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Cnidom variation through distinct developmental stages in the sea anemone Aulactinia marplatensis (Zamponi, 1977) (Cnidaria: Actiniaria)

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Abstract: The cnidae are the exclusive diagnostic structures of phylum Cnidaria. The inventory of all cnidae types of a particular species is called the cnidom. The study of cnidae has been widely addressed in all classes of cnidarians. Particularly in the order Actiniaria (sea anemones), the study of the composition, size and distribution of cnidae is essential to the identification and description of species. In the present study, we examine the cnidom of the sea anemone Aulactinia marplatensis in three different stages of development throughout its life cycle. We found that the composition and abundance patterns are very similar between the adult and juvenile stages, although significant differences in the size capsules were found between both stages and in all cnidae types observed, being bigger those from the adult forms. The planula larvae stage presents a less diverse cnidom in comparison to the juvenile and adult stages; however, it presents an exclusive cnidae type (the mesobasic p-mastigophore) which is the biggest in size of all the cnidae types observed in the species. These results highlight the importance of considering the stage of development when cnidae is used as a diagnostic character, and the particular relevance of the study of the cnidom in larval stages.

Key words: cnidae, cnidocysts, larval cnidom, life cycle, sea anemones.

INTRODUCTION

Cnidae, also known as cnidocysts, are subcellular structures that are exclusive to cnidarians. The inventory of all cnidae types present in a particular species is called the cnidom, which is currently an essential component of most taxonomic descriptions for sea anemones (Fautin 2009).

Studies on cnidae and their distribution and size have been widely developed, particularly in anthozoans (Weill 1934, Carlgren 1940, Cutress 1955, Schmidt 1969, 1972, 1974, Mariscal 1974, 1984, den Hartog 1977, Östman 1988, England 1991, Fautin and Mariscal 1991, Pires and Pitombo 1992, Pires 1997, Acuña et al. 2003, Terrón-Sigler and López-González 2005, Fautin 2009, Picciani et al. 2011, Garese et al. 2016). The analysis of morphometrical data of cnidae from statistical approaches (Thomason 1988, Zamponi and Acuña

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1991, 1994, Allcock et al. 1998, Watts et al. 2000, Ardelean and Fautin 2004, Williams 1996, 1998, 2000, Acuña and Zamponi 1997, Chintiroglou 1996, Chintiroglou et al. 1997, Östman 2000, Francis 2004, Kramer and Francis 2004, Acuña et al. 2004, 2007, 2011, Garese et al. 2016), as well as the study of intraspecific variations of cnidae in response to ecological conditions (Martínez-Baraldés et al. 2014, González-Muñoz et al. 2015, 2017), are among the most studied topics related with the utility of cnidom to taxonomic purposes.

However, studies focused in the variation of cnidom throughout the life cycle of a cnidarian species are very scarce. Chia and Koss (1983) studied the structure of the nematocysts in the planula larvae of Anthopleura elegantissima (Brandt, 1835) and comparing them with those present in adult forms. Holst et al. (2007) described all types of enidocysts occurring in all stages of the life cycle of the scyphozoan Rhizostoma octopus (Linnaeus, 1758), and Reitzel et al. (2009) studied the anatomy and development of Edwardsiella lineata (Verrill, 1873), and included a comparison of the cnidom between the parasitic larvae, the post parasitic larvae and the juvenile form. Nevertheless, none of these studies includes a statistical comparison of the cnidae sizes in the distinct stages throughout the life cycle of the species treated. In this study, we describe and compare the cnidom of the sea anemone species Aulactinia marplatensis (Zamponi, 1977) in three distinct stages of its life cycle (i.e. the planula larvae, and the juvenile and adult forms), and statistically compare the variation in cnidae size ranges between the juvenile and the adult stages.

