCO₃ afforded 11 (92% yield), and 10 was obtained in 71% yield by *N*-amination with NaH and ClNH₂. However, the approach of Snead et al. ¹⁸ is the best one. It is based on a one-pot oxidative Vilsmeier cascade from pyrrole (9) to afford 10 through the intermediacy of 8 and further cyclocondensation of 10 to 12 with formamidine acetate (59% overall yield).

2.2. D-Ribonolactone Moiety. The D-ribono-1,4-lactone (19) derivative **22** used for the first-generation synthesis of Remdesivir was obtained employing the Albright—Goldman oxidation (Ac_2O , DMSO) of lactol **21** (96% yield), ¹⁹ which in turn can be accessed in high yield from D-ribose (**20**). For the second-generation synthesis, lactone **22** was prepared (\sim 95% yield), using a TEMPO-catalyzed NaClO oxidation (Scheme 3).²⁰

Scheme 3

"Reagents and conditions: (a) 1. MeOH, H_2SO_4 , rt (79%); 2. BnBr, NaH, DMF, rt; 3. AcOH, H_2O , 100 °C (95%); (b) DMSO, Ac₂O, rt, 48 h (96%) or NaClO, KBr, K_2HPO_4 , TEMPO, MTBE· H_2O (\sim 4:1), 1 °C (\sim 95%); (c) CCl₃C(NH)OBn, TfOH, dioxane, 0 °C, 3 h (80%); (d) TBDMSCl, imidazole, DMF, 65 °C, 5 h (96%); (e) SOBr₂, Py, CH₂Cl₂, rt, 1 h (93%).

Oxidants such as pyridinium chlorochromate (PCC)²¹ have also been used.²² Given its wide interest, additional syntheses of 22 have been reported, including one which proceeds through the debenzylative lactonization of 23 with SOBr₂.²³ On the other hand, compound 24, used for the third-generation synthesis of Remdesivir, was prepared by conventional silylation of 19.²⁴

2.3. Phosphoramidate Fragment. Three generations of the key phosphoramidate fragment have been reported. The product of the first generation (26) was obtained in two steps and 83% yield by reaction of alanine 2-ethylbutylester hydrochloride (25) with phenyl dichlorophosphate in CH_2Cl_2 at -78 °C (Scheme 4).

Scheme 4

^aReagents and conditions: (a) Cl₂P(O)OPh, Et₃N, CH₂Cl₂, −78 °C → rt, 3 h; (b) 4-NO₂C₆H₄OH, Et₃N, 0 °C → rt, 3 h (66% overall); for **30**: C₆F₅OH, Et₃N, 0 °C → rt, 3 h (**30**); (c) for **28**: *i*-Pr₂O (**28**, 39%) or *i*-PrOAc, heptane, DBU (10% v/v), 0 °C, 21 h; for **30**: *i*-PrOAc, heptane, Et₃N, 0 °C, 10 h.

The second-generation phosphoramidate (28) was accessed by *in situ* exposure of 26 to 4-nitrophenol. This furnished 27 in 80% yield, which was fractionally crystallized from di-isopropyl ether to give (S_p,S) -28 in 39% yield, as proven by single-crystal X-ray diffraction.

A third-generation synthesis of phosphoramidates based on a dynamic kinetic resolution approach was first disclosed in a 2016 patent by Gilead, without informing yields. The method involved preparation of **26** in *i*-PrOAc at -20 °C and its *in situ* conversion into $(R_{\rm p}/S_{\rm p})$ -**27** by reaction with 4-nitrophenol under Et₃N promotion.

The reaction was quenched and subjected to acid and base washings; next, the organic phase was concentrated and then diluted with heptane and treated with 10% of a base like DBU (0 $^{\circ}$ C, 21 h), which caused isomerization of the phosphorus stereocenter. Seeding crystals of (S_p,S) -28 induced selective crystallization of this less soluble diastereomer, driving the isomeric conversion to completion. The yield of this scalable process was not reported.

Analogously, exposure of **26** to per-fluorophenol gave **29**. Kinetic resolution of **29** in hexane-isopropyl acetate, to which Et_3N was added to favor isomerization of the phosphorus stereocenter, provided the (S_p,S) -isomer (**30**).

3. SYNTHETIC ROUTES TOWARD REMDESIVIR

Gilead devised three main generations of the same strategy toward Remdesivir. They were partially reviewed in contributions with different scopes.^{26,27} The approval of the drug for COVID-19 treatment resulted in additional synthetic contributions since the pandemic's outbreak.

