HIGH IMPACT REVIEW IN NEUROSCIENCE, NEUROLOGY OR PSYCHIATRY - REVIEW ARTICLE



Tetracycline repurposing in neurodegeneration: focus on Parkinson's disease

Mariza Bortolanza^{1,2} · Glauce C. Nascimento^{1,2} · Sergio B. Socias³ · Diego Ploper³ · Rosana N. Chehín³ · Rita Raisman-Vozari⁴ · Elaine Del-Bel^{1,2}

Received: 18 April 2018 / Accepted: 3 August 2018 © Springer-Verlag GmbH Austria, part of Springer Nature 2018

Abstract

The prevalence of Parkinson's disease, which affects millions of people worldwide, is increasing due to the aging population. In addition to the classic motor symptoms caused by the death of dopaminergic neurons, Parkinson's disease encompasses a wide range of nonmotor symptoms. Although novel disease-modifying medications that slow or stop Parkinson's disease progression are being developed, drug repurposing, which is the use of existing drugs that have passed numerous toxicity and clinical safety tests for new indications, can be used to identify treatment compounds. This strategy has revealed that tetracyclines are promising candidates for the treatment of Parkinson's disease. Tetracyclines, which are neuroprotective, inhibit proinflammatory molecule production, matrix metalloproteinase activity, mitochondrial dysfunction, protein misfolding/aggregation, and microglial activation. Two commonly used semisynthetic second-generation tetracycline derivative/ neuropsychiatric diseases and no substantial toxicity. Moreover, novel synthetic tetracyclines with different biological properties due to chemical tuning are now available. In this review, we discuss the multiple effects and clinical properties of tetracyclines of tetracyclines regulate inflammasome signaling. Based on their excellent safety profiles in humans from their use for over 50 years as antibiotics, we propose the repurposing of tetracyclines, a multitarget antibiotic, to treat Parkinson's disease.

Keywords Doxycycline · Antibiotic · Drug repurposing · Neuroprotection · Parkinson's disease · Tetracycline

Mariza Bortolanza and Glauce Crivelaro-do-Nascimento contributed equally to the review.

Electronic supplementary material The online version of this article (https://doi.org/10.1007/s00702-018-1913-1) contains supplementary material, which is available to authorized users.

Rita Raisman-Vozari ritaraisman@gmail.com

Elaine Del-Bel eadelbel@usp.br

¹ Department of Morphology, Physiology and Basic Pathology, Faculty of Odontology of Ribeirão Preto, University of São Paulo (USP), Av do Café s/n, São Paulo, Brazil

Introduction

Patients with Parkinson's disease (PD), which is a chronic neurological disorder caused by the loss of dopaminergic neurons in the substantia nigra pars compacta and their terminals in the striatum, exhibit decreased striatal dopamine, which results in the classical PD motor symptoms of bradykinesia, rigidity, and tremor. Nondopaminergic

- ² Center of Interdisciplinary Research on Applied Neurosciences (NAPNA), USP, São Paulo, Brazil
- ³ Instituto de Investigación en Medicina Molecular y Celular Aplicada (IMMCA) (CONICET/UNT/SIPROSA), Pasaje Dorrego, 1080-4000 Tucumán, Argentina
- ⁴ Sorbonne Universite, UPMC Univ Paris 06, INSERM, CNRS, UM75, U1127, UMR 7225, Institut du Cerveau et de la Moelle Epinière, Paris, France

neurons also degenerate during the evolution of the disease and account for the dopamine-resistant symptoms, including olfactory dysfunction, autonomic dysfunction, sleep disorder, pain, and sensory problems (Rodríguez-Violante et al. 2017). Several factors at the cellular level are thought to underlie the neuronal demise in PD. These include oxidative stress, mitochondrial and lysosomal dysfunction, apoptosis, the formation of a pathologic species of aggregated α -synuclein protein, and inflammatory changes (Bi et al. 2013; Olanow 2007). Thus, PD clearly involves multifactorial characteristics.

PD has an inflammatory component, which has been confirmed by postmortem brain studies of patients with PD, serum and cerebrospinal fluid cytokine analyses, and examinations of risk factor associations with cytokine and major histocompatibility complex polymorphisms (Frank-Cannon et al. 2009; Hirsch and Hunot 2009; Lee et al. 2009; Tansey and Goldberg 2010). Epidemiological studies of anti-inflammatory therapies (Hirsch and Hunot 2009; McGeer and McGeer 2008) have indicated a reduced risk of PD among long-term users of nonsteroidal anti-inflammatory drugs (Chen et al. 2003; Ton et al. 2006). However, the results of anti-inflammatory therapies are inconsistent as some nonsteroidal anti-inflammatory drugs have been shown to exacerbate neurodegeneration (Lleo et al. 2007). Moreover, anti-inflammatory efficacy has not been reproduced in all clinical trials (Bartels et al. 2010). These findings suggest that multitarget drugs are more appropriate than treatment with anti-inflammatories alone.

Consistent with the multifactorial origin of PD, pathogenic interactions among the diverse pathologic mechanisms underlying the development of the disease have been proposed to enhance neuronal death. Thus, the release of aggregated α -synuclein from neurons might activate microglia and trigger the production of proinflammatory mediators and neurotoxic factors, which then lead to neuronal damage (Codolo et al. 2013; Santa-Cecilia et al. 2016). Among these proinflammatory mediators is interleukin (IL)-1 beta (IL-1 β), which is one of the most abundant and strongest proinflammatory cytokines (Codolo et al. 2013). The synthesis of IL-1 β is induced by fibrillar α -synuclein through an interaction with the toll-like receptor-2. Furthermore, IL-1ß secretion involves the activation of the nucleotide-binding domain and leucine-rich repeat-containing family, pyrin domain-containing-3 (NLRP3) inflammasome (Codolo et al. 2013).

Inflammasomes are multiprotein complexes that play a central role in inflammatory immune responses. They are primed by the mitogen-activated protein kinase (MAPK) and nuclear factor-kappa B (NF- κ B) pathways and activated by several damage-associated molecular patterns and pathogen-associated molecular patterns (Kauppinen et al. 2013). Inflammasomes are critical in regulating the

maturation of the proinflammatory IL-18 and IL-1 β (Codolo et al. 2013; Lin and Zhang 2017). Interestingly, Codolo et al. (2013) have found that only fibrillar α -synuclein induces the release of IL-1 β by monocytes after activation of the NLRP3 inflammasome.

Unfortunately, only symptomatic treatments are available for PD, and no cure nor disease-modifying drug currently exists (Kowal et al. 2013). Although several pharmacological compounds improve nigrostriatal pathway function and alleviate the motor and nonmotor symptoms of the disease, these compounds do not slow disease progression (Schapira 2005). Indeed, the effects of dopamine replacement therapy with L-3,4-dihydroxyphenylalanine (L-DOPA), which is the most efficient treatment of the PD motor symptoms (Fahn 2008; Olanow 2008), tend to decrease with time. Longer L-DOPA treatments are accompanied by motor and/or psychiatric side effects that are extremely discomforting for the patients. Therefore, future drug research and development must focus on finding compounds that slow dopaminergic neurodegeneration and impede illness progression (LeWitt and Nyholm 2004; Schapira et al. 2006). Despite the huge efforts and resources already invested, the development of new neuroprotective drugs has high rates of failure. In fact, several promising drugs have failed once they reached clinical trials mainly due to safety issues, including unexpected clinical side effects and/or tolerability (Ashburn and Thor 2004; Cha et al. 2018). Therefore, drug repurposing might be useful for overcoming this bottleneck in PD treatment by avoiding future derailments due to appearance of toxicities that were not predicted by the preclinical research.

In addition to their antimicrobial properties, tetracyclines are antibiotics that may protect against neurodegenerative (Blum et al. 2004; Forloni et al. 2009; Noble et al. 2009a, b; Ruzza et al. 2014; Stoilova et al. 2013) and neuropsychiatric (Keller et al. 2013) diseases. These drugs exhibit an array of brain protective functions (Gordon et al. 2012; Moon et al. 2012), including the reduction of neuroinflammatory processes (Gordon et al. 2012; Noble et al. 2009a, b; Nordstrom et al. 1998; Sultan et al. 2013) and prevention of mitochondrial-mediated cytochrome c release, glutamate neurotoxicity, and oxidative stress (Kim and Suh 2009; Mao et al. 2005; Rothstein et al. 2005; Tomiyama et al. 1996a). In addition, they are effective inhibitors of metalloproteinases (Cathcart and Cao 2015; Cho et al. 2009; Lee et al. 2009), tumor progression (Amin et al. 1997), and angiogenesis induction (Furst 1998). Importantly, subantimicrobial doses of tetracyclines have been successfully used for the treatment of acne vulgaris, rosacea, and periodontal disease without serious side effects on the patient's health, which suggests the safety of antibiotic therapy (Skidmore et al. 2003). Interestingly, Egeberg et al. (2016) have reported that increased tetracycline use in rosacea treatment is associated with a small but appreciable reduction in the risk of developing PD,

which suggests that tetracyclines have potential for treating PD.

Clinically, the off-target effects of drugs are critically important. Drug repositioning or repurposing is an attractive alternative (Ashburn and Thor 2004; Johnston et al. 2018; Stock et al. 2013) to de novo drug development. Specifically, drug repositioning reduces development risk because repositioning candidates have usually already been through several stages of clinical development and therefore have well-known safety and pharmacokinetic profiles (Papapetropoulos and Szabo 2018). De novo drug discovery and development, which begins with an idea and continues until the drug is marketed, is a 10–17-year process (Ashburn and Thor 2004; Cha et al. 2018). Thus, drug repurposing may be key for the faster development of neuroprotective treatments against PD.

The aim of the present review was to carefully compile evidence of the potential effects of tetracyclines in the treatment and/or prevention of PD. Here, we briefly examine the chemical structure, mechanisms of action, and properties of tetracyclines that may result in neuroprotection. Finally, we will summarize the available data on the preclinical and clinical testing of tetracyclines.