MATERIALS AND METHODS

Specimens of both juvenile and adult forms of *Aulactinia marplatensis* were collected during February, 2011, at low tides in the rocky intertidal of Punta Cantera, Mar del Plata, Argentina (38°04'

S-57°32'W). Juvenile specimens were identified by the size of its basal diameter, which is lower than 9 mm (Zamponi and Excoffon 1986), and no evidence of gametogenic tissue was observed in any specimen examined at the laboratory. The complete cnidom of 12 juveniles and 12 adult specimens were studied. Squash preparations of small amounts of tissue (approximately 1 mm³) from tentacles, column, mesenterial filaments, actinopharynx, and acrorhagi were made from the 24 collected specimens to study the cnidom. Cnidae terminology follows Östman (2000). From each squash preparation, the length and width of 30 undischarged capsules of each cnidae type, when it was possible, were haphazardly measured using DIC microscopy 1000x oil immersion. For abundance estimations four microscopic view fields were haphazardly taken and each cnidae type counted; then, pie charts were produced. Statistically descriptive parameters (mean, standard deviation, size ranges) were obtained of each data set of cnidae, and the variation of the length sizes between juveniles and adults were statistically compared for each type of cnida. The normality of cnidae length size data was tested with a Shapiro-Wilks test ($\alpha = 0.05$). If normality was confirmed for both data sets to be compared, an ANOVA was carried out. In cases where normal distribution was rejected, a Generalized Linear Model (GLM) was applied, following Garese et al. (2016). The gamma distribution for errors and inverse link function were employed in the model, and it form was:

$$g(\text{length}) = \beta_0 + \beta_1 \text{ stage } + \varepsilon$$

Then, t tests for $\beta 1$ coefficients of the GLM were conducted to evaluate differences between both development stages. All statistical analyses were performed using the R program (R 2008).

For study the cnidom in larval forms, ten adult specimens of *A. marplatensis* were maintained in aquarium with artificial sea water. Several microscope slides were deposited on the floor and the sides of the aquarium. During its reproductive pick (i.e. December to January according to Zamponi and Excoffon 1986), the slides were weekly revised under optic microscope searching for larvae settlement. Three post settlement larvae were obtained and their cnidae identified and measured. Statistic descriptive parameters (mean, standard deviation, size ranges) were obtained for each present cnidae type.

RESULTS

The cnidom of A. marplatensis is composed by spirocysts, basitrichs, holotrichs, microbasic b-mastigophores, microbasic p-mastigophores, and microbasic *p*-amastigophores. Tables I and II show the cnidae found in each tissue and development stage in all specimens examined. The cnidom of the juvenile and adult forms differed only in the presence of spirocysts, although with a negligible abundance, in the mesenterial filaments of the adults forms, which are absent in the juvenile stage; and the presence of two size classes of microbasic *b*-mastigophores in juvenile specimens, while only one size class were found in the adult form. However, the microbasic b-mastigophore from adult forms has a wider size range that could be comparable with both size classes from juvenile specimens together. The patterns of mean abundance of cnidae types on each tissue were also similar in both data sets (Figure 1), except for the microbasic *b*-mastigophores from mesenterial filaments of juvenile specimens, which reach about 42% in abundance, while only the 17% was observed in adult specimens (Figure 1d).

The statistical analyses applied showed a strong significant variation in all cases between the cnidae lengths of both adult and juvenile specimens (Table I). The cnidom of the planula larvae (post settlement) stage was found less diverse than those observed in both the adult and juvenile

stages (Table II). This was composed mainly by spirocysts and basitrichs, and some microbasic *p*-mastigophores and mesobasic *p*-mastigophores, but in lower number (Table II). However, the mesobasic *p*-mastigophores were exclusively found in the larval stage, and its length size range was comparatively the biggest observed among all types of the three stages of development, including those of the holotrichs present in the adult forms (Figure 2). Even though post settlement larvae were here studied, it is expected that have not differences between free living larvae and post settlement larvae. This is supported by the scarce time between the observations of the slides (a week maximum) and taking into account that the larval settlement in the species occurs 96 hs after its origin (Excoffon and Zamponi 1997).

Despite that statistical analyses to compare the cnidae length between the larvae stage with those of the other stages were not carried out due to the low number of samples achieved, their size ranges are somehow coincident to those found in the adult and juvenile stages (Tables I and II).