3.1. First Generation. A New Antiviral Prodrug. The first synthetic route toward Remdesivir was disclosed in 2012 (Scheme 5).²⁸ There, **13** was subjected to a temporary *N*,*N*-bis-

Scheme 5

"Reagents and conditions: (a) n-BuLi, TMSCl, THF, -78 °C (26%) or NaH, n-BuLi, ClSi(Me) $_2$ CH $_2$ CH $_2$ (Me) $_2$ SiCl, THF, -78 °C (60%); (b) TMSCN, TMSOTf, CH $_2$ Cl $_2$, -78 °C, 5 h (65%; β : α = 89:11); (c) BCl $_3$, CH $_2$ Cl $_2$, -78 °C, 1 h (74%); (d) NMI, (MeO) $_3$ P=O, THF, 0 °C (21%); (e) chiral HPLC.

silylation for N-amino protection. Next, metal—halogen exchange in THF at -78 °C and reaction with **22** afforded lactol **31**, as a mixture of anomers. Use of n-BuLi and TMSCl gave a meagre 25% yield of **31**. ¹¹

Therefore, NaH and ClSi(Me)₂CH₂CH₂(Me)₂SiCl were employed for the *N*-amino protection step, and then *n*-BuLi executed the metal—halogen exchange. The modification notably improved the efficiency (up to 60% yield) of this poorly reliable *C*-glycosylation step. Qin et al. demonstrated that replacing NaH with the reagent couple *n*-BuLi/*i*-Pr₂NH afforded 31 (75%), in a hectogram-scale process. ³¹

Cyanation was next performed with TMSOTf and TMSCN at 0 °C, through the intermediacy of an oxocarbenium ion. This gave **32** as an anomeric mixture (β : α = 57:43, 80% yield). A β : α = 89:11 ratio was observed when the reaction was performed for 5 h at -78 °C (65% yield). Interestingly, using BF₃·Et₂O¹ as disclosed by Metobo et al.²⁹ gave 58% yield of **32** (β : α = 85:15) after 5 h at -78 °C. Other approaches toward **32** have been reported.³²

Next, a BCl₃-mediated exhaustive debenzylation gave a diastereoisomeric mixture of nitriles in 74% combined yield, which was separated by preparative reverse-phase HPLC to afford 33 in optically pure form. This set the stage for the coupling with the key fragment 26, in the presence of trimethyl phosphate and *N*-methylimidazole (NMI).

The transformation gave 21% yield of an \sim 1:1 mixture of diastereomers (34), which was resolved by chiral HPLC to afford Remdesivir (1). Overall, the synthetic sequence was straightforward but required several HPLC separations (including chiral HPLC in the final step) and proceeded in low overall yield (0.6–1.5% from 13 and 22).

3.2. Second Generation. The Optically Pure Phosphoramide Alternative. In 2016, a multigram synthesis of the drug was disclosed (Scheme 6), ^{13,33} and a 2017 patent included additional details. ³⁴ This new generation synthesis comprised several improvements, including a TEMPO-catalyzed NaClO oxidation to access 22²⁰ and a *C*-glycosylation which was executed with iodide **18** and Grignard reagents. This combination proved to be superior to that of the first-generation synthesis (based on **13** and *n*-BuLi, NaH, or *n*-BuLi/*i*-PrNH₂). In a modification of this reaction, the LaCl₃·2LiCl complex was added to generate a less basic carbanion intermediate, which avoids deprotonation of lactone **22**. ³⁵

Scheme 6

"Reagents and conditions: (a) TMSCl, PhMgCl, *i*-PrMgCl·LiCl, THF, −20 °C, 1 h (42%); (b) TMSCN, TMSOTf, TfOH, CH₂Cl₂, −78 °C, 2 h (85%); (c) 1. BCl₃, CH₂Cl₂, −40 °C, 2 h; 2. Et₃N, MeOH, −78 °C → rt (86%); (d) 2,2-DMP, H₂SO₄, Me₂CO, rt, 0.5 h; 45 °C, 0.5 h (99%); (e) MgCl₂, DIPEA, MeCN, 50 °C, 0.5 h (70%); (f) 12 N HCl, THF (1:5), rt, 5 h (69%).

The efficiency of this 1,2-addition was improved, and intermediate 31 was obtained in 42% yield, consistently, with less costly reagents and under milder conditions (-20 °C), ensuring scalability. Other lanthanide salts (CeCl₃, NdCl₃, YCl₃) seem to have been tested, without disclosing product yields. However, NdCl₃ was preferred for process scale up (68% yield of 31 from 282 kg of 22). ³⁶

Another modification was the addition of an acid (TfOH, TFA) to the original reagent couple TMSCN/TMSOTf, to carry out the required stereoselective cyanation. This change

improved the access to 32 (85% yield; β : α > 95:5). The exhaustive debenzylation step was refined, being performed at -40 °C to give triol 33 in 86% yield.