The structure-activity relationship of tetracycline

Tetracycline, which is a natural fermentation product of the soil bacterium *Streptomyces aureofaciens* was discovered by Benjamin Duggar in (1948). The first tetracycline to be chemically purified was chlortetracycline in 1954 (Griffin et al. 2010; Sapadin and Fleischmajer 2006). The mechanisms of action underlying the antibiotic properties of tetracyclines are related to their ability to bind to the bacterial 30S ribosomal subunit, which then halts protein translation. Upon binding to the ribosome, tetracyclines allosterically inhibit the binding of the amino acyl-tRNA at the acceptor site, which then prevents assembly of the translational machinery (Connamacher and Mandel 1965; for review, see; Nelson and Levy 2011).

Currently, the following three groups of tetracyclines exist: tetracycline natural products, tetracycline semisynthetic compounds, and chemically modified tetracyclines (CMTs; Golub et al. 1992; Nelson 1998; for review, see; Swamy et al. 2015). All tetracyclines consist of a linearfused tetracyclic nucleus (rings designated a–d in Fig. 1), which is an important feature in its antibacterial activity. The ring structure is surrounded by upper and lower peripheral zones that contain various chemical functional groups (Martin 1985). The dimethylamine group at the C4 carbon on the upper half is necessary for its antimicrobial activity (Fig. 1). Interference with this region reduces or eliminates





Fig. 1 Chemical structure and activity relationship of tetracycline, DOX, MIN and CMT-3. Tetracycline molecules are all comprised by a linear-fused tetracyclic nucleus (rings designated **a**–**d**), an important feature for the antibacterial activity. Modifying the upper peripheral zone (positions C7 through C9 of the D ring) led to the synthesis of molecules with higher activity. The lower peripheral region contains functional groups that are responsible for inhibition of protein aggregation and chelation of metal ions. This feature has an influence on both their antimicrobial and pharmacokinetic properties (Socias et al. 2018; for review see; Bahrami et al. 2012; Chopra and Roberts 2001). Modification of the lower peripheral region reduces both antibiotic and nonantibiotic properties (Nelson 1998)

the effectiveness of the drug as an antibiotic (Golub et al. 1998). The lower peripheral region contains functional groups that are responsible for the chelation of metal ions (Fig. 1; Bahrami et al. 2012).

Chemical modifications have produced the following two newer semisynthetic second-generation tetracyclines: 6-deoxy-5-hydroxytetracycline [doxycycline (DOX)] and 7-dimethylamino-6-demethyl-6-deoxytetracycline [minocycline (MIN)]. DOX and MIN, which are most commonly used clinically as antibiotics, are characterized by reduced toxicity, enhanced antibacterial activity, longer half-lives, superior tissue fluid penetration, easier penetration of the blood-brain barrier (Domercq and Matute 2004), and rapid and complete absorption, even in aging individuals (Sande and Mandell 1985). DOX is indicated for a variety of infections, including anthrax, chlamydia, community-acquired pneumonia, Lyme disease, cholera, syphilis, Yersinia pestis (plague), periodontal infections, severe acne, and malaria. MIN, which is used more often, also displays broad-spectrum efficacy and is indicated for many of the same infections as DOX (Joshi and Miller 1997). However, the effects of MIN and DOX in the central nervous system are not fully understood nor have they been characterized.

A new family of interesting CMT compounds has been chemically modified to eliminate their antimicrobial activities while retaining their anticollagenase activities (Golub et al. 1992, 1998, 1999). Currently, more than eight CMTs are available (Gu et al. 2011). Among them, CMT-1, CMT-3, and CMT-8 have been tested in medical applications. CMT-3 [6-demethyl-6-deoxy-4-de(dimethylamino)-tetracycline] is the only CMT that has been tested in clinical trials on cancer patients (Agnihotri and Gaur 2012). CMTs do not produce major side effects compared to antimicrobial tetracycline therapy, and their administration in experimental animals does not produce tetracycline-resistant microorganisms in the oral and gut flora (Golub et al. 1991).

The pleiotropic properties of CMT-3 provide impressive therapeutic potential for reducing excessive connective tissue breakdown during various pathologic processes, including those in inflammatory diseases (Chu et al. 2007; Dezube et al. 2006; Fingleton 2003; Greenwald 1998). CMT-3 inhibits lipopolysaccharide (LPS)-induced microglia activation and cytokine expression in the brain (Edan et al. 2013). Because CMT-3 is highly lipophilic, it is expected to cross the blood–brain barrier and therefore affect cells within the brain (Chen et al. 2000; Edan et al. 2013).

The nonantimicrobial properties of tetracyclines

A wide spectrum of effects of tetracyclines in the nervous system can be attributed to their nonantibiotic properties (Sapadin and Fleischmajer 2006). These properties are listed below.

Matrix metalloproteinase (MMP) inhibition

MMPs are produced by inflammatory and connective tissue cells. The inhibition of MMPs is beneficial in many pathological conditions in which the MMP-mediated proteolysis of the extracellular matrix contributes to inflammation, which has been shown in animal model studies of stroke, neurodegeneration, neuroimmunity, and neuroinfection (Plane et al. 2010). Tetracyclines are thought to exert their antiproteolytic effects both through the direct inhibition of MMPs activity and the inhibition of their expression (Griffin et al. 2010). Because MMP transcription is induced by numerous proinflammatory cytokines and growth factors,

including IL-1, IL-6, tumor necrosis factor-alpha (TNF- α), and epidermal growth factor, the upstream signaling cascades that induce MMP expression are probably important targets of tetracyclines (Hanemaaijer et al. 1998).

Reactive oxygen species (ROS) scavenging

The increased production of ROS under many pathological conditions results in oxidative destruction or the dysfunction of many cellular constituents. The neuroprotective roles of DOX and MIN have been attributed to their ability to scavenge ROS and free radicals (Bahrami et al. 2012; Garcia-Martinez et al. 2010; Nikodemova et al. 2006; Plane et al. 2006). MIN has been shown to directly scavenge ROS in several cell-free mixed-radical assays (Kraus et al. 2005). Additionally, MIN is very effective for quenching H_2O_2 and scavenging superoxide and peroxynitrite through direct interactions with these free radicals (Kraus et al. 2005; Whiteman and Halliwell 1997).

MIN and DOX inhibit oxidative stress by also attenuating the expression of inducible nitric oxide (NO) synthase (iNOS; Amin et al. 1997). NO reacts with oxygen radicals and forms cytotoxic species, such as peroxynitrite. Tetracyclines can act on this enzyme at the transcriptional and/or translational level, which accounts for the observations of decreased protein levels and specific activity of the enzyme and the subsequent reduction in NO production (Amin et al. 1997; DeClerck et al. 1994; Rifkin et al. 1994).

Antiapoptotic effects

A key event in the execution of the apoptotic cascade is the activation of caspases, which are a family of cysteine proteases. Tetracyclines possess antiapoptotic properties that result in reductions in the expression of caspase-1 and/or caspase-3. In addition, MIN enhances the effects of B-cell lymphoma-2 (Bcl-2), which protects cells against apoptosis (Jordan et al. 2007; Wang et al. 2010).

Anti-inflammatory effects

The downregulated expression of proinflammatory mediators is a well-characterized and common effect of all tetracyclines, especially MIN and DOX (Golub et al. 1998). Reports have shown that tetracycline, DOX, and MIN decrease inflammations of various etiologies (Bahrami et al. 2012). Thus, tetracyclines attenuate both innate and adaptive immune responses (Griffin et al. 2010).

MIN and DOX exert their anti-inflammatory effects in the brain by modulating glial cells. Microglial activation occurs in most neurodegenerative diseases and results in the release of proinflammatory mediators and other injury response factors that compromise cell viability (Domercq and Matute 2004). By reducing microglial activation, tetracyclines reduce the transcription of downstream proinflammatory mediators, such as caspase-1, iNOS, and cyclooxygenase 2, and the subsequent release of IL-1 β , NO, and prostaglandin E₂, which are associated with neuronal cell death (Domercq and Matute 2004; Orsucci et al. 2009). In addition, MIN attenuates the p38 MAPK cascade, which reduces inflammatory cytokine synthesis (Bahrami et al. 2012; Tikka and Koistinaho 2001; Wu et al. 2002; Yrjanheikki et al. 1998).

MIN and, to a lesser extent, DOX inhibit phospholipase A_2 (Pruzanski et al. 1992) and neutrophil migration (Esterly et al. 1984) and adherence (Gabler and Creamer 1991). In addition, DOX reduces mitogen-induced proliferative responses of lymphocytes (Thong and Ferrante 1979).

Furthermore, DOX (Cox et al. 2010) and MINO (Tamargo et al. 1991) inhibit angiogenesis, which occurs also in neurodegenerative diseases (for review see Bradaric et al. 2012).

Regarding the neuroinflammation induced by α -synuclein, Codolo et al. (2013) have suggested that blocking IL-1 β activity may be a valuable alternative to reducing and/or stopping α -synuclein-triggered immune responses because this cytokine is the final product of the NLRP3 inflammasome. Thus, DOX exerts strong anti-inflammatory actions on microglial cells and ceases the production of inflammatory mediators by suppressing the NF- κ B and p38 MAPK pathways. Moreover, DOX strongly restricts the production/ release of IL-1 β and TNF- α (Santa-Cecilia et al. 2016) and diminishes the increase in TNF- α and IL-1 β mRNA transcripts observed in LPS-stimulated BV-2 cells (Cho et al. 2009).

Protein antiaggregation activity

Neurodegenerative diseases can be classified according to their predominant protein aggregates (Maiti et al. 2014). An important hallmark of neurodegenerative disorders, including PD and Alzheimer's disease, is the intraneuronal (tau or α -synuclein) and extracellular [amyloid beta (A β) peptide] accumulation of misfolded proteins. The evidence suggests that α -synuclein is involved in the pathogenesis of several disorders through the promotion of the fibrilization of tau and A β as well as the phosphorylation of tau (Wong and Krainc 2017).