DISCUSSION

The presence of an additional type of cnidae in larval stages has also been observed in other sea anemone species. Chia and Koss (1983) found that cnidom of the larval stage of Anthopleura elegantissima is composed by five types of cnidae, while only four types can be found in the adult stage (Hand 1955). However, they suggested that microbasic *b*-mastigophores, the cnida type solely found in the larval forms, could be actually basitrichs at different phase of development; and these were commonly found in all other stages of development. Reitzel et al. (2009) found holotrichs in the larval stage of Edwardsiella lineata, which are absent in the adult stage. Moreover, the holotrichs have not been reported in the family Edwardsiidae (in which the genus *Edwardsiella* is currently classified)

TABLE I	Cnidom of adult and juvenile specimens of Aulactinia marplatensis
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Tissue	Cnida type	Range (length (mean ± SD)	Range (min-max) length (mean ± SD) x width (mean) [µm]	p value
		Adult ^a	Juvenile	I
Tentacles	basitrich	$16-26 (20.61 \pm 0.52) \ge 2-4 (2.58)$	$13-23 (18.11 \pm 1.92) \ge 2-3 (2.16)$	<0.001*
	spirocyst	$16-26 (21.16 \pm 2.04) \text{ x } 2-4 (2.91)$	13-23 (18.35 ± 2.07) x 2-4 (2.65)	<0.001*
Column	basitrich I	$12-23 (18.71 \pm 1.63) \text{ x } 2-3 (2.44)$	13-21 (17.23 ± 1.57) x 2-3 (2.09)	<0.001*
	basitrich II	7-15 (9.68 \pm 1.54) x 2	7-11 (8.84 \pm 1.05) x 2	<0.001*
	holotrich	23-61 (34.50 \pm 7.55) x 2-5 (3.38)	15-43 (28.74 ± 5.21) x 2-5 (3.24)	<0.001*
Mesenterial	basitrich I	$14-24 (18.36 \pm 1.91) \ge 2-4 (2.38)$	$14-24 (16.88 \pm 1.59) \ge 2-3(2.07)$	<0.001*
filaments	basitrich II	9-14 (11.45 ± 0.93) x 2-3 (2.01)	$8-14 (10.96 \pm 0.97) \ge 2$	<0.001*
	spirocyst	$17-23 (20.36 \pm 1.74) \text{ x } 2-3 (2.91)$		ı
	microbasic <i>b</i> -mastigophore I	$23-40 (32.82 \pm 3.14) \ge 3-7 (4.76)$	22-40 (30.78 ± 3.27) x 3-7 (4.55)	<0.001*
	microbasic b-mastigophore II	I	$13-22 (16.83 \pm 1.46) \ge 2-5 (3.12)$	ı
	holotrich	31-57 (44.78 ± 6.93) x 3-4 (3.47)	$30-34 (34 \pm 5.65) x 4-3 (3.5)$	#
	microbasic <i>p</i> -mastigophore	$17-38 (22.82 \pm 3.57) \text{ x } 4-9 (5.55)$	$15-27 (20.67 \pm 1.90) \text{ x } 3-7 (4.79)$	<0.001*
Actinopharynx	basitrich I	$10-24 (17.84 \pm 2.17) \text{ x } 2-5 (2.34)$	$14-21 (16.68 \pm 1.48) \ge 2-3 (2.12)$	<0.001*
	basitrich II	$21-34 (26.34 \pm 2.02) \text{ x } 3-5 (3.51)$	18-35 (25.2 ± 2.98) x 2-6 (3.42)	<0.001*
	basitrich III	$7-14 (10.85 \pm 1.16) \ge 2-3 (2.01)$	7-13 (10.54 \pm 1.17) x 2-3 (2.01)	<0.001*
	spirocyst	$14-34 (20.29 \pm 3.37) \text{ x } 2-4 (2.83)$	24 x 3	I
	microbasic <i>p</i> -amastigophore	$16-33 (23.40 \pm 3.35) \ge 4-9 (5.56)$	17-32 (22.21 ± 2.92) x 3-10 (4.87)	<0.001*