As proven by Gilead scientists, mild coupling of **28** and **35** did not affect the stereochemical integrity of the phosphorus center, suggesting that coupling of **28** and **33** could have the same outcome. However, in order to overcome the poor yields of the coupling stage between **26** and **33**, the vicinal diol moiety of the latter was protected. Hence, **33** was exposed to 2,2-dimethoxy propane/ H_2SO_4 , giving the acetonide **35** in 99% yield; interestingly, **35** was synthesized earlier in undisclosed yield, under di-*p*-nitrophenylphosphoric acid catalysis. ¹¹

Then, **35** was reacted with the second-generation phosphoramidate S_P -**28**, furnishing **36** (70% yield). The process is a nucleophilic substitution, where MgCl₂ activates the phosphorus center and DIPEA promotes the coupling, with configurational inversion at the *P*-center. Final deprotection of **36** with concentrated HCl in THF gave optically pure Remdesivir (1) in 69% yield. Under the rather mild conditions developed for the coupling, the product exhibited stereochemical integrity of its stereogenic center.

Interestingly, the Gilead 2016 patent¹⁴ also disclosed a *t*-BuMgCl-mediated coupling between unprotected compound 33 and the (R_P/S_P) -diastereomeric mixture of 27 (Scheme 4), which gave (R_P/S_P) -1 in 43% yield. The resulting diastereomers could barely be separated by gradient reverse-phase HPLC (Kinetex C-18 column); however, they were cleanly separated by chiral HPLC [Lux Cellulose-2 column, eluting with MeCN:MeOH (95:5, ν/ν)].

Therefore, the three-step coupling sequence (48% overall yield) for the diastereoselective synthesis of Remdesivir compared favorably with the original coupling-chiral HPLC approach (12.5% overall yield) and the t-BuMgCl-mediated coupling (\sim 21.5% overall yield). Diol protection and use of the enantiopure phosphorylamido derivative (S_P)-28 were key to this notable improvement.

The whole sequence proceeded in only six linear steps from known lactone **22** and iodide **18** and in 14.7% overall yield. This approach avoided costly, bottleneck HPLC separations and enabled preparation of over 200 g of Remdesivir, to support preclinical efficacy and toxicity studies. The preparation of the radiolabeled analogue [¹⁴C]GS-5734 (58.0 mCi/mmol) was carried out with the same strategy, using [¹⁴C]TMSCN.

Kappe et al. developed a small-scale flow chemistry approach of the C-glycosylation step toward 31, which afforded the product in a stable 47% yield, with a total residence time of <1 min and a starting material throughput of 51.8 mmol/h (\sim 690 mg/day). They optimized the reaction stoichiometries to suppress generation of identified impurities arising from onward reaction of the intermediate. The procedure did not provide a significantly higher yield with regard to the original batch conditions (40%), but it offered a marked improvement in processability.

In addition, a manufacturing scale (\sim 200 kg/run) continuous flow chemistry approach for the cyanation of **31** at -30 °C was reported. The process afforded **32** in 78% yield and 99.9% purity, at a rate of 6 mol/h, comparing favorably with a batch process (71%, 99.2% purity). It also provided improved control over the reaction conditions and increased diastereoselectivity [d.r. (β : α) = 96:4].³⁶

Use of large quantities of cyanide derivatives (added in excess) required special conditions for plant design, including detailed written instructions and cyanide detectors as well as

suitable personal training and protective measures. Further, use of cold basic reaction quenching conditions (KOH, H_2O , -10°C) prevented the generation of HCN gas, whereas the waste streams from the process were treated with bleach until the cyanide levels were below the limit of cyanide test strip detection.36

Shen et al.³⁸ recently reported a simplified and efficient Cglycosylation strategy which avoids the formation of overaddition byproducts (Scheme 7). It involves a Weinreb

Scheme 7

^aReagents and conditions: (a) MeONH₂·HCl, i-PrMgCl, THF, 0 °C; (b) TMSCl, MeMgBr, *i*-PrMgCl·LiCl, THF, -10 °C \rightarrow rt (65%

amidation of 22, followed by in situ temporary silvlation of 37, addition of the Grignard reagent derived from 18 (MeMgBr was used instead of PhMgCl), and final cyclization to afford 31 in 65% overall yield at a kilogram scale.

In the absence of additives, it has been shown that the coupling between 26 and the diol 33 gave an ~1:1 mixture of diastereomers. However, efficient organocatalytic asymmetric Pphosphoramidation alternatives to the fragment coupling stage were recently reported by the groups of Zhang³⁹ and Hung.² Their approach involved dynamic kinetic asymmetric transformations (DyKATs) (Scheme 8) with chiral imidazole derivatives (38 and 39) as organocatalysts, mimicking the mechanism of the enzyme glucose-6-phosphatase.4

Scheme 8

^aReagents and conditions: (a) 38 (10 mol %), CH₂Cl₂, 4 Å MS, -40 °C, 48 h (96% conversion; d.r. = 22.1:1) or 1. 39 (20 mol %), -20 °C, 24 h (97%, d.r. = 96.1:3.9); 2. TsOH, MeOH; 3. recrystallization (70%, d.r. = 99.3:0.7).