In the postmortem brains, specifically the substantia nigra and striatum, of patients with PD, increased α -synuclein accumulation has been observed in mitochondria (Subramaniam et al. 2014). In turn, the mitochondrial dysfunction caused by α -synuclein might induce the enhanced production of ROS, which appears to modulate α -synuclein oligomerization and cytotoxicity (Brahmachari et al. 2016). Recently, Socias et al. (2018) have analyzed the antiamyloidogenic effects of different antibiotics on well-known disease-associated proteins. Their results suggested that a specific structural motif in tetracyclines is key for the inhibition of protein amyloid aggregation (Socias et al. 2018) and that the oxidation products of antibiotics with additional hydroxyl groups on their rings further inhibit α -synuclein fibrillation (Fig. 1). DOX also induces the remodeling of α -synuclein oligomers into off-pathway nontoxic and nonseeding species. Interestingly, this remodeling process is only effective on the early species in the aggregation process (González-Lizárraga et al. 2017).

Protection against mitochondrial dysfunction

Mitochondrial dysfunction has also been implicated in multifactorial age-related diseases, including PD (Andreux et al. 2013). Garcia-Martinez et al. (2010) have postulated that mitochondria are pharmacological targets of MIN. When MIN is added to isolated mitochondria, it decreases the mitochondrial inner membrane potential, which might prevent mitochondrial permeability, transition pore opening, and the subsequent release of cytochrome c. In addition, MIN decreases the voltage dependence in a concentrationdependent manner, which alters the permeability of the mitochondrial outer membrane (Garcia-Martinez et al. 2010). These transmembrane potential changes might contribute to the various cytoprotective mechanisms described above.

The pituitary adenylate cyclase-activating polypeptide (PACAP), histone methylation, and poly (ADP-ribose) polymerase-1 (PARP-1): new targets for the neuroprotective effects of tetracyclines?

The pituitary adenylate cyclase-activating polypeptide receptor 1 (PAC1R) is located in the central and peripheral nervous systems, in which it mediates antiapoptotic (Seaborn et al. 2011), anti-inflammatory (Martínez et al. 2006), and neuroprotective (Bourgault et al. 2009) effects. DOX, which is a positive allosteric modulator of PAC1, enhances the activation of this receptor in vivo (Yu et al. 2016). Therefore, PAC1 may be the source of yet another neuroprotective function of DOX, which suggests that the anti-inflammatory effects of DOX interact with the functions of PAC1 (Reglodi et al. 2017).

The activation of PARP-1 by DNA damage promotes both cell death and inflammation. Alano et al. (2006) have reported that the enzymatic activity of PARP-1 is directly inhibited by MIN, DOX, and other tetracycline derivatives that have neuroprotective and anti-inflammatory actions. The neuroprotective and anti-inflammatory effects of tetracycline derivatives may be attributable to PARP-1 inhibition, particularly under conditions in which cell death is mediated primarily by PARP-1 activation. MIN has a beneficial effect on DNA damage, which might be due to its ability to inhibit PARP-1 activation. In addition, MIN might regulate the levels of histone methylation, which is partially explained by its ability to regulate histone methyltransferases or demethylases (Wang et al. 2017).

In summary, it is unclear whether tetracyclines have one or several modes of action. Despite this question and considering the wide range of protective effects reported in different models of various brain diseases, MIN and DOX are considered potential therapeutic agents in the treatment of neurodegenerative disorders (Noble et al. 2009a, b; Yong et al. 2004).

Doxycycline and minocycline in Parkinson's disease

Tetracycline derivatives, which have slowly been recognized as a genre of drugs with pleiotropic properties, have become an alternative form of therapy in neurodegenerative disorders in which inflammation contributes to disease progression (Reglodi et al. 2015; Socias et al. 2018). Indeed, this class of drugs has been reported to exert unique effects on complex pathologies (Griffin et al. 2010). The literature contains 242 clinical trials of MIN and 262 clinical trials of DOX in many types of systemic diseases. In particular, these clinical trials involve patients and/or experimental models related to numerous neurological conditions, including schizophrenia, Alzheimer's disease, amyotrophic lateral sclerosis, Huntington's disease, PD, and autism (Supplementary Table 1; Gordon et al. 2007; Kelly et al. 2015; Loeb et al. 2004; Molloy et al. 2013; Parashos et al. 2014; Pardo et al. 2013; Zhang et al. 2003). Here, we review the evidence for the protective actions of MIN and DOX in PD.

Minocycline

The neuroprotective effects of MIN in PD experimental models have been reported since 2001. In 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) models of PD, MIN prevents dopaminergic nigrostriatal neurodegeneration (Du et al. 2001; He et al. 2001; Wu et al. 2002). In addition, MIN decreases MPTP-mediated nitrotyrosine formation and inhibits MPTP-induced microglial activation, which prevents the production of microglia-derived proinflammatory factors, including IL-1 β , ROS, and NO (Wu et al. 2002). It also inhibits MPP + mediated iNOS expression in vivo and potently blocks NO-induced neurotoxicity in vitro (Du et al. 2001).

The neuroprotective effects of MIN have also been observed after chronic rotenone toxicity in wild-type rodents (Radad et al. 2010), parkin null mice (Casarejos et al. 2006), and PD model *Drosophila* (Faust et al. 2009). In addition, MIN administration has been shown to reduce the number of apomorphine-induced rotations in 6-hydroxydopamine (6-OHDA)-lesioned rats, the loss of tyrosine hydroxylase-positive cells, increasing the size and fiber density of the remaining nigral cells (Quintero et al. 2006). Moreover, MIN exhibits protective effects on nigrostriatal dopaminergic neurodegeneration in the *Weaver* mouse, which has a mutation in the gene encoding for the G-protein-activated inward rectifier potassium channel 2 (Peng et al. 2006).

To investigate the effects of anti-inflammatory treatments on regeneration, Worlitzer et al. (2013) administered MIN for 14 weeks starting 2-3 weeks after injections of 6-OHDA in a mouse model of PD. MIN treatment induced functional regeneration that was dopaminergic neuron activity-dependent. Anti-inflammatory treatment after the degeneration of dopaminergic neurons in the substantia nigra increased the activity of adult neural stem cells in the subventricular zone, which resulted in the generation of neuroblasts that migrated deeply into the lesioned striatum. These newly generated cells differentiated into mature striatal oligodendrocytes. These results suggested that oligodendrogenesis was responsible, at least in part, for the behavioral improvements in the PD symptoms due to the increased stability and efficiency of axonal function in the remaining ipsilateral and/or crossing contralateral dopaminergic neurons (Worlitzer et al. 2013).

Interestingly, MIN prevents *N*-methyl D-aspartate (NMDA) glutamate receptor-induced neuronal death by inhibiting the activation and proliferation of microglia cells in culture (Tikka and Koistinaho 2001). Those authors also showed that MIN inhibits the NMDA-induced activation of p38 MAPK and a specific p38 MAPK inhibitor, while not that of a p44/42 MAPK inhibitor, in microglial cells, which then reduces NMDA toxicity.

Nonetheless, conflicting evidence for the efficacy of MIN has been reported in animal models of PD (Sriram et al. 2006; Yang et al. 2003). MIN has variable and even harmful effects in MPTP models due to the exacerbation of the MPTP-induced damage to dopaminergic neurons both in vitro and in vivo (Yang et al. 2003). Additional uncertainty results from the apparent lack of robust therapeutic efficacy of MIN in clinical trials in which MIN-treated patients with PD exhibited trends for worse Unified Parkinson's Disease Rating Scale scores compared to placebotreated controls (Gordon et al. 2007). In contrast, another trial showed additive neuroprotective effects of MIN when it was combined with creatine, which has been shown to reduce PD progression in patients (NINDS NET-PD, 2006). Thus, several questions on the efficacy of MIN as a viable PD treatment remain.

Doxycycline

DOX is a highly effective and inexpensive antibiotic with a broad therapeutic spectrum and exceptional bioavailability.

Not surprisingly, DOX is included in the World Health Organization's List of Essential Medicines needed for basic health care systems (http://www.who.int/medicines/publi cations/essentialmedicines/en/; Perez-Trallero and Iglesias 2003). DOX was immediately popular after its use was approved by the Food and Drug Administration in 1967 because of its simplified once (or twice)-a-day dosage regimen compared to the four-times-a-day dosing schedule of tetracycline. In addition, DOX presents minimal side effects, even after its long-term administration (Gompels et al. 2006; Langevitz et al. 1992; Smith et al. 2011).

DOX has various effects on central nervous system functions. Because it is already clinically available, it is an obvious and attractive candidate for drug repurposing. Subantimicrobial doses of DOX (20 mg twice daily with serum concentrations in the range of 200–600 ng/mL) have been repeatedly demonstrated as safe with no evidence of posttreatment microbiologic resistance (Payne et al. 2011).

The neuroprotective effects of DOX on dopaminergic neurons have been demonstrated both in vitro and in vivo (Cho et al. 2011; Lazzarini et al. 2013). These effects seem to result from antiapoptotic and anti-inflammatory mechanisms involving the downregulation of MMPs (Cho et al. 2011; Fig. 2). Nevertheless, it has become clear that the use of DOX to control Tet-ON/Tet-OFF systems (DOX-inducible systems) involves risk (Lewandoski 2001) in that the use of DOX per se produces unexpected experimental outcomes. Recently, we showed that the systemic treatment of mice in a 6-OHDA model of PD with DOX protected dopaminergic neurons (Lazzarini et al. 2013) and that this neuroprotection was associated with a reduction in microglial activation. This important discovery happened by accident (serendipity) when 6-OHDA-lesioned mice were fed chow containing DOX. Comparable results were obtained in another group of mice that were later administered DOX subcutaneously (Fig. 2). The 6-OHDA model causes neuroinflammatory responses (Taylor 2013; Tufekci et al. 2012), including reactive astrocytosis (Wachter et al. 2010) and microglial activation (Lazzarini et al. 2013; Marinova-Mutafchieva et al. 2009; Fig. 2). These experimental results strongly suggest that DOX is a promising alternative for the treatment of PD.