CNIDOM VARIATION IN Aulactinia marplatensis

Cnidom of planula larvae of Aulactinia marplatensis.		
	Range	
Cnida type leng	length (mean±sd) x width (mean) [µm]	- N
spirocyst	17-25 (20.5 ± 2.02) x 2-4 (2.95)	24
basitrich	15-24 (19.62 ±1.74) x 2-3 (2.55)	43
microbasic p-mastigophore	17-24 (21 ± 2.59) x 3-7 (5.37)	9
mesobasic p-mastigophore	42-53 (50 ± 4.63) x 9-13 (11.4)	5

 TABLE II

 Cnidom of planula larvae of Aulactinia marplatensis

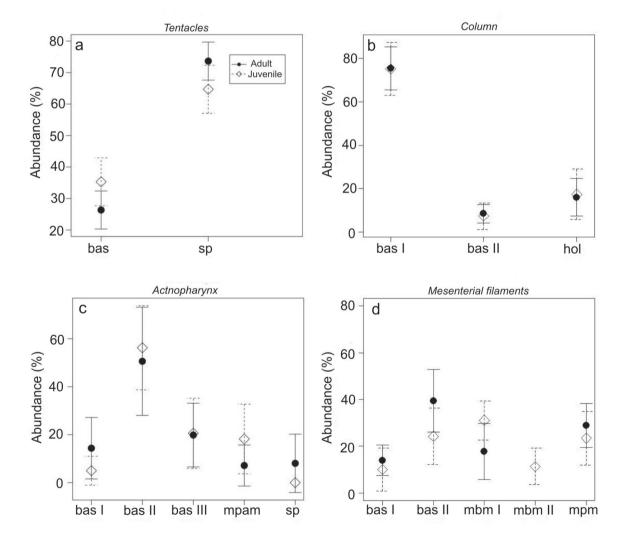


Figure 1 - Mean of abundance of cnida types in juvenile and adults of *Aulactinia marplatensis*. Basitrich (bas), spirocyst (sp), holotrich (hol), microbasic *p*-mastigophore (mpam), microbasic *b*-mastigophore (mbm), microbasic *p*-mastigophore (mpm).



Figure 2 - The exclusive cnidae type of the larval stage of *Aulactinia marplatensis*: mesobasic *p*-mastigophore. Scale = 10μ m.

(Carlgren 1949). Although Gusmão et al. (2016) reports the presence of some rare holotrichs in Edwardsia migottoi, which differ in morphology from holotrichs found in other actiniarians. they attribute their presence to contamination, probably by feeding. Therefore, they decided not to modify the genus diagnosis to include this type of cnidae. The study of larval cnidom can generate some derivations about evolutionary issues and highlighting the importance of its use for phylogenetic and taxonomic information (Reitzel et al. 2009). In addition, although the same cnidom is present in both the juvenile and adult stages of E. lineata, two types of cnidae present in these stages are absent in the larval stage (Daly 2002, Reitzel et al. 2009), similarly as we observed in A. marplatensis, suggesting that the transition from larvae to juvenile stages could be a discrete event (Reitzel et al. 2009).

The significant variation in cnidae size between the juvenile and the adult stages, being bigger those from the adult stage in all cases, suggests that the size of the cnidae have some degree of dependence with the stage of development of the organism, as well as certain dependence on the size the size of the organism. These variables could explain the intraspecific variation in cnidae size usually observed in sea anemones (Garese et al. 2016). Despite that some hypotheses have been evaluated to understand these intraspecific variations (Robson 1988, Zamponi and Acuña 1991, Karalis and Chintiroglou 1997, Francis 2004) only Rietzel et al. (2009) and the present study, have considered the stage of development throughout the life cycle of the organisms. Thus, several of the intraspecific variations documented could have been the result of a non-discretional sampling of specimens of both same stage of development (e.g. adults) and size. Less developed specimens (i.e juvenile) will be naturally smaller than more developed ones (i.e adults). Some studies have analyzed the variation of cnidocyst sizes among adults of different sizes with opposite results. Francis (2004) observed spirocysts of larger sizes in larger individuals of the species Anthopleura elegantissima and A. xhantogrammica; while Acuña et al. (2007) did not found any relation between the cnidae length and the body size (expressed as weight) in their study of the complete cnidom of Oulactis muscosa.