In the procedure devised by Zhang et al., catalyst 38 (10 mol %) was employed under 2,6-lutidine promotion, to diastereoselectively afford the protected intermediate 36 (Scheme 8) after 96% conversion (d.r. = 22.1:1). The improved couplingdeprotection-crystallization sequence of Hung et al. gave 1 in 70% yield (d.r. = 99.3:0.7) under the assistance of 39, which was recovered (83% yield) for reuse.

The solid-state characteristics of Remdesivir were ascertained employing ssNMR, X-ray powder diffraction, calorimetric

(DSC, TGA), and dynamic vapor sorption (DVS) methods. Four polymorphic forms of the drug were differentiated, and a maleate salt was prepared.4

3.3. Third Generation. The Silyl Ether Approach. The sequence was first reported in 2016 without disclosing the yields of any step. 14 There, the silyl lactone 24 was condensed with iodide 18, using the previously developed Grignard reagents strategy (Scheme 9), and the LaCl₃·2LiCl complex was added to

Scheme 9

^aReagents and conditions: (a) 1. i-PrMgCl, TMSCl, 0 °C; 2. i-PrMgCl, -15 °C; 3. LaCl₃·2LiCl, -15 °C; (b) 1. TMSCN, TMSOTf, TfOH, CH₂Cl₂, -30 °C, 15 min; 2. Et₃N, 0 °C (β : α = 3.8:1); (c) 28, MgCl₂, i-Pr₂NEt, THF, 50 °C, 21 h or 30, t-BuMgCl, THF, -10 °C \rightarrow 5 °C, 16 h; (d) 12 N HCl, 20 °C, 72 h or TBAF or KF or Py·HF.

improve access to 40 and avoid side products. In turn, compound 40 is subjected to cyanation with the TMSCN/ TMSOTf reagent system at -30 °C, with the addition of TFA. This selectively affords the monodeprotected nitrile after quenching with Et₃N; however, the product 41 was isolated as a mixture of diastereomers (β : α = 3.8:1), which required a preparative HPLC separation for purification.

Nitrile 41 was further coupled in THF with the enantiomerically pure phosphoramide 28 under MgCl₂/*i*-Pr₂NEt promotion to furnish 42. In an alternative setup, the use of the phosphoramide 30 carrying a pentafluorophenyl leaving group was also proposed, 42 but the coupling was performed with t-BuMgCl in THF at -10 to 5 °C. Incompatibility between the nitro group in 28 with the Grignard reagent may have forced the use of 30. Acid- (HCl) or fluoride- (TBAF, KF, or Py·HF) mediated desilylation finally provided Remdesivir (1).

Since no reaction yields were given, efficiency comparisons with previous syntheses are not possible. However, this route has some clear advantages, such as the selective monodeprotection of the primary silyl ether, which greatly simplifies the last steps (debenzylation, protection, coupling, and deprotection), enabling direct coupling of 41 with the phosphoramide moiety. Another advantage is the milder (room temperature) final deprotection step.

Both the cyanation—monodesilylation step aided by TFA and the use of the pentafluorophenyl derivative **30** for the coupling stage are interesting and novel developments. On the other side, however, the need for preparative HPLC for accessing enantiomerically pure intermediate **41** is a disadvantage.

4. PREPARATION OF (R_p) -REMDESIVIR

The S_P -isomer Remdesivir was originally selected for development due to its greater selectivity and better therapeutic window. ⁴³ However, the R_P -isomer is also active. The enzymes carboxylate esterase 1 (CES1) and cathepsin A (CatA), which have variable expression levels among individual patients, have been identified as the major contributors of the initial activation step inside the cells.

CatA is more widespread and has preference for hydrolysis of (S_p) -Remdesivir, whereas CES1 has a more limited tissue distribution but a strong preference for hydrolysis of the (R_p) -isomer. The new coronavirus affects primarily the lungs, rich in CatA; however, since it attacks multiple tissues, the synthesis of (R_p) -Remdesivir (Scheme 10) becomes relevant.

Scheme 10

^aReagents and conditions: (a) In1W variant of phosphotriesterase from *P. diminuta* (In1W-PTE), HEPES (50 mM, pH 8.0), MeOH- $_{12}$ O (7:3, $_{12}$ V/ $_{12}$ V), 3 h (48.5%, ee > 95%).

This prompted Raushel et al. to prepare the key intermediate 43 through a chemoenzymatic strategy, 44 employing the In1W variant of the phosphotriesterase from *Pseudomonas diminuta* (In1W-PTE). The product was obtained satisfactorily (97% yield with regards to 27, ee > 95%), as assessed by 31P NMR. Other strategies toward this end have been reviewed. 45 This group finally achieved the synthesis of R_P -Remdesivir, using the t-BuMgCl-mediated coupling conditions (38% yield).