Additionally, our group has also reported a direct effect of DOX in the activation of primary microglia in vitro (Santa-Cecília et al. 2016). DOX reduces the LPS-induced activation of microglial cells in a concentration-dependent manner (200–300 μ M) by preventing increase in the expression of Iba-1. Additionally, DOX reduces the expression of inflammatory mediators, such as TNF- α , ROS, and iNOS. This effect was associated with the inhibition of the production of the proinflammatory cytokines TNF- α and IL-1 β by LPS. One possible molecular mechanism underlying these effects is that DOX inhibits the activation of the p38 MAPK and NF- κ B signaling pathways



Fig. 2 Tetracycline and neuronal cell death. Tetracyclines can protect against neuronal cell death by directly suppressing expression of MMPs and activation of glial cells, remodeling α -synuclein early aggregates and scavenging ROS. Indirectly, DOX is able to inhibit mitochondrial dysfunction and subsequent oxidative damage caused by α -synuclein aggregates or production of proinflammatory mediators from glial cell activation, which, in turn, leads to MMP activation. *MMP* matrix metalloproteinase, *ROS* reactive oxygen species, *SYN* α -synuclein

in LPS-stimulated microglial cells. Similarly, Zhang et al. (2015) have thoroughly demonstrated that the effects of DOX in a rat model of PD occur through the inhibition of the LPS-induced degeneration of dopaminergic neurons by the downregulation of the expression of microglial major histocompatibility complex II.

Neuroinflammation and α -synuclein pathology interact synergistically and lead to PD neurodegeneration (Gao and Hong 2008). Neuron-associated proteins, particularly α -synuclein, are modified by oxidation to form nitrated α -synuclein, which misfolds and aggregates to create intracellular inclusions called Lewy bodies (Kosloski et al. 2010). More recently, González-Lizárraga et al. (2017) have reported that DOX reshapes α -synuclein oligomers and inhibits α -synuclein aggregation and the seeding of new oligomers, thus preventing cytotoxicity in dopaminergic cell lines. DOX is clearly promising for preventing and inhibiting dopaminergic cell loss (Sapadin and Fleischmajer 2006) through its anti-inflammatory effects (Joshi and Miller 1997). Importantly, the long-term administration (up to 2 years) of subantimicrobial doses of DOX has not produced antibiotic side effects in clinical trials (Golub et al. 2016; Keijmel et al. 2017). Thus, subantimicrobial doses of DOX may represent a game changer in PD therapy that will benefit a considerable number of patients.

The feasibility of treatment with tetracycline, minocycline, and doxycycline in neurodegenerative diseases

The most common side effects of an antibiotic concentration (200 mg/day) of tetracycline is gastrointestinal upset (Carter 2003), increased photosensitivity, teeth discoloration, and bone formation and growth interference in children under the age of eight (Archer and Archer 2002; Gupta et al. 2006). Other common side effects of this antibiotic dose include nausea, vertigo, and mild dizziness, which are completely reversible upon discontinuation of the drug. Additionally, liver toxicity, pigmentation, and a lupus erythematosus-like syndrome may appear during the use of this drug (Garner et al. 2003).

Although tetracycline has been studied extensively, long-term studies are rare (Smith et al. 2005). Nonetheless clearly a major concern of the use of antibiotics as adjuncts to mechanical debridement procedures is the development of antibiotic side effects (Chopra and Roberts 2001). The administration of tetracyclyne antibiotic doses (200–400 mg/day) is responsible for the antimicrobial effects, which may result in bacterial resistance and alterations in endogenous flora. In contrast, clinical trials have demonstrated that the administration of subantibiotic doses (20–40 mg/day) does not alter bacterial susceptibility to antibiotics and results in anti-inflammatory effects (Payne et al. 2011; NINDS NET-PD Investigators 2006).

Studies on the neuroprotective effects of MIN in experimental models of neurodegeneration have had promising results. However, MIN has shown variable and even contradictory (beneficial and detrimental) effects in various species and models, which suggests an urgent need for the promotion of the publication of negative results (Diguet et al. 2004).

Accordingly, additional studies are required to determine the conditions required for the safe clinical administration of subantibiotic doses during the prolonged treatments necessary in the study of neurodegenerative diseases.

Conclusion and open questions

In this review, we presented the current experimental evidence for the potential use of tetracycline and its derivatives (MIN and DOX) as neuroprotective agents in PD. MIN and DOX, which are already clinically available, have protective actions with long half-lives, minor side effects, increased lipid solubilities, good tolerances, and excellent blood-brain barrier penetrations. The use of these drugs alone or in combination with other agents offers novel therapeutic approaches that target multiple pathways that lead to the degeneration of dopaminergic neurons in PD. Indeed, this class of drugs has been reported to exert unique effects on complex pathologies (Griffin et al. 2010). In brief, both are effective, and each exhibits unique pharmacological properties that may prove to be potentially advantageous.

Because the pathology of PD is very complex, the neuroprotective properties of MIN and DOX seem to be mediated by mechanisms other than their antimicrobial effects (Fig. 2): (1) the protection of dopaminergic cells through decreased levels of intracellular MMPs, which participate in cell death signaling; (2) prevention of the activation of microglia; (3) inhibition of the production of proinflammatory molecules; and 4) inhibition of the aggregation of α -synuclein, among others. These properties are not necessarily limited to MIN and DOX as other tetracyclines have also been shown to have beneficial effects on inflammation and apoptotic cell death.

In conclusion, tetracycline and its derivatives have been increasingly recognized for their anti-inflammatory and neuroprotective potentials. While the mechanisms underlying their benefits are still unclear, this novel mode of action of tetracyclines may help in the development of more specific and effective strategies in the treatment of neurodegenerative disorders. Finally, drug repurposing has potential for quickly bringing medications with known safety profiles to new patient populations.

Neuroprotective antibiotics are a potential treatment for chronic neurological disorders that have few existing treatments. DOX, which is listed on the World Health Organization's list of essential medicines and which is already clinically available, should be considered an excellent candidate in the development of therapeutic strategies for PD.

Acknowledgements This publication was funded by Grants from Fundação de Amparo a Pesquisa do Estado de Sao Paulo (FAPESP), CAPES (Conselho de Aperfeiçoamento de Pessoal) and Conselho Nacional de Pesquisa (CNPq) to EADB. We would like to dedicate this paper to Professor Walter Stuhmer (Director-Max Planck Institute of Experimental Medicine- Göttingen-Germany) who gave us his neverceasing support in his proud struggle for the endurance of science in Latin America.

References

- Agnihotri R, Gaur S (2012) Chemically modified tetracyclines: novel therapeutic agents in the management of chronic periodontitis. Indian J Pharmacol 44:161–167. https://doi. org/10.4103/0253-7613.93841
- Alano CC, Kauppinen TM, Valls AV, Swanson RA (2006) Minocycline inhibits poly(ADP-ribose) polymerase-1 at nanomolar concentrations. Proc Natl Acad Sci USA 103(25):9685–9690. https://doi.org/10.1073/pnas.0600554103
- Amin AR, Patel RN, Thakker GD, Lowenstein CJ, Attur MG, Abramson SB (1997) Post-transcriptional regulation of inducible nitric oxide synthase mRNA in murine macrophages by doxycycline and chemically modified tetracyclines. FEBS Lett 410(2-3):259-264. https://doi.org/10.1016/S0014 -5793(97)00605-4
- Andreux PA, Houtkooper RH, Auwerx J (2013) Pharmacological approaches to restore mitochondrial function. Nat Rev Drug Discov 12:465–483. https://doi.org/10.1038/nrd4023
- Archer JSM, Archer DF (2002) Oral contraceptive efficacy and antibiotic interaction: a myth debunked. J Am Acad Dermatol 46:917–923. https://doi.org/10.1067/mjd.2002.120448
- Ashburn TT, Thor KB (2004) Drug repositioning: identifying and developing new uses for existing drugs. Nat Rev Drug Discov 3:673–683. https://doi.org/10.1038/nrd1468
- Bahrami F, Morris DL, Pourgholami MH (2012) Tetracyclines: drugs with huge therapeutic potential. Mini Rev Med Chem 12(1):44–52. https://doi.org/10.2174/138955712798868977
- Bartels AL, Willemsen AT, Doorduin J, de Vries EF, Dierckx RA, Leenders KL (2010) [11C]-PK11195 PET: quantification of neuroinflammation and a monitor of anti-inflammatory treatment in Parkinson's disease? Parkinsonism Relat Disord 16(1):57–59. https://doi.org/10.1016/j.parkreldis.2009.05.005
- Bi F, Li F, Huang C, Zhou H (2013) Pathogenic mutation in VPS35 impairs its protection against MPP(+) cytotoxicity. Int J Biol Sci 9(2):149–155. https://doi.org/10.7150/ijbs.5617
- Blum D, Chtarto A, Tenenbaum L, Brotchi J, Levivier M (2004) Clinical potential of minocycline for neurodegenerative disorders, Neurobiol Dis 17(3):359–366. https://doi.org/10.1016/j. nbd.2004.07.012
- Bonelli RM, Hödl AK, Hofmann P, Kapfhammer HP (2004) Neuroprotection in Huntington's disease: a 2-year study on minocycline. Int Clin Psychopharmacol 19(6):337–342
- Bourgault S, Vaudry D, Ségalas-Milazzo I, Guilhaudis L, Couvineau A, Laburthe M, Vaudry H, Fournier A (2009) Molecular and conformational determinants of pituitary adenylate cyclase-activating polypeptide (PACAP) for activation of the PAC1 receptor. J Med Chem 52(10):3308–3316. https://doi.org/10.1021/jm900291j
- Bradaric BD, Patel A, Schneider JA, Carvey PM, Hendey B (2012) Evidence for angiogenesis in Parkinson's disease, incidental Lewy body disease, and progressive supranuclear palsy. J Neural Transm 119:59–71
- Brahmachari S, Ge P, Lee SH, Kim D, Karuppagounder SS, Kumar M, Mao X, Shin JH, Lee Y, Pletnikova O, Troncoso JC, Dawson VL, Dawson TM, Ko HS (2016) Activation of tyrosine kinase c-Abl contributes to α-synuclein-induced neurodegeneration. J Clin Invest 126(8):2970–2988. https://doi.org/10.1172/JC185456
- Carter EL (2003) Antibiotics in Cutaneous medicine: an update. Semin Cutan Med Surg 22:196–211. https://doi.org/10.1016/ S1085-5629(03)00046-4
- Casarejos MJ, Menéndez J, Solano RM, Rodríguez-Navarro JA, García de Yébenes J, Mena MA (2006) Susceptibility to rotenone is increased in neurons from parkin null mice is reduced by minocycline. J Neurochem 97(4):934–946. https://doi.org/ 10.1111/j.1471-4159.2006.03777.x