Our results suggest that the composition and the pattern of abundance of the cnidom in A. marplatensis are practically invariable between the juvenile and adult stages, although there is significant variability in cnidae size. In contrast, the larval stage has both a distinct composition and pattern of abundance. Despite the larval cnidom is less diverse, it presents an exclusive cnidae type. There are no others cnidae types which the mesobasic *p*-mastigophore may be confused with, in contrast as suggested by Chia and Koss (1983) for the eventual larval-specific cnidocyst in A. elegantissima. Moreover, this larval-specific cnida (i.e. mesobasic *p*-mastigophore) is the biggest type registered in the species. These observations highlight the need of similar studies in other species to explore if the variability in composition, abundance and size capsules between the cnidae of the larval, juvenile and adult stages are usual or rare in sea anemones and other cnidarians.

The study of the cnidom in the larval stages contributes with valuable information to the study of the biology, ecology, and development of the species, but also could contribute with new characters useful to taxonomic, phylogenetic and evolutionary issues (Chia and Koss 1983, Holst et al. 2007, Reitzel et al. 2009). These appreciations make very interesting the study of the larval cnidom, suggesting that should be incorporated, as much as possible, in the description or re-description of sea anemone species.

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AUTHOR CONTRIBUTIONS

AG and FHA taken the samples. AG obtained the data and made the satistical analysis. AG, RGM and FHA contributed equally to the focus, redaction and discussion of the Manuscript.

REFERENCES

- ACUÑA FH, EXCOFFON AC AND RICCI L. 2007. Composition, biometry and statistical relationships between the cnidom and body size in the sea anemone *Oulactis muscosa* (Cnidaria: Actiniaria). J Mar Biol Ass UK 87: 415-419.
- ACUÑA FH, EXCOFFON AC, ZAMPONI MO AND RICCI L. 2003. Importance of Nematocysts in Taxonomy of Acontiarian Sea Anemones (Cnidaria, Actiniaria): A Statistical Comparative Study. Zool Anz 242(1): 75-81.
- ACUÑA FH, RICCI L AND EXCOFFON AC. 2011. Statistical relationships of cnidocyst sizes in the sea anemone *Oulactis muscosa* (Actiniaria: Actiniidae). Belg J Zool 141(1): 32-37.
- ACUÑA FH, RICCI L, EXCOFFON AC AND ZAMPONI MO. 2004. A novel statistical analysis of cnidocysts in acontiarian sea anemones (Cnidaria, Actiniaria) using generalized linear models with gamma errors. Zool Anz 243: 47-52.
- ACUÑA FH AND ZAMPONI MO. 1997. The use of cnidocysts for ecological races identification from sea anemones populations (Anthozoa, Actiniidae). Iheringia 82: 9-18.
- ALLCOCK AL, WATTS PC AND THORPE JP. 1998. Divergence of nematocysts in two colour morphs of the intertidal beadlet anemone Actinia equina. J Mar Biol Ass UK 78: 821-828.
- ARDELEAN A AND FAUTIN DG. 2004. Variability in nematocysts from a single individual of the sea anemone

Actinodendron arboreum (Cnidaria: Anthozoa: Actiniaria). Hydrobiologia 530/531: 189-197.