5. SUMMARY AND OUTLOOK

This review highlighted how synthetic efforts have improved the synthesis of Remdesivir in terms of cost, yield, and scalability. It also revealed not only how synthetic organic chemistry and its interaction with other disciplines can give specific responses to nowadays urgent needs but also how difficult it still is for science to come out with state-of-the-art suitable pharmaceutical solutions with regard to small organic molecules, even under the pandemic's pressure. Any specific drug against coronavirus should target the virus and its respiratory and cardiovascular effects. While discovery of this magic bullet is underway, the lessons learned during this pandemic will enhance our preparedness for the next time, when urgency will knock the door again.

AUTHOR INFORMATION

Corresponding Authors

Teodoro S. Kaufman – Instituto de Química Rosario (IQUIR, CONICET-UNR) and National University of Rosario (UNR), 2000 Rosario, Argentina; orcid.org/0000-0003-

3173-2178; Phone: +54-341-437-0477; Email: kaufman@iquir-conicet.gov.ar

Enrique L. Larghi — Instituto de Química Rosario (IQUIR, CONICET-UNR) and National University of Rosario (UNR), 2000 Rosario, Argentina; orcid.org/0000-0002-9791-4831; Phone: +54-341-437-0477; Email: larghi@iquir-conicet.gov.ar

Author

Didier F. Vargas – Instituto de Química Rosario (IQUIR, CONICET-UNR) and National University of Rosario (UNR), 2000 Rosario, Argentina

Complete contact information is available at: https://pubs.acs.org/10.1021/acsomega.1c03082

Author Contributions

All authors contributed equally to the manuscript.

Notes

The authors declare no competing financial interest. **Biographies**



Dr. Didier F. Vargas was born in Baraya (Huila, Colombia). He obtained his B.S. in Chemistry in 2014 from the University of Atlántico (Barranquilla, Colombia) and Ph.D. in 2020 from the National University of Rosario, working in Dr. Kaufman's group as a Doctoral fellow of the Argentine National Research Council (CONICET) at the Institute of Chemistry of Rosario (IQUIR, CONICET-UNR). Currently, he is a Postdoctoral research fellow of CONICET, working at IQUIR under the direction of Dr. Larghi. His research interests include the development and optimization of new synthetic methodologies toward nitrogen heterocycles and the synthesis of heterocyclic natural products, their analogues, and relevant derivatives.



Dr. Enrique L. Larghi was born in Rosario (Santa Fe, Argentina). He received his B.S. in Chemistry from the UNR (1997), and his M.Sc.

degree (1999, Professor Claudio C. Silveira) and Ph.D. in Chemistry (2003, Professor Ademir Farias Morel) were awarded by the Universidade Federal de Santa María (Brazil). After returning to Argentine and carrying out a short experience in the pharmaceutical industry as Head of Research and Development, he joined Dr. Kaufman's group at IQUIR as a postdoctoral fellow in 2005. Currently, he is Independent Researcher of the Argentine National Research Council (CONICET) and Adjunct Professor at UNR. His research interests are focused in bioorganic chemistry, especially in the synthesis of heterocyclic natural products and their biological evaluation.



Dr. Teodoro S. Kaufman was born near Moisés Ville (Santa Fe, Argentina). He graduated in Biochemistry (1982) and Pharmacy (1985) at the National University of Rosario (UNR, Rosario, Argentina) and received his Ph.D. (1987) under the guidance of Professor Edmundo A. Rúveda. After a two-year postdoctoral training period at The University of Mississippi (USA), he returned to Rosario. Currently, he is full Professor of the UNR and Superior Researcher of the Argentine National Research Council (CONICET) and is coauthor of over 170 papers. His scientific interests include the synthesis and evaluation of bioactive compounds (especially natural products) and their analogues, with emphasis on nitrogen and oxygen heterocycles.

ACKNOWLEDGMENTS

The authors wish to acknowledge Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) and Agencia Nacional de Promoción Científica y Tecnológica (ANPCyT) for funding our research projects. The authors are indebted to Pharm. Aldana B. Moroni (IQUIR, CONICET-UNR) for revising the text. D.F.V. is also thankful to CONICET for his postdoctoral fellowship.

REFERENCES

- (1) Cho, A.; Saunders, O. L.; Butler, T.; Zhang, L.; Xu, J.; Vela, J. E.; Feng, J. Y.; Ray, A. S.; Kim, C. U. Synthesis and antiviral activity of a series of 1'-substituted 4-aza-7,9-dideaza adenosine C-nucleosides. *Bioorg. Med. Chem. Lett.* **2012**, 22, 2705–2707.
- (2) Siegel, D.; Hui, H. C.; Doerffler, E.; Clarke, M. O.; Chun, K.; Zhang, L.; Neville, S.; Carra, E.; Lew, W.; Ross, B.; Wang, Q.; Wolfe, L.; Jordan, R.; Soloveva, V.; Knox, J.; Perry, J.; Perron, M.; Stray, K. M.; Barauskas, O.; Feng, J. Y.; Xu, Y.; Lee, G.; Rheingold, A. L.; Ray, A. S.; Bannister, R.; Strickley, R.; Swaminathan, S.; Lee, W. A.; Bavari, S.; Cihlar, T.; Lo, M. K.; Warren, T. K.; Mackman, R. L. Discovery and synthesis of a phosphoramidate prodrug of a pyrrolo[2,1-f]triazin-4-amino adenine C-nucleoside (GS-5734) for the treatment of Ebola and emerging viruses. *J. Med. Chem.* 2017, 60, 1648–1661.
- (3) Wang, Z.; Yang, L. GS-5734: FDA potentially approved drug against SARS-Cov-2. New J. Chem. 2020, 44, 12417–12429.