- Cathcart JM, Cao J (2015) MMP inhibitors: past, present and future. Front Biosci (Landmark Ed) 20:1164–1178
- Cha Y, Erez T, Reynolds IJ, Kumar D, Ross J, Koytiger G, Kusko R, Zeskind B, Risso S, Kagan E, Papapetropoulos S, Grossman I, Laifenfeld D (2018) Drug repurposing from the perspective of pharmaceutical companies. Br J Pharmacol 175(2):168–180. https://doi.org/10.1111/bph.13798
- Chen M, Ona VO, Li M, Ferrante RJ, Fink KB, Zhu S et al (2000) Minocycline inhibits caspase-1 and caspase-3 expression and delays mortality in a transgenic mouse model of Huntington disease. Nat Med 6(7):797–801. https://doi.org/10.1038/77528
- Chen H, Zhang SM, Hernán MA, Schwarzschild MA, Willett WC, Colditz GA, Speizer FE, Ascherio A (2003) Nonsteroidal anti-inflammatory drugs and the risk of Parkinson disease. Arch Neurol 60(8):1059–1064. https://doi.org/10.1001/archn eur.60.8.1059
- Cho Y, Son HJ, Kim EM, Choi JH, Kim ST, Ji IJ, Choi DH, Joh TH, Kim YS, Hwang O (2009) Doxycycline is neuroprotective against nigral dopaminergic degeneration by a dual mechanism involving MMP-3. Neurotox Res 16(4):361–371. https://doi.org/10.1007/ s12640-009-9078-1
- Cho DC, Cheong JH, Yang MS, Hwang SJ, Kim JM, Kim CH (2011) The effect of minocy clineon motor neuron recovery and neuropathic pain in a rat model of spinal cord injury. J Korean Neurosurg Soc 49(2):83–91. https://doi.org/10.3340/jkns.2011.49.2.83
- Chopra I, Roberts M (2001) Tetracycline antibiotics: mode of action, applications, molecular biology, and epidemiology of bacterial resistance. Microbiol Mol Biol Rev 65:232–260. https://doi. org/10.1128/MMBR.65.2.232-260.2001
- Chopra I, Hawkey PM, Hinton M, Jordan J, Fernandez-Gomez FJ, Ramos M, Ikuta I, Aguirre N, Galindo MF (2007) Minocycline and cytoprotection: shedding new light on a shadowy controversy. Curr Drug Deliv 4(3):225–331
- Chu QS, Forouzesh B, Syed S, Mita M, Schwartz G, Cooper J, Curtright J, Rowinsky EK (2007) A phase II and pharmacological study of the matrix metalloproteinase inhibitor (MMPI) COL-3 in patients with advanced soft tissue sarcomas. Invest New Drugs 25(4):359–367. https://doi.org/10.1007/s10637-006-9031-6
- Codolo G, Plotegher N, Pozzobon T, Brucale M, Tessari I et al (2013) Triggering of inflammasome by aggregated a–Synuclein, an inflammatory response in synucleinopathies. PLoS ONE 8(1):e55375. https://doi.org/10.1371/journal.pone.0055375
- Connamacher RH, Mandel HG (1965) Binding of tetracycline to the 30S ribosomes and to polyuridylic acid. Biochem Biophys Res Comm 20:98–103
- Cox CA, Amaral J, Salloum R, Guedez L, Reid TW, Jaworski C, John- Aryankalayil M, Freedman KA, Campos MM, Martinez A, Becerra SP, Carper DA (2010) Doxycycline's effect on ocular angiogenesis: an in vivo analysis. Ophthalmology 117(9):1782–1791
- DeClerck YA, Shimada H, Taylor SM, Langley KE (1994) Matrix metalloproteinases and their inhibitors in tumor progression. Ann NY Acad Sci 732:222–232. https://doi. org/10.1111/j.1749-6632.1994.tb24738.x
- Dezube BJ, Krown SE, Lee JY, Bauer KS, Aboulafia DM (2006) Randomized phase II trial of matrix metalloproteinase inhibitor COL-3 in AIDS-related Kaposi's sarcoma: an AIDS Malignancy Consortium Study. J Clin Oncol 24(9):1389–1394. https://doi. org/10.1200/JCO.2005.04.2614
- Diguet E, Gross CE, Tison F, Bezard E (2004) Rise and fall of minocycline in neuroprotection: need to promote publication of negative results. Exp Neurol 189(1):1–4. https://doi.org/10.1016/j.expne urol.2004.05.016
- Dodel R, Spottke A, Gerhard A, Reuss A, Reinecker S, Schimke N et al (2010) Minocycline 1-year therapy in multiple-systematrophy: effect on clinical symptoms and [(11)C] (R)-PK11195

PET (MEMSA-trial). Mov Disord 25(1):97–107. https://doi. org/10.1002/mds.22732

- Domercq M, Matute C (2004) Neuroprotection by tetracyclines. Trends Pharmacol Sci 25(12):609–612. https://doi. org/10.1016/j.tips.2004.10.001
- Du Y, Ma Z, Lin S, Dodel RC, Gao F, Bales KR et al (2001) Minocycline prevents nigrostriatal dopaminergic neurodegeneration in the MPTP model of Parkinson's disease. Proc Natl Acad Sci USA 98(25):14669–14674. https://doi.org/10.1073/pnas.25134 1998
- Duggar BM (1948) Aureomycin: a product of the continuing search for new antibiotics. Ann N Y Acad Sci 51(Art. 2):177–181. https://doi.org/10.1111/j.1749-6632.1948.tb27262.x
- Edan RA, Luqmani YA, Masocha W (2013) COL-3, a chemically modified tetracycline, inhibits lipopolysaccharide-induced microglia activation and cytokine expression in the brain. PLoS ONE 8(2):e57827. https://doi.org/10.1371/journal.pone.00578 27
- Egeberg A, Hansen PR, Gislason GH, Thyssen JP (2016) Exploring the association between rosacea and Parkinson disease: a Danish nationwide cohort study. JAMA Neurol 73(5):529–534. https:// doi.org/10.1001/jamaneurol.2016.0022
- Esterly NB, Koransky JS, Furey NL, Trevisan M (1984) Neutrophil chemotaxis in patients with acne receiving oral tetracycline therapy. Arch Dermatol 120(10):1308–1313. https://doi.org/10.1001/ archderm.1984.01650460048018
- Fahn S (2008) How do you treat motor complications in Parkinson's disease: medicine, surgery, or both? Ann Neurol 64(2):56–64. https://doi.org/10.1002/ana.21453
- Faust K, Gehrke S, Yang Y, Yang L, Beal MF, Lu B (2009) Neuroprotective effects of compounds with antioxidant and anti-inflammatory properties in a Drosophila model of Parkinson's disease. BMC Neurosci 10:109. https://doi.org/10.1186/1471-2202-10-109
- Fingleton B (2003) CMT-3. CollaGenex. Curr Opin Investig Drugs 2003(12):1460–1467
- Forloni G, Salmona M, Marcon G, Tagliavini F (2009) Tetracyclines and prion infectivity. Infect Disord Drug Targets 9(1):23–30. https://doi.org/10.2174/1871526510909010023
- Frank-Cannon TC, Alto LT, McAlpine FE, Tansey MG (2009) Does neuroinflammation fan the flame in neurodegenerative diseases? Mol Neurodegener 4:47. https://doi.org/10.1186/1750-1326-4-47
- Furst DE (1998) Tetracyclines inhibit microglial activation and are neuroprotective in global brain ischemia. Curr Opin Rheumatol 10:123–128
- Gabler WL, Creamer HR (1991) Suppression of human neutrophil functions by tetracycline. J Periodontal Res 26:52–58. https:// doi.org/10.1111/j.1600-0765.1991.tb01626.x
- Gao HM, Hong JS (2008) Why neurodegenerative diseases are progressive: uncontrolled inflammation drives disease progression. Trends Immunol 29:357–365. https://doi.org/10.1016/j. it.2008.05.002
- Garcia-Martinez EM, Sanz-Blasco S, Karachitos A, Bandez MJ, Fernandez-Gomez FJ, Perez-Alvarez S, De Mera RM, Jordan MJ, Aguirre N, Galindo MF, Villalobos C, Navarro A, Kmita H, Jordan J (2010) Mitochondria and calcium flux as targets of neuroprotection caused by minocycline in cerebellar granule cells. Biochem Pharmacol 79:239–250. https://doi.org/10.1016/j. bcp.2009.07.028
- Garner SE, Eady EA, Popescu C, Newton J, Li WA (2003) Minocycline for acne vulgaris: efficacy and safety. Cochrane Database Syst Rev 8:CD002086. https://doi.org/10.1002/14651858.CD002086
- Golub LM, Ramamurthy NS, McNamara TF, Greenwald RA, Rifkin BR (1991) Tetracyclines inhibit connective tissue breakdown: new therapeutic implications for an old family of drugs. Crit Rev Oral Biol Med 2(3):297–321