- CARLGREN O. 1940. A contribution to the knowledge of the structure and distribution of the cnidae in the Anthozoa: CWK Gleerup.
- CARLGREN O. 1949. A survey of the Ptychodactiaria, Corallimorpharia and Actiniaria. Kungl Sven Vetenskapsakad Handl 1: 1-121.
- CHIA FS AND KOSS R. 1983. On the Fine Structure of the Nematocysts in the Planula Larva of a Sea Anemone, *Anthopleura elegantissima*. Biol Bull (Taiwan Nat Norm Univ) 12: 12-24.
- CHINTIROGLOU CC. 1996. Biometric study of *Edwardsia claparedii* (Panceri) cnidome (Actiniaria: Anthozoa). Belg J Zool 126(2): 177-180.
- CHINTIROGLOU CC, CHRISTOU I AND SIMSIRIDOU M. 1997. Biometric investigations on the cnidae of Aegean color morphs of *Actinia equina mediterranea* sensu Schmidt, 1972. Isr J Zool 43: 377-384.
- CUTRESS CE. 1955. An interpretation of the structure and distribution of enidae in Anthozoa. Sys Biol 4: 120-137.
- DALY M. 2002. Taxonomy, anatomy, and histology of the lined sea anemone, *Edwardsiella lineata* (Verrill, 1873) (Cnidaria: Anthozoa: Edwardsiidae). Proc Biol Soc Wash 115: 868-877.
- DEN HARTOG JC. 1977. Descriptions of two new Ceriantharia from the Caribbean region with a discussion of the enidom and of the classification of the Ceriantharia. Zool Meded 51(14): 211-242.
- ENGLAND KW. 1991. Nematocysts of sea anemones (Actiniaria, Ceriantharia and Corallimorpharia: Cnidaria): nomenclature. Hydrobiologia 216/217: 697-691.
- FAUTIN DG. 2009. Structural diversity, systematics, and evolution of cnidae. Toxicon 54: 1054-1064.
- FAUTIN DG AND MARISCAL RN. 1991. Cnidaria Anthozoa. In: Harrison FW and Westfall JA (Eds), Microscopic Anatomy of Invertebrates. Vol. 2 Placozoa, Porifera, Cnidaria, and Ctenophora, New York: Wiley-Liss, USA, p. 267-358.
- FRANCIS L. 2004. Microscaling: Why larger anemones have longer cnidae. Biol Bull 207: 116-129.
- GARESE A, CARRIZO S AND ACUÑA FH. 2016. Biometry of sea anemone and corallimorpharian cnidae: statistical distribution and suitable tools for analysis. Zoomorphology 135(4): 395-404.
- GONZÁLEZ-MUÑOZ R, GARESE A, TELLO-MUSI JL AND ACUÑA FH. 2017. Morphological variability of the "Caribbean hidden anemone" *Lebrunia coralligens* (Wilson, 1890). Zoomorphology 136(3): 287-297.
- GONZÁLEZ-MUÑOZ R, SIMÕES N, MASCARÓ M, TELLO-MUSI JL, BRUGLER MRAND RODRÍGUEZ E. 2015. Morphological and molecular variability of the sea anemone *Phymanthus crucifer* (Cnidaria, Anthozoa,

Actiniaria, Actinoidea). J Mar Biol Assoc UK 95(1): 69-79.

- HAND C. 1955. The sea anemones of central California. Part II. The endomyarian and mesomyarian anemones. Wasmann J Biol 12: 345-375.
- HOLST S, SÖTJE I, TIEMANN H AND JARMS G. 2007. Life cycle of the rhizostome jellyfish *Rhizostoma octopus* (L.) (Scyphozoa, Rhizostomeae), with studies on cnidocysts and statoliths. Mar Biol 151(5): 1695-1710.
- KRAMER AM AND FRANCIS L. 2004. Predation resistance and nematocyst scaling for *Metridium senile* and *M. farcimen*. Biol Bull 207 (2): 130-140.
- MARISCAL RN. 1974. Nematocysts. In: Muscatine L and Lenhoff HM (Eds), Coelenterate Biology: Reviews and New Perspectives. New York: Academic Press, USA, p. 129-178.
- MARISCAL RN. 1984. Cnidaria: Cnidae. In: Bereiter-Hahn J, Matoltsy AG and Richards KS (Eds), Biology of the Integument, vol. 1, Invertebrates. Heidelberg: Springer-Verlag, Berlin, p. 57-68.
- MARTÍNEZ-BARALDÉS I, LÓPEZ-GONZÁLEZ PJ AND MEGINA C. 2014. Application of cnidae composition in phylogenetic analyses of North Atlantic and Mediterranean dendrophylliid corals (Anthozoa: Scleractinia). Invertebr Syst 28(2): 214-230.
- ÖSTMAN C. 1988. Nematocysts as taxonomic criteria within the family Campanulariidae, Hydrozoa. In: Hessinger DA and Lenhoff HM (Eds), The Biology of Nematocysts. Academic Press Inc., San Diego, p. 501-517.
- ÖSTMAN C. 2000. A guideline to nematocyst nomenclature and classification, and some notes on the systematic value of nematocysts. Sci Mar 64: 31-46.
- PICCIANIN, PIRES DO AND SILVA HR. 2011. Cnidocysts of Caryophylliidae and Dendrophylliidae (Cnidaria: Scleractinia): Taxonomic Distribution and Phylogenetic Implications. Zootaxa 3135: 35-54.
- PIRES DO. 1997. Cnidae of Scleractinia. Proc Biol Soc Wash 110: 167-185.
- PIRES DO AND PITOMBO FB. 1992. Cnidae of the Brazilian Mussidae (Cnidaria: Scleractinia) and their value in taxonomy. Bull Mar Sci 51: 231-244.
- REITZELAM, DALY M, SULLIVAN JC AND FINNERTY JR. 2009. Comparative anatomy and histology of developmental and parasitic stages in the life cycle of the lined sea anemone *Edwardsiella lineata*. J Parasitol 95(1): 100-112.