- (4) Food and Drug Administration. Coronavirus (COVID-19) update: FDA issues emergency use authorization for potential COVID-19 treatment; Washington DC, USA, 2020.
- (5) Eastman, R. T.; Roth, J. S.; Brimacombe, K. R.; Simeonov, A.; Shen, M.; Patnaik, S.; Hall, M. D. Remdesivir: A review of its discovery and development leading to emergency use authorization for treatment of COVID-19. *ACS Cent. Sci.* **2020**, *6*, 672–683.
- (6) Gordon, C. J.; Tchesnokov, E. P.; Woolner, E.; Perry, J. K.; Feng, J. Y.; Porter, D. P.; Götte, M. Remdesivir is a direct-acting antiviral that inhibits RNA-dependent RNA polymerase from severe acute respiratory syndrome coronavirus 2 with high potency. *J. Biol. Chem.* **2020**, 295, 6785–6797.
- (7) Vicenti, I.; Zazzi, M.; Saladini, F. SARS-CoV-2 RNA-dependent RNA polymerase as a therapeutic target for COVID-19. *Expert Opin. Ther. Pat.* **2021**, *31*, 325–337.
- (8) Patil, S. A.; Otter, B. A.; Klein, R. S. Synthesis of [2,1-f][1,2,4] triazine congeners of nucleic acid purines via the N-amination of 2-substituted pyrroles. *J. Heterocycl. Chem.* **1994**, 31, 781–786.
- (9) Rai, G. S.; Maru, J. J. Synthetic strategies for pyrrolo[2,1-f] [1,2,4]triazine: the parent moiety of antiviral drug remdesivir. *Chem. Heterocycl. Compd.* **2020**, *56*, 1517–1522.
- (10) Juan, B. V.; Diez, J. A. A.; Albero, M. A. B.; Eastwood, P. R.; Trias, C. E.; Toribio, M. E. L.; Roberts, R. S.; Gispert, L. V.; Rodrigues, J. G.; Cepeda, M. M. New CRTh2 Antagonists. 2013010880 A1, 2013.
- (11) O'Connor, S. J.; Dumas, J.; Lee, W.; Dixon, J.; Cantin, D.; Gunn, D.; Burke, J.; Phillips, B.; Lowe, D.; Shelekhin, T.; Wang, G.; Ma, X.; Ying, S.; Mcclure, A.; Achebe, F.; Lobell, M.; Ehrgott, F.; Iwuagwu, C.; Parcella, K. Pyrrolo[2,1-f][1,2,4]triazin-4-yl amines Igf-lr kinase inhibitors for the treatment of cancer and other hyperproliferative diseases. 2007056170, 2007.
- (12) Zhang, Z.; Shen, M.; Zhao, L. Preparation method of 7-bro mopyrrolo [2.1-f] [1,2,4]-triazene-4-amine. 110845502, 2020.
- (13) Chun, B. K.; Clarke, M. O. H.; Doerffler, E.; Hui, H. C.; Jordan, R.; Mackman, R. L.; Parrish, J. P.; Ray, A. S.; Siegel, D. Gilead Sciences, Inc. Methods for treating filoviridae virus infections. 2016069826 A1, 2016
- (14) Axt, S. D.; Badalov, P. R.; Brak, K.; Campagna, S.; Chtchemelinine, A.; Doerffler, E.; Frick, M. M.; Gao, D.; Heumann, L. V.; Hoang, B.; Lew, W.; Milburn, R. R.; Neville, S. T.; Ross, B.; Rueden, E.; Scott, R. W.; Siegel, D.; Stevens, A. C.; Tadeus, C.; Vieira, T.; Waltman, A. W.; Wang, X.; Whitcomb, M. C.; Wolfe, L.; Yu, C.-Y. Method for the preparation of ribosides. US2016/122356 A, 2016.
- (15) Wang, Y.; Li, Z.; Yin, C.; Li, X. Preparation method of 4-amino-7-iodopyrrolo[2,1-f][1,2,4]triazine. 111423443 A, 2020.
- (16) Jiang, Z.; Jurong, Y. A kind of synthetic method of remdesivir intermediate triazine amine derivative. 112625043 A, 2021.
- (17) dos Santos, J. A.; Pereira, V. R. D; da Silva, A. D.; Amarante, G. W. Klein's remdesivir-nucleobase synthesis revisited: chemoselective cyanation of pyrrol-2-carboxaldehyde. *J. Braz. Chem. Soc.* **2021**, *32*, 1391–1395.
- (18) Agrawal, T.; Burns, J. M.; Cardoso, F. S. P; Cook, D. W.; Gupton, B. F.; Paymode, D. J.; Sieber, J. D.; Snead, D. R.; Stringham, R. W.; Tomlin, J. W. Expanding access to remdesivir via an improved pyrrolotriazine synthesis: supply centered synthesis. *Org. Lett.* **2020**, *22*, 7656–7661.
- (19) Barker, R.; Fletcher, H. G., Jr 2,3,5-Tri-O-benzyl-D-ribosyl and -L-arabinosyl bromides. J. Org. Chem. 1961, 26, 4605–4609.
- (20) Chen, B.; Ye, J.; Li, D.; Zhang, W.; Xu, C.; Jiang, T.; Zhang, L.; Mu, X.; He, Y.; Pang, X.; Cheng, G. Method for preparing 2,3,5-tribenzyloxy-D-ribose-1,4-lactone through oxidation in continuous flow micro-channel reactor. 111548328 A, 2020.
- (21) Beigelman, L.; Wang, G.; Dyatkina, N. Antiviral nucleosides and derivatives thereof. 2020157694 A1, 2020.
- (22) Bernet, B.; Mangholz, S. E.; Briner, K.; Vasella, A. Glycosylidene carbenes Part 31 Glycosylidene diaziridines: Stereoselective addition of ammonia and methylamine to lactone oxime sulfonates. *Helv. Chim. Acta* **2003**, *86*, 1488–1521.
- (23) Delbrouck, J. A.; Tikad, A.; Vincent, S. P. Efficient and regioselective synthesis of γ -lactone glycosides through a novel