- Golub LM, Soummalainen K, Sorsa T (1992) Host modulation with tetracyclines and their chemically modified analogues. Curr Opin Dent 2:80–90. https://doi.org/10.4103/0972-124X.149934
- Golub LM, Lee HM, Ryan ME, Giannobile WV, Payne J, Sorsa T (1998) Tetracyclines inhibit connective tissue breakdown by multiple non-antimicrobial mechanisms. Adv Dent Res 12:12–26. https://doi.org/10.1177/08959374980120010501
- Golub LM, Ramamurthy NS, Llavaneras A, Ryan ME, Lee HM, Liu Y, Bain S, Sorsa T (1999) A chemically modified nonantimicrobial tetracycline (CMT-8) inhibits gingival matrix metalloproteinases, periodontal breakdown, and extra-oral bone loss in ovariectomized rats. Ann N Y Acad Sci 878:290–310. https:// doi.org/10.1111/j.1749-6632.1999.tb07691.x
- Golub LM, Elburki MS, Walker C, Ryan M, Sorsa T, Tenenbaum H, Goldberg M, Wolff M, Gu Y (2016) Non-antibacterial tetracycline formulations: host-modulators in the treatment of periodontitis and relevant systemic diseases. Int Dent J 66(3):127–135. https://doi.org/10.1111/idj.12221
- Gompels LL, Smith A, Charles PJ, Rogers W, Soon-Shiong J, Mitchell A et al (2006) Single-blind randomized trial of combination antibiotic therapy in rheumatoid arthritis. J Rheumatol 33(2):224– 227. https://doi.org/10.1177/08959374980120010501
- González-Lizárraga F, Socías SB, Ávila CL, Torres-Bugeau CM, Barbosa LR, Binolfi A, Sepúlveda-Díaz JE, Del-Bel E, Fernandez CO, Papy-Garcia D, Itri R, Raisman-Vozari R, Chehín RN (2017) Repurposing doxycycline for synucleinopathies: remodelling of α-synuclein oligomers towards non-toxic parallel beta-sheet structured species. Sci Rep 7:41755. https://doi.org/10.1038/srep41755
- Gordon PH, Moore DH, Miller RG, Florence JM, Verheijde JL, Doorish C et al (2007) Efficacy of minocycline in patients with amyotrophic lateral sclerosis: a phase III randomised trial. Lancet Neurol 6(12):1045–1053. https://doi.org/10.1016/S1474 -4422(07)70270-3
- Gordon RA, Mays R, Sambrano B, Mayo T, Lapolla W (2012) Antibiotics used in nonbacterial dermatologic conditions. Dermatol Ther 25(1):38–54. https://doi.org/10.1111/j.1529-8019.2012.01496.x
- Greenwald M (1998) Re: the new macrolide antibiotics: use them carefully. Paediatr Child Health 3(4):281–282
- Griffin MO, Fricovsky E, Ceballos G, Villarreal F (2010) Tetracyclines: a pleitropic family of compounds with promising therapeutic properties. Review of the literature. Am J Physiol Cell Physiol 299(3):C539–C548. https://doi.org/10.1152/ajpcell.00047.2010
- Gu Y, Lee HM, Simon SR, Golub LM (2011) Chemically modified tetracycline-3 (CMT-3): a novel inhibitor of the serine proteinase, elastase. Pharmacol Res 64(6):595–601. https://doi. org/10.1016/j.phrs.2011.05.011
- Gupta AK, Gover MD, Abramovits W, Perlmutter A (2006) Solodyn (Minocycline HCl, USP) extended-release tablets. Skinmed 5(6):291–292
- Hanemaaijer R, Visser H, Koolwijk P, Sorsa T, Salo T, Golub LM, van Hinsbergh VW (1998) Inhibition of MMP synthesis by doxycycline and chemically modified tetracyclines (CMTs) in human endothelial cells. Adv Dent Res 12(2):114–118. https:// doi.org/10.1177/08959374980120010301
- He Y, Appel S, Le W (2001) Minocycline inhibits microglial activation and protects nigral cells after 6-hydroxydopamine injection into mouse striatum. Brain Res 909:187–193. https://doi.org/10.1016/ S0006-8993(01)02681-6
- Hirsch EC, Hunot S (2009) Neuroinflammation in Parkinson's disease: a target for neuroprotection? Lancet Neurol 8(4):382–397. https ://doi.org/10.1016/S1353-8020(11)70065-7
- Johnston TH, Lacoste AMB, Visanji NP, Lang AE, Fox SH, Brotchie JM (2018) Repurposing drugs to treat I-DOPA-induced dyskinesia in Parkinson's disease. Neuropharmacology. https://doi. org/10.1016/j.neuropharm.2018.05.035)

- Jordan J, Fernandez-Gomez FJ, Ramos M, Ikuta I, Aguirre N, Galindo MF (2007) Minocycline and cytoprotection: shedding new light on a shadowy controversy. Curr Drug Deliv 4(3):225–231. https://doi.org/10.2174/156720107781023938
- Joshi N, Miller DQ (1997) Doxycycline revisited. Arch Intern Med 157(13):1421-1428. https://doi.org/10.1001/archi nte.1997.00440340035003
- Kauppinen A, Salminen A, Kaarniranta K (2013) Inflammation as a target of minocycline: special interest in the regulation of inflammasome signaling. Inflammasome 2013:2–14. https:// doi.org/10.2478/infl-2013-0002
- Keijmel SP, Delsing CE, Bleijenberg G, van der Meer JWM, Donders RT, Leclercq M et al (2017) Effectiveness of long-term doxycycline treatment and cognitive-behavioral therapy on fatigue severity in patients with Q fever fatigue syndrome (Qure study): a randomized controlled trial. Clin Infect Dis 64(8):998–1005. https://doi.org/10.1093/cid/cix013
- Keller WR, Kum LM, Wehring HJ, Koola MM, Buchanan RW, Kelly DL (2013) A review of anti-inflammatory agents for symptoms of schizophrenia. J Psychopharmacol 27(4):337–342. https:// doi.org/10.1177/0269881112467089
- Kelly DL, Sullivan KM, McEvoy JP, McMahon RP, Wehring HJ, Gold JM et al (2015) Adjunctive minocycline in clozapinetreated schizophrenia patients with persistent symptoms. J Clin Psychopharmacol 35(4):374–381. https://doi.org/10.1097/ JCP.00000000000345
- Kim H-S, Suh Y-H (2009) Minocycline and neurodegenerative diseases. Behav Brain Res 196(2):168–179. https://doi. org/10.1016/j.bbr.2008.09.040
- Kosloski LM, Ha DM, Stone DK, Hutter JAL, Pichler MR, Reynolds AD, Gendelman HE, Mosley RL (2010) Adaptive immune regulation of glial homeostasis as an immunization strategy for neurodegenerative diseases. J Neurochem 114(5):1261–1276. https://doi.org/10.1111/j.1471-4159.2010.06834.x
- Kowal SL, Dall TM, Chakrabarti R, Storm MV, Jain A (2013) The current and projected economic burden of Parkinson's disease in the United States. Mov Disord 28:311–318. https://doi. org/10.1002/mds.25292
- Kraus RL, Pasieczny R, Lariosa-Willingham K, Turner MS, Jiang A, Trauger JW (2005) Antioxidant properties of minocycline: neuroprotection in an oxidative stress assay and direct radicalscavenging activity. J Neurochem 94(3):819–827. https://doi. org/10.1111/j.1471-4159.2005.03219.x
- Langevitz P, Bank I, Zemer D, Book M, Pras M (1992) Treatment of resistant rheumatoid arthritis with minocycline: an open study. J Rheumatol 19(10):1502–1504
- Lazzarini M, Martin S, Mitkovski M, Vozari RR, Stühmer W, Bel ED (2013) Doxycycline restrains glia and confers neuroprotection in a 6-OHDA Parkinson model. Glia 61(7):1084–1100. https ://doi.org/10.1002/glia.22496
- Lee JK, Tran T, Tansey MG (2009) Neuroinflammation in Parkinson's disease. J Neuroimmune Pharmacol 4(4):419–429. https ://doi.org/10.1007/s11481-009-9176-0
- Lewandoski M (2001) Conditional control of gene expression in the mouse. Nat Rev Gene 2:743-755. https://doi. org/10.1038/35093537
- LeWitt PA, Nyholm D (2004) New developments in levodopa therapy. Neurology 62(1):9–16. https://doi.org/10.1212/ WNL.62.1_suppl_1.S9
- Lin C, Zhang J (2017) Inflammasomes in inflammation-induced cancer. Front Immunol 8:271. https://doi.org/10.3389/fimmu .2017.00271
- Lleo A, Galea E, Sastre M (2007) Molecular targets of non-steroidal anti-inflammatory drugs in neurodegenerative diseases. Cell Mol Life Sci 64(11):1403–1418. https://doi.org/10.1007/s0001 8-007-6516-1