- SCHMIDT H. 1969. Die Nesselkapseln der Aktinien und ihre diferential diagnostische Bedeutung. Helgoländer wiss Meeresunters 19: 284-317.
- SCHMIDT H. 1972. Prodromus zu einer Monographie der mediterranen Aktinien. Zoologica 42(121): 1-146.
- SCHMIDT H. 1974. On evolution in the Anthozoa. In: International Coral Reef Symposium, 2, Great Barrier Reef. Proceedings of the 2nd International Coral Reef Symposium, Great Barrier Reef Committee 1: 533-560.
- TERRÓN-SINGLER A AND LÓPEZ-GONZÁLEZ PJ. 2005. Cnidae variability in *Balanophyllia europaea* and *B. regia* (Scleractinia: Dendrophylliidae) in the NE Atlantic and Mediterranean Sea. Sci Mar 69(1): 75-86.
- THOMASON JC. 1988. The allometry of nematocysts. In: Hessinger DA and Lenhoff HM (Eds), The biology of the nematocysts. San Diego: Academic Press, USA, p. 575-588.
- WATTS PC, ALLCOCK AL, LYNCH SM AND THORPE JP. 2000. An analysis of the nematocysts of the beadlet anemone *Actinia equina* and the green sea anemone *Actinia prasina*. J Mar Biol Ass UK 80: 719-724.
- WEILL R. 1934. Contribution a l'étude des cnidaires et de leurs nématocystes. I. Recherches sur les nematocysts. Trav St Mar Wimereux 10: 1-347.
- WILLIAMS RB. 1996. Measurements of cnidae from sea anemones (Cnidaria: Actiniaria): statistical parameters and taxonomic relevance. Sci Mar 60: 339-351.
- WILLIAMS RB. 1998. Measurements of cnidae from sea anemones (Cnidaria: Actiniaria), II: further studies of differences amongst sample means and their taxonomic relevance. Sci Mar 62: 361-372.
- WILLIAMS RB. 2000. Measurements of cnidae from sea anemones (Cnidaria: Actiniaria), III: ranges and other measures of statistical dispersion, their interrelations and taxonomic relevance. Sci Mar 64: 49-68.
- ZAMPONI MO AND ACUÑA FH. 1991. La variabilidad de los cnidocistos y su importancia en la determinación de clines. Physis 49: 7-18.
- ZAMPONI MO AND ACUÑA FH. 1994. Una metodología para estudios cuantitativos de los cnidocistos (Actiniaria, Actiniidae). Iheringia 76: 9-13.
- ZAMPONI MO AND EXCOFFON AC. 1986. Algunos aspectos de la biología reproductiva de *Bunodactis marplatensis* Zamponi, 1977 (Actiniaria: Actiniidae). Spheniscus 4: 9-18.