- debenzylative cyclization reaction. Chem. Commun. 2018, 54, 9845-9848.
- (24) Attwood, S. V.; Barrett, A. G. M A convenient procedure for the deoxygenation and homologation of D-ribose derivatives. *J. Chem. Soc., Perkin Trans.* 1 1984, 1315–1322.
- (25) Axt, S. D.; Badalov, P. R.; Brak, K.; Campagna, S.; Chtchemelinine, A.; Doerffler, E.; Frick, M. M.; Gao, D.; Heumann, L. V.; Hoang, B.; Lew, W.; Milburn, R. R.; Neville, S. T.; Ross, B.; Rueden, E.; Scott, R. W.; Siegel, D.; Stevens, A. C.; Tadeus, C.; Vieira, T.; Waltman, A. W.; Wang, X.; Whitcomb, M. C.; Wolfe, L.; Yu, C.-Y. Method for the preparation of ribosides. US2020/197422 A1, 2020.
- (26) De Savi, C.; Hughes, D. L.; Kvaerno, L. Quest for a COVID-19 cure by repurposing small-molecule drugs: mechanism of action, clinical development, synthesis at scale, and outlook for supply. *Org. Process Res. Dev.* **2020**, *24*, 940–976.
- (27) Hughes, D. L. Quest for a Cure: Potential Small-Molecule Treatments for COVID-19, Part 2. *Org. Process Res. Dev.* **2021**, 25, 1089–1111.
- (28) Mackman, R. L., Parrish, J. P., Ray, A. S., Theodore, D. A. Methods and compounds for treating paramixoviridae virus infections. 2012/27752 A1, 2012.
- (29) Metobo, S. E.; Xu, J.; Saunders, O. L.; Butler, T.; Aktoudianakis, E.; Cho, A.; Kim, C. U. Practical synthesis of 1'-substituted tubercidin C-nucleoside analogs. *Tetrahedron Lett.* **2012**, *53*, 484–486.
- (30) Clarke, M. O. H.; Doerffler, E.; Mackman, R. L.; Siegel, D. Pyrrolo[1,2-f][1,2,4]triazines useful for treating respiratory syncitial virus infections. 2015/133395 Al, 2015.
- (31) Xue, F.; Zhou, X.; Zhou, R.; Zhou, X.; Xiao, D.; Gu, E.; Guo, X.; Xiang, J.; Wang, K.; Yang, L.; Zhong, W.; Qin, Y. Improvement of the C-glycosylation step for the synthesis of remdesivir. *Org. Process Res. Dev.* **2020**, *24*, 1772–1777.
- (32) Ding, J.; Shi, G.; Chen, D.; Wang, Z.; Liu, Y. Novel compound for preparing key intermediate of remdesivir and preparation method thereof. 111233869 A, 2020.
- (33) Warren, T. K.; Jordan, R.; Lo, M. K.; Ray, A. S.; Mackman, R. L.; Soloveva, V.; Siegel, D.; Perron, M.; Bannister, R.; Hui, H. C.; Larson, N.; Strickley, R.; Wells, J.; Stuthman, K. S.; Van Tongeren, S. A.; Garza, N. L.; Donnelly, G.; Shurtleff, A. C.; Retterer, C. J.; Gharaibeh, D.; Zamani, R.; Kenny, T.; Eaton, B. P.; Grimes, E.; Welch, L. S.; Gomba, L.; Wilhelmsen, C. L.; Nichols, D. K.; Nuss, J. E.; Nagle, E. R.; Kugelman, J. R.; Palacios, G.; Doerffler, E.; Neville, S.; Carra, E.; Clarke, M. O.; Zhang, L.; Lew, W.; Ross, B.; Wang, Q.; Chun, K.; Wolfe, L.; Babusis, D.; Park, Y.; Stray, K. M.; Trancheva, I.; Feng, J. Y.; Barauskas, O.; Xu, Y.; Wong, P.; Braun, M. R.; Flint, M.; McMullan, L. K.; Chen, S.-S; Fearns, R.; Swaminathan, S.; Mayers, D. L.; Spiropoulou, C. F.; Lee, W. A.; Nichol, S. T.; Cihlar, T.; Bavari, S. Therapeutic efficacy of the small molecule GS-5734 against Ebola virus in rhesus monkeys. *Nature* 2016, 531, 381–385.
- (34) Clarke, M. O. H.; Jordan, R.; Mackman, R. L.; Ray, A. S.; Siegel, D. Methods for treating flaviviridae virus infections. WO 2017184668 Al, 2017.
- (35) Conlon, D. A.; Kumke, D.; Moeder, C.; Hardiman, M.; Hutson, G.; Sailer, L. Insights into the cerium chloride-catalyzed grignard addition to esters. *Adv. Synth. Catal.* **2004**, 346, 1307–1315.
- (36) Badalov, P.; Chtchemelinine, A.; Gao, D.; Heumann, L.; Stevens, A. C.; Vieira, T. Development of a large-scale cyanation process using continuous flow chemistry en route to the synthesis of remdesivir. *Org. Process Res. Dev.* **2020**, *24*, 2113–2121.
- (37) von Keutz, T.; Williams, J. D.; Kappe, C. O. continuous flow C-glycosylation via metal-halogen exchange: process understanding and improvements toward efficient manufacturing of remdesivir. *Org. Process Res. Dev.* **2020**, *24*, 2362–2368.
- (38) Xie, Y.; Hu, T.; Zhang, Y.; Wei, D.; Zheng, W.; Zhu, F.; Tian, G.; Aisa, H. A.; Shen, J. Weinreb amide approach to the practical synthesis of a key remdesivir intermediate. *J. Org. Chem.* **2021**, *86*, 5065–5072.
- (39) Wang, M.; Zhang, L.; Huo, X. H.; Zhang, Z. F.; Yuan, Q. J.; Li, P. P.; Chen, J. Z.; Zou, Y. S.; Wu, Z. X.; Zhang, W. B. Catalytic asymmetric synthesis of the anti-COVID-19 drug remdesivir. *Angew. Chem., Int. Ed.* **2020**, *59*, 20814–20819.