- Loeb MB, Molloy DW, Smieja M, Standish T, Goldsmith CH, Mahony J, Smith S, Borrie M, Decoteau E, Davidson W, McDougall A et al (2004) A randomized, controlled trial of doxycycline and rifampin for patients with Alzheimer's disease. J Am Geriatr Soc 52(3):381–387. https://doi.org/10.1111/j.1532-5415.2004.52109 .x
- Maiti P, Manna J, Veleri S, Frautschy S (2014) Molecular chaperone dysfunction in neurodegenerative diseases and effects of curcumin. Biomed Res Int 2014:495091. https://doi. org/10.1155/2014/495091
- Mao J, Barrow J, McMahon J, Vaughan J, McMahon AP (2005) An ES cell system for rapid, spatial and temporal analysis of gene function in vitro and in vivo. Nucleic Acids Res 33(18):e155. https://doi.org/10.1093/nar/gni146
- Marinova-Mutafchieva L, Sadeghian M, Broom L, Davis JB, Medhurst AD, Dexter DT (2009) Relationship between microglial activation and dopaminergic neuronal loss in the substantia nigra: a time course study in a 6-hydroxydopamine model of Parkinson's disease. J Neurochem 110(3):966–975. https://doi.org/10.111 1/j.1471-4159.2009.06189.x
- Martin RB (1985) Tetracyclines and daunorubicin in metal ions in biological systems. In: Sigel H (ed) Antibiotics and their complexes. Marcel Dekker, New York, pp 19–40
- Martínez C, Arranz A, Juarranz Y, Abad C, García-Gómez M, Rosignoli F, Leceta J, Gomariz RP (2006) PAC1 receptor: emerging target for septic shock therapy. Ann N Y Acad Sci 1070:405–410. https://doi.org/10.1196/annals.1317.053
- McGeer PL, McGeer EG (2008) Glial reactions in Parkinson's disease. Mov Disord 23(4):474–483
- Molloy DW, Standish TI, Zhou Q, Guyatt G (2013) A multicenter, blinded, randomized, factorial controlled trial of doxycycline and rifampin for treatment of Alzheimer's disease: the DARAD trial. Int J Geriatr Psychiatry 28(5):463–470. https://doi.org/10.1002/ gps.3846
- Moon A, Gil S, Gill SE, Chen P, Matute-Bello G (2012) Doxycycline impairs neutrophil migration to the airspaces of the lung in mice exposed to intratracheal lipopolysaccharide. J Inflamm 9(1):31. https://doi.org/10.1186/1476-9255-9-31
- Nelson ML (1998) Chemical and biological dynamics of tetracyclines. Adv Dent Res 12:5–11. https://doi.org/10.1177/0895937498 0120011901
- Nelson ML, Levy ST (2011) The history of Tetracyclines Ann. N Y Acad Sci 1241:17–32
- Nikodemova M, Duncan ID, Watters JJ (2006) Minocycline exerts inhibitory effects on multiple mitogen-activated protein kinases and IkappaBalpha degradation in a stimulus-specific manner in microglia. J Neurochem 96:314–323. https://doi.org/10.111 1/j.1471-4159.2005.03520.x
- NINDS NET-PD Investigators (2006) A randomized, double-blind, futility clinical trial of creatine and minocycline in early Parkinson disease. Neurology 66(5):664–671. https://doi. org/10.1212/01.wnl.0000201252.57661.e1
- Noble W, Garwood C, Stephenson J, Kinsey AM, Hanger DP, Anderton BH (2009a) Minocycline reduces the development of abnormal tau species in models of Alzheimer's disease. FASEB J 23(3):739–750. https://doi.org/10.1096/fj.08-113795
- Noble W, Garwood CJ, Hanger DP (2009b) Minocycline as a potential therapeutic agent in neurodegenerative disorders characterized by protein misfolding. Prion 3:78–83. https://doi.org/10.4161/ pri.3.2.8820
- Nordström D, Lindy O, Lauhio A, Sorsa T, Santavirta S, Konttinen YT (1998) Anti-collagenolytic mechanism of action of doxycycline treatment in rheumatoid arthritis. Rheumatol Int 17(5):175–180
- Olanow CW (2007) The pathogenesis of cell death in Parkinson's disease. Mov Disord 22(17):335–342. https://doi.org/10.1002/ mds.21675

- Olanow CW (2008) Levodopa/dopamine replacement strategies in Parkinson's disease–future directions. Mov Disord 23(3):613–622. https://doi.org/10.1002/mds.22061
- Orsucci D, Calsolaro V, Mancuso M, Siciliano G (2009) Neuroprotective effects of tetracyclines: molecular targets, animal models and human disease. CNS Neurol Disord Drug Targets 8(3):222–231. https://doi.org/10.2174/187152709788680689
- Papapetropoulos A, Szabo C (2018) Inventing new therapies without reinventing the wheel: the power of drug repurposing. Br J Pharmacol 175(2):165–167. https://doi.org/10.1111/bph.14081
- Parashos SA, Luo S, Biglan KM, Bodis-Wollner I, He B, Liang GS, Ross GW, Tilley BC, Shulman LM (2014) Measuring disease progression in early Parkinson disease: the National Institutes of Health Exploratory Trials in Parkinson disease (NET-PD) experience. JAMA Neurol 71(6):710–716. https://doi.org/10.1001/ jamaneurol.2014.391
- Pardo CA, Buckley A, Thurm A, Lee LC, Azhagiri A, Neville DM, Swedo SE (2013) A pilot open-label trial of minocycline in patients with autism and regressive features. J Neurodev Disord 5(1):9. https://doi.org/10.1186/1866-1955-5-9
- Payne JB, Golub LM, Stoner JA, Lee HM, Reinhardt RA, Sorsa T, Slepian MJ (2011) The effect of subantimicrobial-dose-doxycycline periodontal therapy on serum biomarkers of systemic inflammation: a randomized, double-masked, placebo-controlled clinical trial. J Am Dent Assoc 142:262–273. https://doi.org/10.14219/ jada.archive.2011.0165
- Peng J, Xie L, Stevenson FF, Melov S, Di Monte DA, Andersen JK (2006) Nigrostriatal dopaminergic neurodegeneration in the weaver mouse is mediated via neuroinflammation and alleviated by minocycline administration. J Neurosci 26(45):11644–11651. https://doi.org/10.1523/JNEUROSCI.3447-06.2006
- Pérez-Trallero E, Iglesias L (2003) Tetracyclines, sulfonamides and metronidazole. Enferm Infecc Microbiol Clin 21(9):520–528. https://doi.org/10.1016/j.eimc.2009.10.002
- Plane JM, Joseph DM, Allan BJ, Ashworth SH, Francisco JS (2006) An experimental and theoretical study of the reactions OIO + NO and OIO + OH. J Phys Chem A 110(1):93–100. https://doi. org/10.1021/jp055364y
- Plane JM, Shen Y, Pleasure DE, Deng W (2010) Prospects for minocycline neuroprotection. Arch Neurol 67(12):1442–1448. https ://doi.org/10.1001/archneurol.2010.191
- Pontieri FE, Ricci A, Pellicano C, Benincasa D, Buttarelli FR (2005) Minocycline in amyotrophic lateral sclerosis: a pilot study. Neurol Sci 26(4):285–287. https://doi.org/10.1007/s1007 2-005-0474-x
- Pruzanski W, Greenwald RA, Street IP, Laliberte F, Stefanski E, Vadas P (1992) Inhibition of enzymatic activity of phospholipases A2 by minocycline and doxycycline. Biochem Pharmacol 44(6):1165–1170. https://doi.org/10.1016/0006-2952(92)90381 -R
- Quintero EM, Willis L, Singleton R, Harris N, Huang P, Bhat N, Granholm AC (2006) Behavioral and morphological effects of minocycline in the 6-hydroxydopamine rat model of Parkinson's disease. Brain Res 1093(1):198–207. https://doi.org/10.1016/j. brainres.2006.03.104
- Radad K, Moldzio R, Rausch WD (2010) Minocycline protects dopaminergic neurons against long-term rotenone toxicity. Can J Neurol Sci 37(1):81–85. https://doi.org/10.1017/S031716710 0009690
- Reglodi D, Maasz G, Pirger Z, Rivnyak A, Balogh D, Jungling A, Fulop B, Mark L et al (2015) Neurochemical changes in different brain regions induced by PACAP—relations to neuroprotection. Springerplus 4(1):L56. https://doi. org/10.1186/2193-1801-4-S1-L56
- Reglodi D, Renaud J, Tamas A, Tizabi Y, Socías SB, Del-Bel E, Raisman-Vozari R (2017) Novel tactics for neuroprotection

🙆 Springer

in Parkinson's disease: role of antibiotics, polyphenols and neuropeptides. Prog Neurobiol 155:120–148. https://doi.org/10.1016/j.pneurobio.2015.10.004

- Rifkin BR, Vernillo AT, Golub LM, Ramamurthy NS (1994) Modulation of bone resorption by tetracyclines. Ann NY Acad Sci 732:165–180. https://doi.org/10.1111/j.1749-6632.1994.tb247 33.x
- Rodríguez-Violante M, Zerón-Martínez R, Cervantes-Arriaga A, Corona T (2017) Who can diagnose Parkinson's disease first? Role of pre-motor symptoms. Arch Med Res 48(3):221–227. https://doi.org/10.1016/j.arcmed.2017.08.005
- Rothstein JD, Patel S, Regan MR, Haenggeli C, Huang YH, Bergles DE et al (2005) β-Lactam antibiotics offer neuroprotection by increasing glutamate transporter expression. Nature 433:73–77
- Ruzza C, Rizzi A, Malfacini D, Cerlesi MC, Ferrari F, Marzola E, Ambrosio C, Gro C et al (2014) Pharmacological characterization of tachykinin tetrabranched derivatives. Br J Pharmacol 171(17):4125–4137. https://doi.org/10.1111/bph.12727
- Sande MA, Mandell GL (1985) Antimicrobial agents, tetracyclines. chloramphenicol, erythromycin, and miscellaneous antibacterial agents. In: Goodman and Gilman's (ed) The pharmacological basis of therapeutics, 7th edn. McGraw-Hill, New York
- Santa-Cecília FV, Socias B, Ouidja MO, Sepulveda-Diaz JE, Acuña L, Silva RL, Michel PP, Del-Bel E, Cunha TM, Raisman-Vozari R (2016) Doxycycline suppresses microglial activation by inhibiting the p38 MAPK and NF-kB signaling pathways. Neurotox Res 29(4):447–459. https://doi.org/10.1007/s1264 0-015-9592-2
- Sapadin AN, Fleischmajer R (2006) Tetracyclines: Nonantibiotic properties and their clinical implications. J Am Acad Dermatol 54:258–265. https://doi.org/10.1016/j.jaad.2005.10.004
- Schapira AH (2005) Present and future drug treatment for Parkinson's disease. J Neurol Neurosurg Psychiatry 76(11):1472–1478. https://doi.org/10.1136/jnnp.2004.035980
- Schapira AH, Bezard E, Brotchie J, Calon F, Collingridge GL, Ferger B, Hengerer B, Hirsch E et al (2006) Novel pharmacological targets for the treatment of Parkinson's disease. Nat Rev Drug Discov 5(10):845–854. https://doi.org/10.1038/nrd2087
- Seaborn T, Masmoudi-Kouli O, Fournier A, Vaudry H, Vaudry D (2011) Protective effects of pituitary adenylate cyclase-activating polypeptide (PACAP) against apoptosis. Curr Pharm Des 17(3):204–214. https://doi.org/10.2174/138161211795049679
- Skidmore R, Kovach R, Walker C, Thomas J, Bradshaw M, Leyden J, Powala C, Ashley R (2003) Effects of subantimicrobial-dose doxycycline in the treatment of moderate acne. Arch Dermatol 139(4):459–464. https://doi.org/10.1001/archderm.139.4.459
- Smith JR, Verwaerde C, Rolling F, Naud MC, Delanoye A, Thillaye-Goldenberg B, Apparailly F, De Kozak Y (2005) Tetracyclineinducible viral interleukin-10 intraocular gene transfer, using adeno-associated virus in experimental autoimmune uveoretinitis. Hum Gene Ther 16(9):1037–1046. https://doi.org/10.1089/ hum.2005.16.1037
- Smith CJ, Sayles H, Mikuls TR, Michaud K (2011) Minocycline and doxycycline therapy in community patients with rheumatoid arthritis: prescribing patterns, patient-level determinants of use, and patient-reported side effects. Arthritis Res Ther 13(5):R168. https://doi.org/10.1186/ar3491
- Socias SB, González-Lizárraga F, Avila CL, Vera C, Acuña L, Sepulveda-Diaz JE, Del-Bel E, Raisman-Vozari R, Chehin RN (2018) Exploiting the therapeutic potential of ready-to-use drugs: repurposing antibiotics against amyloid aggregation in neurodegenerative diseases. Prog Neurobiol. https://doi.org/10.1016/j.pneur obio.2017.12.002
- Sriram K, Miller DB, O'Callaghan JP (2006) Minocycline attenuates microglial activation but fails to mitigate striatal dopaminergic neurotoxicity: role of tumor necrosis factor-alpha. J Neurochem

96(3):706–718. https://doi.org/10.1111/j.1471-4159.2005.03566 .x

- Stock ML, Fiedler KJ, Acharya S, Lange JK, Mlynarczyk GS, Anderson SJ, McCormack GR, Kanuri SH, Kondru NC, Brewer MT, Carlson SA (2013) Antibiotics acting as neuroprotectants via mechanisms independent of their anti-infective activities. Neuropharmacology 73:174–182
- Stoilova T, Colombo L, Forloni G, Tagliavini F, Salmona M (2013) A new face for old antibiotics: tetracyclines in treatment of amyloidoses. J Med Chem 56(15):5987–6006. https://doi.org/10.1021/ jm400161p
- Subramaniam M, Althof D, Gispert S, Schwenk J, Auburger G, Kulik A, Fakler B, Roeper J (2014) Mutant α-synuclein enhances firing frequencies in dopamine substantia nigra neurons by oxidative impairment of A-type potassium channels. J Neurosci 34(41):13586–13599. https://doi.org/10.1523/JNEUR OSCI.5069-13.2014
- Sultan S, Gebara E, Toni N (2013) Doxycycline increases neurogenesis and reduces microglia in the adult hippocampus. Front Neurosci 7:131. https://doi.org/10.3389/fnins.2013.00131
- Swamy DN, Sanivarapu S, Moogla S, Kapalavai V (2015) Chemically modified tetracyclines: the novel host modulating agents. J Indian Soc Periodontol 19(4):370–374. https://doi.org/10.4103/0972-124X.149934
- Tamargo RJ, Bok RA, Brem H (1991) Angiogenesis inhibition by minocycline. Cancer Res 51:672-675
- Tansey MG, Goldberg MS (2010) Neuroinflammation in Parkinson's disease: its role in neuronal death and implications for therapeutic intervention. Neurobiol Dis 37(3):510–518. https://doi.org/10.1016/j.nbd.2009.11.004
- Taylor N (2013) Direct current cardioversion causing ventricular fibrillation in Wolff-Parkinson-White syndrome. Emerg Med Australas 25(6):612–614. https://doi.org/10.1111/1742-6723.12150
- Thong YH, Ferrante A (1979) Inhibition of mitogen-induced human lymphocyte proliferative responses by tetracycline analogues. Clin Exp Immunol 35(3):443–446
- Tikka TM, Koistinaho JE (2001) Minocycline provides neuroprotection against N-methyl-D-aspartate neurotoxicity by inhibiting microglia. J Immunol 166:7527–7533. https://doi.org/10.4049/ jimmunol.166.12.7527
- Tomiyama T, Shoji A, Kataoka K, Suwa Y, Asano S, Kaneko H, Endo N (1996) Inhibition of amyloid beta protein aggregation and neurotoxicity by rifampicin. Its possible function as a hydroxyl radical scavenger. J Biol Chem 271(12):6839–6844
- Ton TG, Heckbert SR, Longstreth WT Jr, Rossing MA, Kukull WA, Franklin GM, Swanson PD, Smith-Weller T, Checkoway H (2006) Nonsteroidal anti-inflammatory drugs and risk of Parkinson's disease. Mov Disord 21(7):964–969. https://doi. org/10.1002/mds.20856
- Tufekci KU, Meuwissen R, Genc S, Genc K (2012) Inflammation in Parkinson's disease. Adv Protein Chem Struct Biol 88:69–132. https://doi.org/10.1016/B978-0-12-398314-5.00004-0
- Wachter B, Schürger S, Rolinger J, von Ameln-Mayerhofer A, Berg D, Wagner HJ, Kueppers E (2010) Effect of 6-hydroxydopamine

(6-OHDA) on proliferation of glial cells in the rat cortex and striatum: evidence for de-differentiation of resident astrocytes. Cell Tissue Res 342(2):147–160. https://doi.org/10.1007/s0044 1-010-1061-x

- Wang X, Cai H, Yan X, Ye Z (2010) Effect of different tidal volume ventilation on apoptosis and the expression of Bax and Bcl-2 in rat lungs. Zhong Nan Da Xue Xue Bao Yi Xue Ban 35(4):351– 357. https://doi.org/10.3969/j.issn.1672-7347.2010.04.012
- Wang W, Sidoli S, Zhang W, Wang Q, Wang L, Jensen ON, Guo L, Zhao X, Zheng L (2017) Abnormal levels of histone methylation in the retinas of diabetic rats are reversed by minocycline treatment. Sci Rep 7:45103. https://doi.org/10.1038/srep45103
- Whiteman M, Halliwell B (1997) Prevention of peroxynitrite-dependent tyrosine nitration and inactivation of alpha1-antiproteinase by antibiotics. Free Radic Res 26:49–56
- Wong YC, Krainc D (2017) α-synuclein toxicity in neurodegeneration: mechanism and therapeutic strategies. Nat Med 23(2):1–13. https ://doi.org/10.1038/nm.4269
- Worlitzer MM, Viel T, Jacobs AH, Schwamborn JC (2013) The majority of newly generated cells in the adult mouse substantia nigra express low levels of Doublecortin, but their proliferation is unaffected by 6-OHDA-induced nigral lesion or Minocycline-mediated inhibition of neuroinflamation. Eur J Neurosci 38:2684– 2692. https://doi.org/10.1111/ejn.12269
- Wu DC, Jackson-Lewis V, Vila M, Tieu K, Teismann P, Vadseth C, Choi DK, Ischiropoulos H, Przedborski S (2002) Blockade of microglial activation is neuroprotective in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson disease. J Neurosci 22:1763–1771
- Yang L, Sugama S, Chirichigno JW, Gregorio J, Lorenzl S, Shin DH, Browne SE, Shimizu Y, Joh TH, Beal MF, Albers DS (2003) Minocycline enhances MPTP toxicity to dopaminergic neurons. J Neurosci Res 74(2):278–285. https://doi.org/10.1002/jnr.10709
- Yong VW, Wells J, Giuliani F, Casha S, Power C, Metz LM (2004) The promise of minocycline in neurology. Lancet Neurol. 3, 744–751. https://doi.org/10.1016/S1474-4422(04)00937-8
- Yrjanheikki J, Keinanen R, Pellikka M, Hokfelt T, Koistinaho J (1998) Tetracyclines inhibit microglial activation and are neuroprotective in global brain ischemia. Proc Natl Acad Sci USA 95:15769–15774
- Yu R, Zheng L, Cui Y, Zhang H, Ye H (2016) Doxycycline exerted neuroprotective activity by enhancing the activation of neuropeptide GPCR PAC1. Neuropharmacology 103:1–15. https://doi. org/10.1016/j.neuropharm.2015.11.032
- Zhang W, Narayanan M, Friedlander RM (2003) Additive neuroprotective effects of minocycline with creatine in a mouse model of ALS. Ann Neurol 53(2):267–270. https://doi.org/10.1002/ ana.10476
- Zhang GB, Feng YH, Wang PQ, Song JH, Wang P, Wang SA (2015) A study on the protective role of doxycycline upon dopaminergic neuron of LPS-PD rat model rat. Eur Rev Med Pharmacol Sci 19(18):3468–3474