- (40) Gannedi, V.; Villuri, B. K.; Reddy, S. N.; Ku, C.-C; Wong, C.-H; Hung, S.-C Practical remdesivir synthesis through one-pot organocatalyzed asymmetric (S)-P-Phosphoramidation. *J. Org. Chem.* **2021**, 86, 4977—4985.
- (41) Brak, K.; Carra, E. A.; Heumann, L. V.; Larson, N. Crystalline forms of (S)-2-ethylbutyl-2-(((S)-(((2R,3S,4R,5R)-5-(4-amino pyrrolo[2,1-f][1,2,4]triazin-7-yl)-5-cyano-3,4-dihydroxy tetrahydro-furan-2-yl)methoxy)(phenoxy)phosphoryl)amino) propanoate. 2018/204198 Al, 2018.
- (42) Hu, T.; Xie, Y.; Liu, Y.; Xue, H.; Zhu, F.; Aisa, H. A.; Shen, J. A convenient and cost efficient route suitable for "one-pot" synthesis of molnupiravir. *ChemRxiv* **2021**, DOI: 10.26434/chemrxiv.
- (43) Liang, C.; Tian, L.; Liu, Y.; Hui, N.; Qiao, G.; Li, H.; Shi, Z.; Tang, Y.; Zhang, D.; Xie, X.; Zhao, X. A promising antiviral candidate drug for the COVID-19 pandemic: a mini-review of remdesivir. *Eur. J. Med. Chem.* **2020**, *201*, 112527.
- (44) Bigley, A. N.; Narindoshvili, T.; Raushel, F. M. A chemoenzymatic synthesis of the (RP)-isomer of the antiviral prodrug remdesivir. *Biochemistry* **2020**, *59*, 3038–3043.
- (45) Liu, Y.; Cheng, J.; Hong, R. Asymmetric synthesis of prodrug nucleotides (ProTides): construction of the P-stereogenic centers. *Youji Huaxue* **2020**, 40, 3237–3248.